

JPET # 265058

Microglia and other myeloid cells in CNS health and disease

Adithya Gopinath, Anthony Collins, Habibeh Khoshbouei, Wolfgang J. Streit

Department of Neuroscience, University of Florida College of Medicine, Gainesville, FL
32610

JPET # 265058

Microglia and other myeloid cells in CNS health and disease

Adithya Gopinath, Anthony Collins, Habibeh Khoshbouei, Wolfgang J. Streit

Department of Neuroscience, University of Florida College of Medicine, Gainesville, FL
32610

Corresponding Author:

Adithya Gopinath

adithya@ufl.edu

Number of Pages (incl. References): 14

Number of Tables: 0

Number of Figures: 2

Number of References: 68

Abbreviations: No non-standard abbreviations used.

Recommended Section Assignment: Special Section Articles

JPET # 265058

Abstract

Mononuclear macrophages derived from the bone marrow (myeloid cells) are key cellular components of the innate immune system in different organs. In this mini-review, we are focused on both brain and blood macrophages known as microglia and monocytes, respectively. We provide a succinct summary of the cells' functions under both normal and pathological conditions, with particular reference to common neurodegenerative disorders, Alzheimer's and Parkinson's disease.

Significance Statement

In this mini-review we aim to summarize available literature on microglial and myeloid involvement in CNS disease, directing the reader towards relevant and translatable interpretations of myeloid cell function in CNS health and neurodegeneration.

Introduction

Mononuclear macrophages are essential cellular components of the innate immune system, and as such they are present in all organs. They are called 'mononuclear' to distinguish them from polymorphonuclear macrophages, also known as neutrophils, which are residents of blood tissue along with multiple other lineages of immune cells, both innate and adaptive. The term 'myeloid' refers to the cells' origin from the bone marrow and ultimately from the yolk sac from which the earliest macrophage progenitors arise. These primordial, so-called fetal macrophages (or early myeloid progenitors) are present throughout the embryo and fetus beginning during early developmental stages. Myeloid cells arise as the result of distinct hematopoietic waves in the embryonic yolk sac, followed by the embryonic liver and finally bone marrow resident stem cells (Goldmann et al., 2016). Microglia, the resident myeloid cells in the central nervous system (CNS), arise from primitive yolk-sac macrophages that engraft the developing neuroectoderm destined to become the CNS parenchyma. As the embryo develops, subsequent hematopoietic events generate additional populations of both tissue-resident and circulating macrophages (Kierdorf et al., 2019).

In adults, depending on which organ they occupy mononuclear phagocytes may assume characteristic morphologies that reflect a given organ's or tissue's unique cytoarchitecture. A good example of tissue-specific morphology are microglial cells, which reside in the CNS parenchyma, and like all neural cells there, they are process-bearing (ramified) cells. In the blood, monocytes represent the second most prevalent group of macrophages in addition to neutrophils. Unlike microglia, monocytes are rounded similar to all other blood leukocytes. We use the term 'myeloid cells' to refer collectively to microglia, other CNS-associated macrophages, and monocytes as they are closely related cell types. They are distinguished most easily by their locations in the healthy state: microglia reside in the CNS parenchyma, other CNS-associated macrophages reside outside the CNS parenchyma but inside the skull, and monocytes reside in the bloodstream.

CNS-associated macrophages and CNS-resident microglia

We only briefly describe CNS-associated macrophages, directing the interested reader to comprehensive recent reviews on these cells (Mrdjen et al., 2018; Kierdorf et al., 2019). In essence, CNS-associated macrophages refer to cells located within structures that border the CNS parenchyma, i.e. the perivascular space, the leptomeningeal (subarachnoid) space, and the choroid plexus. The perivascular and leptomeningeal spaces are compartments containing cerebrospinal fluid (CSF), produced by the choroid plexus, and they serve as drainage and barriers systems of the CSF. They are somewhat analogous to lymphatic channels in other tissues. Their existence serves important protective roles for the CNS in that they afford a means for eliminating waste products while also providing an immunological border preventing entry of infectious microorganisms or blood borne leukocytes into the CNS parenchyma. They are an essential component of CNS border security, so to speak. The security guards are the macrophages that reside there, which are the perivascular macrophages, leptomeningeal macrophages, and choroid plexus macrophages (epiplexus or Kolmer cells). Further

details and sub-classifications of these guardian cells are described elsewhere (Kierdorf et al., 2019).

In the context of CNS disease or injury, other CNS macrophages can present CNS antigens or may serve as entry points for peripheral immune cell infiltration. Microglia, the CNS-resident macrophages, participate in maintaining parenchymal homeostasis and respond to insult or injury, also acting as gatekeepers for entry of other peripheral immune subtypes (Kierdorf et al., 2019; Van Hove et al., 2019). CNS myeloid cells are also present in the form of choroid plexus macrophages. As choroid plexus ependymal cells are responsible for maintaining and filtering CSF, choroid plexus-associated macrophages and dendritic cells are ideally positioned to encounter CNS antigens and cytokines released into CSF (Ransohoff and Engelhardt, 2012). Additionally, CNS antigen presentation in cervical lymph nodes, even in the absence of dedicated lymphatic vessels in the CNS, may occur via CSF drainage to the nasal mucosa. Finally, while choroid plexus-associated macrophages have not been shown to traffic to cervical lymph nodes for antigen presentation, both choroid plexus macrophages and dendritic cells have been shown to present CNS antigens during CNS disease such as multiple sclerosis (Ransohoff and Engelhardt, 2012).

The meninges, composed of three distinct tissue layers, play host to resident meningeal macrophages and also to a variety of other peripheral leukocytes (Wilson et al., 2010). Bathed in CSF, meningeal macrophages in the subarachnoid space are key antigen presenting cells in the context of pathological conditions and participate in local control of CNS pathogens (Wilson et al., 2010; Van Hove et al., 2019). Perivascular macrophages are particularly specialized to the CNS microenvironment, arising from an embryonic hematopoietic wave distinct from that giving rise to microglia. Similar to choroid plexus and meningeal macrophages, perivascular macrophages are bathed in CSF; however, unlike other CNS-associated macrophage populations, perivascular cells are very close to most CNS regions being separated from the parenchyma only by a basement membrane (Graeber and Streit, 1990b). They are ideally positioned to sample the changing local CNS milieu.

Mononuclear macrophages, both CNS and peripheral, balance between removal and non-removal of damaged or injured cells in efferocytosis – the balance between “eat-me” and “don’t-eat-me” signals (Lagasse and Weissman, 1994; Li, 2012; Weiskopf et al., 2016). Dysfunction in these and associated pathways may induce immune dysregulation which could contribute to CNS diseases and their peripheral correlates.

Peripheral myeloid cell subtypes

In contrast to both CNS-resident microglia and CNS-associated macrophages, peripheral myeloid cells (monocytes), are continuously repopulated by hematopoietic stem cells (HSCs) in the bone marrow niche. Thus monocytes have a much shorter lifespan than microglia in the CNS parenchyma, which are thought to be long-lived (Lawson et al., 1992; Askew et al., 2017). Normally, monocytes reside in the blood stream for relatively short periods before migrating into various other organs and tissues where they differentiate into tissue-specific macrophages. A notable exception to this is the CNS, which does not allow for entry of monocytes under normal conditions. This can change dramatically in acute pathological situations, such as CNS trauma and stroke (Streit et

al., 1998; Gelderblom et al., 2009; Hu et al., 2012), but also in chronic diseases, notably in multiple sclerosis where a massive influx of mononuclear cells (both monocytes and lymphocytes) from the bloodstream occurs. This type of immense and rapid mononuclear cell infiltration causes pathology; it represents an autoimmune attack directed against certain myelin antigens that, for reasons unknown, have become encephalitogenic, i.e. causative of encephalitic neuroinflammation. Mononuclear cell infiltration in multiple sclerosis represents the best example of a CNS autoimmune disease, and this kind of autoantigen-induced encephalitic neuroinflammation causes debilitating demyelination. There is no evidence to suggest that this type of autoimmune neuroinflammatory response occurs in neurodegenerative diseases (see below).

Neuroimmune cells in the healthy brain

Monocytes and other blood leukocytes are notably absent from the normal CNS parenchyma. Instead, the CNS has evolved to harbor its own specialized immune cells called microglia (Graeber and Streit, 1990a). Microglia are distributed ubiquitously throughout the entire CNS and they continuously monitor the parenchymal microenvironment on the lookout for perturbations that may disrupt homeostasis (Figure 1A). As mononuclear phagocytes they are the primary source of endogenous brain macrophages, and as such their key role is clearance of debris which facilitates reorganization of neuronal circuits and trigger repair (Figure 1B,C) (Neumann et al., 2009). Thus phagocytosis of debris serves not only the purpose maintaining a clean microenvironment, but it is also important for contributing to normal neuronal functioning allowing changes in neuronal connectivity and synaptic plasticity (Graeber, 2010; Paolicelli et al., 2011). Perhaps the most important and essential function of microglia is neuroprotection (Streit, 2002), which is no different from that of other neuron-supporting glial cells. Phagocytosis of microorganisms in the parenchyma is one way for microglia to be neuroprotective. However, viruses and bacteria do not easily get past the aforementioned ‘border security’ of meninges, CSF, and other CNS-associated macrophages. Thus, the CNS is well protected by different layers of structural and immunological barriers, and brain infections are not very common in adults with healthy immune systems. However, these barriers break down when individuals become immunocompromised, HIV/AIDS being probably the best example.

Neuroimmune cell function in the diseased brain

Microglial activation occurs when perturbations of CNS homeostasis occur, usually in the form of acute traumatic or ischemic insults or onset of disease (Kreutzberg, 1996). Microglial activation represents the first and quintessential cellular response to CNS injury, and as such, the first step in the wound healing process that ensues over subsequent days and weeks (Figure 1B). Inflammation is defined as the cellular response to injury (Streit et al., 2014), thus microglial activation represents a neuroinflammatory response of CNS endogenous cells. Exogenous (blood-borne) immune cells may also become involved and enter into the damaged CNS if indeed the injury involves BBB breakdown (Figure 1D), which allows blood leukocytes to migrate into the parenchyma. Endogenous neuroinflammation in the CNS is also known as ‘reactive gliosis’, a term that

refers to both microglial and astrocytic mobilization after injury or disease (Figure 1B). In terms of chronic CNS neurodegenerative disease, it is worth noting that both AD and PD have an endogenous neuroinflammatory component as part of their neuropathology, but neither of these conditions are inflammatory diseases, meaning inflammation is not the central cause of degenerative phenomena. In both AD and PD, microglia exhibit macrophage responses, i.e. in AD microglia become activated in response to amyloid aggregation (foreign body response, Figure 2B) (Streit et al., 2018); in PD microglia become activated in response to dying dopaminergic neurons (necroptosis) (Figure 2C) and release of neuromelanin pigment in the SN (Langston et al., 1999; Imamura et al., 2003; Gordon et al., 2018).

Alzheimer's Disease

Our discussion here will be restricted to post-mortem findings from human beings with late-onset Alzheimer's Disease (LOAD), which is the most common form of AD and accounts for more than 95% of all cases. It is beyond the scope of the current review to discuss the many transgenic AD mouse models that have been developed because these models attempt to model certain molecular aspects of AD but do not recapitulate human disease progression. With regard to LOAD, it is important to realize that the condition is marked by a prolonged preclinical phase during which individuals are non-symptomatic but develop lesions identical to those seen in clinical, symptomatic disease although to a lesser extent (Price et al., 2009; Braak and Del Tredici, 2015). Recent work focused on delineating the sequence of pathological lesion development during the preclinical phase of LOAD in non-selected individuals (Streit et al., 2018) has clearly shown that appearance of neurofibrillary degeneration precedes amyloid β protein (A β) deposition confirming the results of others generated in much larger human cohorts (Braak et al., 2011). Our work has also shown that microglial activation (neuroinflammation) occurs only after both neurofibrillary degeneration (NFD) and amyloid aggregates are already present (Figure 2B). Thus, as mentioned above, it is clear that onset of NFD is not the result of neuroinflammation, as claimed in the past (McGeer and McGeer, 2001; Hardy and Selkoe, 2002; Heneka et al., 2015), but that the inflammatory component of LOAD occurs only after aggregation of soluble A β into insoluble amyloid has occurred (Streit et al., 2018). This transformation of A β into amyloid represents the formation of a foreign body (Gray et al., 1990) that attracts the attention of CNS microglia because their job as mononuclear phagocytes is to remove undesirable material, including foreign bodies, from the extracellular space as shown in Figure 2B. The formation of insoluble amyloid aggregates causes microglia to become activated and form phagocytic cell clusters around amyloid deposits producing the characteristic inflammatory neuropathology described in the literature for over thirty years (McGeer et al., 1987; Itagaki et al., 1989; Perlmuter et al., 1990; Sasaki et al., 1997; Rogers et al., 2002). Importantly, the neuroinflammatory component of LOAD is limited to involvement of microglia and does not involve parenchymal infiltration of other CNS-associated macrophages or peripheral monocytes. Microglial activation is directed towards amyloid elimination, and not towards neurodegeneration. That said, one cannot exclude the possibility that myeloid cells other than microglia may be affected by LOAD pathophysiology. The literature in human tissues, as well as studies in cultured human cells, indicates that blood brain barrier (BBB)

function in AD patients is compromised (Alafuzoff et al., 1983; Wada, 1998; Farrall and Wardlaw, 2009). Peripheral monocytes and macrophages in AD patients show altered phagocytic and chemotactic activity, when exposed to amyloid-beta aggregates *in vitro* (Fiala et al., 2005). In this context, defective BBB function might conceivably allow peripheral myeloid cells to infiltrate the CNS, but an invasion of peripheral immune cells has not been demonstrated in humans with AD.

What is the relationship between microglia and neurofibrillary degeneration in LOAD? It has been shown that areas in the human AD brain containing high levels of NFD do not show presence of activated microglia (Streit et al., 2009). Instead, these regions are characterized by abundant presence of so-called dystrophic microglia (Figure 2B), which are cells that display an abnormal morphology thought to reflect cell senescence (Streit et al., 2004). Microglial dystrophy increases with normal aging but becomes highly prevalent in LOAD, as well as in Down syndrome (Xue and Streit, 2011), where it is prominently co-localized with NFD. In fact, it appears that during preclinical and clinical LOAD the extent of microglial dystrophy largely parallels the extent of NFD, suggesting that both degenerative phenomena may have a common underlying cause related to advanced age, the greatest risk factor of LOAD (Streit, 2020).

Parkinson's Disease

Our discussion of myeloid cell function and dysregulation in Parkinson's disease (PD) will focus on potential myeloid cell interactions with the midbrain dopaminergic regions, with relevant forays into secreted chemical mediators that may provide clues to altered peripheral immune states in PD patients and animal models of PD-like degeneration. In some human studies as well as in rodent models of PD, immunological changes are observable in the microglial secretome, microglial morphology and both CNS and peripheral cytokine and chemokine concentrations (Vawter et al., 1996; Hurley et al., 2003; Gordon et al., 2018). Post-mortem analysis of human brain tissues shows altered cytokine and chemokine secretion profiles compared to healthy, age-matched controls (Blum-Degen et al., 1995; Vawter et al., 1996; Nagatsu et al., 2000; Mount et al., 2007). Given, however, that secreted signaling factors can have varying effects on immune cells, depending on tissue-specific cues and cytokine combinations and concentrations, the presence of elevated cytokines in post-mortem PD brain tissues does not necessarily imply an inflammatory scenario (Mantovani et al., 2004; Lucin and Wyss-Coray, 2009). Indeed, the myeloid cell response to insult, injury or cell death varies depending on the milieu chemical cues in the microenvironment. The terms "activation" and "inflammation", often used to describe pathological changes in myeloid cells, fail to convey the complexity of the extremes of monocyte and microglial function ranging from adaptive inflammation to injury resolution and healing (Streit et al., 2014; Lively and Schlichter, 2018).

Degenerative conditions affecting the CNS such as AD (above) and PD induce changes in a wide range of cytokines with pleiotropic effects – cytokine milieus that induce either proinflammatory or regenerative microglial phenotypes have both been repeatedly shown over the last three decades to be altered in Parkinson's disease, with studies often contradicting one another. Reflecting complexity of the condition, the combinations of cytokines present naturally depend on the nature of the insult. In the context of PD, the insult is chronic and would result in a distinct cytokine milieu (Blum-Degen et al., 1995;

Mount et al., 2007). Adding interest to the story is the fact that the BBB, traditionally thought to separate CNS from periphery into discrete compartments, in fact acts as an educational gate (Park et al., 2016) which allows CNS-originating signaling molecules, and even antigens, to enter systemic circulation with the potential to influence the peripheral immune system (Figure 2C). That CNS molecules can exit the parenchyma, enter peripheral circulation, and retain the ability to modulate peripheral immune homeostasis in much the same way that cytokines modulate microglial response raises intriguing questions for neurodegeneration. Indeed, studies within the last 5 years have begun to investigate such changes in peripheral myeloid cell populations in PD, along with the beginnings of mechanistic studies in animal models. While the infiltration of peripheral myeloid cells into the CNS in PD remains largely unexplored, PD patients have been shown to exhibit elevated circulating classical monocytes along with a concurrent increase in CCL2 (Grozdanov et al., 2014), a chemokine which promotes myeloid cell egress from bone marrow (Fujimura et al., 2015). Further investigation revealed that inhibition of the CD95/CD95L axis, commonly considered an apoptosis pathway (Park et al., 2003), resulted in an elevation in of non-classical monocytes (Grozdanov et al., 2014), possibly restoring a more-healthy immune profile. Gao and colleagues went on to demonstrate that both genetic and pharmacological inhibition of this pathway is protective against MPTP-induced degeneration (Gao et al., 2015). Parallel studies to assess stem cell proliferation reveal that circulating HSCs from PD patients, both idiopathic and familial, have the potential to generate monocytes more robustly and with greater expression of chemokine receptor CCR2 than healthy controls, along with dramatically elevated CCR2 expression (Funk et al., 2013). In other words, expression of both chemokine (CCL2) and receptor (CCR2) are elevated in suggesting peripheral monocyte dysregulation at a hematopoietic level. These alterations in the basic characteristics of myeloid cell genesis in Parkinson's disease, the implications of which remain unexplored, warrant detailed investigation.

Resident microglia in Parkinson's disease have been pointed out as culprits in dopaminergic neuron degeneration (Lecours et al., 2018), but evidence is often contradictory. In an attempt to address this question, Kim et al studied LRRK2 mutant model systems and human peripheral myeloid cells derived from Parkinson's patients to determine whether phagocytic activity differed from healthy controls. Guided by genetic studies in flies, it was demonstrated that murine microglia and human macrophages exhibited increased phagocytic capacity, which was abrogated by knockdown of LRRK2 (Kim et al., 2018). These data suggest that, rather than inducing inflammation and contributing to neuronal degeneration, microglia may elevate their phagocytic activity in PD possibly as an attempt to keep up with to ongoing necroptosis (Figure 2C). A separate study of gene expression sought to use peripheral myeloid cells as a proxy for CNS myeloid cells, considering likely differences between mononuclear phagocytes as a result of residence in different tissue compartments. Inspired by GWAS studies which indicated the potential for an inflammatory state in PD (Nalls et al., 2011), differential gene expression in PD patients and healthy control subjects did not reveal such an association (Schlachetzki et al., 2018). This study was not, however, adequately powered to detect a difference between PD patients and controls; a larger cohort study could reveal different results and show dysregulated myeloid cell function.

In the discussion of CNS and peripheral myeloid cells, it is important to note that the innate immune system *may* cascade into adaptive immune activation and should be at least considered in the context of CNS and peripheral myeloid cells in Parkinson's disease. Groups exploring this area have studied PD patients (Sulzer et al., 2017), post-mortem tissue (Gonzalez et al., 2013) and in vitro systems to explore possible mechanisms of interaction between myeloid cells and T cells in PD (Sommer et al., 2016). Sulzer et al. have elaborated on the identification of antigen-specific T cells in PD patients (Sulzer et al., 2017), while another group has shown evidence of CD4+ T cell infiltration in post-mortem PD midbrain tissue (Gonzalez et al., 2013). However, much work remains to be done to identify the role, if any, played by these cells in PD, as T cells are known to patrol the CNS compartment in homeostasis (Ransohoff and Engelhardt, 2012) and during acute injury/infection (Russo and McGavern, 2015). Experimental conditions using in vivo and in vitro overexpression systems (Sommer et al., 2016) have shown that T-cell-mediated suppression of microglial phagocytosis occurs in the presence of alpha synuclein burden, and that genetic ablation of T-cell development appears to resolve this. One should note that alpha synuclein overexpression does not represent the human condition and is of limited translational value – results should be interpreted as such. Given that T-cell infiltration into the midbrain in Parkinson's disease is a rare occurrence, T-cell mediated suppression of microglial function in the physiological condition would be limited and likely would not impede microglial function.

Conclusions

A diverse and well-documented set of microglial responses occur in the CNS of patients with degenerative disease. In more recent years the peripheral myeloid cell response in Alzheimer's and Parkinson's disease has been more extensively characterized, in which complex and yet-to-be-understood changes occur. Such a complex mix of phenotypic, chemotactic, secretory and functional myeloid cell responses suggests a level of complexity in neurodegenerative conditions that cannot be described simply as "inflammation." Immune dysregulation would be a better term to capture the complexity of immunological changes in various diseases.

Acknowledgements

We would like to thank Dr. Eduardo Candelario-Jalil (University of Florida, Department of Neuroscience) for the generous gift of MCAO stroke tissues, and Dr. Fatemeh Shaerzadeh (University of Florida, Department of Neuroscience) for assistance with imaging. We acknowledge funding from UF Research Developmental Fund (Moonshot) from the Fixel Institute for Neurological Diseases, and from NIH Grants 2R01NS071122-07A1 to H. Khoshbouei, and shared instrumentation grant S10OD020026, grant 8AZ19 awarded to W.J. Streit from the Florida Department of Health, and multi-PI 5R21NS103108-02 awarded to H. Khoshbouei and W.J. Streit. We also acknowledge funding from the National Center for Advancing Translational Sciences of the National Institutes of Health under University of Florida Clinical and Translational Science Awards TL1TR001428 and UL1TR001427. The content is solely the

JPET # 265058

responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

JPET # 265058

Authorship Contributions:

Participated in research design: Adithya Gopinath, Habibeh Khoshbouei, Wolfgang J. Streit

Conducted experiments: Adithya Gopinath, Anthony Collins, Habibeh Khoshbouei, Wolfgang J. Streit

Contributed new reagents or analytic tools:

Performed data analysis: Adithya Gopinath

Wrote or contributed to the writing of the manuscript: Adithya Gopinath, Habibeh Khoshbouei, Wolfgang J. Streit

References

- Alafuzoff I, Adolfsson R, Bucht G and Winblad B (1983) Albumin and immunoglobulin in plasma and cerebrospinal fluid, and blood-cerebrospinal fluid barrier function in patients with dementia of Alzheimer type and multi-infarct dementia. *J Neurol Sci* **60**:465-472.
- Askew K, Li K, Olmos-Alonso A, Garcia-Moreno F, Liang Y, Richardson P, Tipton T, Chapman MA, Riecken K, Beccari S, Sierra A, Molnar Z, Cragg MS, Garaschuk O, Perry VH and Gomez-Nicola D (2017) Coupled Proliferation and Apoptosis Maintain the Rapid Turnover of Microglia in the Adult Brain. *Cell Rep* **18**:391-405.
- Blum-Degen D, Muller T, Kuhn W, Gerlach M, Przuntek H and Riederer P (1995) Interleukin-1 beta and interleukin-6 are elevated in the cerebrospinal fluid of Alzheimer's and de novo Parkinson's disease patients. *Neurosci Lett* **202**:17-20.
- Braak H and Del Tredici K (2015) The preclinical phase of the pathological process underlying sporadic Alzheimer's disease. *Brain : a journal of neurology* **138**:2814-2833.
- Braak H, Thal DR, Ghebremedhin E and Del Tredici K (2011) Stages of the pathologic process in Alzheimer disease: age categories from 1 to 100 years. *Journal of neuropathology and experimental neurology* **70**:960-969.
- Farrall AJ and Wardlaw JM (2009) Blood-brain barrier: ageing and microvascular disease--systematic review and meta-analysis. *Neurobiol Aging* **30**:337-352.
- Fiala M, Lin J, Ringman J, Kermani-Arab V, Tsao G, Patel A, Lossinsky AS, Graves MC, Gustavson A, Sayre J, Sofroni E, Suarez T, Chiappelli F and Bernard G (2005) Ineffective phagocytosis of amyloid-beta by macrophages of Alzheimer's disease patients. *J Alzheimers Dis* **7**:221-232; discussion 255-262.
- Fujimura N, Xu B, Dalman J, Deng H, Aoyama K and Dalman RL (2015) CCR2 inhibition sequesters multiple subsets of leukocytes in the bone marrow. *Sci Rep* **5**:11664.
- Funk N, Wieghofer P, Grimm S, Schaefer R, Buhning HJ, Gasser T and Biskup S (2013) Characterization of peripheral hematopoietic stem cells and monocytes in Parkinson's disease. *Mov Disord* **28**:392-395.
- Gao L, Brenner D, Llorens-Bobadilla E, Saiz-Castro G, Frank T, Wieghofer P, Hill O, Thiemann M, Karray S, Prinz M, Weishaupt JH and Martin-Villalba A (2015) Infiltration of circulating myeloid cells through CD95L contributes to neurodegeneration in mice. *J Exp Med* **212**:469-480.
- Gelderblom M, Leyboldt F, Steinbach K, Behrens D, Choe CU, Siler DA, Arumugam TV, Orthey E, Gerloff C, Tolosa E and Magnus T (2009) Temporal and spatial dynamics of cerebral immune cell accumulation in stroke. *Stroke* **40**:1849-1857.
- Goldmann T, Wieghofer P, Jordao MJ, Prutek F, Hagemeyer N, Frenzel K, Amann L, Staszewski O, Kierdorf K, Krueger M, Locatelli G, Hochgerner H, Zeiser R, Epelman S, Geissmann F, Priller J, Rossi FM, Bechmann I, Kerschensteiner M, Linnarsson S, Jung S and Prinz M (2016) Origin, fate and dynamics of macrophages at central nervous system interfaces. *Nat Immunol* **17**:797-805.
- Gonzalez H, Contreras F, Prado C, Elgueta D, Franz D, Bernales S and Pacheco R (2013) Dopamine receptor D3 expressed on CD4+ T cells favors neurodegeneration of dopaminergic neurons during Parkinson's disease. *J Immunol* **190**:5048-5056.
- Gordon R, Albornoz EA, Christie DC, Langley MR, Kumar V, Mantovani S, Robertson AAB, Butler MS, Rowe DB, O'Neill LA, Kanthasamy AG, Schroder K, Cooper MA and Woodruff TM (2018) Inflammasome inhibition prevents alpha-synuclein pathology and dopaminergic neurodegeneration in mice. *Sci Transl Med* **10**.
- Graeber MB (2010) Changing face of microglia. *Science* **330**:783-788.
- Graeber MB and Streit WJ (1990a) Microglia: immune network in the CNS. *Brain pathology* **1**:2-5.
- Graeber MB and Streit WJ (1990b) Perivascular microglia defined. *Trends in neurosciences* **13**:366.

JPET # 265058

- Gray F, Vinters HV, Le Noan H, Salama J, Delaporte P and Poirier J (1990) Cerebral amyloid angiopathy and granulomatous angiitis: immunohistochemical study using antibodies to the Alzheimer A4 peptide. *Hum Pathol* **21**:1290-1293.
- Grozdanov V, Bliederhaeuser C, Ruf WP, Roth V, Fundel-Clemens K, Zondler L, Brenner D, Martin-Villalba A, Hengerer B, Kassubek J, Ludolph AC, Weishaupt JH and Danzer KM (2014) Inflammatory dysregulation of blood monocytes in Parkinson's disease patients. *Acta Neuropathol* **128**:651-663.
- Hardy J and Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* **297**:353-356.
- Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM, Herrup K, Frautschy SA, Finsen B, Brown GC, Verkhratsky A, Yamanaka K, Koistinaho J, Latz E, Halle A, Petzold GC, Town T, Morgan D, Shinohara ML, Perry VH, Holmes C, Bazan NG, Brooks DJ, Hunot S, Joseph B, Deigendesch N, Garaschuk O, Boddeke E, Dinarello CA, Breitner JC, Cole GM, Golenbock DT and Kummer MP (2015) Neuroinflammation in Alzheimer's disease. *The Lancet Neurology* **14**:388-405.
- Hu X, Li P, Guo Y, Wang H, Leak RK, Chen S, Gao Y and Chen J (2012) Microglia/macrophage polarization dynamics reveal novel mechanism of injury expansion after focal cerebral ischemia. *Stroke* **43**:3063-3070.
- Hurley SD, O'Banion MK, Song DD, Arana FS, Olschowka JA and Haber SN (2003) Microglial response is poorly correlated with neurodegeneration following chronic, low-dose MPTP administration in monkeys. *Experimental Neurology* **184**:659-668.
- Imamura K, Hishikawa N, Sawada M, Nagatsu T, Yoshida M and Hashizume Y (2003) Distribution of major histocompatibility complex class II-positive microglia and cytokine profile of Parkinson's disease brains. *Acta Neuropathol* **106**:518-526.
- Itagaki S, McGeer PL, Akiyama H, Zhu S and Selkoe D (1989) Relationship of microglia and astrocytes to amyloid deposits of Alzheimer disease. *Journal of neuroimmunology* **24**:173-182.
- Kierdorf K, Masuda T, Jordao MJC and Prinz M (2019) Macrophages at CNS interfaces: ontogeny and function in health and disease. *Nat Rev Neurosci* **20**:547-562.
- Kim KS, Marcogliese PC, Yang J, Callaghan SM, Resende V, Abdel-Messih E, Marras C, Visanji NP, Huang J, Schlossmacher MG, Trinkle-Mulcahy L, Slack RS, Lang AE, Canadian Lrrk2 in Inflammation T and Park DS (2018) Regulation of myeloid cell phagocytosis by LRRK2 via WAVE2 complex stabilization is altered in Parkinson's disease. *Proc Natl Acad Sci U S A* **115**:E5164-E5173.
- Kreutzberg GW (1996) Microglia: a sensor for pathological events in the CNS. *Trends in neurosciences* **19**:312-318.
- Lagasse E and Weissman IL (1994) bcl-2 inhibits apoptosis of neutrophils but not their engulfment by macrophages. *J Exp Med* **179**:1047-1052.
- Langston JW, Forno LS, Tetrad J, Reeves AG, Kaplan JA and Karluk D (1999) Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann Neurol* **46**:598-605.
- Lawson LJ, Perry VH and Gordon S (1992) Turnover of resident microglia in the normal adult mouse brain. *Neuroscience* **48**:405-415.
- Lecours C, Bordeleau M, Cantin L, Parent M, Paolo TD and Tremblay ME (2018) Microglial Implication in Parkinson's Disease: Loss of Beneficial Physiological Roles or Gain of Inflammatory Functions? *Front Cell Neurosci* **12**:282.
- Li W (2012) Eat-me signals: keys to molecular phagocyte biology and "appetite" control. *J Cell Physiol* **227**:1291-1297.

JPET # 265058

- Lively S and Schlichter LC (2018) Microglia Responses to Pro-inflammatory Stimuli (LPS, IFN γ +TNF α) and Reprogramming by Resolving Cytokines (IL-4, IL-10). *Front Cell Neurosci* **12**:215.
- Lucin KM and Wyss-Coray T (2009) Immune activation in brain aging and neurodegeneration: too much or too little? *Neuron* **64**:110-122.
- Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A and Locati M (2004) The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* **25**:677-686.
- McGeer PL, Itagaki S, Tago H and McGeer EG (1987) Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. *Neuroscience letters* **79**:195-200.
- McGeer PL and McGeer EG (2001) Inflammation, autotoxicity and Alzheimer disease. *Neurobiology of aging* **22**:799-809.
- Mount MP, Lira A, Grimes D, Smith PD, Faucher S, Slack R, Anisman H, Hayley S and Park DS (2007) Involvement of interferon-gamma in microglial-mediated loss of dopaminergic neurons. *J Neurosci* **27**:3328-3337.
- Mrdjen D, Pavlovic A, Hartmann FJ, Schreiner B, Utz SG, Leung BP, Lelios I, Heppner FL, Kipnis J, Merkler D, Greter M and Becher B (2018) High-Dimensional Single-Cell Mapping of Central Nervous System Immune Cells Reveals Distinct Myeloid Subsets in Health, Aging, and Disease. *Immunity* **48**:380-395 e386.
- Nagatsu T, Mogi M, Ichinose H and Togari A (2000) Changes in cytokines and neurotrophins in Parkinson's disease. *J Neural Transm Suppl*:277-290.
- Nalls MA, Plagnol V, Hernandez DG, Sharma M, Sheerin UM, Saad M, Simon-Sanchez J, Schulte C, Lesage S, Sveinbjornsdottir S, Stefansson K, Martinez M, Hardy J, Heutink P, Brice A, Gasser T, Singleton AB and Wood NW (2011) Imputation of sequence variants for identification of genetic risks for Parkinson's disease: a meta-analysis of genome-wide association studies. *Lancet* **377**:641-649.
- Neumann H, Kotter MR and Franklin RJ (2009) Debris clearance by microglia: an essential link between degeneration and regeneration. *Brain : a journal of neurology* **132**:288-295.
- Paolicelli RC, Bolasco G, Pagani F, Maggi L, Scianni M, Panzanelli P, Giustetto M, Ferreira TA, Guiducci E, Dumas L, Ragozzino D and Gross CT (2011) Synaptic pruning by microglia is necessary for normal brain development. *Science* **333**:1456-1458.
- Park DR, Thomsen AR, Frevert CW, Pham U, Skerrett SJ, Kiener PA and Liles WC (2003) Fas (CD95) induces proinflammatory cytokine responses by human monocytes and monocyte-derived macrophages. *J Immunol* **170**:6209-6216.
- Park HS, Park MJ and Kwon MS (2016) Central Nervous System-Peripheral Immune System Dialogue in Neurological Disorders: Possible Application of Neuroimmunology in Urology. *Int Neurourol J* **20**:S8-14.
- Perlmuter LS, Barron E and Chui HC (1990) Morphologic association between microglia and senile plaque amyloid in Alzheimer's disease. *Neuroscience letters* **119**:32-36.
- Price JL, McKeel DW, Jr., Buckles VD, Roe CM, Xiong C, Grundman M, Hansen LA, Petersen RC, Parisi JE, Dickson DW, Smith CD, Davis DG, Schmitt FA, Markesbery WR, Kaye J, Kurlan R, Hulette C, Kurland BF, Higdon R, Kukull W and Morris JC (2009) Neuropathology of nondemented aging: presumptive evidence for preclinical Alzheimer disease. *Neurobiology of aging* **30**:1026-1036.
- Ransohoff RM and Engelhardt B (2012) The anatomical and cellular basis of immune surveillance in the central nervous system. *Nat Rev Immunol* **12**:623-635.
- Rogers J, Strohmeyer R, Kovelowski CJ and Li R (2002) Microglia and inflammatory mechanisms in the clearance of amyloid beta peptide. *Glia* **40**:260-269.
- Russo MV and McGavern DB (2015) Immune Surveillance of the CNS following Infection and Injury. *Trends Immunol* **36**:637-650.

JPET # 265058

- Sasaki A, Yamaguchi H, Ogawa A, Sugihara S and Nakazato Y (1997) Microglial activation in early stages of amyloid beta protein deposition. *Acta neuropathologica* **94**:316-322.
- Schlachetzki JCM, Prots I, Tao J, Chun HB, Saijo K, Gosselin D, Winner B, Glass CK and Winkler J (2018) A monocyte gene expression signature in the early clinical course of Parkinson's disease. *Sci Rep* **8**:10757.
- Sommer A, Fadler T, Dorfmeister E, Hoffmann AC, Xiang W, Winner B and Prots I (2016) Infiltrating T lymphocytes reduce myeloid phagocytosis activity in synucleinopathy model. *J Neuroinflammation* **13**:174.
- Streit WJ (2002) Microglia as neuroprotective, immunocompetent cells of the CNS. *Glia* **40**:133-139.
- Streit WJ, Braak H, Del Tredici K, Leyh J, Lier J, Khoshbouei H, Eisenloffer C, Muller W and Bechmann I (2018) Microglial activation occurs late during preclinical Alzheimer's disease. *Glia* **66**:2550-2562.
- Streit WJ, Braak H, Xue QS and Bechmann I (2009) Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in Alzheimer's disease. *Acta neuropathologica* **118**:475-485.
- Streit WJ, Khoshbouei H, Bechmann I. (2020) Dystrophic microglia in late-onset Alzheimer's Disease. *Glia* **in press**.
- Streit WJ, Sammons NW, Kuhns AJ and Sparks DL (2004) Dystrophic microglia in the aging human brain. *Glia* **45**:208-212.
- Streit WJ, Semple-Rowland SL, Hurley SD, Miller RC, Popovich PG and Stokes BT (1998) Cytokine mRNA profiles in contused spinal cord and axotomized facial nucleus suggest a beneficial role for inflammation and gliosis. *Experimental neurology* **152**:74-87.
- Streit WJ, Xue QS, Tischer J and Bechmann I (2014) Microglial pathology. *Acta neuropathologica communications* **2**:142.
- Sulzer D, Alcalay RN, Garretti F, Cote L, Kanter E, Agin-Liebes J, Liong C, McMurtrey C, Hildebrand WH, Mao X, Dawson VL, Dawson TM, Oseroff C, Pham J, Sidney J, Dillon MB, Carpenter C, Weiskopf D, Phillips E, Mallal S, Peters B, Frazier A, Lindestam Arlehamn CS and Sette A (2017) T cells from patients with Parkinson's disease recognize alpha-synuclein peptides. *Nature* **546**:656-661.
- Van Hove H, Martens L, Scheyltjens I, De Vlaminck K, Pombo Antunes AR, De Prijck S, Vandamme N, De Schepper S, Van Isterdael G, Scott CL, Aerts J, Berx G, Boeckxstaens GE, Vandenbroucke RE, Vereecke L, Moechars D, Guillems M, Van Ginderachter JA, Saeys Y and Movahedi K (2019) A single-cell atlas of mouse brain macrophages reveals unique transcriptional identities shaped by ontogeny and tissue environment. *Nat Neurosci* **22**:1021-1035.
- Vawter MP, Dillon-Carter O, Tourtellotte WW, Carvey P and Freed WJ (1996) TGFbeta1 and TGFbeta2 concentrations are elevated in Parkinson's disease in ventricular cerebrospinal fluid. *Exp Neurol* **142**:313-322.
- Wada H (1998) Blood-brain barrier permeability of the demented elderly as studied by cerebrospinal fluid-serum albumin ratio. *Intern Med* **37**:509-513.
- Weiskopf K, Jahchan NS, Schnorr PJ, Cristea S, Ring AM, Maute RL, Volkmer AK, Volkmer JP, Liu J, Lim JS, Yang D, Seitz G, Nguyen T, Wu D, Jude K, Guerston H, Barkal A, Trapani F, George J, Poirier JT, Gardner EE, Miles LA, de Stanchina E, Lofgren SM, Vogel H, Winslow MM, Dive C, Thomas RK, Rudin CM, van de Rijn M, Majeti R, Garcia KC, Weissman IL and Sage J (2016) CD47-blocking immunotherapies stimulate macrophage-mediated destruction of small-cell lung cancer. *J Clin Invest* **126**:2610-2620.
- Wilson EH, Weninger W and Hunter CA (2010) Trafficking of immune cells in the central nervous system. *J Clin Invest* **120**:1368-1379.
- Xue QS and Streit WJ (2011) Microglial pathology in Down syndrome. *Acta neuropathologica* **122**:455-466.

JPET # 265058

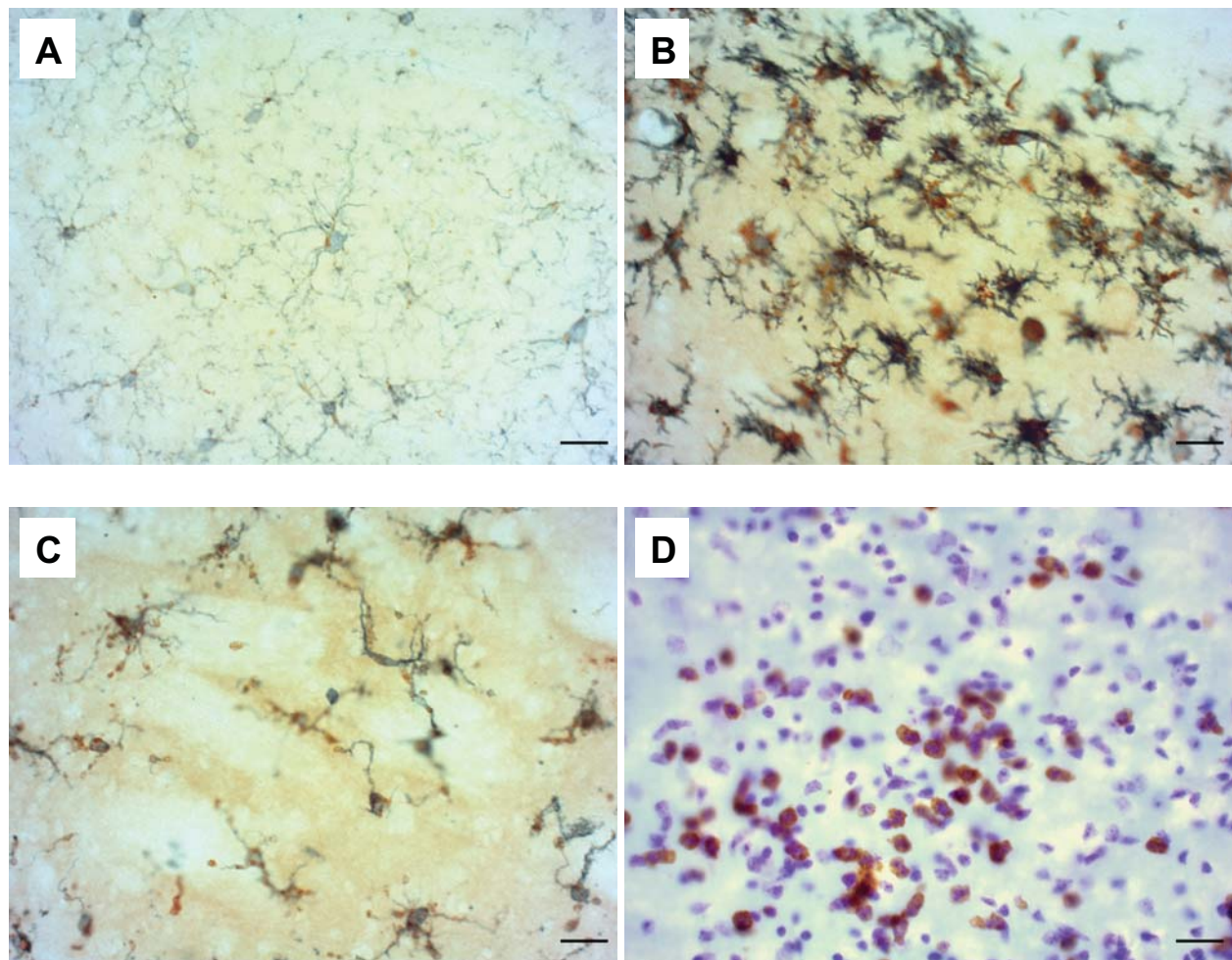
Legends for Figures

Figure 1. Microglia at resting, activated and dystrophic states are seen in MCAO unilateral stroke model brain tissue, accompanied by extensive peripheral myeloid cell infiltration. Within a coronal brain section from a mouse at 24 hours following the stroke in the contralateral hemisphere, A) resting microglia are labeled with myeloid cell marker IBA1 (grey), and a marker of phagocytic and lysosomal function CD68 (brown). B) In the ipsilateral hemisphere, activated microglia at the edge of the lesion display a characteristic phenotype with increased IBA1 and CD68 expression, indicating active phagocytosis. C) Dystrophic, fragmenting microglia can be observed within the stroke core. In addition to microglial activation, lesioned CNS tissue displays D) extensive peripheral myeloid cell infiltration shown by Ly6B.2 immunostaining, specific to infiltrating peripheral myeloid cells. Identification of myeloid cell subtypes can be made using nuclear Nissl counterstain, as monocytes (mononuclear) or neutrophils (polymorphonuclear). (40x magnification images, scale bar represents 20 microns)

Figure 2. Microglia and myeloid cell states in health and disease. While at homeostasis (A) microglial activation in the CNS and alterations in peripheral blood myeloid cells are seldom seen, dramatic changes in immune activation are seen in degenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD). B) In AD, microglia become activated in response to amyloid formation and form phagocytic clusters attempting to remove this insoluble material. Neurofibrillary degeneration does not elicit microglial activation but coincides with microglial dystrophy; C) in PD microglial activation, phagocytic activity, and secretion of soluble signaling molecules are associated with response to dopaminergic neurons undergoing necroptosis in the midbrain. Secreted chemical factors also induce changes in peripheral myeloid populations, altering peripheral immune function and potentially inducing peripheral myeloid participation in dopaminergic degeneration.

JPET # 265058

Figure 1



JPET # 265058

Figure 2

