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Title page

The minimal pharmacophore for silent agonism of $\alpha 7$ nAChR

Roger L. Papke, Kinga Chojnacka, and Nicole A. Horenstein

Department of Pharmacology and Therapeutics, University of Florida, PO Box 100267
Gainesville, FL 32610-0267 (RLP)

Department of Chemistry, University of Florida, PO Box 117200, Gainesville, FL 32611-
7200 (KC, NAH)

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*To whom correspondence should be addressed:

Name: Roger L. Papke

Phone: 352-392-4712

Fax: 352-392-9696

E-mail: rlpapke@ufl.edu

Address: Department of Pharmacology and Therapeutics

University of Florida

P.O. Box 100267

Gainesville, FL 32610-0267

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Abbreviations: ACh, acetylcholine; nAChR, nicotinic acetylcholine receptor; PAM, positive allosteric modulator; D_s, PAM-sensitive closed state ; D_i, PAM-insensitive closed state;

Structure-based names: tetMA, tetramethylammonium; EtriMA, ethyltrimethylammonium; diEdiMA, diethyldimethylammonium; tetEA, tetraethylammonium; (2OHE)-EdiMA, (2-hydroxyethyl)-ethyldimethylammonium; (2OHE)-diEMA, (2-hydroxyethyl)-diethylmethylammonium; (2OHE)-triEA, (2-hydroxyethyl)-triethylammonium; BtriMA, benzyltrimethylammonium; BEdiMA, benzylethyldimethylammonium; BdiEMA, benzyldiethylmethylammonium; diMPyrr, dimethylpyrrolidinium; EMPyrr, ethylmethylpyrrolidinium; diEPyrr, diethylpyrrolidinium; diMPip, dimethylpiperidinium; EMPip, ethylmethylpiperidinium; diEPip, diethylpiperidinium; diMHHA, dimethylhexahydroazepinium; EMHHA, ethylmethylhexahydroazepinium; diEHHA, diethylhexahydroazepinium; diMPP,

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dimethylphenylpiperazinium; EMPP, ethylmethylphenylpiperazinium; diEPP,
diethylphenylpiperazinium

Abstract

The minimum pharmacophore for activation of the human $\alpha 7$ nicotinic acetylcholine receptor (nAChR) is the tetramethylammonium cation. Previous work demonstrated that larger quaternary ammonium compounds such as diethyldimethylammonium or 1-methyl quinuclidine were $\alpha 7$ -selective partial agonists, but additional increase in the size of the ammonium cation or the quinuclidine N-alkyl group by a single carbon to an N-ethyl group led to a loss of efficacy for ion channel activation. We report that, while such compounds are ineffective at inducing the normal channel open state, they nonetheless regulate the induction of specific conformational states normally considered downstream of channel activation. We synthesized several panels of quaternary ammonium nAChR ligands that systematically varied the size of the substituents bonded to the central positively charged nitrogen atom. In these molecular series we found a correlation between the molecular volume of the ligand and/or charge density, and the receptor's preferred distribution amongst conformational states including the closed state, the active state, a non-conducting state that could be converted to an activated state by a positive allosteric modulator (PAM), and a PAM-insensitive non-conducting state. We hypothesize that the changes of molecular volume of an agonist's cationic core subtly impact interactions at the subunit interface constituting the orthosteric binding site in such a way as to regulate the probability of conversions among the conformational states. We define a new minimal pharmacophore for the class of compounds we have termed as "silent agonists", which are able to induce allosteric modulator-dependent activation but not the normal activated state.

Introduction

Binding of ligands to allosteric proteins such as nicotinic acetylcholine receptors (nAChR) regulates the probability with which the protein shifts among various conformational states (Papke, 2014). In the case of ligand-gated ion channels, it is customary to focus on the conformational change associated with channel activation. Agonists are drugs which promote conversion to the channel-activated state(s). In a specific experimental context such as the evaluation of macroscopic currents from cells heterologously expressing a specific receptor subtype, drugs which are less effective at producing channel activation than a reference agonist, such as acetylcholine in the case of nicotinic receptors, are classified as partial agonists.

The structural dissection of a series of compounds to define a receptor's pharmacophore typically addresses efficacy first, defining the molecular elements required to activate the receptor. A second property to be considered is selectivity, defining what is required to activate one receptor subtype but not other closely related subtypes. However, by considering the potential for differential induction of alternative conformational states, including non-conducting (desensitized) states, we can generate expanded and refined pharmacophores. The $\alpha 7$ -type homomeric nicotinic acetylcholine receptor has an especially interesting array of conducting and nonconducting conformational states, regulated by both orthosteric and allosteric ligands (Williams et al., 2011a). It has recently been proposed that the $\alpha 7$ receptor may mediate both channel activation-dependent and channel activation-independent forms of signaling (de Jonge and Ulloa, 2007; Skok, 2009; Kabbani et al., 2013). Therefore, understanding the pharmacophore for selectively inducing specific nonconducting states may be of equal interest and importance as understanding the pharmacophore for inducing open channel states.

The minimal pharmacophore for activating the ion channel of neuronal type nAChR is tetramethylammonium (tetMA, see Figure 1), and the minimal structure for selectively activating $\alpha 7$ nAChR is diethyldimethylammonium (diEdiMA) (Horenstein et al., 2008), with the selectivity arising from replacement of a methyl group with an ethyl group, since ethyltrimethylammonium (EtriMA) is not $\alpha 7$ -selective. In this paper we return to the minimal structures to define the requirements for producing what we have referred to as "silent agonists" (Chojnacka et al., 2013), molecules which have little or no efficacy as orthosteric agonists but can work in concert with positive allosteric modulators (PAMs) to produce channel activation through the induction of modulator-sensitive desensitized states. PAMs therefore serve to reveal the existence of these allosteric-potentiated open states, which otherwise might go undetected.

When activated by the binding of agonist alone, the openings of $\alpha 7$ receptors are extremely brief ($<100 \mu\text{s}$ on average), and appear in isolation (Williams et al., 2012; Pesti et al., 2014). However, openings of receptors with both agonist and PNU-120596 bound occur as protracted bursts with hundreds of intraburst closed events (Williams et al., 2011a), which suggested evidence for an agonist-bound closed state kinetically coupled with a PAM-dependent open state and detectable only if the PAM is also bound. We hypothesized that such a PAM-sensitive closed state (D_s) could also contribute to the population of desensitized receptors under control conditions (Williams et al., 2011a). Comparing the probability of the channel being open during these bursts to the steady-state probability of channels being open under the control condition (same population of channels in the very same patch), the effect of the PAM is to produce large (100,000-fold) intermittent increases in the probability that a given channel will be open. However, the PAM effect on single channels is temporary, and there also exists a more stable closed state that we associate with a PAM-insensitive closed state (D_i). Note that it is not likely that the long closed times between PAM-dependent bursts are strictly due to unbinding of ligands (although in some cases they must be), because at high concentrations of the agonist and PAM, the frequency of bursts decreases and the durations of the interburst closed times increase, associated with decreased macroscopic current (Williams et al., 2011a). In the present study we evaluate macroscopic current responses evoked by several series of structurally related compounds, either applied alone or in combination with PNU-120596 at a concentration which we determined in previous experiments to give us optimal potentiation of ACh-evoked currents (Williams et al., 2011a). The macroscopic currents we record reflect the integrated outcome of the dynamic induction of conducting and nonconducting states (Papke, 2010). The nonconducting states may encompass unliganded states as well as desensitized and possibly blocked states. We identify the minimal structural requirements for silent agonists, compounds which induce only nonconducting states in the absence of the PAM but produce significant currents in the presence of the PAM.

Methods

Commercial reagents

Acetylcholine chloride (ACh), atropine, tetMA chloride, EtriMA iodide, diEdiMA hydroxide solution, tetEA chloride, diMPP iodide, and choline chloride were purchased from Sigma-Aldrich Chemical Company (St. Louis, MO). General reagents for chemical synthesis were purchased from Fischer Scientific or Sigma-Aldrich Chemical Company.

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Cell culture supplies were purchased from Life Technologies (Grand Island, NY). triEMA was purchased from Tokyo Chemical Industry (TCI America, Portland OR). Fresh ACh stock solutions were made in Ringer's solution each day of experimentation. Stock solutions of the test drugs were made in Ringer's solution and kept at 4 °C and used within two days. Working solutions were prepared freshly at the desired concentration from the stored stock.

Synthesis of ammonium compounds

All compounds displayed satisfactory ¹H- and ¹³C-NMR spectra and were found to be >95% purity by NMR methods. (supplemental material)

Quaternary ammonium salts were prepared by reacting commercially available methyl or ethyl amines with methyl iodide or ethyl iodide in tetrahydrofuran or ethanol and purified by recrystallization.

(2-hydroxyethyl)-ethyltrimethylammonium iodide [(2OHE)-EdiMA] (Calas et al., 2000): From 2-dimethylaminoethanol and ethyl iodide. White hygroscopic solid.

(2-hydroxyethyl)-diethylmethylammonium iodide [(2OHE)-diEMA] (Green et al., 2009): From 2-diethylaminoethanol and methyl iodide. White solid.

(2-Hydroxyethyl)-triethylammonium iodide [(2OHE)-triEA] (Kasuga et al., 1969): From 2-diethylaminoethanol and ethyl iodide. White solid.

Benzyltrimethylammonium iodide [BtriMA] (Ito et al., 2005): From dimethylbenzylamine and methyl iodide. White solid.

Benzylethyltrimethylammonium iodide [BEdiMA] (Short and Biermacher, 1962): From dimethylbenzylamine and ethyl iodide. White hygroscopic solid.

Benzyl-diethylmethylammonium iodide [BdiEMA] (Wempe, 2001): From ethylmethylbenzylamine and ethyl iodide. White solid.

Dimethylpyrrolidinium iodide [diMPyrr] (MacFarlane et al., 1999): From 1-methylpyrrolidine and methyl iodide. White solid.

Ethylmethylpyrrolidinium iodide [EMPyrr] (MacFarlane et al., 1999): From 1-methylpyrrolidine and ethyl iodide.

Diethylpyrrolidinium iodide [diEPyrr] (Huenig, 1957): From 1-ethylpyrrolidine and ethyl iodide. White solid.

Dimethylpiperidinium iodide [diMPip] (Gamal-Eldin and Macartney, 2013): From 1-methylpiperidine and methyl iodide. White solid.

Ethylmethylpiperidinium iodide [EMPip] (von Braun and Buchman, 1931): From 1-methylpiperidine and ethyl iodide. White solid.

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Diethylpiperidinium iodide [diEPip] (Lowe and Rendall, 1971): From 1-ethylpiperidine and ethyl iodide. White solid.

Dimethylhexahydroazepinium iodide [diMHHA] (Huenig, 1957): From 1-methylhexahydroazepine and methyl iodide. White solid.

Ethylmethylhexahydroazepinium iodide [EMHHA] (Huenig, 1957): From 1-methylhexahydroazepine and ethyl iodide. White solid.

Diethylhexahydroazepinium iodide [diEHHA] (Huenig, 1957): From 1-ethylhexahydroazepine and ethyl iodide. White solid.

1-methylhexahydroazepine was made from hexahydroazepine by the Eschweiler-Clarke reaction (formalin and formic acid) (Eschweiler, 1905; Clarke et al., 1933), and 1-ethylhexahydroazepine was prepared from hexahydroazepine using potassium carbonate and ethyl iodide.

Ethylmethylphenylpiperazinium iodide [EMPP]: From 1-ethyl-4-phenylpiperazine and methyl iodide. White solid. ¹H NMR (DMSO-d₆, 500 MHz): δ 7.29-7.26 (m, 2H); 7.02 (d, 2H, 8.1 Hz); 6.87 (t, 1H, 7.3 Hz); 3.61-3.50 (m, 8H); 3.45-3.40 (m, 2H); 3.11 (s, 3H); 1.29 (t, 3H, 7.1 Hz). ¹³CNMR (DMSO-d₆, 125 MHz): δ 149.2, 129.0, 119.8, 115.6, 58.4, 58.3, 45.1, 41.8, 7.1.

Diethylphenylpiperazinium iodide [diEPP]: From 1-ethyl-4-phenylpiperazine and ethyl iodide. White solid. ¹H NMR (DMSO-d₆, 500 MHz): δ 7.28 (t, 2H, 7.9 Hz); 7.01 (d, 2H, 8.3 Hz); 6.87 (t, 1H, 7.1 Hz, 7.5 Hz); 3.55-3.44 (m, 12H); 1.22 (t, 6H, 7.0 Hz, 7.2 Hz). ¹³CNMR (DMSO-d₆, 125 MHz): δ 149.3, 129.0, 119.8, 115.5, 56.4, 51.8, 41.5, 6.8.

Heterologous expression of nAChRs in Xenopus laevis oocytes

Human nAChR clones were obtained from Dr. J. Lindstrom (University of Pennsylvania, Philadelphia, PA). The human resistance-to-cholinesterase 3 (RIC-3) clone, obtained from Dr. M. Treinin (Hebrew University, Jerusalem, Israel), was co-injected with α7 to improve the level and speed of α7 receptor expression without affecting the pharmacological properties of the receptors (Halevi et al., 2003). Subsequent to linearization and purification of the plasmid cDNAs, cRNAs were prepared using the mMessage mMachine in vitro RNA transfection kit (Ambion, Austin, TX).

Oocytes were surgically removed from mature *Xenopus laevis* frogs (Nasco, Ft. Atkinson, WI) and injected with appropriate nAChR subunit cRNAs as described previously (Papke and Stokes, 2010). Frogs were maintained in the Animal Care Service facility of the University of Florida, and all procedures were approved by the University of Florida Institutional Animal Care and Use Committee. In brief, the frog was first

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anesthetized for 15-20 min in 1.5 L frog tank water containing 1 g of 3-aminobenzoate methanesulfonate buffered with sodium bicarbonate. The harvested oocytes were treated with 1.25 mg/ml collagenase (Worthington Biochemicals, Freehold, NJ) for 2 h at room temperature in a calcium-free Barth's solution (88 mM NaCl, 1 mM KCl, 2.38 mM NaHCO₃, 0.82 mM MgSO₄, 15 mM HEPES, and 12 mg/l tetracycline, pH 7.6) to remove the follicular layer. Stage V oocytes were subsequently isolated and injected with 50 nl of 5-20 ng nAChR subunit cRNA. Recordings were carried out 1-7 days after injection.

Two-electrode voltage clamp electrophysiology

Experiments were conducted using OpusXpress 6000A (Molecular Devices, Union City, CA) (Papke and Stokes, 2010). Both the voltage and current electrodes were filled with 3 M KCl. Oocytes were voltage-clamped at -60 mV. The oocytes were bath-perfused with Ringer's solution (115 mM NaCl, 2.5 mM KCl, 1.8 mM CaCl₂, 10 mM HEPES, and 1 μ M atropine, pH 7.2) at 2 ml/min for $\alpha 7$ receptors and at 4 ml/min for other subtypes. To evaluate the effects of experimental compounds compared to ACh-evoked responses of various nAChR subtypes expressed in oocytes, baseline conditions were defined by two initial applications of ACh made before co-applications of experimental compounds with the control ACh. The agonist solutions were applied from a 96-well plate via disposable tips, and the test compounds were applied alone, co-applied with ACh, or co-applied with PNU-120596. For the concentration-response study, drug applications alternated between ACh controls and experimental compounds. Unless otherwise indicated, drug applications were 12 s in duration followed by a 181 s washout period for $\alpha 7$ receptors and 6 s with a 241 s washout for other subtypes. A typical recording for each oocyte constituted two initial control applications of ACh, an experimental compound application, and then a follow-up control application of ACh to determine the desensitization or rundown of the receptors. The control ACh concentrations were 300 μ M for $\alpha 7$ or 60 μ M when followed up with PNU-120596 application, 100 μ M for $\alpha 3\beta 4$, and 30 μ M for $\alpha 4\beta 2$. The responses of $\alpha 4\beta 2$ and $\alpha 3\beta 4$ -expressing cells were measured as peak current amplitudes, and the $\alpha 7$ data were calculated as net charge, as previously described (Papke and Papke, 2002).

Data were collected at 50 Hz, filtered at 20 Hz, analyzed by Clampfit 9.2 (Molecular Devices) and Excel 2003 (Microsoft, Redmond, WA), and normalized to the averaged peak current or net-charge response of the two initial ACh controls (Papke and Papke, 2002). Data were expressed as means \pm SEM from at least four oocytes for each experiment and plotted by Kaleidagraph (Abelbeck Software, Reading, PA).

It should be noted that in our analyses of PNU-120596 potentiated currents, the test compounds were co-applied with 10 μ M PNU-120596, a concentration we previously found to produce maximal potentiation with a single application (Williams et al., 2011a). Although the kinetics of the binding of the putative orthosteric ligands would be expected to be more rapid (especially at high concentrations), than the binding of the PAM, which may need to reach a binding site in the transmembrane domains (Young et al., 2008), contributions to the macroscopic response due to the binding of the orthosteric ligands alone would be small compared with amplified currents generated when the PAM sites are also bound. However, the faster binding of the orthosteric ligands at high concentrations may have contributed to the induction of PAM-insensitive desensitization (D_i), accounting for reduced PAM-potentiated currents at high concentrations and inverted "U" concentration-response functions.

Results

Responses of neuronal nAChR to series 1 quaternary amines

In order to expand our previous study of quaternary amines (Papke et al., 1996; Horenstein et al., 2008), we tested tetMA and the other series 1 amines on $\alpha 4\beta 2$, $\alpha 3\beta 4$, and $\alpha 7$ nAChR-expressing cells and compared those responses to control ACh-evoked responses. The $\alpha 4\beta 2$ -expressing cells only responded to tetMA with peak current responses $\leq 5\%$ the ACh controls, while $\alpha 3\beta 4$ receptors responded to both tetMA and EtriMA, and $\alpha 7$ receptors also responded well to diEdiMA (Figure 2) and weakly to triEMA, at our limit of detection. None of the subtypes tested gave responses to the application of 100 μ M tetEA that were larger than stimulus artifacts associated with the application of Ringer's solution alone. When co-applied with ACh, tetEA functioned as a competitive antagonist for $\alpha 7$, decreasing currents evoked by 60 μ M ACh with an IC_{50} of 80 ± 5 μ M (not shown).

The quaternary amines were then co-applied to $\alpha 7$ -expressing cells with the efficacious type II PAM, PNU-120596, and all of them evoked significant responses. The revelation of tetEA-induced expression of PAM-sensitive desensitization (D_s) identifies the structure of tetEA as the minimal pharmacophore for an $\alpha 7$ silent agonist.

We tested the panel of series 1 quaternary amines on $\alpha 7$ across of range of concentrations, applied alone or in combination with PNU-120596 (Figure 3). When applied alone, EtriMA was most efficacious, more so than ACh, but only at very high concentrations, so that the data were not well fit with the Hill equation. tetMA and diEdiMA were effectively full agonists relative to ACh (see Table 1 for EC_{50} values), while triEMA was a partial agonist, and tetEA had negligible efficacy when applied

alone. All of the series 1 amines synergized effectively with PNU-120596, although tetMA and EtriMA had pronounced inverted-U concentration-response functions, presumably related to the preferential induction of PAM-insensitive desensitization (D_i) (Williams et al., 2011a) at higher concentrations. EtriMA and diEdiMA appeared notably more potent for generating PNU-120596-potentiated currents than when used alone. The efficacy of tetEA as an orthosteric agonist was so low that it was effectively an antagonist of ACh-evoked $\alpha 7$ responses (Figure 3F) with an IC_{50} of $80 \pm 5 \mu M$. These data are summarized in Table 1.

The series 1 quaternary amines were also tested at a concentration of 100 μM for their antagonist activity on $\alpha 3\beta 4$ and $\alpha 4\beta 2$ receptors (Table 2). Although the large amines did not activate $\alpha 4\beta 2$ receptors, they produced more than 40% inhibition of the $\alpha 4\beta 2$ ACh-evoked response.

Structural analogs of the series1 quaternary amines

Since we determined in the series of simple amines that there was a critical size between triEMA and tetEA where orthosteric but not allosteric activity was lost, we tested the hypothesis that the same size criteria might apply to quaternary amines of more complex structure. Therefore, we synthesized five additional sets of compounds (Figure 1), which progressively substituted ethyl for methyl groups onto the key charged nitrogen. The base of the Series 2 compounds was choline. The root for the third series was benzyltrimethylamine. The remaining three sets had the charged nitrogen incorporated into 5, 6, or 7 member rings (Figure 1). The Connolly solvent-excluded volumes for each of these quaternary ammonium compounds were calculated using Chem3D and are provided in Figure 1.

Choline and related compounds

The concentration-response relationships for currents evoked in $\alpha 7$ nAChR-expressing oocytes by choline and related compounds (Figure 1, series 2) applied alone, or in combination with 10 μM PNU-120596, are shown in Figure 4. As previously reported (Papke and Papke, 2002), choline is less potent than ACh but equally efficacious. Interestingly, choline appeared more potent when used in combination with PNU-120596. Similar results were obtained with (2OHE)-EdiMA, except that this compound was reduced in potency (Table 1) when applied alone and more effective than choline at synergizing with the PAM. The diethylmethyl choline analog was further reduced in potency compared to the smaller compounds, with data that could not be fit well to the Hill equation. Responses to (2OHE)-diEMA were significantly increased by

co-application with PNU-120596 without a change in potency. Applications of (2OHE)-triEA at concentrations greater than 100 μ M produced responses that were unlike normal α 7-mediated responses, which rapidly decay to baseline (see Figure 2A). The responses associated with the application of (2OHE)-triEA were not significantly increased when the compound was co-applied with PNU-120596 (note that the right and left-hand scales in Figure 4D are identical). To determine whether the (2OHE)-triEA responses were α 7-mediated, (2OHE)-triEA was applied to oocytes that had not been injected with α 7 RNA, and similar responses were observed (Figure 4D insert). Since (2OHE)-triEA responses were not receptor dependent, data from this compound was excluded from further pharmacophore analyses.

The choline compounds were tested at a concentration of 1 mM on α 3 β 4– and α 4 β 2-expressing cells. They did not activate receptors but, to varying degrees, produced some inhibition of ACh-evoked responses (Table 2).

Benzylc quaternary amines

Shown in Figure 5 are the concentration-response studies for the third series of compounds illustrated in Figure 1. BtriMA was a reasonably potent and efficacious α 7 partial agonist (Table 1) compared to the other compounds in the series. All of the compounds in this series except BtriEA synergized effectively with PNU-120596.

The benzylc compounds were tested at a concentration of 100 μ M on α 3 β 4– and α 4 β 2-expressing cells. They did not activate receptors but, to varying degrees, produced some inhibition of ACh-evoked responses (Table 2).

Pyrrolidinium compounds

Three compounds were tested with the core nitrogen contained within a pyrrolidinium ring (Figure 1, series 4). As shown in Figure 6, the dimethyl and ethylmethyl derivatives were full agonists, with diMPyrr being more potent than EMPyrr (Table 1). These compounds also synergized effectively with PNU-120596, and while the potentiated diMPyrr currents were shifted in potency compared to the orthosteric currents, the potency was similar for orthosteric and allosterically potentiated currents evoked by EMPyrr. The orthosteric and allosteric potency and the efficacy of diEPyrr were reduced compared to the other compounds in this series.

The pyrrolidinium compounds were tested at a concentration of 100 μ M on α 3 β 4– and α 4 β 2-expressing cells. They did not activate receptors but, to varying degrees, produced some inhibition of ACh-evoked responses (Table 2).

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Piperidinium compounds

Three compounds were tested with the core nitrogen contained within a piperidinium ring (Figure 1, series 5). As shown in Figure 7, only the dimethyl derivative was an efficacious agonist (Table 1). To varying degrees, all three compounds synergized with PNU-120596. Co-applications of EMPip or diEPip with ACh significantly reduced the ACh-evoked responses of $\alpha 7$ -expressing cells, as shown in Figure 7D (IC₅₀ values in Table 1).

The piperidinium compounds were tested at a concentration of 100 μ M on $\alpha 3\beta 4$ - and $\alpha 4\beta 2$ -expressing cells. They did not activate receptors but, to varying degrees, produced some inhibition of ACh-evoked responses (Table 2).

Hexahydroazepinium compounds

Three compounds with the core nitrogen contained within a hexahydroazepinium ring were tested (Figure 1, series 6). As shown in Figure 8, these compounds only evoked significant responses at concentrations greater than 100 μ M when applied alone. The three compounds synergized with PNU-120596 although their efficacy was progressively reduced as the size of the compounds was increased, and only the dimethyl compound showed significant currents in combination with PNU-120596 at any concentration less than 300 μ M. Note that while 1 mM diEHHA generated significant responses when co-applied with 10 μ M PNU-120596, the responses to 3 mM diEHHA co-applied with 10 μ M PNU-120596 were much smaller (Figure 8C).

The hexahydroazepinium compounds were tested at a concentration of 100 μ M on $\alpha 3\beta 4$ - and $\alpha 4\beta 2$ -expressing cells. They did not activate receptors but, to varying degrees, produced some inhibition of ACh-evoked responses (Table 2).

Constraints on orthosteric and allosteric activation of $\alpha 7$ nAChR

Our analysis of series 1 quaternary amines (Figures 1-3) identified tetEA as a minimal pharmacophore for silent agonism of $\alpha 7$. Some critical differences between triEMA and tetEA led to the loss of orthosteric efficacy, yet with the retention of an ability to bind to the receptor and induce a nonconducting conformation that was sensitive to allosteric activation by PNU-120596. This was effectively an endpoint in a trend that was present through the first series of compounds tested, and similar trends were observed in our analyses of the additional compounds studied. Two features of the compounds were systematically altered in six compound sets, size and shielding of electrostatic interactions. With the substitution of ethyl for methyl groups, the compounds become bulkier, and the charge center of the nitrogen becomes shielded and

presumably less effective for forming the cation-pi bond interactions believed important for receptor activation (Zhong et al., 1998). While the electrostatic effects will be relatively consistent from one series to the next, each series started out with a different bulk that was then increased. We evaluated the potential relationships between the estimated Connolly solvent excluded volumes (Figure 1) and the orthosteric and allosteric activity of all of the compounds by taking a cross section of the data at 100 μM , allowing both potency and efficacy to be reflected in the data (Figure 9). We found that there was a marked trend for orthosteric activity to fall off for compounds with a volume greater than 130 \AA^3 . The potential for allosteric activation was retained in compounds up to a size 180 \AA^3 . The data therefore suggest that agonists may be tuned for $\alpha 7$ selectivity and preferential induction of PAM-sensitive desensitized states by designing their core pharmacophoric elements to be between 130 and 180 \AA^3 .

Tuning the selectivity and activity profile of the ganglionic agonist diMPP

Dimethylphenylpiperazinium (diMPP), a ganglionic agonist first described in 1951 (Chen et al., 1951), is more efficacious than ACh for stimulating $\alpha 3\beta 4$ receptors and is also a full agonist for $\alpha 7$ receptors. We created new analogs of diMPP by substituting ethyl groups for one or both of the diMPP methyl groups (Figure 10A). As expected (Horenstein et al., 2008), the ethyl substituted diMPP analogs showed no efficacy for $\alpha 3\beta 4$ nAChR. Compared to diMPP, EMPP and, to a greater degree, diEPP had reduced efficacy and potency for activating $\alpha 7$. However, although diEPP was significantly compromised as an orthosteric agonist, it was relatively potent and efficacious in combination with PNU-120596. As shown in Figure 10, 30 μM diEPP was ineffective when applied alone but activated net-charge responses in combination with PNU-120596 that were 50-fold larger than the 60 μM ACh controls. Note that all of these compounds have estimated solvent-excluded volumes larger than the 180 \AA^3 cut-off for allosteric activation suggested by the study of the more simple compounds (Figure 9). However, approximately 70 \AA^3 of this space is accounted for by the phenyl group of the extended structure, suggesting that this part of the molecules is accommodated in space outside that which is critical for the core activity, such as in the cleft between the subunits, as suggested for the phenyl groups of benzylidene anabaseines (Hibbs et al., 2009). When the estimated volume of the phenyl groups is subtracted, sizes of these compounds fall well within the ranges illustrated in Figure 9.

Channel block as a possible limiting factor for the efficacy of silent agonists.

At high concentrations, ACh (Auerbach and Akk, 1998) and other agonists (Carter and Oswald, 1993) block muscle-type nAChR. Additionally, tetEA is known to be a blocker of voltage-gated potassium channels (Armstrong and Hille, 1972). Therefore, it is likely that, to varying degrees, the small charged molecules used in these experiments could act as channel blockers, and such activity might be a limiting factor, especially when the drugs are used at high concentration. In order to evaluate the degree to which channel block limited the $\alpha 7$ responses to tetEA when co-applied with PNU-120596, we made applications of increasing concentrations of tetEA during steady-state activation of receptors by the bath application of 60 μ M choline plus 10 μ M PNU-120596, a protocol previously used to evaluate the channel-blocking activity of benzylidene anabaseines (Papke et al., 2009). Within four minutes of starting the bath application, steady-state currents were established that were 22.1 ± 7.1 ($n=4$) times larger than initial peak current responses to 60 μ M ACh, and since choline has relatively low potency, the currents remained relatively stable throughout the course of the experiment. As shown in Figure 11A, applications of tetEA produced transient augmentations of the currents. Only when tetEA was applied at 1 mM were the augmented currents preceded by a brief decrease in current likely to be associated with channel block. The inhibition produced by tetEA was small compared to that produced by 100 μ M mecamylamine. Note that once the bath application of 60 μ M choline and PNU-120596 was terminated, the channel remained primed for potentiation, so that a follow-up application of 60 μ M ACh alone produced a large response. These results with tetEA were quite different from those obtained with 100 μ M - 3 mM diEHHA, which when applied under similar conditions produced only inhibition of the steady-state currents stimulated by 60 μ M choline and 10 μ M PNU-120596 (Figure 11B).

We also evaluated the degree to which the responses to tetEA and diEHHA co-applied with PNU-120596 could be limited by voltage-dependent channel block. The drugs were co-applied at 3 mM with 10 μ M PNU-120596 at a holding potential of +50 mV and compared to data obtained at the normal holding potential of -60 mV. Consistent with the biphasic effects of 1 mM tetEA on the steady-state currents, the results suggest that voltage-dependent channel block may be a factor limiting the potentiated responses to tetEA. When measured relative to the ACh controls, the PNU-120596-potentiated outward currents at +50 mV had peak amplitudes 2.8 ± 0.5 -fold larger and net charge values 4.6 ± 0.8 -fold larger than the responses at -60 mV (Figure 11C). When responses to 3 mM diEHHA plus 10 μ M PNU-120596 were compared at the two voltages, the currents were relatively small compared to those evoked by 3 mM tetEA, but that effect of voltage was greater. The peak amplitudes were 26 ± 9 -fold larger and net charge

values 18 ± 5 -fold larger than the responses at -60 mV (Figure 11D). While these data suggest that noncompetitive inhibition is an important factor limiting the diEHHA responses at high concentration, when 100 μ M diEHHA was co-applied with ACh, its effects were most consistent with competitive activity since the inhibition of the net charge responses to 60 μ M ACh (70 ± 2 % inhibition, $n = 7$) was greater ($p < 0.05$) than the inhibition of 300 μ M ACh responses (25 ± 13 % inhibition, $n = 5$), and in each case 100 μ M diEHHA had the effect of increasing peak current amplitudes (Papke, 2005). Therefore, while high concentrations of diEHHA clearly have mixed silent agonist/antagonist activity and a large portion of the antagonist activity is due to channel block, intrinsically weak silent agonism may also be a limiting factor.

Discussion

The discovery of the role of $\alpha 7$ nAChR in vagal-mediated cholinergic anti-inflammatory response (Borovikova et al., 2000; van Westerloo et al., 2006; Pavlov et al., 2007; Rosas-Ballina et al., 2009; Rosas-Ballina and Tracey, 2009) has provided exciting impetus to discover drugs that might be used to treat sepsis, other inflammatory diseases, and various types of inflammation-related pain. This discovery provided a compelling motivation to reconsider our view of $\alpha 7$ and other nAChR strictly as mediators of transmembrane signals relying on channel-mediated ion flux. The non-neuronal cells which mediate $\alpha 7$'s control of inflammation have not been shown to generate $\alpha 7$ -mediated currents, and, moreover, some $\alpha 7$ -targeting ligands which can effectively control inflammation have little or no efficacy as ion channel activators (van Maanen et al., 2009; Thomsen and Mikkelsen, 2012; Clark et al., 2014).

Even the $\alpha 7$ agonists which are most efficacious for producing channel activation produce only brief and infrequent ion channel currents and are far more effective at inducing and maintaining the receptors in non-conducting states, which have traditionally been dismissed as desensitized and functionally unimportant (Williams et al., 2011a). The time has come to discard the prejudice that the ligand-bound non-conducting states of nAChR are all functionally unimportant. Proteomic analyses have indicated a large number of intracellular protein partners for $\alpha 7$ (Paulo et al., 2009), many of which are mediators of signal transduction. It has been proposed that desensitization is associated with an uncoupling of the amino terminal ligand binding domain from the pore-lining second transmembrane domain (Zhang et al., 2013). This effect may be specifically affected by PAM binding consistent with the proposed binding site for PNU-120596 being associated with a transmembrane site (Young et al., 2008). Just as conformational changes promoted by ligand binding spread through the transmembrane domains, they

must also pass through to the intracellular domain and likely regulate signal transduction processes in both neuronal and nonneuronal cells.

Our ability to probe the conformational landscape of $\alpha 7$ receptors has been greatly aided by the discovery of $\alpha 7$ -selective PAMs. The selectivity of these PAMs for $\alpha 7$ appears to arise from their specific effects on the class of desensitized states which are unique to $\alpha 7$ and are induced at levels of agonist occupancy higher than those which most effectively lead to ion channel activation (Williams et al., 2011a). Based on the use of highly effective PAMs such as PNU-120596, we have been able to use ion channel currents to detect and distinguish two classes of agonist-induced states which are non-conducting in the absence of the PAM. One functionally unique class of these desensitized states (D_s), is destabilized by the PAM and can convert to a novel conducting state, while the other class (D_i) is PAM-insensitive. Figure 11 provides a graphic representation of the thermodynamic relationship between the conformation states of $\alpha 7$ as regulated by the binding of agonists, silent agonists and PAMs (Williams et al., 2011a) (for further discussion of such models see (Williams et al., 2011b; Papke, 2014)). Receptors that have bound neither orthosteric nor allosteric ligands are most stable in the resting closed state (C). Upon binding an efficacious agonist such as tetMA, the energy barrier to the open state (O^*) is sufficiently reduced so that some channels may open. However, for $\alpha 7$ this remains a low probability event (Williams et al., 2011a) and receptors are more likely to enter D_s than to open. Upon binding a silent agonist such as tetEA, the barrier to the open state remains high so that the receptors are more likely to enter one of the desensitized states than to open. However, the binding of PNU-120596 couples the D_s state to a PAM-dependent open state (O') (Williams et al., 2011a), allowing both tetMA and tetEA to stimulate bursts of channel activation.

Our data indicate that, while it is a common feature for all efficacious agonists to induce both D_s and D_i to varying degrees depending on their applied concentration, ligands with such low efficacy that they are functional antagonists of normal channel activation can, in the case of compounds we classify as silent agonists, induce the D_s state and only appear efficacious when co-applied with a PAM or applied to receptors primed by previous exposure to PAMs. One of the first such compounds to be identified was NS-6740 (Thomsen and Mikkelsen, 2012), and we have also characterized the compounds KC-1 (Chojnacka et al., 2013) and R-47 (Clark et al., 2014) (CTI-15072 (van Maanen et al., 2009)), as silent agonists. These compounds are large and structurally diverse, so here we start with the smallest $\alpha 7$ agonists in order to find the most rudimentary structure capable of silent agonism, which is that of tetEA.

Through our systematic analysis of additional quaternary ammonium compounds, we identified unique size constraints on the effectiveness of ligands as orthosteric activators or as silent agonists, such that ligands with estimated solvent-excluded volumes around the cationic center between 130 and 180 Å³ show a tendency toward silent agonism. While this correlation with molecular volume appears to be valid, the underlying molecular interactions remain to be elucidated. In the simplest way, the larger volume could induce and or preclude certain conformational configurations of the complex. Further, when the central ammonium core is surrounded by increased volume associated with larger alkyl groups, the effective charge density at the alkyl ammonium group is lower, potentially lessening cation- π interactions, which could also have functional significance.

Previous studies of an agonist series binding to and activating the muscle-type nAChR suggested that the steric bulk of an experimental agonist affected association and disassociation, while the ionic character of ligand with comparable size more specifically affected opening and closing rates (Papke et al., 1988). With the present set of compounds, both size and charge density are changed in parallel, so it would not be possible, even with single-channel data, to separate these factors. An alternative hypothesis is that binding of the PAM opens up the orthosteric binding site, allowing access to larger ligands. However, the silent agonist BdiEMA acts as an antagonist of orthosteric activation (Table 1) with approximately the same potency with which it generates currents in co-application with PNU-120596 (Figure 5), suggesting that access to the orthosteric binding site is similar in the absence and presence of the PAM.

We have previously shown that the structural motifs which account for the $\alpha 7$ selectivity of specific agonists can be installed onto other non-selective agonists to make new $\alpha 7$ -selective drugs (Horenstein et al., 2008). Our identification of triEA as a minimal element for being an $\alpha 7$ silent agonist encouraged us to generate diEPP, a modified form of the highly efficacious $\alpha 3\beta 4$ agonist diMPP, that lost all activity for $\alpha 3\beta 4$, showed reduced orthosteric activity for $\alpha 7$, and yet had good allosteric activity for $\alpha 7$. This indication that the minimal silent agonist pharmacophore might be moved on to other $\alpha 7$ -selective agonists such as quinuclidines or benzylidene anabaseines, large molecules with structural elements that are accommodated in the extended binding site, provides a promising new idea for potential drug development. Since it has been shown that silent agonists are promising agents for signal transduction, the development of such new compounds has the potential for more effective therapeutics selectively targeted to the roles of $\alpha 7$ in non-neuronal cells.

We have used PNU-120596 to distinguish silent agonists from simple antagonists of orthosteric activation, and we hypothesize that even in the absence of the PAM, receptors in the D_s and/or D_i states are potential mediators of signal transduction. Some of the best support for this hypothesis comes from the documented anti-inflammatory activity of NS-6740 (Thomsen and Mikkelsen, 2012).

It is interesting to note the qualitative differences in the concentration-response functions for the PNU-120596-potentiated currents of the various agents. While there are plateaus in the response data for some agents such as diEdiMA, tetEA, diMPyrr, EMPyrr, EMPip, diEPip, and diMPP, other agents such as BEdiMA and tetMA show well-potentiated responses over a very narrow range of concentration. These data suggest that the dynamic interconversion of receptors between the D_s state, associated with the potentiated currents, and the D_i state, which limits such responses, varies depending on the specific ligand. If, in fact, these non-conducting states can be associated with signal transduction, it is unclear whether such signaling would preferentially arise from induction of D_s or D_i or some other state that is independent of what can be detected by the use of the PAM.

The question also remains whether PAMs themselves induce novel conformational states which may also be effective modulators of signal transduction in non-neuronal cells. Some recent studies (Freitas et al., 2013) demonstrating analgetic effects of PNU-120596 suggest that this may be the case. However, the ubiquitous presence of choline in all *in vivo* and most *in vitro* experiments makes it impossible to conclude that such effects are due to the activity of the PAM alone. Therefore, the challenge remains to better define the relationship between $\alpha 7$ conformational states and the role of allosteric modulation in signal transduction by the receptor that does not rely on channel-mediated ionic currents.

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Authorship Contributions

Participated in research design: Papke, Chojnacka, Horenstein

Conducted experiments: Chojnacka

Performed data analysis: Chojnacka, and Papke

Provided experimental materials: Chojnacka, Horenstein

Wrote or contributed to the writing of the manuscript: Papke, Chojnacka, Horenstein

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Footnotes

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Figure legends

Figure 1. Structures of the quaternary ammonium compounds and their Connolly solvent-excluded volumes. The progressive replacement of methyl groups with ethyl groups, which characterized the first series of amines, was used to systematically generate five additional sets of compounds. The structure-based names and abbreviations used in the text are as follows: tetMA, tetramethylammonium; EtriMA, ethyltrimethylammonium; diEdiMA, diethyldimethylammonium; tetEA, tetraethylammonium; (2OHE)-EdiMA, (2-Hydroxyethyl)-ethyldimethylammonium; (2OHE)-diEMA, (2-Hydroxyethyl)-diethylmethylammonium; (2OHE)-triEA, (2-Hydroxyethyl)-triethylammonium; BtriMA, benzyltrimethylammonium; BEdiMA, benzylethyldimethylammonium; BdiEMA, benzyldiethylmethylammonium; diMPyrr, dimethylpyrrolidinium; EMPyrr, ethylmethylpyrrolidinium; diEPyrr, diethylpyrrolidinium; diMPip, dimethylpiperidinium; EMPip, ethylmethylpiperidinium; diEPip, diethylpiperidinium; diMHHA, dimethylhexahydroazepinium; EMHHA, ethylmethylhexahydroazepinium; diEHHA, diethylhexahydroazepinium.

Figure 2. **A)** Drug-evoked responses of human nAChR to ACh and series 1 quaternary amines. Cells expressing $\alpha 4\beta 2$, $\alpha 3\beta 4$, or $\alpha 7$ were first stimulated with ACh (30 μ M, 100 μ M, or 60 μ M, respectively) to establish a basis for comparisons, and all representative evoked responses displayed have been scaled relative to the ACh responses from the individual cells. Each trace is 200 s in duration. The bottom row of traces shows the responses of $\alpha 7$ receptors to each of the compounds at 100 μ M, co-applied with 10 μ M of the $\alpha 7$ PAM PNU-120596. Responses to the various agents were first scaled relative to the ACh controls initially recorded from the same cells and then reduced by a factor of twenty for purposes of display. **B)** The three groups of columns on the left show the average responses of $\alpha 4\beta 2$ -, $\alpha 3\beta 4$ -, and $\alpha 7$ -expressing cells to 100 μ M applications of the series 1 quaternary amines, expressed relative to ACh control responses recorded in the same cells. The $\alpha 4\beta 2$ and $\alpha 3\beta 4$ data are for peak current, and the $\alpha 7$ data are for net charge. The group of columns on the right gives the average net-charge responses to co-applications of the amines plus 10 μ M PNU-120596 to $\alpha 7$ -expressing cells, expressed relative to the net-charge responses to 60 μ M ACh alone. Each bar shows the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 3. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of $\alpha 7$

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nAChR by series 1 quaternary amines, tetMA (**A**), EtriMA (**B**), diEdiMA (**C**), triEMA (**D**), and tetMA (**E**). In plots A-E the left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles), normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to the drugs applied alone in panels A-D were fit to the Hill equation, and the fit parameters are given in Table 1. F) Since tetEA did not evoke significant currents when applied alone, we tested its potency as an antagonist by co-applying it with 60 μ M ACh. The tetEA IC₅₀ is reported in Table 1. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 4. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of α 7 nAChR by choline (**A**) and related compounds, (2OHE)-EdiMA (**B**), (2OHE)-diEMA (**C**), and (2OHE)-triEA (**D**). The left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles), normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to the application of the drugs when applied alone in panels A-C were fit to the Hill equation, and the fit parameters are given in Table 1. Note that when (2OHE)-triEA was applied at concentrations of 300 μ M or higher, there were relatively large responses that were slower than normal α 7-mediated responses. These responses were not significantly increased when the drug was co-applied with PNU-120596. Similar responses were to (2OHE)-triEA were observed in oocytes that had not been injected with α 7 RNA (insert), indicating that these were not α 7-nAChR-mediated currents. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 5. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of α 7 nAChR by the benzylic quaternary amines BtriMA (**A**), BEdiMA (**B**), BdiEMA (**C**), and BtriEA (**D**). The left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles),

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normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to BtriMA when applied alone (panel A) were fit to the Hill equation, and the fit parameters are given in Table 1. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 6. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of $\alpha 7$ nAChR by the pyrrolidinium compounds, diMPyrr (**A**), EMPyrr (**B**), and diEPyrr (**C**). The left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles), normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to the application of the drugs when applied alone in panels A-C were fit to the Hill equation, and the fit parameters are given in Table 1. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 7. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of $\alpha 7$ nAChR by the piperidinium compounds, diMPip (**A**), EMPip (**B**), and diEPip (**C**). The left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles), normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to the application of the drugs when applied alone in panels A-C were fit to the Hill equation, and the fit parameters are given in Table 1. Each point is the average normalized response of at least 4 cells (\pm S.E.M.). **D**) We tested the potency of EMPip and diEPip as antagonists by co-applying them with 60 μ M ACh. The IC₅₀ values are reported in Table 1.

Figure 8. Concentration-response studies for the orthosteric (drug applied alone) and allosterically potentiated (drug co-applied with 10 μ M PNU-120596) activation of $\alpha 7$ nAChR by the hexahydroazepinium compounds, diMHHA (**A**), EMHHA (**B**), and diEHHA (**C**). The left-hand Y-axes apply to the net-charge responses recorded when the drugs were applied alone (open circles), normalized to the responses to 300 μ M ACh

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applied prior to the experimental application. The right-hand Y-axes apply to the net-charge responses when the drugs were co-applied with PNU-120596 (filled circles), normalized to average net-charge responses to two prior applications of 60 μ M ACh. The responses to the application of the drugs when applied alone in panel A were fit to the Hill equation, and the fit parameters are given in Table 1. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 9. Relationships between the size (estimated Connolly solvent-excluded volume) of the test compounds and the net-charge responses of $\alpha 7$ -expressing cells stimulated by application of the agents at the concentration of 100 μ M, normalized to the responses of the same cells to stimulation with 300 μ M ACh (upper panel) and the net-charge responses application of 100 μ M of the test compounds plus 10 μ M PNU-120596 (lower panel) normalized to the responses of the same cells to control applications of 60 μ M ACh alone.

Figure 10. **A)** Dimethylphenylpiperazinium (diMPP) and ethyl-substituted analogs. **B)** Responses of oocytes expressing human $\alpha 3\beta 4$ receptors to the phenylpiperazinium compounds. Peak-current responses were measured and compared to prior control applications of 100 μ M ACh. Data were then normalized to ACh maximum by adjusting for the ratio of 100 μ M ACh control responses to that of the ACh maximum responses determined in previous experiments (Stokes and Papke, 2012). **C)** Responses of oocytes expressing human $\alpha 7$ receptors to the phenylpiperazinium compounds. Net-charge responses were measured and compared to prior control applications of 300 μ M ACh, which correspond to the ACh maximum determined in previous experiments (Papke and Papke, 2002). **D)** Net-charge responses of oocytes expressing human $\alpha 7$ receptors to the phenylpiperazinium compounds co-applied with 10 μ M PNU-120596. Responses were calculated relative to the average of two initial control responses to 60 μ M ACh. Each cell was treated with a single concentration of a phenylpiperazinium compound in combination with PNU-120596. Each point is the average normalized response of at least 4 cells (\pm S.E.M.).

Figure 11. Effects of silent agonists on steady-state activation in the presence of PNU-120596. In order to achieve a relatively steady level of $\alpha 7$ receptor activation, 10 μ M PNU-120596 was bath applied with 60 μ M of the low-potency agonist choline. This procedure produced currents that were increased from the original baseline about 20 times the peak current amplitudes of the initial control response to 60 μ M ACh ($22.1 \pm$

7.1 for the experiments represented by trace A and 19.3 ± 4.9 for the experiments represented by trace B, $n = 4$ in both experiments). **A)** A representative trace illustrating the effects of increasing concentrations of tetEA, which produced additional increases in the currents to peak amplitudes 12.8 ± 2.7 , 9.3 ± 2.8 , and 22.3 ± 5.0 times larger than initial ACh controls for 10 μ M, 100 μ M, and 1 mM tetEA, respectively. The application of 100 μ M mecamylamine (Mec) reduced the steady-state current to its original baseline. After the bath perfusion was stopped, the primed channels responded to 60 μ M ACh with peak currents 82 ± 33 times larger than the original controls. The increased current in response to 1 mM tetEA was preceded by small decreases in the steady-state current that was 1.5 ± 0.3 the amplitude of the initial ACh controls. This initial decrease is visible in the insert, which is amplified 5-fold relative to the main trace. **B)** A representative trace illustrating the effects of increasing concentrations of diEHHA, which produced progressive decreases in the activation, measured as net charge, to 50 ± 8 , 160 ± 16 , and 230 ± 35 times larger than initial ACh controls for 100 μ M, 1 mM, and 3 mM diEHHA, respectively. The applications of 1 mM diEHHA, 3 mM diEHHA, and 100 μ M mecamylamine (Mec) all reduced the steady-state current to the original baseline, with the mecamylamine effect comparable to that of 3 mM diEHHA (251 ± 32 , relative to the ACh control net charge response). After the bath perfusion was stopped, the PNU-120596-primed receptors responded to 60 μ M ACh with peak currents 167 ± 27 times larger than the original controls. **C)** Representative traces illustrating the effects on holding potential on the potentiated currents evoked by 3 mM tetEA. Responses measured were 4.8 ± 0.7 and 9.9 ± 1.4 (peak current and net charge, respectively, $n = 7$) relative to control responses to ACh alone at the standard holding potential of -60 mV, and the absolute value of the outward currents relative to the ACh controls were 13.1 ± 1.8 and 45 ± 10 (peak current and net charge, respectively, $n = 6$) at the depolarized potential of +50 mV. **D)** Representative traces illustrating the effects of holding potential on the potentiated currents evoked by 3 mM diEHHA. Responses measured were 0.27 ± 0.11 and 0.72 ± 0.21 (peak current and net charge, respectively, $n = 7$) relative to control responses to ACh alone at the standard holding potential of -60 mV, and the absolute value of the outward currents relative to the ACh controls were 7.1 ± 2.4 and 13 ± 4 (peak current and net charge, respectively, $n = 6$) at the depolarized potential of +50 mV. The traces in C and D were scaled to have the same amplitude as the original ACh control responses.

Figure 12. Hypothetical energy landscapes for the conformational changes of $\alpha 7$ nAChR as affected by the binding the efficacious agonist tetMA or the silent agonist tetEA, with

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or without the effects of PNU-120596. These models are based on our previous studies which identified the existence of two forms of $\alpha 7$ desensitization (D_s and D_i), in addition to a resting closed state (C), a low probability open state (O^*), and the existence of a PAM dependent open state (O') which can be coupled to the D_s state. The models illustrated are those proposed for intermediate levels of ligand binding at both the orthosteric (agonist) and allosteric (modulator) sites, conditions which most effectively promote channel opening (Williams et al., 2011a). For each landscape the absolute free energy of the various states is represented by the vertical displacement of the states, while the rate constants for transitions between the states would be inversely related to the height of the energy barriers, represented by the lines connecting the states (lower barriers predicting faster rate constants). In the absence of ligands, the C state is most stable and transitions from that state are thermodynamically unlikely. Once drugs are applied, the thermodynamic landscapes are changed and since prior to the drug applications all channels would be predicted to be in the C state there is an initial phase of conformational change during which channels are to some degree synchronized as they move with highest probability from the C state to whatever state is connected to the state by the lowest energy barrier. Note that in the absence of the PAM, the $\alpha 7$ channel opening probability is low, even for an efficacious agonist like ACh (Williams et al., 2012).

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Table 1
Effects $\alpha 7$ nAChR

		Drug alone		Plus 10 μ M PNU-120596*
	I_{\max}	EC_{50} μ M	IC_{50} μ M	Normalized Response
tetMA	\pm	30 ± 3		96 ± 28
EtriMA	> 4	> 300		41 ± 8
diEdiMA	0.9 ± 0.01	31 ± 1		55 ± 10
triEMA	$0.15 \pm .01$	107 ± 43		68 ± 11
tetEA			80 ± 5	5.6 ± 2.5
choline	1.1 ± 0.1	270 ± 30		23 ± 7
(2OHE)-EdiMA	1.5 ± 0.3	1500 ± 300		173 ± 50
(2OHE)- diEMA			$>10,000$	24 ± 9
BtriMA	0.47 ± 0.02	39 ± 5		38 ± 20
BEdiMA			67 ± 2	27 ± 8
BdiEMA			42 ± 4	25 ± 12
BtriEA			26 ± 1	0.18 ± 0.07
diMPyrr	1.0 ± 0.1	18 ± 3		16 ± 3
EMPyrr	0.9 ± 0.1	50 ± 1		46 ± 11
diEPyrr	0.38 ± 0.03	180 ± 33	350 ± 100	24 ± 6
diMPip	0.8 ± 0.1	24 ± 2		49 ± 10
EMPip			350 ± 100	31 ± 4
diEPip			200 ± 70	5.1 ± 1.9
diMHHA	0.29 ± 0.01	183 ± 7		15 ± 3.5
EMHHA			159 ± 36	0.14 ± 0.06
diEHHA			40 ± 8	0.23 ± 0.15

* Data for choline and related compounds were obtained at 1 mM plus 10 μ M PNU-120596; data for all other compounds were obtained at 100 μ M plus 10 μ M PNU-120596. Values are for net charge relative to responses evoked by 60 μ M ACh applied alone to the same cells.

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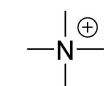
Table 2
Effects on other neuronal nAChR

	agonism	$\alpha 3\beta 4$ antagonism	IC ₅₀ μ M	$\alpha 4\beta 2$ antagonism
tetMA	+	-		+
EtriMA	+	-		+
diEdiMA	-			++
triEMA	-	+		++
tetEA	-	+		++
choline	-	-		-
(2OHE)-EdiMA	-	-		+
(2OHE)-diEMA	-	+		++
BtriMA	-	+		-
BEdiMA	-	++	124 \pm 13	-
BdiEMA	-	++	92 \pm 33	-
BtriEA	-	++	39 \pm 17	+
diMPyrr	-	-		++
EMPyrr	-	+		-
diEPyrr	-	++	59 \pm 3	+
diMPip	-	+		++
EMPip	-	++	90 \pm 4	-
diEPip	-	++	51 \pm 1	+
diMHHA	-	++	111 \pm 14	-
EMHHA	-	++	55 \pm 4	+
diEHHA	-	+++	21 \pm 1.5	+

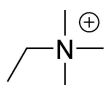
Single concentration tests were conducted with the compounds applied alone to determine agonism or co-applied with ACh to determine antagonism. Compounds scored as positive for agonism produced responses that were above our limit of detection and > 5% the amplitude of ACh controls. Compounds scored as positive (+ or ++) for antagonism reduced ACh responses so that the co-application responses were < 80% (+) or < 40 % (++) the amplitude of ACh controls. The compounds in the choline series were tested at 1 mM, and all other compounds were tested at 100 μ M.

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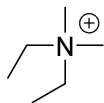
Series 1



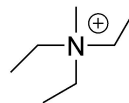
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94.0 Å³



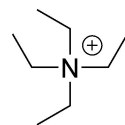
EtriMA
111.3 Å³



diEdiMA
128.4 Å³

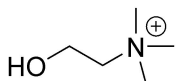


triEMA
146.0 Å³

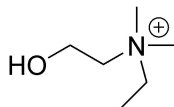


tetEA
162.8 Å³

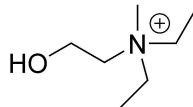
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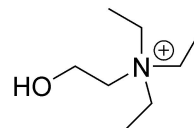
choline
115.3 Å³



(2OHE)-EdiMA
132.5 Å³

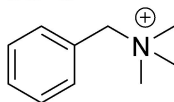


(2OHE)- diEMA
149.7 Å³

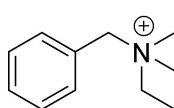


(2OHE)-triEA
166.6 Å³

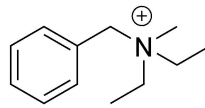
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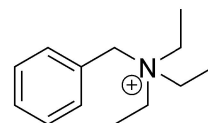
BtriMA
151.6 Å³



BEdiMA
165.8 Å³



BdiEMA
181.2 Å³

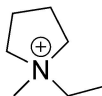


BtriEA
195.4 Å³

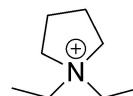
Series 4



diMPyrr
112.4 Å³

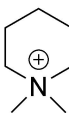


EMPyrr
129.0 Å³

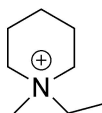


diEPyrr
142.2 Å³

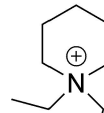
Series 5



diMPip
130.0 Å³

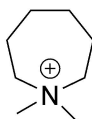


EMPip
145.7 Å³

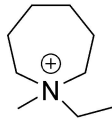


diEPip
160.1 Å³

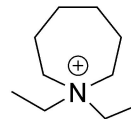
Series 6



diMHHA
151.4 Å³



EMHHA
168.5 Å³



diEHHA
185.8 Å³

Figure 1

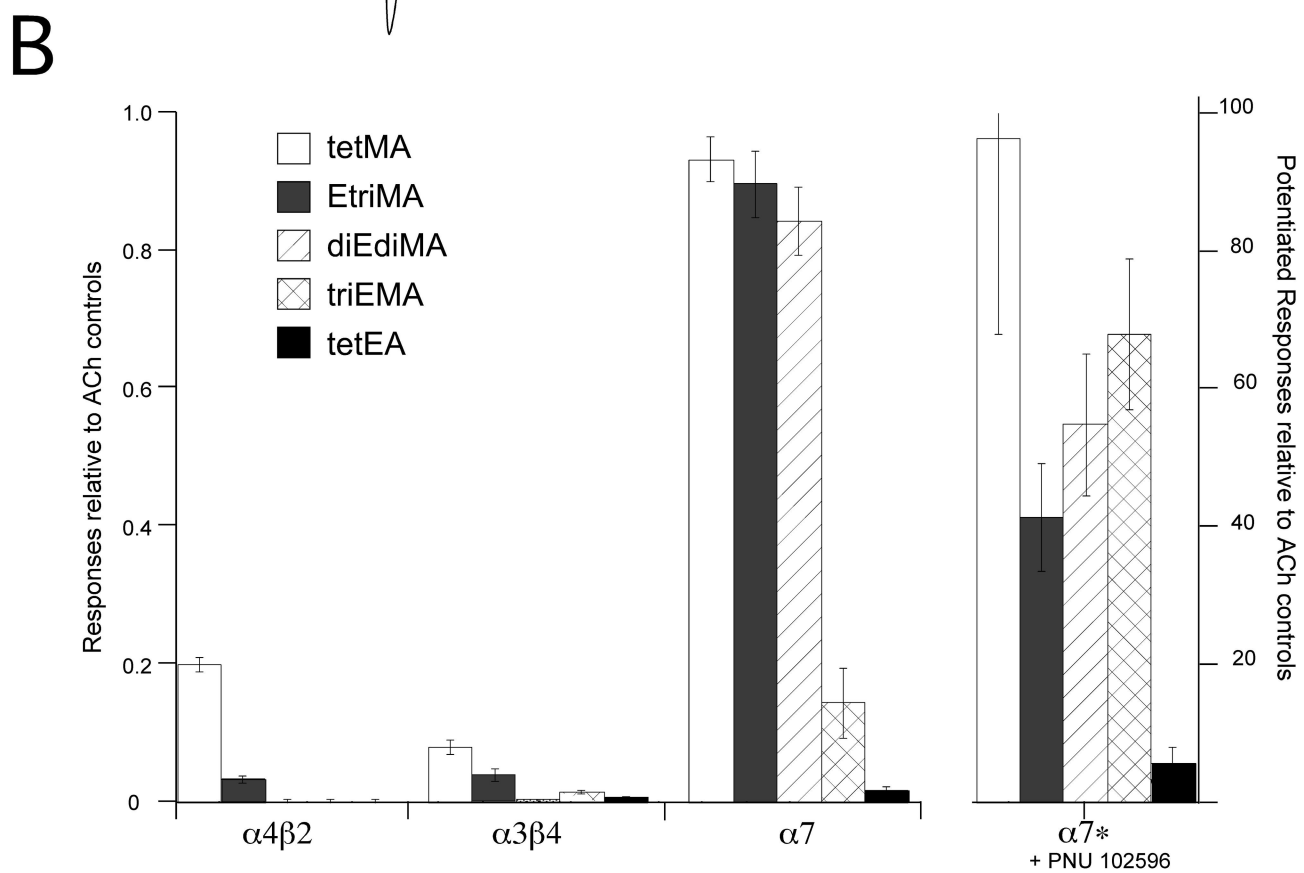
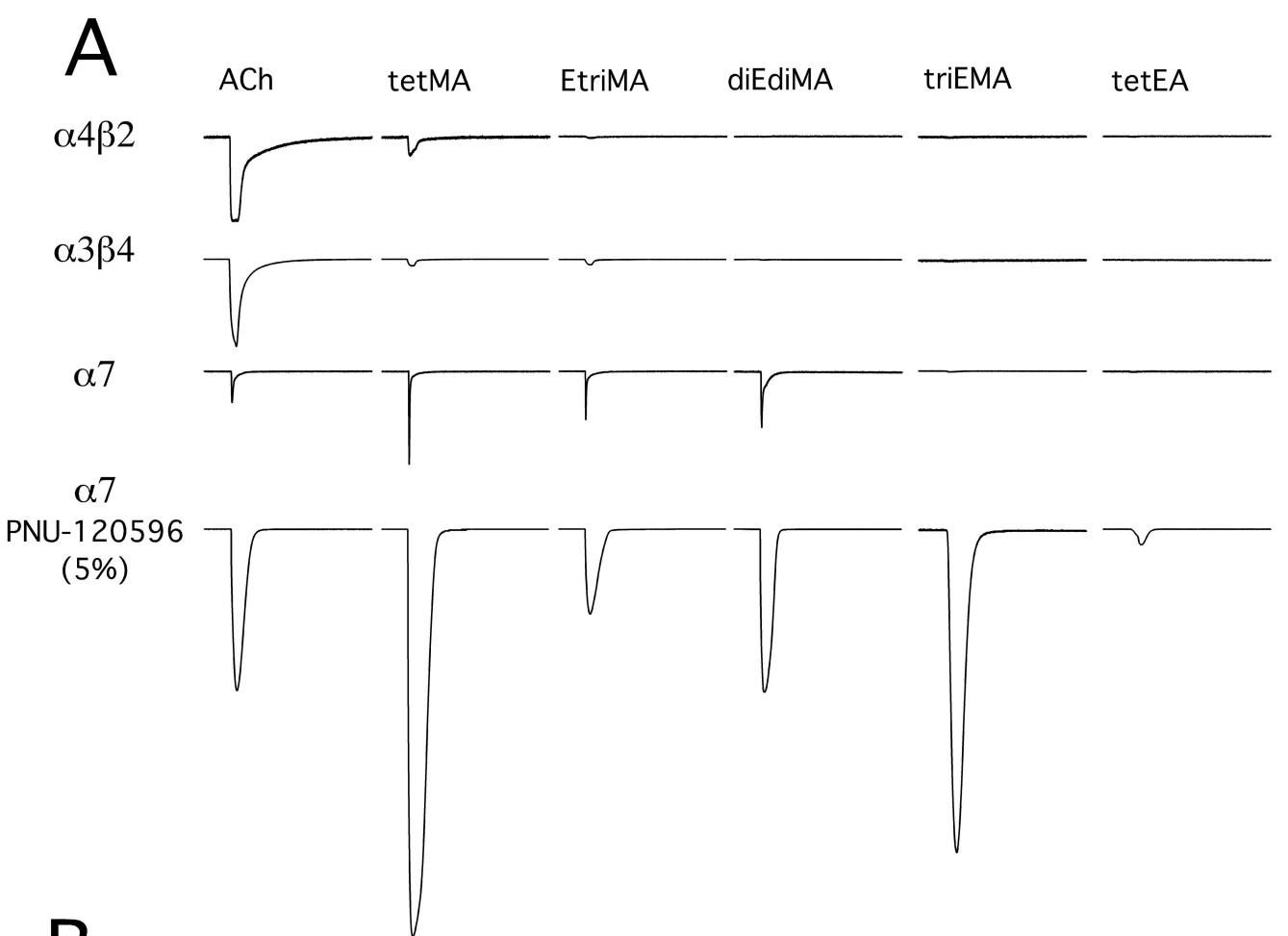


Figure 2

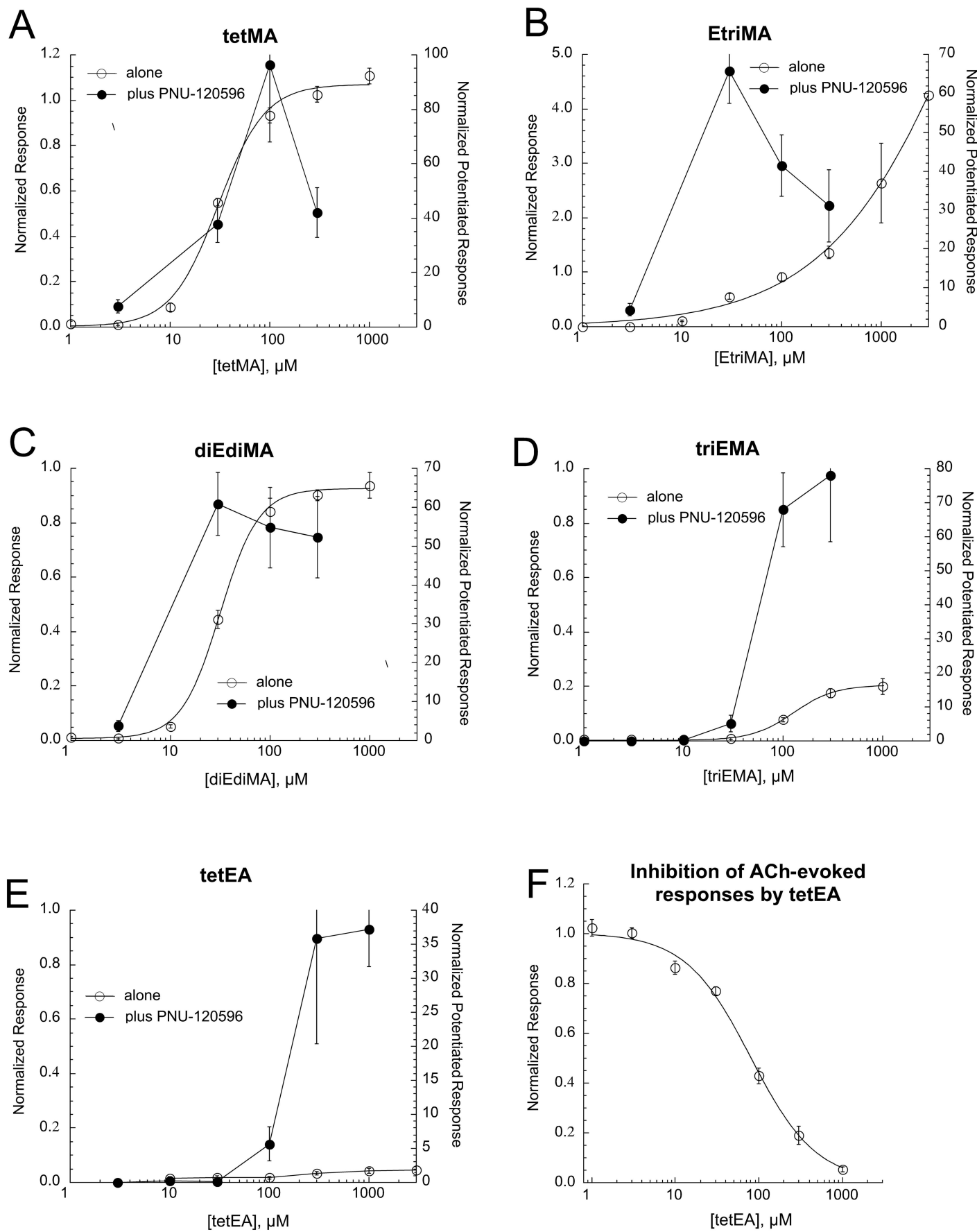


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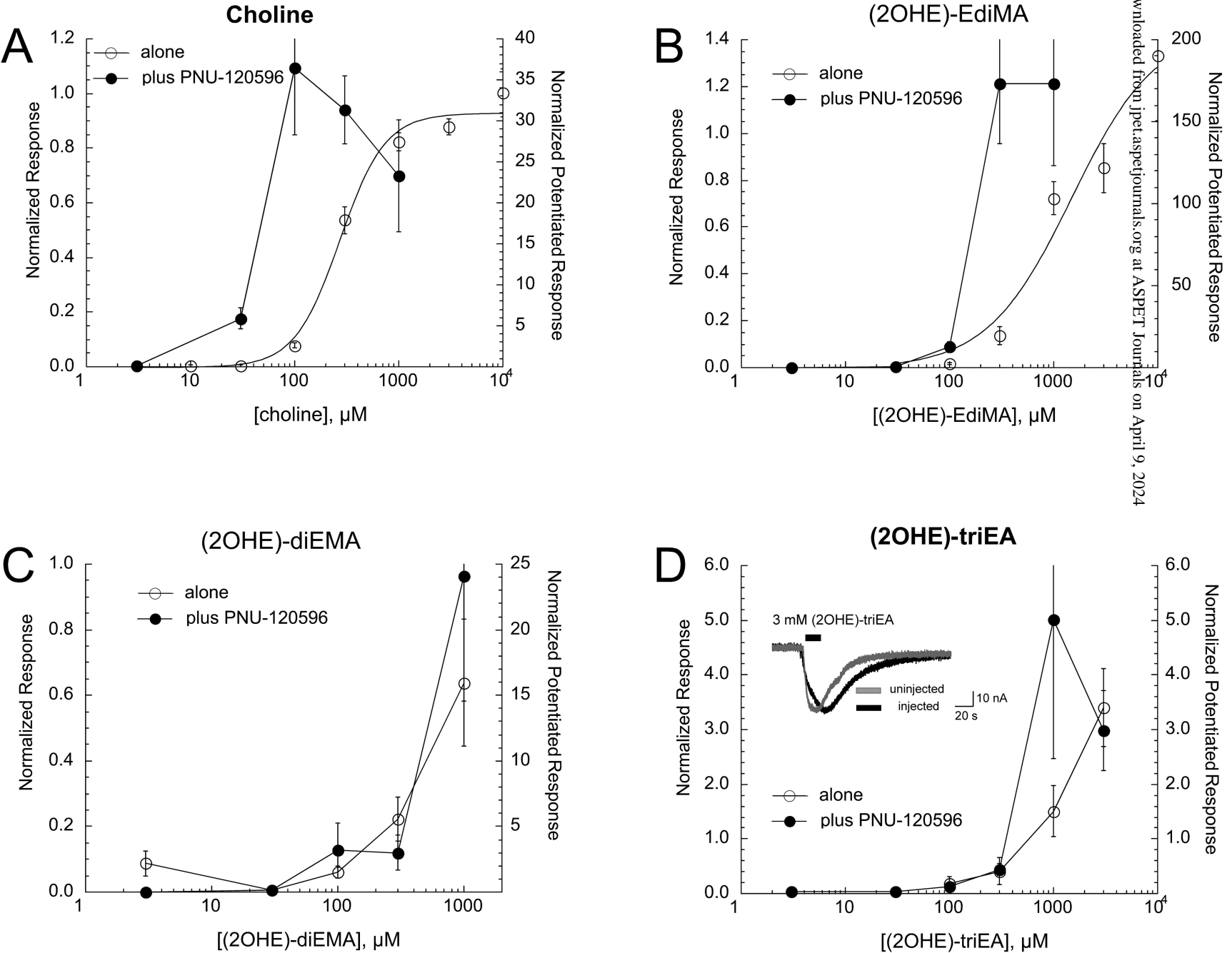


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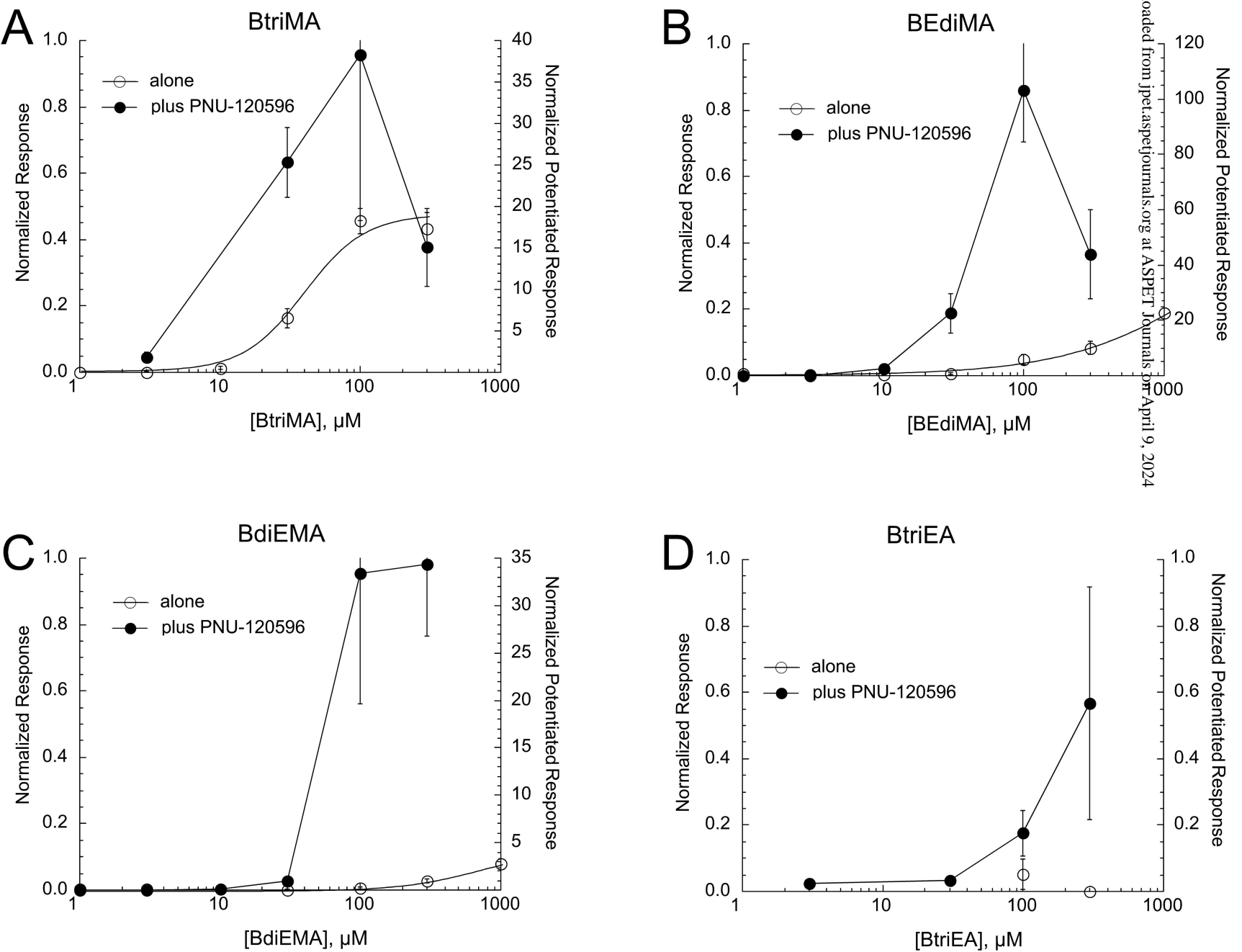


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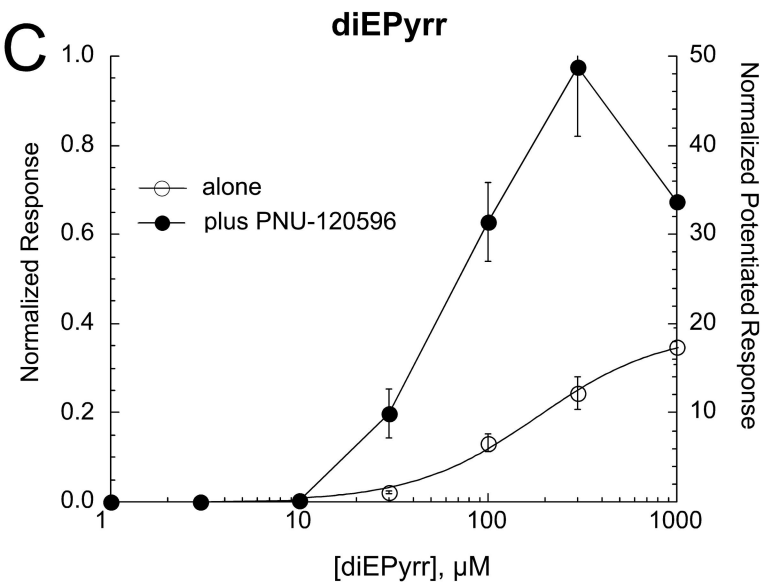
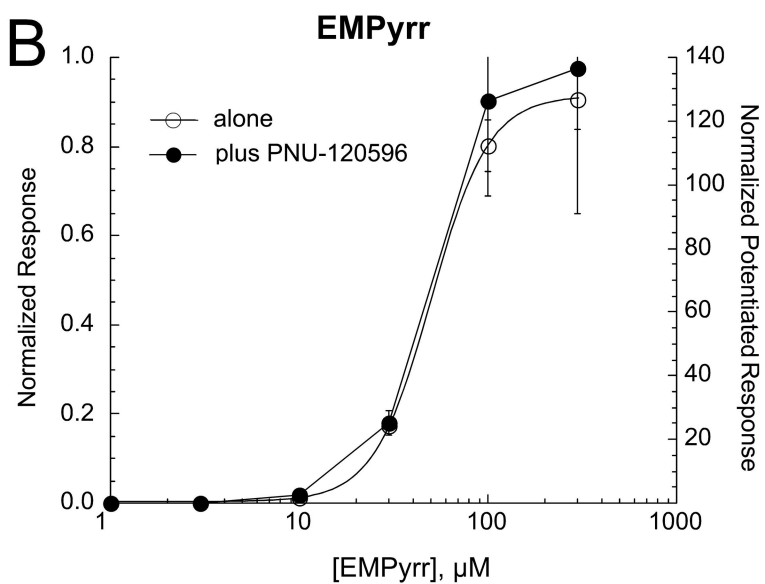
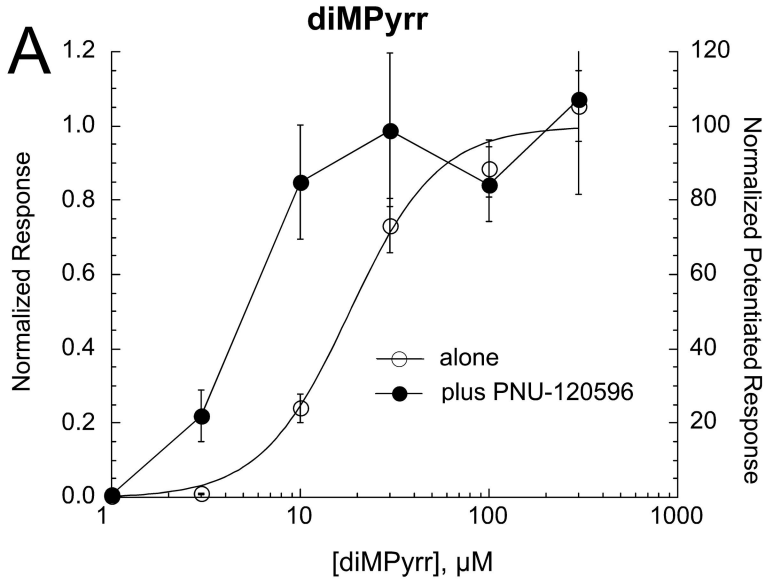


Figure 6

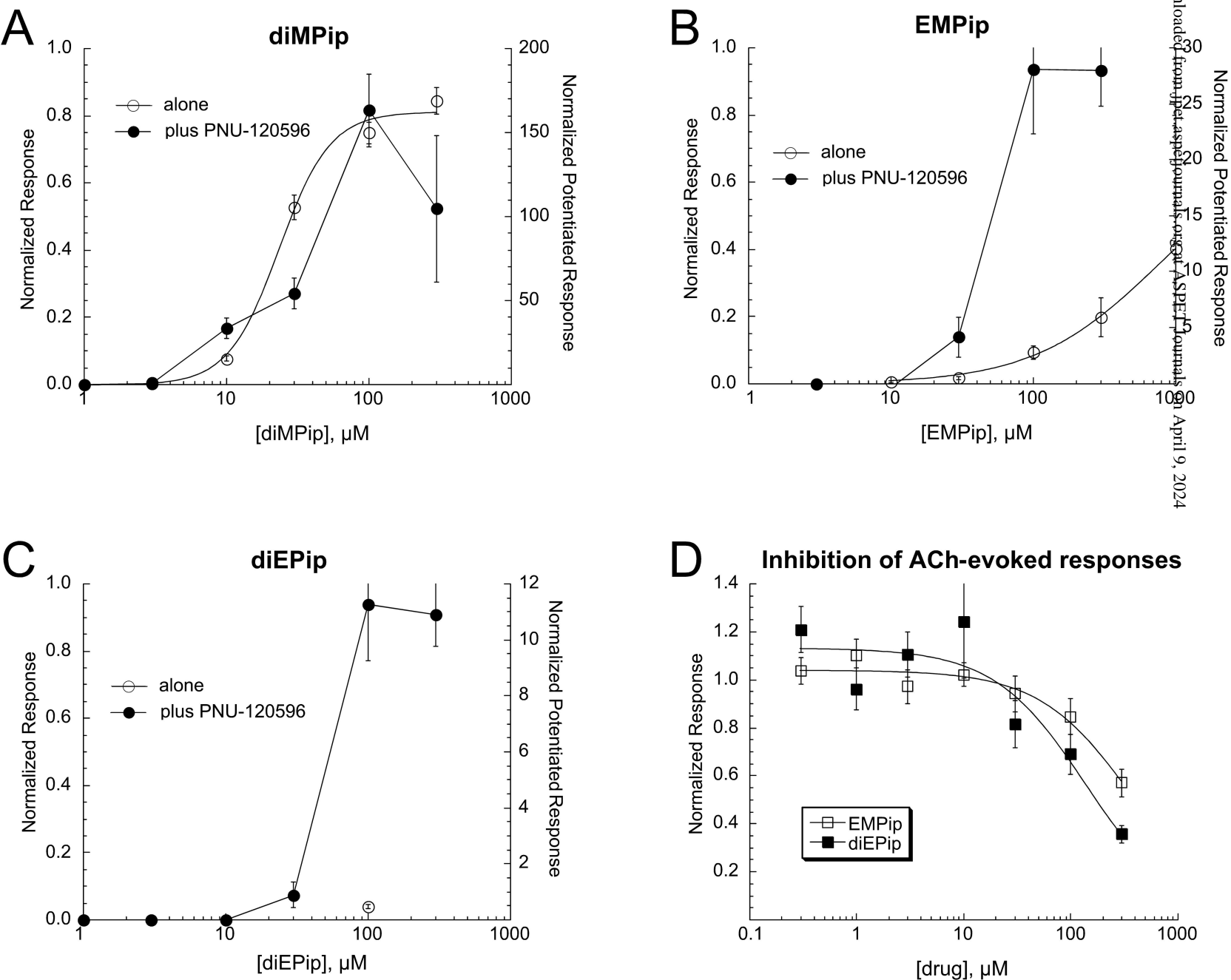


Figure 7

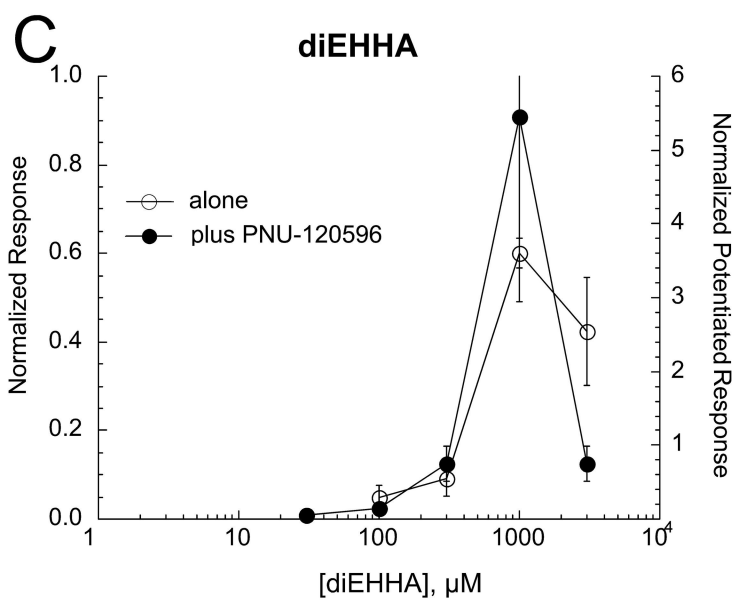
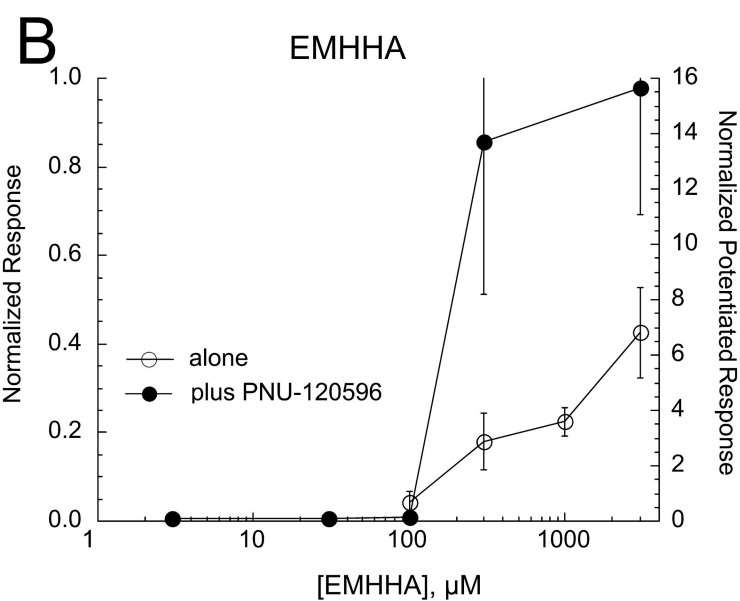
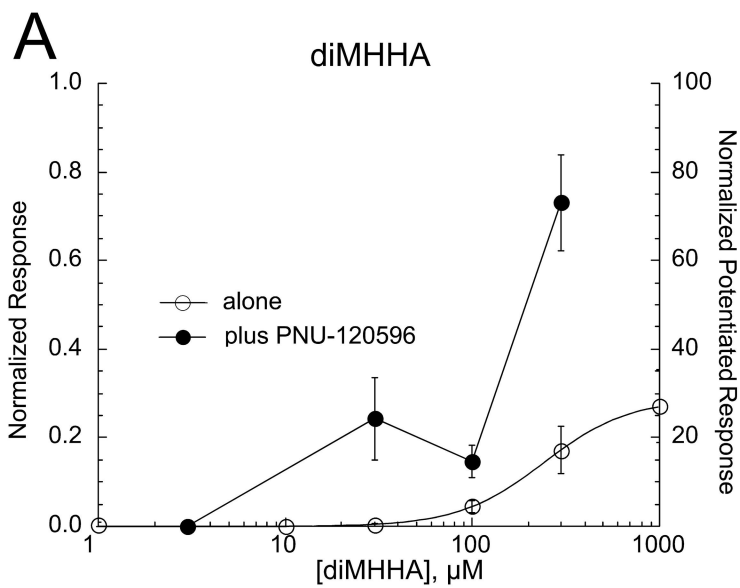


Figure 8

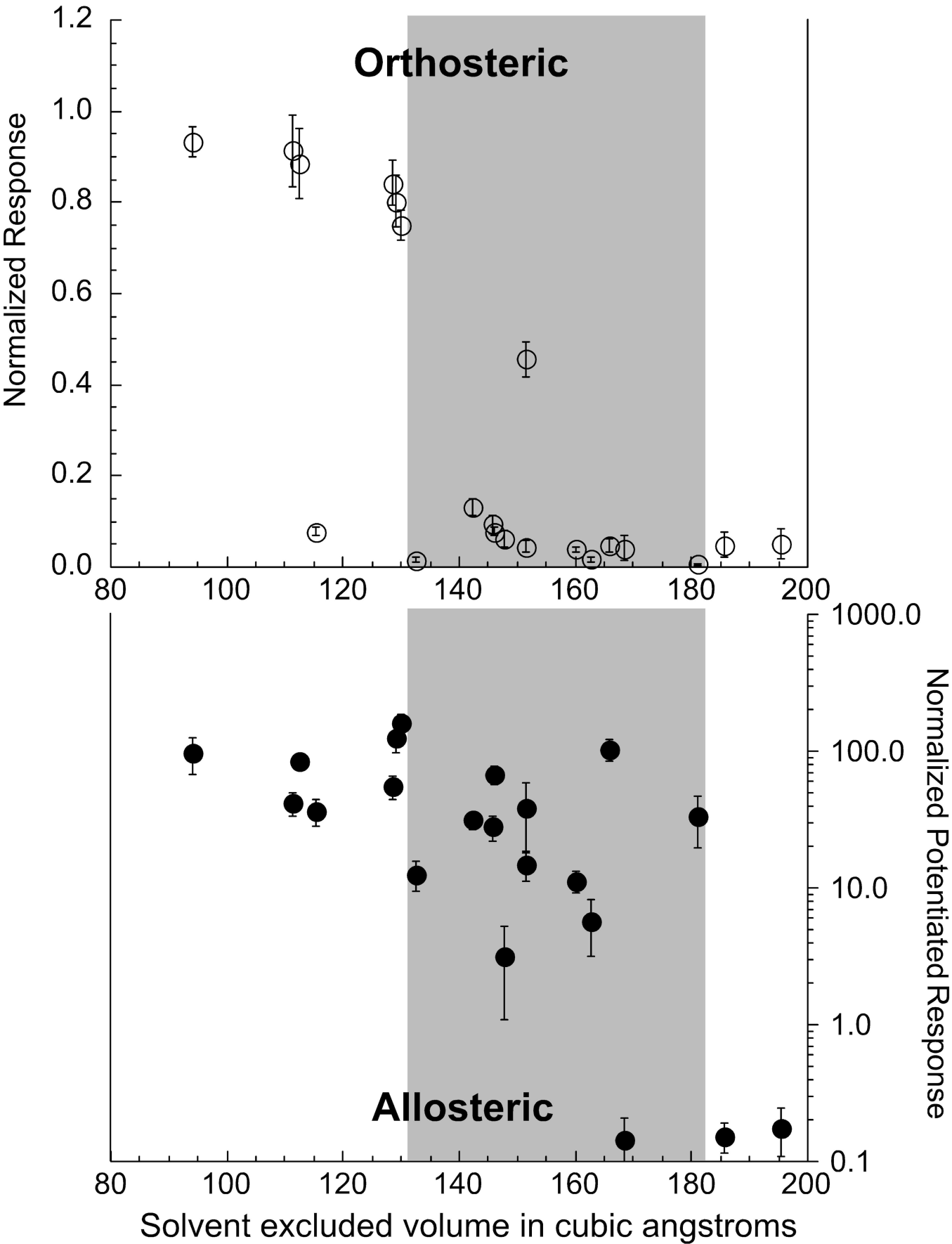


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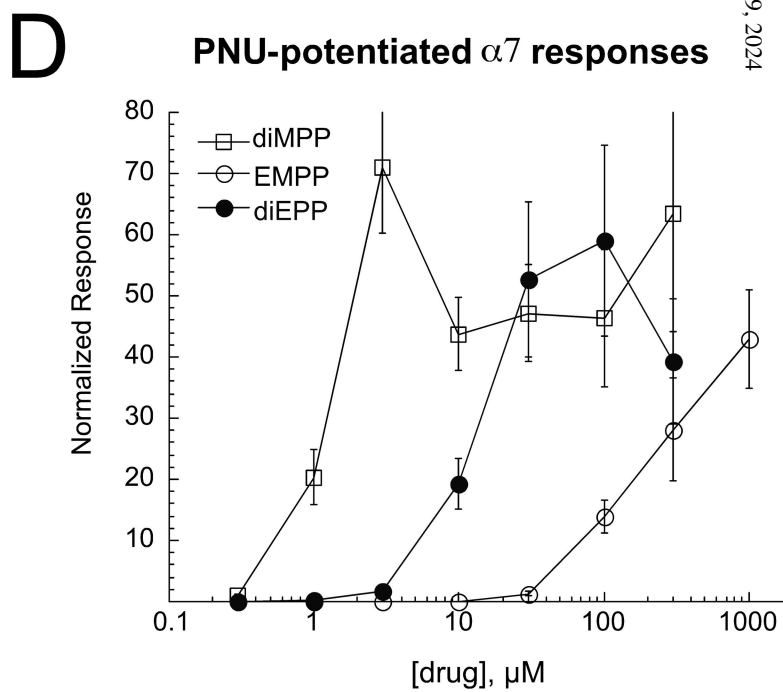
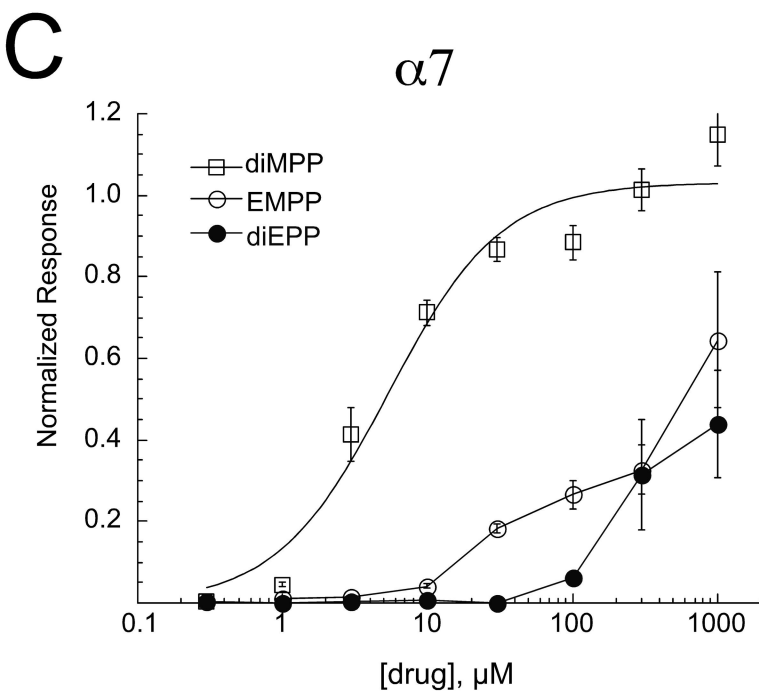
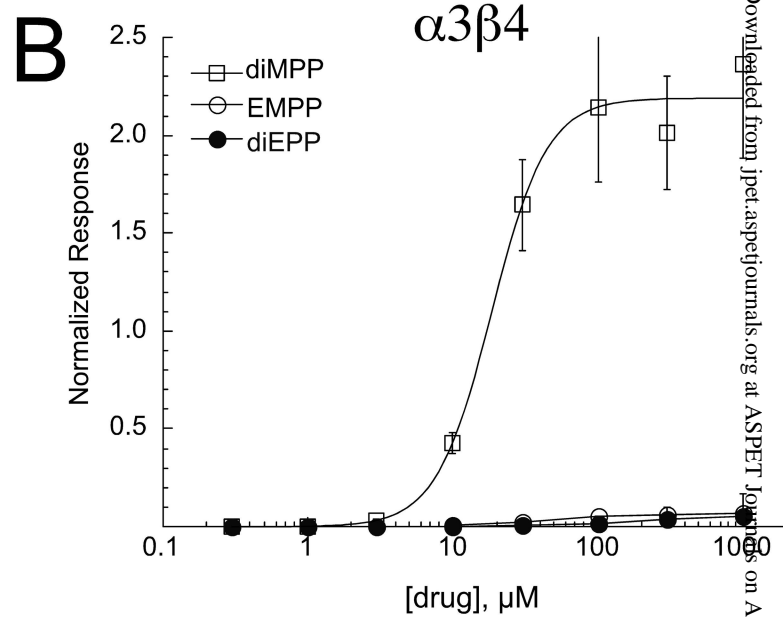
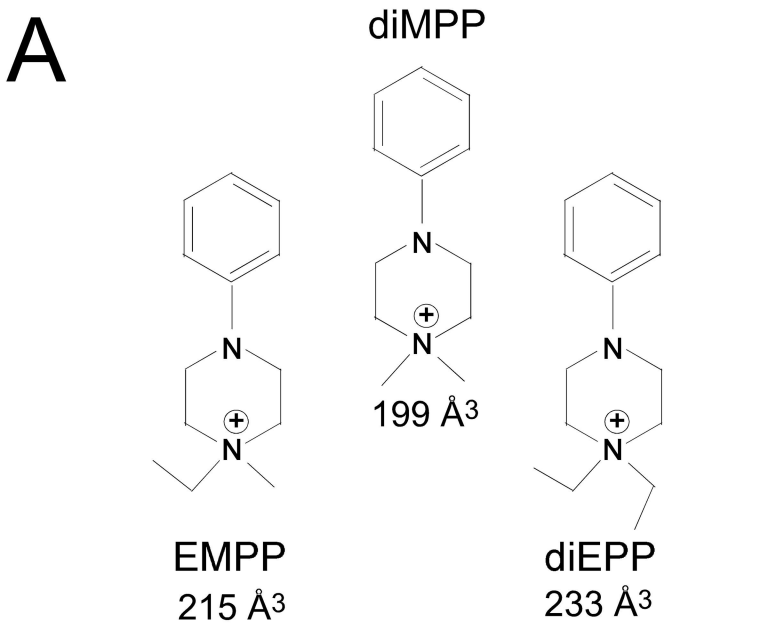


Figure 10

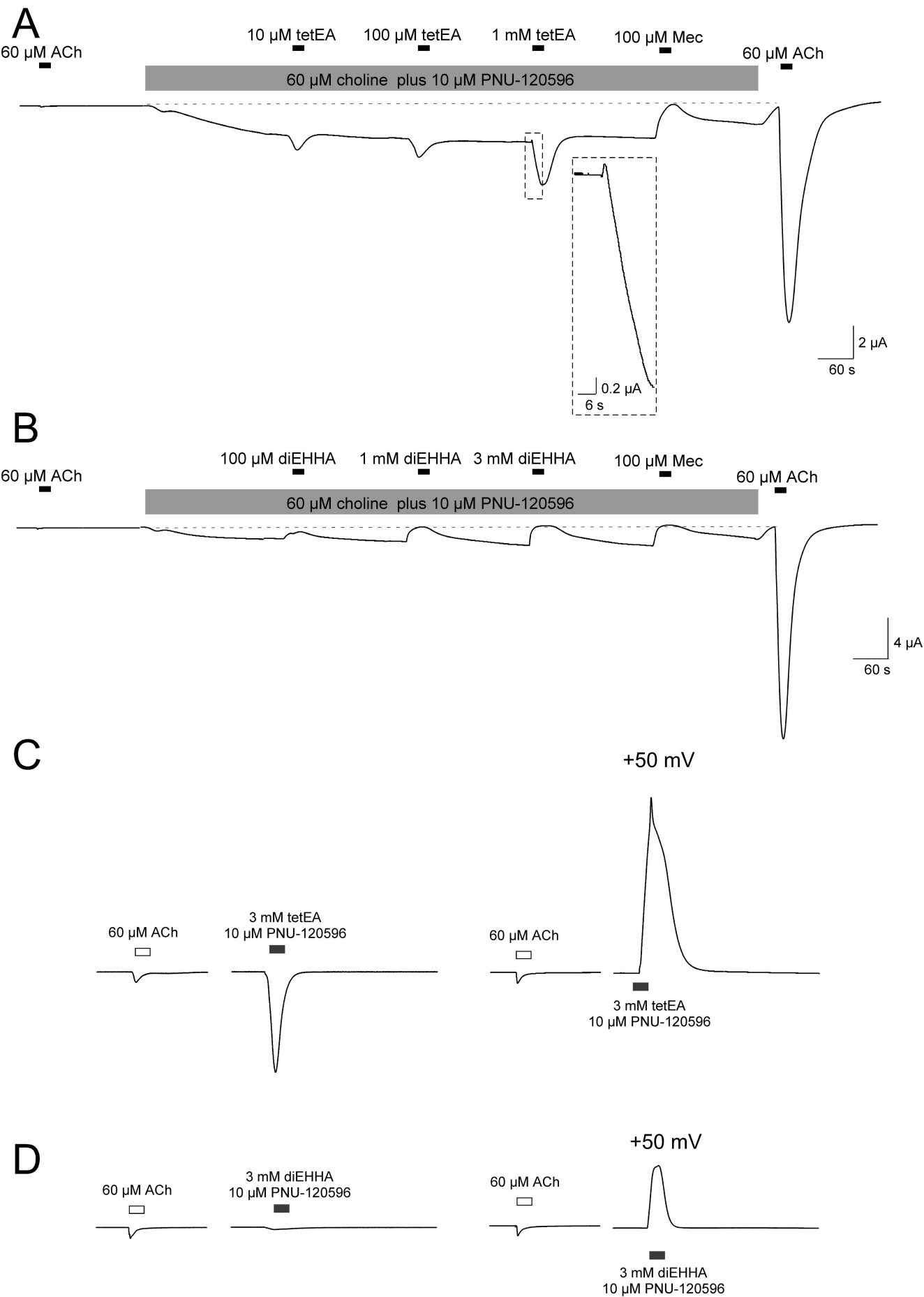


Figure 11

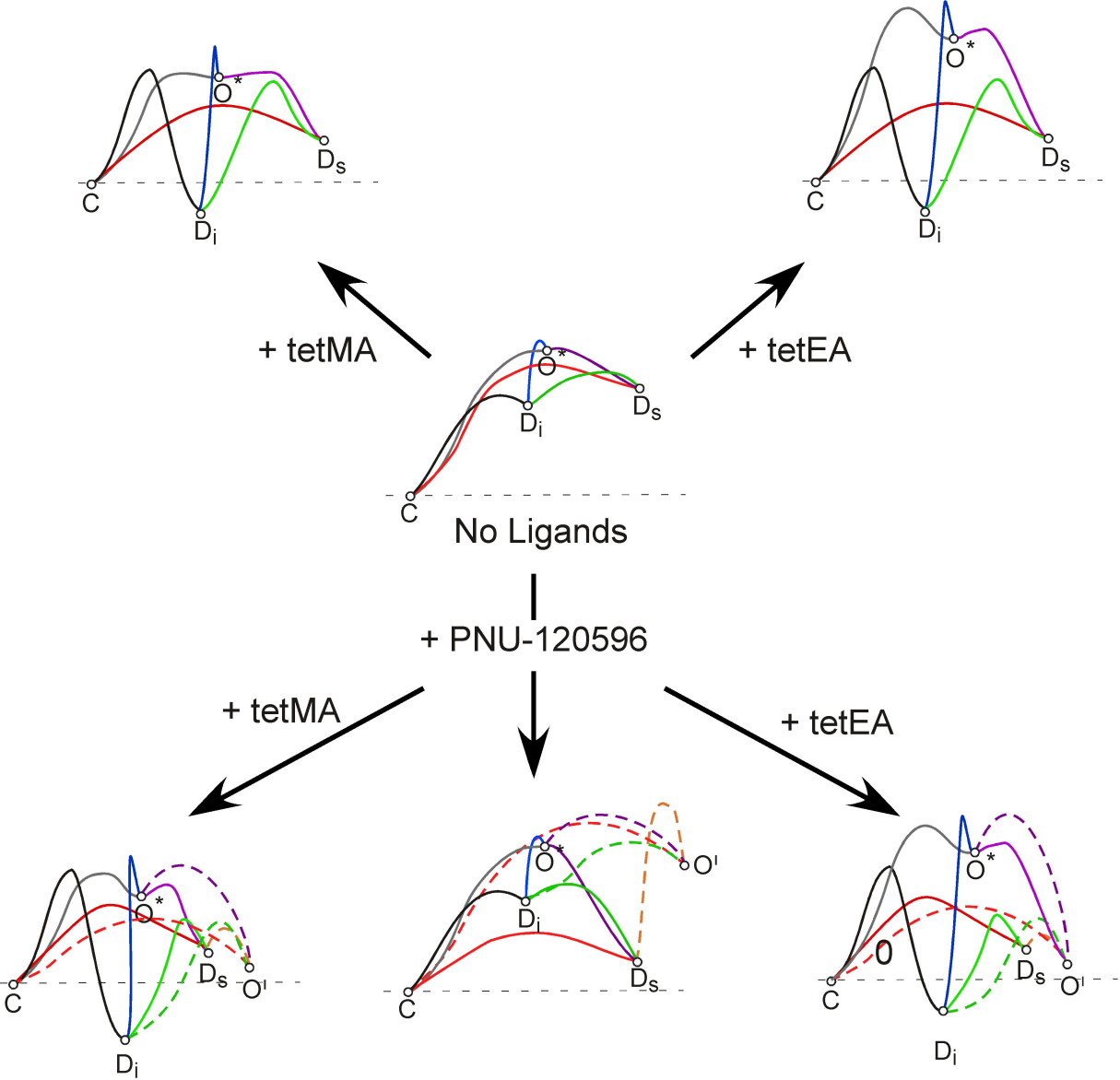


Figure 12