Pharmacokinetics and Pharmacodynamic Effects of Nemvaleukin Alfa, a Selective Agonist of the Intermediate-Affinity IL-2 Receptor, in Cynomolgus Monkeys

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ABSTRACT
Nemvaleukin alfa (nemvaleukin, ALKS 4230) is a novel cytokine created by the fusion of circularly permuted interleukin-2 (IL-2) to the IL-2Rα subunit of the IL-2 receptor (IL-2R) complex that confers selectivity for the intermediate-affinity IL-2R expressed on CD8+ T cells and natural killer (NK) cells. The pharmacokinetics and selective pharmacodynamic properties of nemvaleukin have been demonstrated in vitro and in vivo mouse models. The pharmacokinetic/pharmacodynamic effects of nemvaleukin on immune cell subtypes were evaluated in cynomolgus monkeys after intravenous and subcutaneous administration to inform dose selection and predict pharmacodynamic effects in humans. Male drug-naïve cynomolgus monkeys (N = 15) were administered either single-dose (0.3 mg/kg i.v.; 0.3 mg/kg or 1.0 mg/kg s.c.) or repeated doses (0.1 mg/kg i.v. on days 1–5 or 0.5 mg/kg s.c. on days 1 and 4) of nemvaleukin. Serial blood samples were collected for pharmacokinetic assessment, immunophenotyping by flow cytometry, and profiling of serum cytokines. Repeat-dose subcutaneous administration of nemvaleukin with less frequent dosing resulted in total systemic exposure and trough serum concentrations comparable to those seen with intravenous administration, with lower peak serum concentrations. Transient elevation of interferon-γ and IL-6 peaked at 2 and 8 hours after intravenous and subcutaneous administration, respectively. Selective expansion of immunoprotective central memory, effector memory, terminal effector CD8+ T cells, and CD56+ NK cells, and minimal expansion of immunosuppressive CD4+CD25+FoxP3+ regulatory T cells was observed after both intravenous and subcutaneous administration. These data support the ongoing clinical evaluation of intravenous and subcutaneous nemvaleukin.

SIGNIFICANCE STATEMENT
Administration of the novel interleukin-2 receptor agonist nemvaleukin alfa to cynomolgus monkeys resulted in selective expansion of immune effector cells, including CD8+ T and natural killer cells with minimal effects on immunosuppressive CD4+ regulatory T cells, confirming the design of nemvaleukin and highlighting its potential as a cancer immunotherapy. Subcutaneous administration of nemvaleukin achieved systemic exposure and immunostimulatory effects similar to those observed after more frequent intravenous dosing and may represent a practical alternative in a clinical setting.

Introduction
Interleukin-2 (IL-2) was originally discovered as a T cell growth factor based on its ability to promote the growth of lymphocytes in vitro (Boymann and Sprent, 2012). However, it is now appreciated that IL-2 plays a much larger role in maintaining immune homeostasis, given that it is required for the development and function of immunosuppressive CD4+CD25+CD127int/FoxP3+ regulatory T cells (CD4+ Treg), as well as the function of immune effector cells, including conventional CD4+ T cells, CD8+ T cells, natural killer (NK) cells, and B cells (Waldmann, 2006; Boyman and Sprent, 2012; Choudhry et al., 2018). Presentation of tumor antigens by mature dendritic cells to antigen-specific CD4+ and CD8+ T cells in secondary lymphoid tissues results in the expression of high-affinity IL-2R complexes and the local production of IL-2, supporting the expansion of tumor antigen–specific T cells (Boymann and Sprent, 2012). NK cells, which are cytotoxic for major histocompatibility complexes class I–deficient tumor cells, also express high- and intermediate-affinity IL-2R complexes. However, high-affinity IL-2R complexes are constitutively expressed on CD4+ Treg (Boymann and Sprent, 2012). Thus, by activating and expanding CD4+ Treg, as well as conventional CD4+ T cells, CD8+ T cells, and NK cells, IL-2 exerts...
dual regulatory (e.g., on immunosuppressive CD4+ Tregs) and stimulatory (e.g., on immune effector CD8+ T cells and NK cells) effects on the immune cells that are integral to the cancer immunity cycle. Recombinant IL-2 (aldesleukin) was among the first immunotherapies to be approved for the treatment of advanced/metastatic renal cell carcinoma and melanoma. Paradoxically, the high doses required for antitumor activity have been associated with activation and expansion of high-affinity IL-2R–expressing CD4+ Tregs that are hypothesized to hinder antitumor efficacy. Furthermore, widespread use of high-dose IL-2 has been limited due to its association with potentially life-threatening acute toxicities (e.g., vascular leak syndrome that can lead to liver cell damage and renal failure), which may be explained by the expression of the high-affinity IL-2R complex on endothelial cells (Atkins et al., 1999; Boyman and Sprent, 2012; Sim et al., 2014; Amaria et al., 2015; Choudhry et al., 2018).

Leveraging the immunostimulatory effects of IL-2, while at the same time mitigating the protumor activation and expansion of CD4+ Tregs and avoiding the toxicity associated with high doses, is hypothesized to enhance the immunotherapeutic potential of IL-2 (Ahmadzadeh and Rosenberg, 2006; Skrombolas and Frelinger, 2014). This strategy can be achieved through selective targeting of the intermediate-affinity IL-2R, comprising IL-2Rβ and common γ chains expressed on immune effector cells, including CD8+ T cells and NK cells, while avoiding activation of the high-affinity IL-2R, comprising IL-2Rα (also known as CD25), IL-2Rβ, common γ chains expressed on CD4+ Tregs, and endothelial cells (Boyman and Sprent, 2012).

Nemvaleukin alfa (nemvaleukin, ALKS 4230) is a novel, engineered cytokine that was created by fusing circularly permuted IL-2 to the extracellular domain of the IL-2Rα subunit of the IL-2R complex (Fig. 1A). In comparison with IL-2, nemvaleukin selectively engages the intermediate-affinity IL-2R complex that is expressed on subsets of CD8+ T cells and NK cells and is sterically occluded from binding to the high-affinity IL-2R complex that is expressed preferentially on CD4+ Tregs (Fig. 1B) (Antony and Restifo, 2005; Sun et al., 2019; Lopes et al., 2020). Data from in vitro human studies and in vivo mouse studies have shown that, compared with recombinant human IL-2, nemvaleukin selectively activates and expands circulating NK cells and memory-phenotype CD8+ T cells with negligible effects on CD4+ Tregs (Lopes et al., 2020). This selective expansion of immune effector cells with antitumor function(s) has promising therapeutic potential, and clinical studies of nemvaleukin, as monotherapy or in combination with the anti–programmed death-1 checkpoint inhibitor pembrolizumab, in patients with advanced solid tumors are ongoing (ARTISTRY-1 (NCT02799085); ARTISTRY-2 (NCT03861793); ION-01 nemvaleukin (NCT04144517)).

The present study examines the pharmacokinetics (PK) and pharmacodynamic (PD) effects of nemvaleukin on multiple immune cell subtypes (including circulating CD8+ T cells, NK cells, CD4+ T cells, and CD4+ Tregs) in cynomolgus monkeys after intravenous and subcutaneous routes of administration. The PK-PD relationship in cynomolgus monkeys is expected to inform dose selection and predict PD effects in humans.

Materials and Methods

Animals

Male drug-naïve cynomolgus monkeys (N = 15) (Covance Laboratories, Inc., Madison, WI and Covance Research Products, Inc., Alice, TX) aged 2–4 years were housed in stainless steel cages at ambient temperatures under a 12-hour light/dark cycle and fed a certified pristine diet with water ad libitum. After acclimation to study conditions over a 1- to 4-day period, animals were selected for testing based on their overall health and body weight, which ranged from 2.4 kg to 3.4 kg at baseline. All procedures were conducted in compliance with the Animal Welfare Act Regulations (9 CFR 3).

Study Design and Dosing Schedule

All dosing and sample collections were conducted at Covance Laboratories. Individual doses of nemvaleukin were calculated based on body weight as measured on the day of dosing in the single-dose study.
and the first day of dosing in the repeat-dose study. Animals were not fasted prior to dosing. In the single-dose study, nemvaleukin was administered intravenously at 0.3 mg/kg or subcutaneously at 0.3 or 1.0 mg/kg (three animals per dose group; Supplemental Fig. 1A). In the repeat-dose study, nemvaleukin was administered intravenously at 0.1 mg/kg once daily for 5 consecutive days on days 1–5 (group 1) or subcutaneously at 0.5 mg/kg on day 1 and day 4 (group 2; three animals per dose group; Supplemental Fig. 1B). The intravenous dose was administered via a saphenous vein using a needle and catheter set as a slow push (over ≥1 minute). The subcutaneous dose was administered via syringe and needle in the midscapular region. Blood samples to determine serum concentrations of nemvaleukin and selected cytokines (approximately 2 ml) were collected from the femoral vein into tubes without anticoagulant at predefined time points (Supplemental Fig. 1). Whole blood samples for immunophenotyping (approximately 2 ml) were collected into tubes containing potassium EDTA anticoagulant at predefined time points (Supplemental Fig. 1).

Pharmacokinetic Data Analysis

Serum concentrations of nemvaleukin were analyzed by an independent bioanalytical laboratory (Covance, Chantilly, VA) via electrochemiluminescence assay using a sandwich immunoassay format on the Meso Scale Discovery (MSD) (Meso Scale Diagnostics, LLC, Rockville, MD) platform. The capture antibody, anti-human IL-2 Rα monoclonal antibody (mAb; clone 24204, mouse IgG1, catalog number MAB623, R and D Systems Inc., Minneapolis, MN), was coated onto a single-spot 96-well standard bind MSD plate and incubated for at least 12 hours at 2°C–8°C. The plate was washed and blocked before the sample was added. Nemvaleukin in the sample was bound to the capture antibody immobilized on the working electrode surface and was detected using the SULFO-TAG anti-hIL-2 antibody MAB202 (R and D Systems Inc., catalog number MAB202, clone #5334). After a 1-hour incubation, the plates were washed, and MSD read buffer (Meso Scale Diagnostics) was added. The plate was read in an MSD Sector Imager 6000 (Meso Scale Diagnostics) instrument, and data were analyzed using analysis of variance (ANOVA) with Tukey’s HSD test weighting in Watson software version 7.4.1 (range of quantitation (LLOQ) of 1.0 ng/ml and an upper limit of quantitation (ULOQ) of 2.5 ng/ml), medium QC (5.0 ng/ml), high QC (11.0 ng/ml), and D Systems Inc., catalog number MAB202, clone #5334). After a 1-hour incubation, the plates were washed, and MSD read buffer (Meso Scale Diagnostics) was added. The plate was read in an MSD Sector Imager 6000 (Meso Scale Diagnostics) instrument, and data were analyzed using analysis of variance (ANOVA) with Tukey’s HSD test weighting in Watson software version 7.4.1 (range of quantitation (LLOQ) of 1.0 ng/ml and an upper limit of quantitation (ULOQ) of 2.5 ng/ml), medium QC (5.0 ng/ml), high QC (11.0 ng/ml), and ULOQ (15.0 ng/ml). Please refer to the Supplemental Methods for the validation data. Nemvaleukin was stable in monkey serum for up to 26 hours at room temperature and up to six freeze/thaw cycles, for up to 72 hours refrigerated at 2°C–8°C, and for 3 months (99 days) at −60°C to −80°C.

PK parameters of nemvaleukin were calculated by a standard non-compartmental analysis method using Phoenix WinNonlin v6.4 (Pharsight, a Certara Company, Mountain View, CA). The following parameters were estimated after single-dose administration of nemvaleukin: Cmax, time to Cmax (Tmax), terminal elimination half-life (t1/2), area under the concentration-time curve from time 0 to the time of the last measurable serum concentration estimated by the linear trapezoidal rule (AUC), AUC from time 0 to infinity (AUC∞); calculated as AUC∞ = AUC1 + C0/t1/2, where C0 is the last measurable serum concentration and t1/2 is the terminal elimination rate constant estimated using log-linear regression of the terminal elimination phase of the concentration-time curve), mean residence time (MRT; calculated as AUMC/AUC∞, where AUMC is the area under the moment curve from 0 to infinity), total body clearance (CLc, calculated as dose/AUC∞), volume of distribution at steady state (Vdss, calculated as (AUC∞/AUC∞) × CLc), and absolute bioavailability after subcutaneous administration (%F, calculated as ([mean AUC{sub}i(i) × dose{sub}i(i)])/[mean AUC{sub}i(i) × dose{sub}i(i)]) × 100).

The following PK parameters were estimated after repeated-dose administration of nemvaleukin: Cmax, Tmax, t1/2, AUC∞, and total AUC over the entire dosing period (AUCtotal). For subcutaneous dosing, this was calculated as AUC{sub}i(i) + AUC{sub}d(i)i, and for intravenous dosing, this was estimated to lie between AUC{sub}t(1)(i) × AUC{sub}t(2)(i) × AUC{sub}t(3)(i) × AUC{sub}t(4)(i) × AUC{sub}t(5)(i) (assuming AUC{sub}t(1)(i) on days 2, 3, and 4 = AUC{sub}t(2)(i) on day 1) and AUC{sub}t(1)(i) × AUC{sub}t(2)(i) × AUC{sub}t(3)(i) × AUC{sub}t(4)(i) × AUC{sub}t(5)(i) (assuming AUC{sub}t(1)(i) on days 2, 3, and 4 = AUC{sub}t(4)(i) on day 5).

Immune Cell Analyses by Flow Cytometry

Assessment of Nemvaleukin in Vivo Potency on Primary Leukocytes. Leukocytes obtained from cynomolgus monkey whole blood after standard red blood cell lysis were counted, resuspended in X-VIVO 10 media, and incubated in the presence of decreasing concentrations of nemvaleukin (4-fold dilutions beginning at 150 nM for samples intended for analysis of memory CD8+ T cells and 3-fold dilutions beginning at 20 nM for samples intended for analysis of CD4+ Treg, NK, and CD8+ T cell surface markers). After surface marker staining, cells were washed and permeabilized for subsequent intracellular staining steps before sample acquisition on a BD LSRSpectra X-20 (BD Biosciences, San Jose, CA). Antibodies and fluorescent dyes used are described in Supplemental Table 1. Markers used to identify immune cell populations are detailed in Supplemental Table 2.

Immunophenotyping of Whole Blood. Blood sample processing and immunophenotypic data acquisition were performed at Alkermes, Inc. (Waltham, MA) approximately 24 hours after sample collection. Leukocytes obtained from cynomolgus monkey whole blood after standard red blood cell lysis were counted, resuspended in X-VIVO 10 media, and plated. Cells were then resuspended in viability stain buffer and incubated at room temperature, washed, Fc-blocked, and surface-stained with fluorescently conjugated antibodies. Cells were washed and permeabilized for subsequent intracellular staining steps prior to sample acquisition on a BD LSRSpectra X-20 using a high-throughput sampler. Antibodies and fluorescent dyes used are listed in Supplemental Table 3. CD4+ Treg, NK, and naive/memory CD4+ and CD8+ T cell populations were identified using the markers described in Supplemental Table 4.

Quantification of Serum Concentrations of Proinflammatory Cytokines

The V-PLEX Proinflammatory Panel 1 NHP Kit (Meso Scale Diagnostics) was used to quantify concentrations of IL-1β, IL-6, IL-8, IL-10, and interferon-γ (IFNγ) in monkey serum samples using an MSD SQ120 instrument.

Results

In Vitro Potency of Nemvaleukin on Activating Lymphocyte Populations. The in vitro potency of nemvaleukin on activating selected lymphocyte subsets, including CD16+ NK cells, memory subsets of CD8+ T cells, and CD4+ Treg, in terms of the EC50, were similar between cynomolgus monkeys and humans (Table 1), suggesting that cynomolgus monkey is a pharmacologically relevant species for predicting immunologic activity in humans. Potencies for NK cells and Treg, were similar to each other and between the monkey and human samples, with EC50 values within the 0.5 nM range, whereas EC50 values for memory CD8+ T cell subsets were in the 1 to 2 nM range.

Pharmacokinetics of Nemvaleukin after Single-Dose Administration. After a single intravenous bolus dose of 0.3 mg/kg, serum nemvaleukin concentrations declined in a
In notable increases in CD8 doses of intravenous and subcutaneous nemvaleukin resulted. Concentrations and AUCtotal after two subcutaneous doses repeated intravenous doses of 0.1 mg/kg, trough serum concentrations of nemvaleukin after subcutaneous 8 hours after the second dose on day 4. Although peak serum CL and mean Vdss were 10.6 ml/h/kg and 135 ml/kg, respectively. In contrast, approximately 2-fold increases in CD4 two dosing regimens, with 6- and 4-fold increases, respectively (Table 2). After a single subcutaneous dose of 0.3 mg/kg or 1 mg/kg, serum concentrations of nemvaleukin from 0.3 mg/kg to 1.0 mg/kg (Table 2). As a result, subcutaneous bioavailability was estimated to be 45% at 0.3 mg/kg and 28% at 1.0 mg/kg of nemvaleukin.

**Pharmacokinetics of Nemvaleukin after Repeat-Dose Administration.** After subcutaneous administration of nemvaleukin at 0.5 mg/kg, Cmax of nemvaleukin was reached at 8 hours after the first dose on day 1 and at 2 or 8 hours after the second dose on day 4. Although peak serum concentrations of nemvaleukin after subcutaneous doses of 0.5 mg/kg were 2- to 3-fold lower than those after repeated intravenous doses of 0.1 mg/kg, trough serum concentrations and AUCtotal after two subcutaneous doses of 0.5 mg/kg (on days 1 and 4) were comparable to those after five daily intravenous doses of 0.1 mg/kg (on days 1–5; Fig. 2B, Table 3).

**Pharmacodynamic Effects of Nemvaleukin.** Repeat doses of intravenous and subcutaneous nemvaleukin resulted in notable increases in CD8 T cells and NK cells, peaking on days 6–8, whereas only modest increases in CD4 Tregs were observed (Fig. 3, A and B). Elevated numbers of CD8 T cells and CD56 NK cells were sustained until day 14, at which point cell numbers returned to predose levels. The expansion of CD8 T cells and CD56 NK cells was similar between the two dosing regimens, with 6- and 4-fold increases, respectively. In contrast, approximately 2-fold increases in CD4 Tregs were observed (Fig. 3B). Expression of the intracellular proliferation marker Ki-67 was also measured. Increased expression of Ki-67 was observed starting on day 3, and peaked at greater than 80% on day 6 for CD8 T cells and CD56 NK cells after intravenous administration (Fig. 3C). The peak of Ki-67 expression was not captured after subcutaneous administration, because several blood samples collected on days 6 and 7 froze during shipment, precluding their analyses. Increased Ki-67 expression was also observed in CD4 Tregs, but to a lesser extent (~40%), consistent with the modest 2-fold increases in cell numbers.

To assess whether particular subsets of CD8 T cells responded more markedly to nemvaleukin, CD8 T cells were categorized as naïve, central memory, effector memory, and terminal effector phenotype CD8 populations according to the surface markers listed in Supplemental Table 4. Increased cell numbers after intravenous and subcutaneous administration.

### Table 1

**Potency of nemvaleukin on selected lymphocyte subsets from cynomolgus monkey and human samples (Lopes et al., 2020)**

<table>
<thead>
<tr>
<th>Lymphocyte Population</th>
<th>Monkey</th>
<th>Human*</th>
<th>EC50</th>
<th>EC50</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>EC50</td>
<td>Mean ± S.D.</td>
<td>EC50</td>
<td>Mean ± S.D.</td>
</tr>
<tr>
<td>NK cells</td>
<td>0.47 ± 0.22</td>
<td>0.45 ± 0.09</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Naïve CD8+ T cells</td>
<td>1.4 ± 0.45</td>
<td>2.2 ± 1.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Central/transitional memory CD8+ T cells</td>
<td>1.3 ± 0.4</td>
<td>1.1 ± 0.1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Effector memory CD8+ T cells</td>
<td>2.0 ± 1.0</td>
<td>1.25 ± 0.4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Terminal effector CD8+ T cells</td>
<td>1.7 ± 0.8</td>
<td>0.93 ± 0.3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tregs</td>
<td>0.59 ± 0.14</td>
<td>0.59 ± 0.2</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Data generated from three separate experiments, each performed in triplicate, and the error represents S.D.

### Table 2

**PK parameters of nemvaleukin in male cynomolgus monkeys after a single intravenous (0.3 mg/kg) or subcutaneous (0.3 or 1.0 mg/kg) dose**

<table>
<thead>
<tr>
<th>Dose Route</th>
<th>Dose Cmax</th>
<th>Tmax</th>
<th>AUC0–12h</th>
<th>t1/2</th>
<th>MRT</th>
<th>CL</th>
<th>Vdss</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intravenous</td>
<td>0.3</td>
<td>6119 ± 1188</td>
<td>0.083 (0.083, 0.083)</td>
<td>28,460 ± 1230</td>
<td>49.5 ± 5.6</td>
<td>12.8 ± 1.9</td>
<td>10.6 ± 0.46</td>
</tr>
<tr>
<td>Subcutaneous</td>
<td>0.3</td>
<td>549 ± 208</td>
<td>8 (8, 8)</td>
<td>12,931 ± 3893</td>
<td>61.9 ± 4.6</td>
<td>31.6 ± 4.0</td>
<td>—</td>
</tr>
<tr>
<td>Subcutaneous</td>
<td>1.0</td>
<td>1035 ± 133</td>
<td>8 (8, 8)</td>
<td>26,878 ± 2106</td>
<td>37.3 ± 19.4</td>
<td>21.7 ± 2.8</td>
<td>—</td>
</tr>
</tbody>
</table>

N = 3 per treatment group.

Mean (± S.D.) for all parameters except Tmax [median (min, max)].
of nemvaleukin were observed for all CD8⁺ T cell subsets (Fig. 4A). Numbers of terminal effector and effector memory CD8⁺ T cells were increased to a greater extent than of central memory and naïve CD8⁺ T cells, by approximately 20- and 30-fold compared with 10- and 4-fold, respectively (Fig. 4B).

Changes in the number of central memory CD4⁺ T cells after repeat-dose administration of nemvaleukin (Supplemental Fig. 2) were generally similar to the observed changes in CD4⁺ Treg.

Serum levels of proinflammatory cytokines were also measured, and may correlate with responses by particular immune cell populations. Transient elevation of IL-6 and IFNγ was observed after both intravenous and subcutaneous administration of nemvaleukin and peaked at 2 hours (intravenous) and 8 hours (subcutaneous) postdosing (Supplemental Fig. 3). No notable increases in levels of IL-1β, IL-8, or IL-10 were observed after intravenous or subcutaneous administration of nemvaleukin (data not shown).

**Discussion**

The amplification of antitumor T cell responses is a key component of the cancer immune cycle and forms the basis of the rationale for the use of high-dose IL-2 in patients with metastatic melanoma or advanced renal cell carcinoma (Chen and Mellman, 2013; Clark et al., 2017; Buchbinder et al., 2019; Fishman et al., 2019). However, high-dose IL-2 therapy has been shown to induce expansion of both antitumor effector cell and Treg populations, with greater expansion of ICOsTreg correlating with poor outcome of high-dose IL-2 therapy (Sim et al., 2014). In addition, maximal efficacy of recombinant IL-2 therapy in humans requires an intensive high-dose intravenous administration regimen, as demonstrated in a head-to-head phase III study comparing intravenous dosing regimens (McDermott et al., 2015). Nemvaleukin is designed to selectively activate antitumor T cell and NK cell populations over Treg, as was observed in mice and with human peripheral blood mononuclear cells in vitro. In addition, equivalent antitumor efficacy was observed after intravenous and subcutaneous administration of nemvaleukin in tumor-bearing mice (Lopes et al., 2020). The pathologic effects of recombinant IL-2 in monkeys are known to be predictive of pathogenesis in humans (Harada and Yahara, 1993; Gillies et al., 2011). As the in vitro potency for nemvaleukin on immune cell subsets found in peripheral blood were similar for cynomolgus monkeys and humans, the PK-PD relationship in cynomolgus monkeys is expected to inform dose selection and predict PD effects in humans. These studies conducted with cynomolgus monkeys set out to compare the PK and PD effects of single or repeated doses of nemvaleukin after intravenous or subcutaneous administration. An additional goal was to ascertain whether subcutaneous administration of nemvaleukin could achieve similar or more favorable PD responses versus intravenous administration in cynomolgus monkeys.

PK analysis showed that subcutaneous administration of nemvaleukin with less frequent dosing can achieve total systemic exposure and trough serum concentrations that are comparable to those achieved with daily intravenous administration, albeit with lower peak serum concentrations. Analysis of the PD effects of nemvaleukin on multiple immune cell subtypes (including circulating CD4⁺ T cells, CD8⁺ T cells, CD56⁺ NK cells, and Treg) revealed the selective expansion of potentially immunoprotective central memory, effector memory, and terminal effector CD8⁺ T cell and CD56⁺ NK cell populations after both intravenous and subcutaneous administration. It is worth noting that, in the current study, the overall expansion of CD8⁺ T cell and NK cell populations was similar between intravenous (once daily on days 1–5) and subcutaneous (on day 1 and day 4) dosing regimens, which correlated with similar total systemic exposure to nemvaleukin between the two dosing regimens.

Ki-67 protein expression was used as a marker of immune cell proliferation and, as such, has been validated and is widely used as a means of quantifying the dividing fraction of a given cell population (Scholzen and Gerdes, 2000; Sun and Kaufman, 2018). Our data show that the expanded populations of CD8⁺ T cells and NK cells that result from nemvaleukin administration are actively undergoing proliferation, whereas the population of Treg showed less pronounced levels of proliferation. The comparatively modest expansion of total Treg after administration of intravenous or subcutaneous nemvaleukin further supports its immunotherapeutic potential and distinguishes it from other IL-2–based therapies. The selective expansion of CD8⁺ T cells and NK cells by nemvaleukin, with limited impact on Treg expansion, observed in mice (Lopes et al., 2020) and here in cynomolgus monkeys, is in marked contrast to the Treg-biased immune profile induced by recombinant IL-2 in mice (Lopes et al., 2020), monkeys (Bell et al., 2015), and humans (Sim et al., 2014). Indeed, the propensity of low-dose recombinant IL-2 to elicit the expansion of Treg has prompted its use as a treatment of autoimmune diseases, such as rheumatoid arthritis, which are characterized by deficiencies of Treg (Ye et al., 2018).

Analysis of serum concentrations of proinflammatory cytokines showed transient elevation of both IL-6 and IFNγ that peaked at 2 hours and 8 hours after intravenous and subcutaneous administration of nemvaleukin, respectively. After

### Table 3

<table>
<thead>
<tr>
<th>Dose route</th>
<th>Dose</th>
<th>Day</th>
<th>Cmax</th>
<th>Tmax</th>
<th>Ctrough</th>
<th>AUC₂₄h,day₁</th>
<th>AUC₂₄h,day₅</th>
<th>AUCt,day₁</th>
<th>AUCt,day₅</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intravenous</td>
<td>0.1</td>
<td>1</td>
<td>2757 ± 176</td>
<td>0.083 (0.083, 0.083)</td>
<td>9.62 ± 1.39</td>
<td>5159 ± 301</td>
<td>9035 ± 602</td>
<td>41,753 ± 2213</td>
<td>30,124 ± 1112</td>
</tr>
<tr>
<td>Intravenous</td>
<td>0.5</td>
<td>1</td>
<td>911 ± 432</td>
<td>8 (8, 8)</td>
<td>&lt;LLOQ</td>
<td>17,656 ± 6700</td>
<td>17,656 ± 6700</td>
<td>33,402 ± 9204</td>
<td></td>
</tr>
<tr>
<td>Intravenous</td>
<td>1</td>
<td>1</td>
<td>1006 ± 119</td>
<td>2 (2, 8)</td>
<td>10.0 ± 1.25</td>
<td>15,214 ± 2529</td>
<td>15,746 ± 2601</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Subcutaneous</td>
<td>0.1</td>
<td>1</td>
<td>2096 ± 80</td>
<td>0.083 (0.083, 0.083)</td>
<td>9.62 ± 1.39</td>
<td>5159 ± 301</td>
<td>9035 ± 602</td>
<td>41,753 ± 2213</td>
<td>30,124 ± 1112</td>
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<td>15,214 ± 2529</td>
<td>15,746 ± 2601</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*N = 3 per treatment group, LLOQ = 1.00 ng/ml.

*Mean (± S.D.) for all parameters except Cmax (median, min, max).

*Calculated based on AUC₂₄h,day₁ = AUCt,day₁.

*Calculated based on AUC₂₄h,day₁ + AUC₂₄h,day₅ = AUCt,day₁ + AUCt,day₅.

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Repeat dosing, levels of IL-6 trended higher after intravenous administration relative to subcutaneous administration, which could be attributed to the higher observed Cmax for nemvaleukin after intravenous administration compared with subcutaneous administration. These cytokine data might suggest better tolerability with a subcutaneous treatment regimen, given the association between IL-6 and loss of endothelial barrier function and, by implication, vascular leakage (Alsaffar et al., 2018; Narazaki and Kishimoto, 2018). IFNγ production by immune effector cells is associated with antitumor efficacy (Ivashkiv, 2018); therefore, the induction of IFNγ suggests that nemvaleukin might promote the activation of cells associated with antitumor immune responses after both intravenous and subcutaneous administration.

In conclusion, subcutaneous administration of nemvaleukin can achieve total systemic exposure similar to that of intravenous administration, with less frequent dosing and a lower Cmax, resulting in similar expansion of CD8+ T cells and

Fig. 3. Effect of repeated intravenous (0.1 mg/kg for 5 consecutive days) or subcutaneous (0.5 mg/kg on days 1 and 4) administration of nemvaleukin on (A) total CD8+ T cells and CD56+ NK cells and (B) CD4+ Tregs, and (C) the proportion of cells expressing the cell proliferation marker Ki-67. Data shown are the mean ± S.E. of the mean (S.E.M.) with N = 3 per treatment group. Fold changes were calculated relative to predose levels for individual monkeys.
CD56⁺ NK cells with minimal expansion of CD4⁺ Treg. The PK and PD data in monkeys reported here support clinical evaluation of intravenous and subcutaneous administration of nemvaleukin in humans with advanced solid tumors. In addition to the ongoing phase I/II ARTISTRY-1 first-in-human study of intravenous nemvaleukin (NCT02799095), the PD, PK, safety, and efficacy of subcutaneous administration of nemvaleukin are currently under investigation in the phase I/II ARTISTRY-2 study (NCT03861793). The selection of subcutaneous nemvaleukin dose and dosing frequency in ARTISTRY-2 was determined from predictions (manuscript in development) based on the observed PK-PD relationship from ARTISTRY-1 and in monkeys after intravenous and subcutaneous routes of administration. In the ARTISTRY-2 study, subcutaneous nemvaleukin is given as a single agent or in combination with pembrolizumab to patients with selected advanced or metastatic solid tumors. The subcutaneous route of administration might provide an alternative administration option for patients.

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Participated in research design: Lopes, Sun, Losey.
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References
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