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Antiemetic Effects of Cannabinoid Agonists in Nonhuman Primates

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Δ^9-THC, Δ^9-tetrahydrocannabinol; LiCl, lithium chloride; mAEA, methanandamide,
(R)-(+)-arachidonyl-1'-hydroxy-2'-propylamide; rimonabant, SR141716A, 5-(4-
chlorophenyl)-1-(2,4-dichloro-phenyl)-4-methyl-N-(piperidin-1-yl)-1-H-pyrazole-3-
carboxamide

ABSTRACT

Attenuating emesis elicited by both disease and medical treatments of disease remains a critical public health challenge. Although cannabinergic medications have been used in certain treatment-resistant populations, FDA-approved cannabinoid antiemetics are associated with undesirable side effects, including cognitive disruption, that limit their prescription. Previous studies have shown that a metabolically stable analog of the endocannabinoid anandamide, methanandamide (mAEA), may produce lesser cognitive disruption than that associated with the primary psychoactive constituent in cannabis, Δ^9-tetrahydrocannabinol (Δ^9-THC), raising the possibility that endocannabinoids may offer a therapeutic advantage over currently used medications. The present studies were conducted to evaluate this possibility by comparing the antiemetic effects of Δ^9-THC (0.032-0.1 mg/kg) and mAEA (3.2-10.0 mg/kg), against nicotine- and lithium chloride (LiCl)-induced emesis and prodromal hypersalivation in squirrel monkeys. Pretreatment with 0.1 mg/kg Δ^9-THC blocked nicotine-induced emesis and reduced hypersalivation in all subjects and blocked LiCl-induced emesis and reduced hypersalivation in 3 of 4 subjects. Pretreatment with 10 mg/kg mAEA blocked nicotine-induced emesis in 3 of 4 subjects and LiCl-induced emesis in 1 of 4 subjects, and reduced both nicotine- and LiCl-induced hypersalivation. Antiemetic effects of Δ^9-THC and mAEA were reversed by rimonabant pretreatment, providing verification of CB_1 receptor-mediation. These studies systematically demonstrate for the first time the antiemetic effects of cannabinoid agonists in nonhuman primates. Importantly,

although Δ^9-THC produced superior antiemetic effects, the milder cognitive effects of mAEA demonstrated in previous studies suggests that it may provide a favorable treatment option under clinical circumstances in which antiemetic efficacy must be balanced against side-effect liability.

SIGNIFICANCE STATEMENT

Emesis has significant evolutionary value as a defense mechanism against ingested toxins; however, it is also one of the most common adverse symptoms associated with both disease and medical treatments of disease. The development of improved anti-emetic pharmacotherapies has been impeded by a paucity of animal models. The present studies systematically demonstrate for the first time the antiemetic effects of the phytocannabinoid Δ^9-tetrahydrocannabinol and endocannabinoid-analog methanandamide in nonhuman primates.

INTRODUCTION

Cannabinergic drugs are currently under investigation as pharmacotherapies for a variety of medical conditions. However, prior to the approval of Epidiolex for treatment-resistant seizures in 2018, the Food and Drug Administration (FDA) had approved only two cannabinoid pharmaceuticals – Dronabinol (Marinol), a synthetic Δ^9-tetrahydrocannabinol (Δ^9-THC), and nabilone (Cesamet), a structurally distinct synthetic cannabinoid agonist (Bedi et al, 2013). Both cannabinoids were initially approved explicitly for the treatment of refractory emesis and nausea secondary to chemotherapy for cancer, and their prescription is still limited to gastrointestinal disturbances during severe chronic illness (Seamon, 2006).

Although 30-50% of patients receiving highly emetogenic chemotherapy will experience refractory vomiting despite guideline-directed prophylaxis (Cohen et al., 2007; Tamura et al., 2017), the FDA-approved cannabinoids are not recommended as first-line antiemetics and appear sparsely as adjunctive therapies in clinical guidelines (Garcia and Shamliyan, 2018). This is because dronabinol and nabilone are associated with a higher rate of side effects than other antiemetics, including attention and memory impairment and dysphoria (Mathai et al., 2018; Schussel et al., 2017; Tafefsky et al., 2016; Wesnes et al., 2010). Despite their unfavorable side-effect profile, several studies have indicated that patients prefer cannabinoids over other antiemetics such as dopamine and serotonin antagonists (Ahmedzai et al., 1983; Einhorn et al., 1981; Smith et al., 2015). The reasons underlying this preference are unclear; however,

there is evidence that cannabinoids are more effective at also attenuating nausea (Abrahamov, et al., 1995; Meiri et al., 2007). Collectively, these studies confirm the medicinal utility of cannabinoids as antiemetic pharmacotherapies and suggest that the development of novel cannabinergic drugs with a reduced side-effect profile would be clinically beneficial.

In addition to novel cannabinoid agonists, the development of improved cannabinergic pharmacotherapies has increasingly focused on enhancing endogenous activity. Indeed, emesis in shrews and ferrets can be blocked with endocannabinoids such as anandamide and 2-arachidonoylglycerol or by targeting catabolic enzymes (fatty acid amide hydrolase and monoacyglycerol lipase) to increase circulating endocannabinoid levels (Darmani, 2002; Parker et al., 2009; Sharkey et al., 2007; Sticht et al., 2013). In addition, separate studies have provided evidence that elevation of endocannabinoid activity has fewer cognition-impairing effects than the administration of synthetic agonists or phytocannabinoids such as Δ^9-THC (Kangas et al., 2016; Mechoulam and Parker, 2013). Thus, increasing endocannabinoid activity might provide a novel avenue for development of cannabinergic antiemetic pharmacotherapies with fewer adverse effects, especially related to cognition.

Unfortunately, the development of improved cannabinergic antiemetic pharmacotherapies has been impeded by a paucity of animal models. This is due to the fact that several of the most common laboratory animals, including the mouse, rat, guinea pig, and rabbit, are physically incapable of vomiting due to a complex array of neural and anatomical constraints (Horn et al., 2013). Most

preclinical research in this area thus has been restricted to other species such as the house musk shrew (Parker et al., 2004; 2009; Rock et al., 2016; Sticht et al., 2013), least shrew (Darmani, 2002; Ray et al., 2009), and ferret (Sharkey et al., 2007; Simoneau et al., 2001). Studies conducted in these subjects have provided important insights into emetic mechanisms. However, the shrew and ferret are relatively atypical laboratory animals that have not been extensively used for *in vivo* pharmacological studies. Thus, in the absence of data on possible side effects, it is difficult to assess the potential clinical value of novel antiemetics in these species.

Surprisingly, there are no published reports regarding antiemetic effects of cannabinoids in nonhuman primates. This is a curious gap considering the limitations of rodent subjects in emesis research and substantial *in vivo* cannabinoid research that has been conducted in nonhuman primates. In particular, squirrel monkeys in which cannabinoids have been extensively studied (e.g., Branch et al., 1980; Desai et al., 2013; Justinova et al., 2003; 2013; Kangas and Bergman, 2012; Kangas et al., 2013; 2016; Leonard et al., 2017; Solinas et al., 2007; Tanda et al., 2000) and which have an emetic response are highly suitable for evaluating the antiemetic effects of cannabinoids. The present studies therefore examined Δ^9-THC, the primary psychoactive constituent in cannabis, and methanandamide (mAEA), a metabolically stable analog of the endocannabinoid anandamide, for their ability to block emesis and prodromal hypersalivation in the squirrel monkey. The antiemetic abilities of these drugs were examined by pretreating subjects prior to pharmacological challenges using

two common emetic agents, nicotine and lithium chloride (LiCl). Finally, pretreatment with rimonabant, the selective CB_1 receptor antagonist, was assessed for its ability to reverse the antiemetic effects of Δ^9-THC and mAEA to determine if the observed effects were CB_1-mediated.

METHODS

Subjects

Five adult male squirrel monkeys (*Saimiri sciureus*) served in the present studies (one subject that served in the nicotine group did not serve in the LiCl group; nicotine, *n*=4; LiCl, *n*=4). Four subjects were experimentally and drug naïve at the start of the study. The fifth subject previously served in a behavioral study examining opioid agonists but had not received drug treatment for 6 months prior to the present study. Subjects were housed in a temperature- and humidity-controlled vivarium with a 12-hr light/dark cycle (lights on at 7 a.m.), and environmental enrichment was provided daily. Subjects had unlimited access to water in the home cage and were maintained at approximate free-feeding weights by daily feedings of fresh fruit and nutritionally balanced high protein biscuits (Purina Monkey Chow, St. Louis, MO). Experimental sessions were conducted 5 days a week (M-F). Subjects were fed approximately 2 hrs following each experimental session. The protocol for the present studies was approved by the Institutional Animal Care and Use Committee at McLean Hospital in a facility licensed by the United States Department of Agriculture and in accordance with guidelines provided by the Committee on Care and Use of Laboratory Animals of

the Institute of Laboratory Animals Resources, Commission on Life Sciences
(National Research Council, 2011).

Apparatus

A custom designed dual-compartment observation chamber was used to
monitor two subjects simultaneously (Wooldridge and Kangas, 2019). Two clear
Plexiglas cubes (25x25x25 cm) separated by an opaque Plexiglas divider resided
in a light- and sound-attenuating ventilated enclosure (75x60x50 cm). Mirrors
were affixed to the walls and floor of the enclosure to provide a view of orofacial
and abdominal movements when subjects were facing away from the observer.
White noise was present in the experimental room to provide masking sound.
Subjects were leashed but otherwise unrestrained within the observation
chamber.

Experimental Procedures

Subjects were monitored continuously in the observation chamber during
experimental sessions conducted at approximately the same time each day
according to a preset plan. Instances of licking lasting longer than 2 sec, chewing,
drooling, foaming, and emesis were recorded as quantal measures (presence or
absence) during each 1-min bin. When hypersalivation was observed, duration
was calculated by summing the total number of 1-min bins in which at least one
of the following responses occurred: licking, chewing, drooling, and foaming.

Effects of Δ^9-THC and mAEA on nicotine- and LiCl-induced emesis and hypersalivation

Drug testing sessions were conducted no more than once per week. Control sessions, in which 0.1-0.3 ml of saline was administered, were conducted during intervening days to preclude the development of conditioned responses to injections or to the observation chamber.

The antiemetic effects of Δ^9-THC and mAEA were studied in subjects treated with emetic doses of nicotine and LiCl, both of which have been extensively used as emetic challenges in previous studies with shrews (Parker et al., 2004; 2009) and ferrets (Billig et al., 2001; du Sert et al., 2012). The doses of nicotine (0.32 mg/kg) and LiCl (200 mg/kg), as well as the duration of observation periods (20 and 60 min, respectively) and Δ^9-THC and mAEA pretreatment times (30 min), were based on previous studies of nicotine, LiCl, and cannabinoid agonists in squirrel monkeys (Justinova et al., 2013; Kangas et al., 2013; 2016; Leonard et al., 2017; Wooldridge and Kangas, 2019). First, the ability of Δ^9-THC and mAEA to block nicotine-induced emesis and hypersalivation were determined by administering Δ^9-THC (0.032 or 0.1 mg/kg), mAEA (3.2 or 10.0 mg/kg), or vehicle 30 min prior to treatment with saline or, in separate test sessions, a reliably emetic dose of nicotine (0.32 mg/kg). Next, the doses of Δ^9-THC and mAEA that were most effective in preventing nicotine-induced emesis (0.1 and 10 mg/kg, respectively) were examined further for their ability to modify LiCl-induced emesis. The antiemetic effects of Δ^9-THC against nicotine and LiCl

were tested in all subjects before those of mAEA. Finally, the involvement of CB_1 receptor activity in the antiemetic effects of Δ^9-THC and mAEA were studied by determining whether an antagonist dose of rimonabant in squirrel monkeys (0.32 mg/kg; Kangas et al., 2013; Schindler et al., 2016), administered 30 min prior to Δ^9-THC or mAEA, could block the cannabinoid's effects. In these last experiments, the effects of rimonabant alone, and prior to each agonist without nicotine, also were assessed in separate test sessions by administering the CB_1 antagonist and replacing injections of Δ^9-THC, mAEA, and/or nicotine with injections of saline.

Drugs

(-)-Nicotine hydrogen tartrate was purchased from Sigma-Aldrich (St. Louis, MO) and was prepared in a 0.9% saline solution. The pH of the resulting solution was adjusted to ~7.0 with the addition of 0.1N sodium hydroxide as needed. LiCl was purchased from Fisher Scientific (Hampton, NH) and was prepared in sterile water. Δ^9-THC and rimonabant were provided by the National Institutes of Health National Institute on Drug Abuse Drug Supply Program (Rockville, MD). mAEA was synthesized by the present authors (LJ, YL, SPN, AM) at the Center for Drug Discovery at Northeastern University (Boston, MA) following previously described procedures (Abajji et al., 1994). The identity of the compound was confirmed via NMR spectra of mAEA recorded in $CDCl_3$ on Varian 500 (^{1}H at 500 MHz) and Bruker 400 (^{13}C at 100 MHz) NMR spectrometers using previously reported procedures (Liu et al., 2018). mAEA

purity was determined via LC/MS analysis using a Waters MicroMass ZQ system electrospray ionization with Waters-2525 binary gradient module coupled to a photodiode array detector (Waters-2996) and ELS detector (Waters-2424) using a XTerra MS C18 (5μm, 4.6×50 mm column and acetonitrile/water) and was found to be >97%. Δ^9-THC, mAEA, and rimonabant were prepared in a 20:20:60 mixture by volume of 95% ethanol, Tween-80, and saline. All drugs and vehicle were administered via intramuscular (i.m.) injection in volumes of 0.4 ml/kg or less. Drug concentrations are expressed in terms of their free base.

RESULTS

Effects of Δ^9-THC on nicotine- and LiCl-induced emesis and hypersalivation

Figure 1 presents the effects of Δ^9-THC pretreatment (0.032 and 0.1 mg/kg) on drug-induced emesis (Fig. 1A, 1C) and hypersalivation (Fig. 1B, 1D). Administration of vehicle, 0.032 mg/kg Δ^9-THC, or 0.1 mg/kg Δ^9-THC alone did not produce emesis in any subject. Pretreatment with 0.032 mg/kg Δ^9-THC blocked nicotine-induced emesis in 1 out of 4 subjects and pretreatment with 0.1 mg/kg Δ^9-THC blocked nicotine-induced emesis in all 4 subjects (Fig. 1A). Administration of Δ^9-THC alone produced minimal hypersalivation. Nicotine administration produced hypersalivation in each subject, with a mean duration of 8.8 (±1.55) min. Nicotine-induced hypersalivation was dose-dependently attenuated by Δ^9-THC – 0.032 mg/kg Δ^9-THC decreased the mean duration of hypersalivation to 6.5 (±1.94) min and 0.1 mg/kg Δ^9-THC decreased the mean duration to 3.0 (±1.73) min (Fig. 1B).

The most effective antiemetic dose of Δ^9-THC tested against nicotine-induced emesis (0.1 mg/kg) was subsequently studied for its ability to attenuate LiCl-induced emesis (Fig. 1C) and hypersalivation (Fig. 1D). Pretreatment with 0.1 mg/kg Δ^9-THC blocked LiCl-induced emesis in 2 out of 4 subjects (Fig. 1C). LiCl alone produced a mean duration of hypersalivation of 6.3 ($\pm$2.59) min and pretreatment with 0.1 mg/kg Δ^9-THC reduced the duration to 1.8 ($\pm$0.48) min (Fig. 1D).

Effects of mAEA on nicotine- and LiCl-induced emesis and hypersalivation

Figure 2 presents the effects of mAEA pretreatment (3.2 and 10 mg/kg) on drug-induced emesis (Fig. 2A, 2C) and hypersalivation (Fig. 2B, 2D). Administration of vehicle, 3.2 mg/kg mAEA, or 10 mg/kg mAEA did not produce emesis in any subject. Pretreatment with 3.2 mg/kg mAEA blocked nicotine-induced emesis in 1 out of 4 subjects and pretreatment with 10 mg/kg mAEA blocked nicotine-induced emesis in 3 out of 4 subjects (Fig 2A). A higher dose of 17 mg/kg mAEA was tested in the fourth subject and also failed to block nicotine-induced emesis (data not shown). Pretreatment with mAEA also reduced nicotine-induced hypersalivation (Fig. 2B). Vehicle, 3.2 mg/kg mAEA, or 10 mg/kg mAEA did not produce hypersalivation when administered alone; however, pretreatment with 3.2 mg/kg mAEA reduced nicotine-induced hypersalivation from a mean duration of 8.8 ($\pm$1.55) min to 5.0 ($\pm$2.04) whereas 10 mg/kg mAEA reduced hypersalivation to 4.8 ($\pm$ 2.06) min. In subsequent studies, pretreatment with 10 mg/kg mAEA blocked LiCl-induced emesis in only one of four subjects

(Fig. 2C), and reduced LiCl-induced hypersalivation from a mean duration of 6.3 (±2.59) min to 4.3 (±3.59) min (Fig. 2D).

Effects of rimonabant pretreatment on Δ^9-THC and mAEA antiemesis

Administration of 0.32 mg/kg rimonabant did not produce emesis or hypersalivation when administered alone or when administered before either 0.1 mg/kg Δ^9-THC (Fig. 3A) or 10 mg/kg mAEA (Fig. 3C). However, this dose of rimonabant antagonized the previously observed antiemetic effects of 0.1 mg/kg Δ^9-THC in all subjects (Fig. 3A) and 10 mg/kg mAEA in 3 of 4 subjects (Fig. 3C) against nicotine-induced emesis. In addition, rimonabant pretreatment reversed the reductions in nicotine-induced hypersalivation following Δ^9-THC from 3.0 (±1.73) min to 9.8 (±2.39) min (Fig. 3B), and following mAEA from 4.8 (±2.06) to 9.0(±1.6) (Fig. 3D).

DISCUSSION

The present studies compared the ability of Δ^9-THC and mAEA to block nicotine- or LiCl-induced emesis and hypersalivation in the squirrel monkey. Δ^9-THC was able to block nicotine-induced emesis and hypersalivation in all subjects tested and LiCl-induced emesis in some, but not all, subjects. Like Δ^9-THC, mAEA was able to block nicotine- and LiCl-induced emesis and reduce hypersalivation. However, these effects were not evident in all subjects, regardless of whether the emetic agent was nicotine or LiCl. Finally, rimonabant pretreatment reversed the antiemetic effects of both Δ^9-THC and mAEA,

providing evidence that their antiemetic effects are mediated via CB_1-receptor mechanisms. These findings are consistent with previous work demonstrating CB_1-mediated antiemetic effects of Δ^9-THC and mAEA against a variety of emetic stimuli in the least shrew (Darmani, 2002) and ferret (Van Sickle et al., 2001).

Notably, Δ^9-THC and mAEA were more effective at blocking nicotine-induced than LiCl-induced emesis. Both emetics have been shown to reliably produce emesis in several species, including the squirrel monkey, but may act via different mechanisms (Beleslin and Krstic, 1987; Billig et al., 2001; Lee et al., 1978, Parker et al., 2004; 2009; Wooldridge and Kangas, 2019). Nicotine primarily acts at nicotinic receptors in the area postrema, or the *chemoreceptor trigger zone*, of the central nervous system. The blood-brain barrier in this region of the medulla is relatively permeable, permitting the detection of circulating emetogens in the bloodstream (Beleslin and Krstic, 1987). Although the mechanisms by which LiCl produces emesis are less thoroughly understood, they are thought to involve both central and peripheral actions. Centrally, LiCl is thought to act in the area postrema (Fox et al., 1990; Spencer et al., 2012) and, via elevation of serotonin release, in the interoceptive insular cortex (Limebeer et al., 2018), a region implicated in nausea in humans (Napadow et al., 2013; Sclocco et al., 2016; Penfield and Falulk, 1955) and rats (Contraeras et al., 2007; Sticht et al., 2016). Peripherally, LiCl is thought to act at the splanchnic and vagus nerves in the gut (Horn et al., 2014; Yamamoto et al., 1992). In this regard, previous work in the least shrew has suggested that Δ^9-THC may more potently

block centrally-mediated than peripherally-mediated emesis (Darmani and Johnson, 2004). Thus, it is possible that the greater effectiveness of both cannabinoid agonists against nicotine-induced than LiCl-induced emesis in the present studies is related to ineffectiveness against LiCl's peripheral actions. The evaluation of higher doses that might also block such peripheral actions could address this possibility.

The antiemetic effects of both Δ^9-THC and mAEA in the present study were reversed by pretreatment with the selective CB_1 receptor antagonist rimonabant, indicating that their effects are mediated by CB_1 receptors. This result is consistent with previous studies demonstrating that Δ^9-THC and mAEA act at CB_1 receptors to block the emetic reflex initiated in the brainstem (Van Sickle et al., 2001; 2005). Indeed, cannabinoid receptors are ubiquitous throughout both the gastrointestinal tract and the brainstem areas responsible for the production of emesis (Darmani, 2010). While CB_2 receptor activation is also associated with the antiemetic effects of certain cannabinoid agonists (Rock et al., 2016; Van Sickle et al., 2005), the selective blockade of CB_2 receptors with AM630 or SR144528 has been shown to be insufficient to block the antiemetic effects of Δ^9-THC or anandamide in the ferret (Van Sickle et al., 2005) and least shrew (Darmani et al., 2007).

Both Δ^9-THC and, to a lesser extent, mAEA also reduced the hypersalivation that accompanied nicotine or LiCl-induced emesis and, consistent with the involvement of CB_1 receptor mechanisms, these cannabinergic effects could be blocked by rimonabant. Hypersalivation is thought

to be a prodromal sign that often accompanies and worsens the subjective experience of emesis (Kenward et al., 2015; Sanger and Andrews 2016; Wooldridge and Kangas, 2019). The ability of cannabinoid agonists to abate hypersalivation may therefore be a means of alleviating such distress and, consequently, reflect a desirable feature of their medicinal value.

In these studies, Δ^9-THC consistently produced a more robust antiemetic effect than did mAEA. The difference in antiemetic activity may reflect a difference in CB_1 receptor efficacy; that is, while Δ^9-THC and mAEA are both generally considered CB_1 receptor partial agonists, Δ^9-THC may have greater CB_1 efficacy than either mAEA or anandamide (Brodkin and Moerschbaecher, 1997; Desai et al., 2013; Järbe et al., 1998). Alternatively, the engagement of non-cannabinoid neurotransmitter systems may account for the difference in antiemetic effects of the two cannabinoid agonists. For example, Δ^9-THC primarily acts at cannabinoid receptors (de Petrocellis et al., 2011), whereas the endocannabinoids, including anandamide, have other targets, including the capsaicin-sensitive transient receptor potential-vanilloid type 1 receptor (TRPV1), another channel associated with the production of emesis (Andrews et al., 2000; Andrews and Bhandari, 1993; Chu et al., 2010; Ross, 2003; Yamakuni et al., 2002).

As the antiemetic effects of Δ^9-THC were examined first in all subjects, it is possible that the difference in effectiveness reflects tolerance to the effects of the cannabinoid agonists. However, tolerance to the physiological, rate-decreasing, and cognition-impairing effects of cannabinoids in laboratory animals typically

requires several consecutive days of high-dose treatment (reviewed in González et al., 2005), whereas cannabinergic drugs in the present studies were administered no more than once per week over a period of several months. Subchronic or chronic treatment with cannabinergic drugs has not yet been examined in animal models of emesis, but tolerance to nabilone or dronabinol has not been reported over a typical course of chemotherapy in clinical studies (May and Glode, 2016; Meiri et al., 2007; Ware et al., 2008). Nevertheless, further studies are needed to systematically evaluate the development of tolerance over the treatment periods that would be necessary in subchronic or chronic regimens in clinical settings.

Finally, although a complete antiemetic effect was not achieved with doses of mAEA tested here against either emetic, this compound – or related endocannabinoid derivatives – may offer some translational advantage over Δ^9-THC, in particular with regards to cognition-impairing side effects. For example, Kangas et al. (2016) compared the cognition-impairing effects of several cannabinoid agonists, including Δ^9-THC and mAEA, in similarly-aged adult male squirrel monkeys and found that antiemetic doses of Δ^9-THC identified in the present study produced considerably more pronounced cognition-impairing effects than did the most effective dose of mAEA identified here (see Table 1). Specifically, 0.1 mg/kg Δ^9-THC disrupted performance across a battery of cognitive tasks designed to assay learning (repeated acquisition), cognitive flexibility (discrimination reversal), and working memory (delayed matching-to-sample). In contrast, 10 mg/kg mAEA was not associated with any cognition-

impairing effects yet, in the present study, produced moderate antiemetic effects. Collectively, these data suggest that mAEA (or, possibly, other endocannabinoid derivatives) may offer a balance between moderate yet clinically beneficial antiemetic efficacy independent of cognition-impairing effects. Such effects are particularly important to consider in the development of antiemetics, as many of the conditions for which novel antiemetic treatments are needed, most notably chemotherapy, also are associated with distressing disruptions in cognitive function – colloquially referred to as *chemo brain* or *chemo fog* (Asher, 2011; Janelsins et al., 2017).

In summary, the present studies systematically demonstrate for the first time that the phytocannabinoid Δ^9-THC and the endocannabinoid analog mAEA produce antiemetic effects in nonhuman primates. Future studies are necessary to confirm the utility of cannabinergic antiemetics against chemotherapy-induced nausea and vomiting, as well as to evaluate the possibility of tolerance to their antiemetic effects over treatment periods that would be clinically necessary. Such studies should also establish the ability of these cannabinergic compounds to limit anticipatory nausea and vomiting, which often develops during the course of emetic chemotherapy. Finally, methods of engaging the endocannabinoid system, such as exogenous administration of other endocannabinoids (e.g., 2-arachidonoylglycerol) or by inhibition of their metabolic enzymes (e.g., fatty acid amide hydrolase, monoacylglycerol lipase), should be examined, ideally to identify compounds with maximal antiemetic and anti-nausea effects and minimal cognition-impairing side effects.

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AUTHORSHIP CONTRIBUTIONS

Participated in research design: Wooldridge, Bergman, and Kangas.

Conducted experiments: Wooldridge.

Contributed new reagents or analytic tools: Ji, Liu, Nikas, Makriyannis.

Performed data analysis: Wooldridge and Kangas.

Wrote or contributed to the writing of the manuscript: Wooldridge, Bergman, and Kangas.

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FOOTNOTES

LEGENDS FOR FIGURES

Figure 1. Upper panels: Number of subjects to exhibit an emetic episode following administration of vehicle, 0.032, or 0.1 mg/kg Δ^9-THC alone and 30 min before nicotine (A) or LiCl (B). **Lower panels:** Mean (± SEM) duration of hypersalivation (min) following vehicle and Δ^9-THC alone and 30 min before nicotine (C) or LiCl (D). *n*=4.

Figure 2. Upper panels: Number of subjects to exhibit an emetic episode following administration of vehicle, 3.2, or 10 mg/kg mAEA alone and 30 min before nicotine (A) or LiCl (B). **Lower panels:** Mean (± SEM) duration of hypersalivation (min) following vehicle and mAEA alone and 30 min before nicotine (C) or LiCl (D). *n*=4.

Figure 3. Upper panels: Effects of 0.32 mg/kg rimonabant (SR) on the ability of 0.1 mg/kg Δ^9-THC (A) or 10 mg/kg mAEA (B) to block nicotine-induced emesis. **Lower panels:** Effects of 0.32 mg/kg rimonabant (SR) on the ability of 0.1 mg/kg Δ^9-THC (A) or 10 mg/kg mAEA (B) to reduce nicotine-induced hypersalivation. Rimonabant was administered 60min before nicotine, and Δ^9-THC or mAEA were administered 30min before nicotine. *n*=4.

	Maximally Effective Antiemetic Dose	Lowest Dose that Disrupted Cognitive Behavior			
		RA	DR	DMTS	PVT
Δ^9-THC	0.1 mg/kg	0.1 mg/kg	0.1 mg/kg	0.1 mg/kg	0.32 mg/kg
mAEA	10 mg/kg	>32 mg/kg	>32 mg/kg	17.8 mg/kg	32 mg/kg

Table 1. Comparison of the maximally effective antiemetic doses of Δ^9-THC and

mAEA derived from present findings, and cognition-impairing doses of Δ^9-THC

and mAEA on touchscreen-based assays of learning (repeated acquisition [RA]),

cognitive flexibility (discrimination reversal [DR]), short-term memory (delayed

matching-to-sample [DMTS]), and sustained attention (psychomotor vigilance

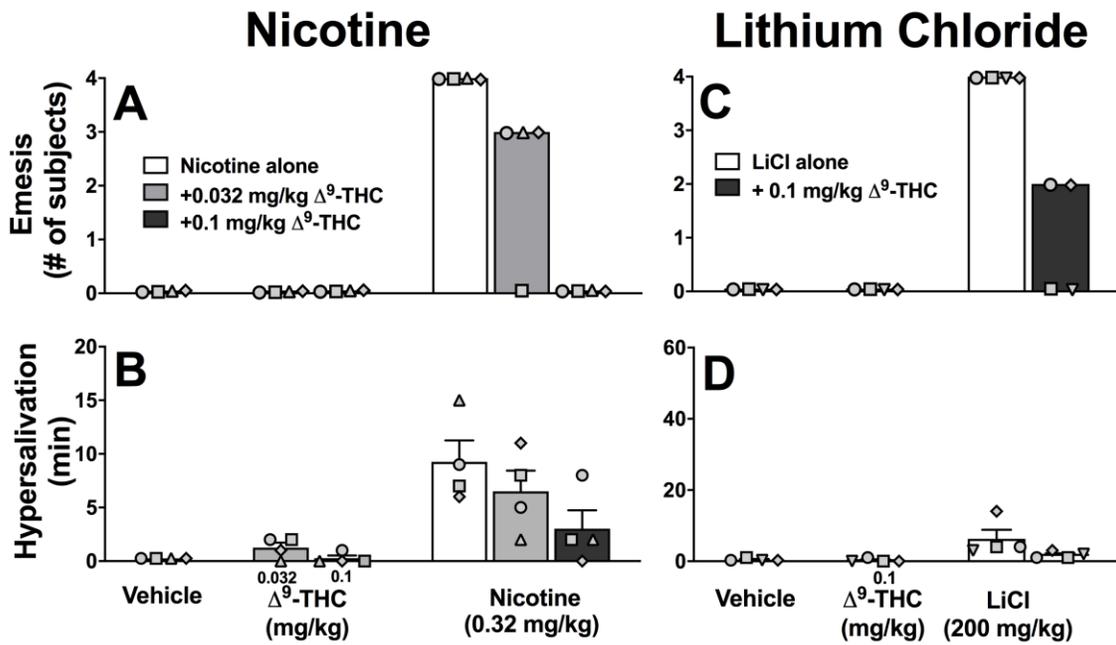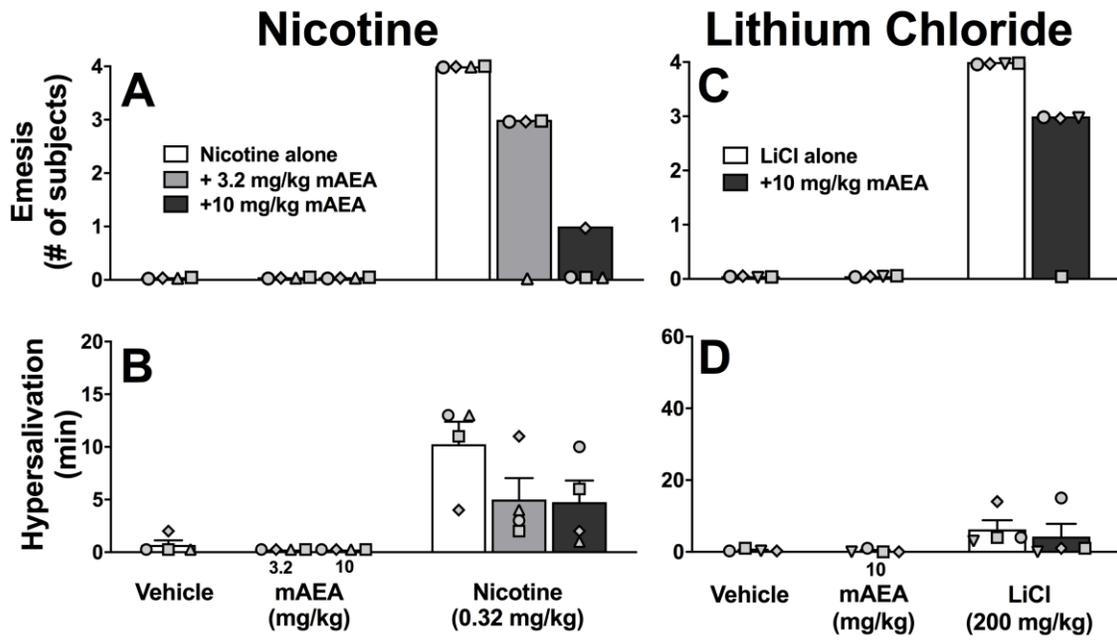[PVT]) derived from Kangas et al. (2016).

Fig. 1

Fig. 2

Fig. 3

Δ^9-THC

mAEA