Ginsenoside Ro Ameliorates High-Fat Diet–Induced Obesity and Insulin Resistance in Mice via Activation of the G Protein–Coupled Bile Acid Receptor 5 Pathway

Lin-shan Jiang1, Wei Li1, Tong-xi Zhuang, Jie-jing Yu, Shuai Sun, Zheng-cai Ju, Zheng-tao Wang, Li-li Ding*, and Li Yang*

Shanghai Key Laboratory of Complex Prescription and MOE Key Laboratory for Standardization of Chinese Medicines, Institute of Chinese Materia Medica (L.J., W.L., T.Z., J.Y., S.S., Z.J., Z.W., L.D., L.Y.), and Institute of Interdisciplinary Integrative Medicine Research (L.J., J.Y., L.Y.), Shanghai University of Traditional Chinese Medicine, Shanghai, China; and Shanghai R&D Center for Standardization of Traditional Chinese Medicine, Shanghai, China (L.J., W.L., T.Z., J.Y., S.S., Z.J., Z.W., L.D., L.Y.)

Received November 24, 2020; accepted March 26, 2021

ABSTRACT

Obesity, a well known risk factor in multiple metabolic diseases, is dramatically increasing worldwide. Ginsenosides extracted from ginseng have been reported against obesity and the associated metabolic disorders. As a subtype of ginsenoside, ginsenoside Ro is a critical constituent of ginseng. However, its specific effects on obesity remain unknown. G protein–coupled bile acid receptor 5 (TGR5) (also known as GPBAR1) is a bile acid membrane receptor, widely expressed in human tissues contributing to various metabolic processes to confer the regulations of glucose and lipid homeostasis. TGR5 has displayed potential as a therapeutic target for the treatment of metabolic disorders. Here, we explore the antiobesity effect of ginsenoside Ro with TGR5 activation screened by a library of natural products. Our results showed that the ginsenoside Ro (90mg/kg) treatment ameliorated body weight and lipid accumulation in multiple metabolic organs of high-fat diet–induced obese (DIO) mice without affecting food intake and improved oral glucose tolerance tests, intraperitoneal insulin tolerance tests, and fasting serum glucose. We also found that triglyceride and total cholesterol in serum and liver were significantly decreased after ginsenoside Ro treatment. Then we used Tgr5 knockout mice to explore the role of Tgr5 in the antiobesity effect of ginsenoside Ro. Our results further demonstrated that ginsenoside Ro promoted glucagon-like peptide 1 (GLP-1) secretion and energy expenditure in wild-type DIO mice. However, the stimulation of ginsenoside Ro on GLP-1 secretion and energy expenditure were restrained in the Tgr5 knockout mice. In conclusion, our findings demonstrated that ginsenoside Ro ameliorates obesity and insulin resistance in DIO mice via activating TGR5, indicating a potential therapeutic role of ginsenoside Ro to treat obesity and its associated metabolic diseases.

SIGNIFICANCE STATEMENT

Obesity is dramatically increasing worldwide, and it contributes to multiple metabolic diseases. G protein–coupled bile acid receptor 5 (TGR5) is a potential therapeutic target for the treatment of metabolic disorders. Ginsenoside Ro, as an oleanane-type ginsenoside, ameliorates obesity and insulin resistance, promotes glucagon-like peptide 1 secretion, and increases energy expenditure via activating TGR5. Ginsenoside Ro could be a potential leading compound for treating obesity and its associated metabolic diseases.

Introduction

With the accelerated pace of life, high-calorie diets have been becoming more popular and result in a high prevalence of patients that are overweight or obese (Netto Candido et al., 2018). The incidence of obesity has been greatly increased worldwide during recent years (Bluher, 2019). For example, around 19.5% of adults were obese in 2015 in the Organisation for Economic Cooperation and Development (OECD) countries, and the prevalence of obesity could be increased up to 47% by 2030 as suggested by a forecast (2017). Obesity is a major risk factor in multiple metabolic diseases including diabetes, fatty liver, cardiovascular diseases, stroke, and cancer. A recent study reported that people with obesity have increased risk of severe COVID-19 (Stefan et al., 2020). Behavior and genetics have been recognized as the main factors influencing obesity...
Currently, bariatric surgery is the most effective treatment of morbid obesity and other complications (Arterburn et al., 2020). However, numerous studies were also conducted to identify therapeutic targets for treating obesity. G protein–coupled bile acid receptor 1 (TGR5) (also known as GPBAR1) is a bile acid membrane receptor and is expressed in many tissues including intestines, brown adipose tissue, spleen, and macrophages (Maruyama et al., 2002). Accumulating evidence suggests that the activation of TGR5 can affect glucose and lipid metabolism in a direct or indirect way. TGR5 was shown to play a crucial role in the vertical sleeve gastrectomy–mediated metabolic improvement [6]. In brown adipose tissue (BAT), TGR5 promotes intracellular thyroid hormone activation and induces energy expenditure via regulating the expression of mitochondrial uncoupling protein (UCP) 1 and the activation of CAMP-dependent iodothyronine deiodinase 2 (Watanabe et al., 2006). Besides, a TGR5 selective agonist, INT-777, was found to promote the release of glucagon-like peptide 1 (GLP-1) secreted by enteroendocrine L cells of the intestinal epithelium. INT-777 also improves insulin sensitivity and lipid-loading in obese mice (Kumar et al., 2016). In a mouse model, TGR5 was found to be activated by vertical sleeve gastrectomy to sustain weight loss and improve fatty liver and insulin resistance (Ding et al., 2016). Meanwhile, bariatric surgery reveals cholic acid 7-sulfate, a gut-restricted TGR5 agonist, to alleviate obesity and type 2 diabetes (Chaudhari et al., 2021). Therefore, it’s well recognized that TGR5 can be therapeutically targeted for obesity and its associated morbidities by developing its ligands.

In our previous study, we demonstrated that the activation of TGR5 could be increased by ginsenoside Ro in vitro. As the major active ingredient in ginseng, ginsenosides is a triterpenoid saponin mainly extracted from Panax ginseng C. A. Mey. Ginsenoside has diverse pharmacological activities such as antitumor, anti-inflammatory, and antioxidant (Kiefer and Pantuso, 2003; Wang et al., 2016; Gao et al., 2017b). Previous studies have revealed that ginseng extract and ginsenoside can improve obesity, metabolic syndrome, and type 2 diabetes (Chen et al., 2019). Dammarane-type saponins of ginsenosides such as Rb1, Rg1, and Rg3 show the effects of regulating glucose and lipid metabolism and promoting energy expenditure (Bai et al., 2018; Zhou et al., 2019; Lee et al., 2020). A recent study showed that Changbai Mountain Ginseng extract, whose major compound is ginsenoside Ro, improved exercise performance and energy utilization in mice (Ma et al., 2017). Moreover, the extract affects glucose and triacylglycerol levels and increases the mass of BAT in mice (Ma et al., 2017). As a typical oleanane-type saponin, the antitumor and anti-inflammatory effects (Kim et al., 2015b; Zheng et al., 2019) as well as the protection against platelet aggregation of ginsenoside Ro have been reported (Kwon, 2019). However, its specific impact on obesity and metabolic diseases remains unknown. In this study, we explored the effects and underlying mechanisms of ginsenoside Ro on the glucose and lipid metabolism in high-fat diet–induced obese (DIO) mice.

**Materials and Methods**

**Drug and Reagents**

Ginsenoside Ro (purity: ≥ 98%) was purchased from Chengdu Bio-purity Phytochemicals Ltd, Sichuan, China. High-fat diet (HFD) (D12492i, contains 60% fat, 20% protein, and 20% carbohydrate) and control diet (also named low-fat diet, D12450J, contains 10% fat, 20% protein, and 70% carbohydrate) were purchased from Research Diets, Inc. (New Brunswick, NJ).

**Animal Procedures**

Male C57BL/6 wild-type mice between 6 and 8 weeks old were purchased from the SLAC Laboratory (Shanghai, China). Tgr55−/− mice, Tgr5−/− mice, and Villin-Cre mice in C57BL/6 background were constructed by the facility of Shanghai Biomed Model Organism Science and Technology Development. To generate intestine-specific Tgr5−null mice (Tgr55AN), homozygous Tgr5−/− mice were crossed with mice harboring the cre recombinase under the control of the villin promoter [Villin-Cre mice, from Shanghai Biomed Model Organism Science and Technology Development Co., Ltd. (stock number Jax-021504-B6.Cg-Tg [Vill1-cre] 1000GumJ), Shanghai, China]. All mice were housed under a 12-hour light/dark cycle at 18–22°C in the facility of Shanghai Biomed Model Organism Science and Technology Development. All wild-type mice, Tgr55−/−, and Tgr55AN mice were fed with a high-fat diet for 6 weeks (body weight of HFD-induced obese mouse was 35.675±2.275 g) and were randomly divided into HFD group and HFD supplemented with ginsenoside Ro powder group at a dose of 90mg/kg (HFD + Ro-H group) or 5mg/kg (HFD + Ro-L group) based on food intake (n = 8 per group). The doses were determined based on previous in vivo studies (Matsuda et al., 1990; Matsuda et al., 1991) of ginsenoside Ro and our pre-experiments, and the safety was examined (Supplemental Fig. 1). Simultaneously, the control group was fed on a low-fat diet (lean group, n = 8). After oral administration with ginsenoside Ro for 8 weeks, mice from each group were administered oral glucose tolerance tests (OGTTs), intraperitoneal insulin tolerance tests (IPITTs), and GLP-1 secretion assay. Age- and body weight–matched animals were used. After mice were euthanized by CO2, serum, liver tissues, white adipose tissue (WAT), and BAT were collected and snap frozen in liquid nitrogen for RNA extracts or biochemistry studies. All studies were performed according to protocols approved by Shanghai University of Traditional Chinese Medicine’s Animal Care and Use Committee.

**OGTT, IPITT, and Homeostasis Model Assessment of Insulin Resistance**

OGTT: mice were fasted for 14 hours prior to OGTT, and blood was sampled from the tail vein before and 15, 30, 60, and 120 minutes after an oral administration of D-glucose at 2.0 g/kg body weight. Blood glucose level (mmol/l) was measured by a blood glucose meter. IPITT: mice were fasted for 4 hours prior to IPITT, and blood was collected from the tail vein before and 15, 30, 60, and 120 minutes after an intraperitoneal injection of insulin at 0.75 U/kg body weight. Blood glucose levels (mmol/l) were measured using a blood glucose meter. The index of the homeostasis model assessment of insulin resistance (HOMA-IR) was calculated according to the following formula: (fasted insulin (μIU/mL) × fasted glucose (mM))/22.5.

**Serum Biochemistry Measurements**

Serum was gathered from blood collected through cardiac puncture. Total serum cholesterol and triglyceride levels were examined by an automatic blood biochemical instrument (2700; Olympus, Tokyo, Japan).

**GLP-1 Release Assay**

After 14 hours of fasting, mice were orally administered Sitagliptin at 0.3 mg/g body weight 60 minutes prior to oral treatment with D-glucose at 2.0 g/kg body weight. Plasma was collected from the medial canthus vein of the eye before and 15 and 30 minutes after D-glucose administration. Plasma active GLP-1 was measured by using Millipore GLP-1 active ELISA kit (Millipore).
Indirect Calorimetry

Energy expenditure was evaluated by an indirect calorimetry system of 16 identical, independent metabolic cages equipped for the continuous monitoring of ambulatory activity and ad libitum access to HFD and water (Columbus Instruments Comprehensive Laboratory Animal Monitoring System). Temperature and humidity levels were regulated tightly. O₂ and CO₂ levels were monitored constantly. CO₂ consumption and O₂ production of each mouse were measured at 10-minute intervals for 1 minute. Energy expenditure is calculated utilizing the Weir equation: Kcal/h = 60 * (0.003941 * VO₂ + 0.001106 * VCO₂).

Serum Bile Acid Profile Analysis

Chromatographic conditions: chromatographic column used Cortec UPLC C18 1.6 μm, 2.1 × 100 mm column. Mobile phase A was a: 0.01% formal acid, b: acetonitrile. Mobile phase A was a gradient: 0.1 minute: 5% B, 1.0 minute: 5% B, 6.0 minutes: 25% B, 13.0 minutes: 30% B, 20 minutes: 40% B, 24.0 minutes: 75% B, 26.0 minutes: 75% B, 26.5 minutes: 95% B, 27.0 minutes: 95% B. Flow rate was 0.3 ml/minute. Injection volume was 5 μL. Ion source temperature was 600°C, and collision gas at medium.

Mass spectrometry conditions: electrospray ionization source, ion source temperature was 120°C, capillary voltage 2.6 kV, taper hole voltage 55 V, desolvent gas (N₂) flow rate was 600 l/h, taper hole gas (N₂) flow rate was 50 l/h; desolvent temperature was 350°C; negative ion monitoring mode; scanning mode for full scan, scanning range (m/z) 50–1000. 1500.

Samples and standards solution preparation: For analyzing bile acid in liver, we took the appropriate amount of liver samples homogenized with ultrapure water, then added methanol and centrifuged. We kept the supernatant, blow-dried with N₂, and added methanol for re-dissolution. For serum solution preparation, we took the appropriate amount of serum and mixed with interior label solution and methanol for dissolution. For standard solu-

Cyclic AMP Release Assay

HEK293T cell line was transiently transfected with mTGR5 and pCRE-luc plasmids and treated with LCA and ginsenoside Ro for 30 minutes in Krebs’ ringer buffer without serum added with 100 mM RO 20-1274 and 500 mM 3-Isobutyl-1-methylxanthine (IBMX) (Sigma, St. Louis, MO). According to the instructions of cAMP-Glo kit (Promega), the cell lysates were used to determine the content of cAMP.

Real-Time PCR

At 8 weeks after treatment, the mouse liver was obtained and subjected to total RNA extraction using Trizol reagent. Relative levels of amplifiers were identified utilizing SYBR Green quantitative real-time PCR (qPCR) mix (Applied Biosystems) on Applied Biosystems Real-Time PCR System (Table 1).

Histologic Analysis of Liver and Adipose Tissue

H&E staining: liver and adipose tissues were fixed in 4% paraformaldehyde, embedded in paraffin, and sectioned at 5 mm onto poly-l-lysine–coated slides. Tissue sections were stained in Mayer’s hematoxylin. Oil red O staining: liver tissue was embedded in Tissue-Tek OCT cryostat molds and frozen at −80°C. Liver tissue was used to generate 10-mm thick sections in a cryostat, and tissue sections were stained in 0.5% oil red O and then counterstained with Mayer’s hematoxylin. Images were quantified by image J 1.51k.

Statistics

All data were expressed as mean ± S.E.M. All statistical analyses were carried out by one-way analysis of variance with Dunn’s posttest Graphpad Prism 7.0 (7.0 version; GraphPad, La Jolla, CA). P value < 0.05 was considered statistically significant.

Results

Ginsenoside Ro Improves HFD-Induced Obesity. It has been demonstrated that the activation of TGR5 inhibits HFD-induced obesity. Given our previous finding that ginsenoside Ro activates TGR5 in vitro, we hypothesized that ginsenoside Ro might have an antiobesity effect. The C57BL/6 wild-type mice were randomly divided into lean group (low-fat diet), HFD group (60 Kcal% high-fat diet), HFD + Ro-L group (60 Kcal% high-fat diet with 45 mg/kg ginsenoside Ro), and HFD + Ro-H group (60 Kcal% high-fat diet with 90 mg/kg ginsenoside Ro). The HFD group was compared with lean group, and both HFD + Ro-L and HFD + Ro-H group were compared with HFD group.

First, we tested the activation effect of ginsenoside Ro on TGR5 in vitro by dual-luciferase reporter assay and cAMP release assay. The results showed that treatment with ginsenoside Ro (20 μM), which is a typical oleanane-type sapoin (Fig. 1A), increased luminescence signal and the cAMP release (Fig. 1, B and C).

<table>
<thead>
<tr>
<th>Gene</th>
<th>Sense Primer</th>
<th>Antisense Primer</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gapdh</td>
<td>GGCCGGAATGCGAGAAGCTGT</td>
<td>ACATACTGAGACGGGCTCA</td>
</tr>
<tr>
<td>Tgr5</td>
<td>AGGTTGCTCATCCGAGTGCTT</td>
<td>CATTGGCTACTGTTGAGG</td>
</tr>
<tr>
<td>Dio2</td>
<td>CGTCTCAATCCCTGAAATCA</td>
<td>AAGTCAAGAAGTGGCATTT</td>
</tr>
<tr>
<td>Ucp1</td>
<td>GAAACCTGCTCCCTCCTC</td>
<td>ACCCTCAGCCCTCCTGTA</td>
</tr>
<tr>
<td>Ucp3</td>
<td>ACCCGATACTACGAGACCTCC</td>
<td>CACAATCTTTGTTGAGGGCCG</td>
</tr>
<tr>
<td>Cpt1β</td>
<td>AAGAGACAGACTGTCTACAG</td>
<td>TAGAGCCAGACCTTGAAGA</td>
</tr>
</tbody>
</table>
We then recorded the body weight and food intake of DIO mice and examined the histopathology of BAT and WAT by H&E staining. The results showed that high-dose ginsenoside Ro significantly reduced the body weight and fat mass (Fig. 1, D–G) of the mice without affecting the food intake (Fig. 1H) compared with the HFD group. However, low-dose ginsenoside Ro had no effect on either body weight or fat mass. In addition, histologic examination showed a substantial decrease of adipocyte volume in BAT and inguinal white adipose tissue (iWAT) (Fig. 1, I and J) in both high- and low-dose ginsenoside Ro–treated mice compared with those in the HFD group.

**Ginsenoside Ro Improves the Glucose Metabolism of DIO Mice.** To examine the effect of ginsenoside Ro on glucose metabolism, glucose tolerance, insulin secretion, insulin tolerance, and fasting serum glucose levels were tested. High-dose ginsenoside Ro significantly improved the glucose tolerance and insulin tolerance (Fig. 2, A–C) in DIO mice. Both high- and low-dose ginsenoside Ro had no effect on fasting insulin (Fig. 2 E). Besides, high-dose but not the low-dose ginsenoside Ro markedly descended the fasting serum glucose and HOMA-IR index (Fig. 2, D and F).
Ginsenoside Ro Improves Lipid Metabolism of DIO Mice. To explore the effect of ginsenoside Ro on lipid metabolism, the lipid accumulation in the liver was visualized by oil red O staining and H&E staining, and serum triglycerides (TG), total cholesterol (TC), high-density lipoprotein cholesterol (HDL-c), and low-density lipoprotein cholesterol (LDL-c) were tested. Meanwhile, the contents of TC and TG in the liver were also measured. Lipid accumulation was observed to be reduced in the liver of both high- and low-dose treated DIO mice (Fig. 3, A, B, and I). The contents of TC and TG in serum and liver were significantly reduced by the treatment of high-dose ginsenoside Ro (Fig. 3, C–F). The serum HDL-c level was also markedly raised in response to the treatment of high-dose ginsenoside Ro (Fig. 3G). However, no significant difference in the effects on serum TC, TG, or LDL-c was seen between the low-dose ginsenoside Ro–treated group and the HFD group. Unexpectedly, low-dose ginsenoside Ro treatment significantly decreased liver TC but increased serum HDL-c levels (Fig. 3, E and G) compared with the HFD group. High-dose ginsenoside Ro significantly improved lipid metabolism, and low-dose ginsenoside Ro also had partial improvement effect.

Ginsenoside Ro Alters Bile Acids Profile of DIO Mice. Bile acids primarily serving as emulsifiers to facilitate fat absorption have been considered crucial signaling molecules involved in glucose and lipid metabolism. To explore the underlying mechanisms of ginsenoside Ro on regulating glucose and lipid metabolism, we analyzed the bile acids profile with ginsenoside Ro treatment. Total bile acids in serum and liver were both upregulated in high-dose ginsenoside Ro–treated mice (Fig. 4, A and B). It has been demonstrated that farnesoid X receptor (FXR) is an important bile acid nuclear receptor involved in the regulation of bile acid circulation and synthesis, and the inhibition of liver FXR promotes the synthesis and secretion of bile acids. We thus measured the hepatic mRNA levels of FXR, Shp, Cyp7a1, and Cyp7b1 to test
whether ginsenoside Ro treatment disturbs the synthesis pathway of bile acids. As showed in Fig. 4C, ginsenoside Ro treatment suppressed mRNA expression of FXR and Shp in DIO mice, but the levels of Cyp7a1 and Cyp7b1 were promoted, which indicates that ginsenoside Ro regulates bile acid synthesis in the liver by modulating the transcription of genes. Besides, we also found that, compared with the HFD group, the concentration of serum glycine–conjugated bile acids GUDCA, GCA, and GCDCA increased in the ginsenoside Ro–treated group (Fig. 4D). However, ginsenoside Ro treatment did not affect the concentration of hepatic glycine–conjugated bile acids (Fig. 4E). Furthermore, taurine-conjugated bile acids levels in serum and liver were significantly raised after ginsenoside Ro treatment (Fig. 4F and G). The concentration of free bile acids including serum β-MCA, serum UDCA (Fig. 4H), and hepatic β-MCA were also increased significantly after ginsenoside Ro treatment (Fig. 4I).
Ginsenoside Ro Promotes GLP-1 Secretion via Activating Intestinal TGR5. We further tested the mRNA level of intestinal Tgr5 in DIO mice and found that mRNA expression is enhanced by ginsenoside Ro (Fig. 5A). Intestinal TGR5 activation was reported to promote GLP-1 secretion, which is involved in the regulation of glucose and lipid metabolism. We subsequently explored whether GLP-1 is regulated by ginsenoside Ro using intestinal Tgr5 knockout mice. Our results revealed that ginsenoside Ro treatment significantly increased GLP-1 level in WT mice but not in Tgr5⁻/⁻ mice (Fig. 5, B and D). In addition, fasting serum glucose levels, glucose tolerance, and insulin tolerance were not increased in Tgr5⁻/⁻ mice either (Fig. 5, C, E, and F).

The Metabolic Benefits of Ginsenoside Ro Treatment Are Absent in Tgr5⁻/⁻ Mice. We then investigated whether ginsenoside Ro could affect the phenotype of Tgr5⁻/⁻ mice. First of all, we measured and found that the body weights of Tgr5⁻/⁻ mice were not affected by ginsenoside Ro.
treatment (Fig. 6A), but the serum TC level still decreased in Tgr5−/− mice after ginsenoside Ro treatment (Fig. 6B). Furthermore, the effect of ginsenoside Ro on serum TG was restrained by Tgr5 knockout (Fig. 6C), and the enhancement of mRNA expressions of Dio2, Ucp1, Ucp3, and Cpt1b observed in BAT of WT mice was not shown in Tgr5−/− mice (Fig. 6D). Consistently, the improvements of ginsenoside Ro-mediated lipid accumulation in BAT and iWAT in WT mice were abolished in Tgr5−/− mice (Fig. 6E).

Ginsenoside Ro Promotes Energy Expenditure via TGR5. Finally, we evaluated the energy expenditure by an indirect calorimetry system of 16 identical, independent metabolic cages. As expected, ginsenoside Ro promoted oxygen consumption, carbon dioxide output, and energy consumption during the day and night in WT mice (Fig. 6, F, H, and J). However, the energy metabolism was not affected by ginsenoside Ro in Tgr5−/− mice (Fig. 6, G, I, and K).

Discussion
As a traditional Chinese medicine, Panax ginseng has long been used against metabolic diseases such as type 2 diabetes and obesity (Bai et al., 2018). Ginsenosides, a main active ingredient in ginseng, have been reported to modulate glycolipid metabolism including gluconeogenesis reduction, glucose transport, and lipid regulation (Bai et al., 2018). The regulation of ginsenosides on energy metabolism has also been illustrated (Li and Ji, 2018). Based on the structure differentiation of the sapogenins, ginsenosides are classified into two main types including oleanane-type saponin and dammarane-type saponin (Supplemental Fig. 2). Compared with dammarane-type saponins, the pharmacological study of oleanane-type ginsenosides is limited. Ginsenoside Rb1 has protective effects on obesity, hyperglycemia, and diabetes through regulating mitochondrial energy metabolism and improving

Fig. 5. Ginsenoside Ro promotes GLP-1 secretion via TGR5. (A) Relative mRNA expression of Tgr5 in the intestine of WT mice. (B) Active GLP-1 release between 0 and 30 minutes was measured after administration with 2g/kg D-glucose in WT mice. (C) Fasting serum glucose in intestinal Tgr5 knockout mice after 8 weeks of ginsenoside Ro treatment. (D) Active GLP-1 release in Tgr5−/− mice. (E and F) The OGTT and IPITT of Tgr5−/− mice were measured at 6 weeks and 7 weeks after ginsenoside Ro treatment, respectively. The area under the curve (AUC) measurements of OGTT or IPITT between 0 and 120 minutes were calculated. Data are represented as mean ± S.E.M. *P < 0.05 versus lean, **P < 0.05 versus HFD controls by one-way analysis of variance with Graphpad Prism 7.0.
Fig. 6. Effects of ginsenoside Ro on Tgr5−/− mice and energy expenditure. Tgr5−/− mice were fed with a regular high-fat diet or the high-fat diet containing ginsenoside Ro for 8 weeks. (A) Final body weight of Tgr5−/− mice. (B and C) Serum cholesterol and triglycerides of Tgr5−/− mice. (D) Relative mRNA expression of Dio2, Ucp1, Ucp3, and Cpt1β. (E) H&E staining of BAT and WAT. Magnification bar: 100 μM. (F-G) Oxygen production volume of the period over 24 hours. (H and I) Carbon dioxide consumption volume of the period over 24 hours. (J and K) Energy expenditure of the period over 24 hours. Data are represented as mean ± S.E.M. ####P < 0.001 versus lean, *P < 0.05, **P < 0.01 versus HFD controls by one-way analysis of variance with Graphpad Prism 7.0.
insulin resistance (Zhou et al., 2019). Ginsenoside Rg1 reduced gluconeogenesis through increased adenosine 5'-monophosphate-activated protein kinase (AMPK) expression and decreased FOXO1 activity (Liu et al., 2017). It also improved lipid metabolism, antiapoptotic, and anti-inflammatory properties by inhibiting the JNK pathway (Tian et al., 2017). Ginsenoside Rg1 and Rg3 can reduce intestinal glucose uptake by inhibiting SGLT1 (Wang et al., 2015; Gao et al., 2017a). Rg3 affected lipid regulating dependent on the STAT5-PPARγ pathway (Lee et al., 2017). However, ginsenoside Ro only has been reported to increase the secretion of lipase activity in 3T3-L1 cells (Masuno et al., 1996). In the present work, we showed that ginsenoside Ro reduced body weight and lipid accumulation of peripheral adipose tissue in obese mice without affecting the food intake. Meanwhile, ginsenoside Ro improved insulin resistance and fatty liver. Our data suggest that ginsenoside Ro plays a similar role as many other dammarane-type saponins in contributing to the antiobesity effects of ginseng by ameliorating glucose and lipid metabolism. Furthermore, ginsenoside Ro promoted GLP-1 secretion and energy expenditure in HFD-induced obese mice, indicating that the antiobesity effects of ginsenoside Ro were likely due to promoting GLP-1 release and energy consumption. However, some ginsenoside Ro treatment–induced metabolic improvements were absent in Tgr5<sup>−/−</sup> mice or Tgr5<sup>5/5N</sup> mice, indicating that ginsenoside Ro improves metabolism through TGR5 activation.

TGR5 is an important bile acid receptor involved in the regulation of glucose, lipid, and energy metabolism. There is growing evidence that activated TGR5 can be a promising target for treating obesity, type 2 diabetes, and nonalcoholic steatohepatitis (Duboc et al., 2014). TGR5 transmits its signal by rising intracellular concentrations of cAMP, causing rapid phosphorylation of downstream kinases (van Nierop et al., 2017). Bile acids are internal activators of TGR5, but complex enterohepatic bile acid cycling limits the exposure of some of these target tissues to the receptor ligand. To investigate how to use TGR5 as a therapeutic target, a number of TGR5 abiotic agonists including selective TGR5 agonists INT-777, MN6, intestinal TGR5 agonist (Chen et al., 2018), and a newly reported TGR5 agonist RDX8940 that function in metabolic diseases have been identified and studied recently (Finn et al., 2019). On the other hand, several natural products, especially triterpenoids such asoleanolic acid and betulinic acid, were also proven to affect the activity of TGR5 (Lo et al., 2016; Liu et al., 2019). In the present study, we demonstrated that ginsenoside Ro activated TGR5 in vitro and increased the mRNA expression of Tgr5 in vivo, suggesting the effects of ginsenoside Ro on obesity and glycolipid metabolism through TGR5. We next clarified whether the endogenous TGR5 ligands or bile acids were altered because of the lower bioavailability of ginsenoside Ro (Qi et al., 2013). We examined the content and composition of bile acids in ginsenoside Ro–treated mice, and the results showed that both of them were affected. Ginsenoside Ro raised the concentration of various bile acids, such as chenodeoxycholic acid (CDCA),ursodeoxycholic acid (UDCA), and its conjugated bile acids. Deoxycholic acid (DCA), CDCA, UDCA, and its conjugated bile acids are reported to be TGR5 agonists (Fiorucci and Distritti, 2019). Together, these results suggested that the regulation through bile acids is likely one of the potential mechanisms of how ginsenoside Ro improves obesity.

TGR5 contributing metabolic improvements are mainly attributable to GLP-1 secretion in ileum and energy expenditure in adipose tissue. GLP-1 plays a critical role in maintaining blood glucose homeostasis, and both secretion and sensitivity of insulin can be promoted by GLP-1. Simultaneously, GLP-1 inhibits the release of glucagon (Nadkarni et al., 2014). In a type 2 diabetes mellitus (T2DM) model, ginsenoside Rg3 increased GLP-1 secretion and decreased hyperglycemia through a sweet taste receptor–mediated signal transduction pathway (Kim et al., 2015a). Ginseng total saponins and ginsenoside Rb1 exhibit antidiabetic effects by promoting GLP-1 secretion in cultured NCI-H716 cells (Liu et al., 2014). Compound K, a major metabolite of ginsenosides, induced GLP-1 secretion in NCI-H716 cells via TGR5 activation (Kim et al., 2014). The promoting effect on GLP-1 secretion is one of the major mechanisms of TGR5 treatment against obesity. The selective TGR5 agonist improves obesity, glucose tolerance, insulin, GLP-1 secretion, and insulin sensitivity in both liver and muscle of TGR5-Tg mice with a high-fat diet, but not in TGR5 knockout (Tgr5<sup>−/−</sup>) mice. TGR5 overexpression or selective TGR5 agonist treatment increases energy expenditure and reduces hepatic steatosis and obesity. In the present study, ginsenoside Ro increased GLP-1 secretion in WT mice but not the intestinal Tgr5 knockout mice, suggesting that ginsenoside Ro enhanced GLP-1 secretion via TGR5 activation.

Besides, activation of TGR5 increases energy expenditure by regulating the activity of type 2 iodothyronine deiodinase and the subsequent activation of thyroid hormone in BAT and muscles. In the present study, ginsenoside Ro increased energy expenditure in the WT but not the Tgr5 knockout mice. Moreover, ginsenoside Ro had no effects on the body weight, serum TG level, and lipid in the Tgr5 knockout mice, suggesting that ginsenoside Ro ameliorated obesity and improved energy expenditure by activating TGR5.

**Conclusion**

Ginsenoside Ro treatment ameliorates obesity and insulin resistance in diet-induced obese mice. The antioesity activity of ginsenoside Ro is mediated through TGR5. The findings of this study provide new insights into the mechanisms on improving metabolic disease by ginsenoside Ro or herbs containing ginsenoside Ro and indicate ginsenoside Ro a potential leading compound for treating obesity and its associated metabolic disease.

**Authorship Contributions**

**Participated in research design:** Wang, Ding, Yang.
**Conducted experiments:** Jiang, Li, Zhuang, Yu, Sun, Ju.
**Performed data analysis:** Jiang, Li.
**Wrote or contributed to the writing of the manuscript:** Jiang, Li, Ding.

**References**


Ginsenoside Ro Ameliorates Obesity via the TGR5 Pathway


Address correspondence to: Dr. Li-li Ding, Institute of Traditional Chinese Materica Medica, Shanghai University of Traditional Chinese Medicine, Cai Lun Rd. 1260, Zhangjiang, Shanghai 201203, China. E-mail address: naili320196159@163.com; or Dr. Li Yang, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China. E-mail: yangli7951@hotmail.com; yil7@shutcmed.edu.cn
**Ginsenoside Ro ameliorates high fat diet-induced obesity and insulin resistance in mice via activation of the TGR5 pathway**

Lin-shan Jiang#, Wei Li#, Tong-xi Zhuang, Jie-jing Yu, Shuai Sun, Zheng-cai Ju, Zheng-tao Wang, Li-li Ding*, Li Yang*

*Journal of pharmacology and experimental therapeutics*

**Supplementary Figure 1. Effects of Ginsenoside Ro on hepatic and renal functions.** After the treatment of ginsenoside Ro for 6 weeks, serum ALT, AST, BUN and CREA levels of Lean, HFD group, HFD supplemented with ginsenoside Ro high dose group (90mg/kg, HFD+Ro-H) and ginsenoside Ro low dose group (45mg/kg, HFD+Ro-L) mice were measured. (A) Serum ALT level. (B) Serum AST level. (C) Serum BUN level. (B) Serum CREA level. Data are represented as mean ± SEM.

**Supplementary Figure 2. Chemical structures of oleanane and dammarane type saponins.** Ginsenosides are classified into two main types: oleanane-type saponin and dammarane-type saponin, based on the structure differentiation of the sapogenins. (A) Structures of oleanane aglycone and representative chikusetsu saponin IVa. (B) Structures of dammarane aglycone and representative ginsenoside Rg1, Rb1, Rg3.