Special Section on Drug Delivery Technologies—Minireview

Ocular Drug Delivery: Present Innovations and Future Challenges

Vrinda Gote, Sadia Sikder, Jeff Sicotte, and Dhananjay Pal

ABSTRACT

Ocular drug delivery has always been a challenge for ophthalmologists and drug-delivery scientists due to the presence of various anatomic and physiologic barriers. Inimitable static and dynamic ocular barriers not only exclude the entry of xenobiotics but also discourage the active absorption of therapeutic agents. Designing an ideal delivery scheme should include enhanced drug bioavailability and controlled release of drug at the site of action, which can overcome various ocular barriers. Conventional ophthalmic medications include the use of topical eye drops and intravitreal injections of anti-vascular endothelial growth factor agent for treatment of anterior and posterior segment disorders, respectively. Current inventions for anterior ocular segment disorders such as punctum plugs, ocular implants, drug-eluting contact lenses, and ocular iontophoresis represent state-of-the-art inventions for sustained and controlled drug release. Parallel efforts for ocular drug delivery technologies for back of the eye disorders have resulted in the approval of various intravitreal implants. Novel drug-delivery technologies, including nanoparticles, nanomicelles, dendrimer, micro-needles, liposomes, and nanowafers, are increasingly studied for anterior and posterior disorders. To achieve patient compliance for back of the eye disorders, novel approaches for noninvasive delivery of potent therapeutic agents are on the rise. In this review article, we discuss past successes, present inventions, and future challenges in ocular drug-delivery technologies. This expert opinion also discusses the future challenges for ocular drug-delivery systems and the clinical translatable potential of nanotechnology from benchtop to bedside.

Introduction

In the past two decades, the arena of ocular drug-delivery technologies has dynamically advanced and resulted in newer therapeutic interventions for chronic ocular disorders. The primary objectives of any ocular drug-delivery system are to maintain therapeutic drug concentrations at the target site, reduce dosage frequency, and overcome various dynamic and static ocular barriers. Most importantly, the drug-delivery system should cause no adverse ocular reactions and aim to achieve enhanced drug bioavailability. Ocular pathologic disorders are generally described as anterior segment and posterior segment disorders. Clinicians treat anterior segment disorders such as dry eye disease, cataract, and allergic conjunctivitis by topical eye drops. The major disadvantage of topically applied ophthalmic formulations is relatively low ocular bioavailability. This can be attributed to high tear-fluid turnover rates and high nasolacrimal drainage. Novel ocular drug-delivery systems include nanoparticles, nanomicelles, drug-eluting contact lenses, ocular inserts, and ocular devices that allow enhanced peroneal residence and enhance the bioavailability of the therapeutic agents (Achouri et al., 2013; Fangeiro et al., 2016).

Ocular pathologic conditions involving the posterior segment generally result in vision loss due to damage to the retina. Hyperglycemia for a prolonged period of time can cause damage to the retinal endothelial cells, causing back of the eye disorders such as diabetic retinopathy (DR), diabetic macular edema (DME), and retinal vein occlusion (RVO). High oxidative stress, endoplasmic reticulum stress, and aging can damage the retinal pigmented epithelial cells (RPE) and Bruch’s membrane in the macular region, leading to the death of the photoreceptors. Such pathologic conditions can cause retinal degenerative disorders such as age-related macular degeneration (AMD) (Yasukawa et al., 2004; Janoria et al., 2007). Retinal and choroidal neovascularization (CNV), evident in back of the eye disorders, is primarily due to overexpression of vascular endothelial growth factor (VEGF) receptor. Before the invention of anti-VEGF agents, the gold standard treatment of these disorders was the...
application of laser photocoagulation to lower overall oxygen demand of the retina. This therapy allowed suppression of CNV and retinal neovascularization. Since then, clinicians have introduced a plethora of anti-VEGF agents in the market, including pegaptanib, bevacizumab (off-label), ranibizumab, and aflibercept, for the treatment of back of the eye disorders with neovascularization. Clinicians administer these agents as intravitreal injections, which has drawbacks such as retinal hemorrhage and retinal detachment. Moreover, intravitreal injections lack patient compliance. Novel ocular drug-delivery technologies such as nanoformulations, implants, and other ocular devices allow enhanced drug residence time at the target tissue along with improvements in pharmacological response (Peyman et al., 2009).

In this article, we present a comprehensive and detailed review of past successes, current inventions, and future challenges in anterior and posterior ocular drug-delivery systems. Developments in novel drug-delivery technologies can ultimately improve pharmacological action of drugs at the target tissue by elevating the concentrations and ocular bioavailability of the required therapeutic agent.

**Fig. 1. Ocular anatomic barriers and routes of drug administration.** Ocular barriers to topical administration (iv) of therapeutic agents to the anterior surface of the eye and to the posterior segment are illustrated. These include (A) tear film barrier; (B) corneal barrier; (C) vitreous barrier; (D) blood–retinal barrier and (E) blood–aqueous barrier. Various methods for drug delivery to the eye include; (I) intravitreal injection, (II) subconjunctival injection, (III) subretinal injection and (IV) topical administration. Topical administration of eye drops is one of the non-invasive route of administration and has minimum side effects. Intravitreal injections on the other hand are invasive, can cause retinal damage but can easily bypass all ocular barriers. While subconjunctival and subretinal injections can bypass some of the ocular barriers and are less invasive. (Alqawlaq et al., 2012).
10-Å molecules, with a higher drug-distribution coefficient limiting the barrier for hydrophobic drugs. Therefore, drug absorption requires overcoming corneal epithelia efficiently. Decrease in transcorneal diffusion of drug through the aqueous humor and expression of efflux transporters on the plasma membrane of corneal cells are major restrictions for drug delivery to the targeted ocular tissues. The use of prodrugs, permeation enhancers, and recent use of nanomicelles can enhance permeability of the drug through the corneal barriers (Cholkar et al., 2012; Huang et al., 2018).

While in the posterior segment of the eye, the scleral, choroidal, and retinal epithelial and the blood-retinal barrier account for limiting ocular drug bioavailability. The sclera provides higher trans-scleral permeability than the cornea for hydrophilic compounds diffusing through the collagen network. Permeation through the sclera is largely dependent on molecular weight, molecular radius, and charge. Macromolecules exhibit lower penetration through scleral pores than small molecules. This is the reason why macromolecules, including anti-VEGF agents, exhibit low diffusion through the sclera and are administered by intravitreal injections (Huang et al., 2018). The choroid is a vascular-natured dynamic barrier, which impedes drug delivery by the trans-scleral pathway (Tsai et al., 2018). The retina is a significant limiting factor for diffusion of molecules with a larger radius and a molecular mass greater than 76 kDa (Jackson et al., 2003). The inner limiting membrane of the retina severely confines the passage of macromolecules over 150-kDa molecular mass (Mordenti et al., 1999; Jackson et al., 2003; Tao et al., 2007). Moreover, the inner limiting membrane progressively restricts molecules with a larger radius. Retinal pigmented epithelia and choriocapillaries collaboratively produce Bruch’s membrane. The thickness of Bruch’s membrane increases with age, inhibiting drug transport into tissues and draining hydrophobic drugs through systemic circulation (Cholkar et al., 2012). The blood-retinal barrier (BRB) comprises two subdivisions, an outer BRB and an inner BRB. Both the outer BRB and inner BRB are permeation barriers between the blood and the retina having tight junction proteins between the cells (Kamei et al., 1999; Achouri et al., 2013). The BRB also exhibits efflux transporters, which reduce bioavailability of several therapeutic agents (Mitra, 2009; Vadlapatla et al., 2014). The blood-aqueous barrier consists of an epithelial and an endothelial barrier. The permeability of drugs through the blood-aqueous barrier is determined by osmotic pressure and physical-chemical characteristics of drug molecules (Dubald et al., 2018). Ocular drug delivery presents a unique challenge due to its incredibly specialized tissue barriers that act as obstacles to therapies (Gaudana et al., 2010). Table 1 summarizes present routes of ocular therapy administration, and Fig. 1 details the anatomic makeup, indicating how each therapy travels to its active site.

### Past Successes in Ocular Drug-Delivery Technologies

#### Drug Delivery to the Anterior Segment of the Eye

Topical delivery of ophthalmic formulations is the most preferred route for the delivery of therapeutic agents to the anterior segment of the eye. Ocular formulations (solutions, suspensions, emulsions, gels, and ointments) are most commonly used to treat common anterior segment disorders such as dry eye diseases, allergic conjunctivitis, and glaucoma (Kaur and Kanwar, 2002). Topical ocular administration gains merit over systemic ocular administration. This is because topical administration is (i) relatively non-invasive, (ii) minimizes systemic side effects of the drug, (iii) avoids first pass metabolism, (iv) reduces drug dosage due to localized drug delivery (v) and increases patient compliance due to ease of topical administration. Factors limiting absorption of topically applied ophthalmic formulations are high tear turnover rate (1 μl/ml), loss of drug due to rapid blinking, reflex tear production, and limited absorption due to the tear-film barrier (Lee and Robinson, 1986; Schoenwald, 1990; Cholkar et al., 2013). To enhance the drug bioavailability, ophthalmic formulation requires a higher precorneal residence

<table>
<thead>
<tr>
<th>Route</th>
<th>Benefits</th>
<th>Obstacles</th>
<th>Diseases/Disorders Treated</th>
</tr>
</thead>
<tbody>
<tr>
<td>Topical</td>
<td>Patient compliance is high; self-administration and noninvasive nature</td>
<td>Corneal barrier difficult to penetrate; dilution and efflux via tears is high</td>
<td>Conjunctivitis, keratitis, uveitis, episcleritis, scleritis, blepharitis</td>
</tr>
<tr>
<td>Intravitreal</td>
<td>Direct delivery to retinal and vitreal structures; drug has high bioavailability</td>
<td>Patient compliance low; risk of retinal detachment, hemorrhage, development of endophthalmitis or cutaaneous</td>
<td>AMD, BRVO, CRVO, DME, CMV retinitis</td>
</tr>
<tr>
<td>Sub-Tenon</td>
<td>Relatively noninvasive, decreased risk of comorbidity compared with intravitreal delivery, maintains high vitreal drug levels</td>
<td>Retinal pigment epithelium is a barrier; subconjunctival hemorrhage, chemosis</td>
<td>DME, AMD, RVO, uveitis</td>
</tr>
<tr>
<td>Posterior juxtascleral</td>
<td>Advantageous for drug depository; avoids intraocular damage, and macula can sustain drug level for 6 mo</td>
<td>Retinal pigment epithelium barrier, and surgical procedure required</td>
<td>AMD, risk of endophthalmitis</td>
</tr>
<tr>
<td>Systemic/oral</td>
<td>Promotes patient compliance, noninvasive mode of delivery</td>
<td>Retinal and blood-aqueous barriers; low bioavailability leading to systemic toxicity</td>
<td>Scleritis, epikeratitis, CMV retinitis, posterior uveitis</td>
</tr>
<tr>
<td>Intracameral</td>
<td>Reduces systemic and corneal side effects vs. topical steroid use; high anterior chamber drug concentration</td>
<td>Toxic endothelial cell destruction syndrome and toxic anterior segment syndrome pose major risks to patients</td>
<td>Anesthesia, prevention of endophthalmitis, inflammation, pupil dilation</td>
</tr>
<tr>
<td>Subconjunctival</td>
<td>Anterior and posterior delivery method, ideal for depot formation</td>
<td>Chronoidal and conjunctival circulation of therapies increases toxicity</td>
<td>Glaucoma, CMV retinitis, AMD</td>
</tr>
<tr>
<td>Retrobulbar</td>
<td>Minimal IOP involvement, ideal for high local anesthetic administration</td>
<td>Respiratory arrest, retrobulbar hemorrhage, globe perforation</td>
<td>Anesthesia</td>
</tr>
</tbody>
</table>

BRVO, branched retinal vein occlusion; CMV retinitis, cytomegalovirus retinitis; CRVO, central retinal vein occlusion; IOP, intraocular pressure.
time and an enhanced drug penetration. Therefore, a drug-delivery system offering longer retention and a sustained release of the drug molecule to pass through these barriers is essential (Khar et al., 2010; Reimondex-Trottiño et al., 2015). Novel drug-delivery technologies utilizing cyclodextrins, prodrugs, and colloidal systems such as nanoparticles, liposomes, and nanomicelles have been studied extensively (Tirucherai and Mitra, 2003; Gunda et al., 2006; Vaka et al., 2008). Conventional eye drops in the form of solutions, suspensions, and emulsions have been used over a long period of time to treat anterior segment disorders. The following section describes topical ophthalmic formulations in detail.

**Ophthalmic Solutions.** Topical eye drop solutions are patient-compliant, noninvasive, immediate-acting drug formulations. Eye drop solutions are instilled in the cul-de-sac, which is followed by a rapid first-order absorption into the corneal and conjunctival tissues. An increase in drug permeation and drug bioavailability can be attained by modifying the drug properties or properties of the drug-delivery system.

**Modification of drug properties by utilizing prodrug strategy.** Drug molecules require appropriate lipophilic and hydrophilic properties to overcome the ocular tear barrier and to reach the corneal membrane. The prodrug approach modifies the physiochemical properties of the drug for better absorption of the drug by passive or active diffusion (Mandal et al., 2016a,b). Once the prodrug reaches the corneal tissue, cellular enzymes cleave it into the active drug. Dipivefrine (Propine, Allergan) is an ester prodrug of epinephrine, that demonstrates a 17-fold higher corneal permeation, resulting in 10 times higher epinephrine bioavailability in the corneal tissues than the unmodified drug. Cyclosporine-A is a lipophilic drug, which poses a challenge for formulation development and corneal permeation. UNILO888 ((1R,2R,E)-1-((25S,3S,14S,17S,20S,23S,26S,29S,32S)-5-ethyl-11,17,26,29-tetraisobutyl-14,32-diisopropyl-1,7,10,16,20,23,25,28,31-nonamethyl-3,6,9,12,15,18,21,24,27,30,33-undecaexo-1,4,7,10,13,16,19,22,25,28,31-undecazacyclotritracontan-2-yl)-2-methylhex-4-en-1-yl N-((1-acetoxyethoxy)carbonyl)-O-phosphono-L-seryl)-N-methylglycinate) is a hydrophilic prodrug of cyclosporine-A, demonstrating 25,000 times higher solubility than the parent drug at pH 7 (Lallemand et al., 2005). Another prodrug of cyclosporine-A (OPPH008) was characterized, and its efficacy in the treatment of dry eye disease was determined. OPPH008 achieved higher tissue concentrations as compared with a cyclosporine-A ophthalmic emulsion (Restasis, Allergan) in rabbit ocular conjunctivitis (Rodriguez-Aller et al., 2012). Prodrug strategy is also useful for reducing the dose of drugs with a narrow therapeutic index. Latanoprost is an ester prodrug of prostaglandin used to treat glaucoma. It has a higher bioavailability than the parent compound at lower doses (Kompella et al., 2010; Alm, 2014).

**Modification of formulation properties.**

**I. Cyclodextrins for solubilizing hydrophobic drugs.** Cyclodextrins are cyclic oligosaccharides arranged in a truncated cone-like structure. Cyclodextrins allow hydrophobic drugs to form complexes, enhancing drug solubility and bioavailability. Such complexation also improves corneal residence time and reduces local tissue inflammation (Achouri et al., 2013). Cyclodextrin complexation permits aqueous formulation of various drugs, including dexamethasone, chloramphenicol, and corticosteroids, for ocular disorders (Loftsson and Jarvinen, 1999; Loftsson and Stefansson, 2002; Sigurdsson et al., 2007). A study by Saari et al. (2006) concluded that 0.7% dexamethasone-cyclodextrin eye drops demonstrated significantly higher safety and efficacy as an anti-inflammatory medication for postcataract inflammation than 0.1% dexamethasone sodium phosphate eye drops.

**II. Viscosity and permeation enhancers.** Ophthalmic formulations traditionally use viscosity enhancers to improve precorneal residence time of the drug. Various viscosity enhancers, such as hydroxypropyl methylcellulose, polyalcohol, sodium carboxyl methylcellulose, and hydroxyethyl methylcellulose, improve drug retention time and absorption. Permeability of ophthalmic drugs is elevated by the addition of permeation enhancers (Achouri et al., 2013). Such agents temporarily adjust the corneal and conjunctival surface to facilitate rapid drug penetration. Ophthalmic preservatives such as benzalkonium chloride, surfactants such as polyethylene glycol, ethers, EDTA, chelating agents, and bile salts are a few examples of permeation enhancers that raise drug bioavailability (Burgalassi et al., 2001; van der Bijl et al., 2001, 2002; Hornof and Bernkop-Schnurch, 2002). Despite the various advantages offered by penetration enhancers, these agents can cause tissue irritation and damage the corneal and conjunctival tissues (Achouri et al., 2013).

**Suspensions.** Ocular suspensions are a dispersion of finely divided insoluble drug particles suspended in an aqueous medium containing dispersing and solubilizing agents. The precorneal cavity retains drug particles in suspension, enhancing the contact time of the drug. The particle size of the drug determines the time required for the absorption of the drug molecules into corneal tissue, thus ultimately affecting the drug bioavailability. TobraDex ST, Alcon, Inc. is a suspension of (0.3%) tobramycin and (0.05%) dexamethasone indicated for bacterial ocular infections (Scoper et al., 2008). TobraDex ST was developed from TobraDex to overcome the high viscosity of the initial formulation. TobraDex ST demonstrated higher tissue concentrations of the drugs tobramycin and dexamethasone in rabbits along with improvements in formulation quality and pharmacokinetic parameters. Clinical studies also showed similar results with higher concentrations of dexamethasone in the aqueous humor after TobraDex ST administration as compared with TobraDex. Yet another US Food and Drug Administration (FDA)—approved ophthalmic suspension is Besivance (Bausch & Lomb) Besivance is a suspension of 0.6% besifloxacin and is prescribed to treat bacterial conjunctivitis. A multicenter, randomized, double-masked, vehicle-controlled clinical trial in adults and children demonstrated that administration of 0.6% besifloxacin ophthalmic suspension twice daily resulted in reduction of signs and symptoms of bacterial conjunctivitis (Silverstein et al., 2011). In another phase III study, 2% rebamipide suspension (OPC-12759) was effective for treatment of dry eye disease as compared with the control group (NCT00885079). Also, the formulation was well tolerated and demonstrated high efficacy for the treatment of dry eye disease (Diestelhorst et al., 1998; Kinoshita et al., 2012).

**Emulsions.** An emulsion is a biphasic system composed of two immiscible phases. Ophthalmic emulsions can offer advantages in improvement of drug solubility and bioavailability of previously water-insoluble drugs. Pharmaceutical emulsions can be widely categorized as water in oil and oil in water (w/o). Ophthalmic formulations widely use the w/o system, which consists of a hydrophobic drug mixed in oil and dispersed in an aqueous medium. An o/w emulsion is preferred over a water in oil emulsion for the reasons of better ocular tolerability and lower ocular irritation due to the external aqueous phase. Some examples of marketed ophthalmic eye drops are Restasis (Allergan), AzaSite (Akorn), Refresh Endura (Allergan), and Durezol (Alcon). Restasis is
a 0.05% emulsion of cyclosporine-A indicated for the treatment of dry eye disease. AzaSite is a 1% azithromycin ophthalmic emulsion used to treat bacterial conjunctivitis and various other ocular infections, while Refresh Endura is a nonmedicated emulsion for dry eye disease (Opitz and Harthan, 2012). Durezol is an emulsion of difluprednate, an anti-inflammatory corticosteroid used to treat anterior ocular uveitis. Studies have demonstrated that Durezol can be applied to treat DME and for the management of postoperative ocular pain and inflammation (Korenfeld et al., 2009; Kang-Mieler et al., 2014). Emulsions can sustain drug release, improve corneal drug absorption, and prolong the formulation residence time in the precorneal cavity. This helps in enhancing the bioavailability of lipophilic drugs for the treatment of anterior segment disorders (Liang et al., 2008).

Drug Delivery to Back of the Eye

**Intravitreal Injections of Anti-VEGF Agents.** The first indication of VEGF in ophthalmology can be traced back to 1940, when a group of scientists proposed that a diffusible factor was responsible for normal vasculature development. Imbalance in the particular factor resulted in neovascularization evident in proliferative DR. By the late 1990s, VEGF was identified as a potential mediator of choroidal neovascularization and intraocular neovascularization for patients suffering from AMD (Amin et al., 1994; Lopez et al., 1996). Proof-of-concept studies established that VEGF blockage resulted in inhibition of neovascularization in various animal models (Aiello et al., 1995; Zhu et al., 1999; Campochiaro and Hackett, 2003) and indicated VEGF blockage can be a potential new approach to overcome retinal disorders involving neovascularization (Adamis et al., 1996).

Forty years after cloning of VEGF, a humanized monoclonal antibody, bevacizumab (148 kDa), was developed as a VEGF-specific antibody. Bevacizumab was approved for treatment of various cancers, but soon its effectiveness in choroidal neovascularization was recognized. Currently, Avastin, Genentech (bevacizumab) is a realistic off-label treatment of wet AMD and DR. Pegaptanib sodium (Macugen, Bausch & Lomb) was the first antiangiogenic VEGF aptamer approved by the US FDA for the treatment of wet or nonvascular AMD in 2004. An intravitreal injection of pegaptanib sodium (pegylated anti-VEGF aptamer) alleviated the conditions of wet AMD and reduced vision loss (Gragoudas et al., 2004; Ng et al., 2006). Subsequently, an F(ab) fragment of bevacizumab, ranibizumab (49 kDa), was developed by Genentech (Presta et al., 1997). Ranibizumab demonstrated a higher binding affinity than pegaptanib to VEGF and better penetration into the retinal layers as compared with bevacizumab (Mordenti et al., 1999). Due to prior success of earlier clinical trials (phase I and phase II) of ranibizumab intravitreal injection, a phase III trial (MARINA) was conducted with 716 patients as a treatment of wet AMD. More than 94% of the patients in the phase III trial (MARINA) was conducted with 716 patients as a treatment of wet AMD once every month (LUCENTIS, 2006). The most recent approved monoclonal antibody for the treatment of wet AMD is aflibercept (97 kDa), a recombinant fusion protein. Eylea (Regeneron Pharmaceuticals) (aflibercept, an intravitreal injection) acts by blocking the action of VEGF and inhibiting neovascularization. Aflibercept has revealed approximately 200 times higher binding affinity to VEGF as compared with ranibizumab. While ranibizumab only binds to the VEGF-A isoform (Zarwar et al., 2016; Zhang et al., 2017), aflibercept binds to various isoforms of VEGFs (VEGF-A and VEGF-B) and placentational growth factor. Binding of aflibercept to various growth factors suppresses all of the actions of VEGF and blocks many pathways, such as cell migration, cell proliferation, and cellular differentiation, leading to neovascularization. Both Eylea and Lucentis are biotech drugs extensively used in the form of intravitreal injections and now serve as the gold standard for the treatment of wet AMD and DME (Chang et al., 2012; Rodrigues et al., 2018).

**Recent Inventions for Ocular Drug-Delivery Technologies**

**Anterior Segment Ocular Drug-Delivery Technologies**

**Punctum Plugs.** Punctum plugs are biocompatible devices inserted in the tear ducts to block tear drainage. These are also known as occluders or lacrimal plugs, which have a size of 2–5 mm. Punctum plugs are noninvasive and can provide controlled drug release to the anterior segment of the eye. Construction of such ocular inserts is possible from nonbiodegradable and biodegradable materials. Nonbiodegradable punctum plug delivery systems (PPDS) are made from silicone, polycaprolactum, and hydroxethyl methacrylate, which is intended to provide controlled drug release up to 180 days. After this period, the insert is removed. Recently, a PPDS (SmartPlug, Medennium Inc.) was developed from a thermosensitive hydrophobic acrylic polymer for the treatment of dry eye disease. The thermosensitive PPDS undergoes modification from rigid solid to a soft gel-like structure after insertion into the eye (http://www.eyeconsultant.info/pdfs/smartplug.pdf). Ocular Therapeutix (Bedford, MA) has developed OTX-TP (travoprost punctum plug insert) to deliver travoprost to the ocular tissues for 90 days. Currently, a phase III clinical trial is set for evaluating the safety and efficacy of OTX-TP for reduction of intraocular pressure (IOP) and ocular hypertension (NCT02914509). Recently, Ocular Therapeutix also completed a phase III clinical study for the safety and efficacy of OTX-DP (dexamethasone punctum plug insert) for the treatment of chronic allergic conjunctivitis and for treatment of inflammation after cataract surgery as compared with a placebo punctum plug (NCT02988882, NCT02736175). High efficiency and safety of OTX-DP led to the US FDA approval of Dextenza (dexamethasone insert; Ocular Therapeutix) for the treatment of pain following ophthalmic surgery (Dextenza, 2018). The company has also developed OTX-TP2 (a prostaglandin trap), which can be used for the treatment of glaucoma and postoperative ocular care (Kang-Mieler et al., 2014).

Several clinical trials have been conducted to investigate the effectiveness of PPDS for the treatment of open-angle glaucoma, glaucoma, and ocular hypertension (NCT00650702, NCT01845038).

**Subconjunctival/Episcleral Implants.** Ocular implants can be inserted into the anterior segment of the eye for controlled drug delivery for a prolonged period. Such implants can be surgically inserted into the subconjunctival region, aqueous humor, and episcleral region. These implants provide
the advantage of sustained localized drug delivery and higher patient compliance as compared with topical eye drops. An insertion is made on the conjunctiva for the insertion of the implants. While some inserts are implanted in the junction between the conjunctiva and the sclera (Nicoli et al., 2009), others are inserted into the aqueous humor (Mohokhia et al., 2013). Surodex (Allergan Inc.) is an example of an anterior segment insert, which is inserted into the anterior ocular segment post cataract surgery to alleviate post-surgery inflammation. Surodex is a rod-shaped biodegradable insert consisting of the drug dexamethasone using polymers such as poly lactide-co-glycolide (PLGA) and hydroxypropyl methyl cellulose, allowing sustained drug release for 7–10 days (Tan et al., 1999, 2001). A study demonstrated that a 7-day drug release with Surodex achieved higher concentrations as compared with maximum peak drug concentrations after topical treatment with dexamethasone eye drops (Tan et al., 1999). Lux Biosciences developed a silicone-based episcleral implant (LX201) for delivery of cyclosporine-A to the anterior ocular tissues for a period of 1 year. In a phase III clinical study, Lux Biosciences also evaluated the effectiveness of LX201 to prevent corneal graft rejection (NCT00447642).

**Cul-de-sac Implants.** The cul-de-sac of the eye is a pocket-like depression where the bulbar and palpebral conjunctiva meet in the upper or lower eyelid. Ocular devices such as Lacrisert (Bausch & Lomb) and Ocusert (Akorn) are examples of cul-de-sac implants designed for drug delivery to the anterior segment of the eye. These devices are safer and less invasive than the conjunctival and episcleral implants. Lacrisert (Bausch & Lomb) is a hydroxypropyl cellulose implant inserted into the inferior cul-de-sac. The implant is suitable for patients with moderate to severe dry eye disease (McDonald et al., 2009). Lacrisert decreased corneal sensitivity, recurrent corneal erosions, and exposure to keratitis. It is also effective for the treatment of conjunctival hyperemia (Lacrisert, 1988). Lacrisert releases cellulose, allowing maintenance of tear film integrity. The implant acts as a lubricant and helps to protect the ocular surface. However, Lacrisert can cause discomfort. It causes foreign body sensation, ocular irritation, hypersensitivity, hyperemia, and blurry vision. Ocusert is a drug-eluting implant delivering pilocarpine over a period of 7 days and directed for the treatment of glaucoma. However, pilocarpine in the insert caused unwanted side effects, such as eyebrow ache and miosis. This resulted in removal of Ocusert from the market (Pollack et al., 1976). Yet another cul-de-sac implant is DSP-Visulex (Aciont Inc., Salt Lake City, UT), which has completed a phase II clinical trial for the treatment of anterior uveitis (NCT02309385). DSP-Visulex contains dexamethasone and is inserted into the bulbar conjunctiva (Papangkorn et al., 2018).

**Drug-Eluting Contact Lenses.** Drug-eluting contact lenses (CLs) are light-transparent corneal dressings acting as drug reservoirs and sustaining drug discharge near the post lens tear fluid for the treatment of anterior ocular disorders. Drug-loaded soft contact lenses are an innovative drug-delivery system to not only prolong and sustain drug release but also enhance drug penetration across the corneal epithelium as compared with conventional eye drops. Contact lenses can increase bioavailability of the drug by increasing the contact time of the drug (Mandal et al., 2017a). Various soft contact lenses have been developed for antifungal agents, which can prolong drug delivery up to 21 days (Phan et al., 2014). A clinical trial was conducted for evaluation of the safety and efficacy of drug-eluting contact lenses for the management of glaucoma. The contact lenses are loaded with timolol maleate and dorzolamide HCl along with vitamin E as an additive for achieving sustained drug release (NCT0252057). Various technologies have been used to load drugs on contact lenses instead of just soaking the lens with the drug. Recently, Gulsen and Chauhan (2004) advanced a novel drug-eluting contact lens, which embedded lidocaineladen nanoparticles. The investigators studied the drug release from the formulation and observed a sustained lidocaine release in vitro over 7 to 8 days (Gulsen and Chauhan, 2004). Similarly, Ciolino et al. (2009) fabricated a drug-eluting contact lens using a polymer-embedded matrix for ciprofloxacin and econazole. The in vitro data demonstrated a zero-order drug-release profile, which can sustain drug release up to 1 month (Ali et al., 2007; Ciolino et al., 2009). Figure 2 depicts the advantage of soft drug-loaded contact lenses over conventional eye drops.

Contact lenses offer the highest drug bioavailability as compared with other noninvasive ophthalmic medications due...
Ocular Iontophoresis. Ocular iontophoresis is a method for active drug delivery utilizing mild electric charges for effective delivery through the ocular barriers. Iontophoresis enhances ocular drug delivery by utilizing electroportation (electrical field–induced ocular tissue structure alteration and pore formation), electrophoresis (direct application of electric field), and electro-osmosis (convective solvent flow through an applied electric potential). Iontophoresis is a noninvasive method having advantages over invasive techniques requiring surgical interventions. This technique of drug permeation can be used for anterior and posterior ocular disorders by utilizing trans-corneal and trans-scleral routes, respectively. Trans-corneal iontophoresis can be used for treatment of anterior segment disorders such as corneal ulcers, dry eye disease, ocular inflammation, keratitis, and ocular uveitis. Trans-corneal iontophoresis is unsuitable for posterior segment delivery due to the presence of barriers such as the lens diaphragm and iris-ciliary. However, the trans-scleral pathway allows drug transport at the back of the eye due to avoidance of anterior segment barriers (Morokhia et al., 2013). The success of iontophoresis-mediated drug delivery depends on several factors, such as charge density of the intended molecule, electron current applied, duration of treatment application, and position of electrode placement (Morokhia et al., 2007; Gratieri et al., 2017).

Eyegate Pharmaceuticals Inc. has developed trans-scleral iontophoresis for delivering drugs in the intended target tissues. The company conducted several clinical trials on the safety and efficacy of a dexamethasone phosphate (EFG-437) formulation for distribution through the EyeGate II Delivery System for the treatment of dry eye disease, anterior uveitis, cataract, postoperative pain, anterior chamber inflammation, and anterior scleritis (NCT01129856, NCT02517619, NCT03180255, NCT01059955). EGF-437 delivered through the EyeGate II Delivery System resulted in reduction of dose frequency as compared with standard dexamethasone eye drops. The US FDA has granted an orphan drug designation for the delivery of EGF-437 through the EyeGate II Delivery System as a treatment option for corneal graft rejection. Iontophoresis is a valuable treatment option for patients who are nonresponsive to eye drop therapy (Kompella et al., 2010). The treatment also resulted in fewer incidences of increased IOP and controlled drug delivery with lower iontophoresis dose (mA-min) (http://www.eyegatepharma.com/technology/iontophoresis-delivery-system/). Visulex-P (Aciont Inc.) and OcuPhor (Iomed Inc., Salt Lake City, UT) are ocular iontophoresis systems currently under investigation for trans-scleral iontophoresis.

Iontophoresis has certain advantages over other ocular drug-delivery modalities, including injections and topical drops. It can achieve higher bioavailability and reduced clearance as compared with topical eye drops. Treatment with the iontophoresis method usually has better patient compliance as compared with ocular injections. Nonetheless, certain patients screened for ocular iontophoresis experienced some discomfort and burning sensation (Parkinson et al., 2003). Posterior segment ocular disorders such as AMD, DR, DME, and central retinal vein occlusion (CRVO) require sustained drug delivery at higher doses. Aciont Inc. has evaluated the potential of ocular iontophoresis for the treatment of AMD by a Visulex-I-noninvasive ocular drug device for the delivery of Avastin (bevacizumab) and Lucentis (ranibizumab) through the trans-scleral route (https://www.sbir.gov/sbirsearch/detail/1070943) (Pescina et al., 2010). Table 2 summarizes currently available ocular drug-delivery devices in clinical trials for the management of anterior segment disorders.

Posterior Segment Ocular Drug-Delivery Technologies

Novel drug-delivery systems, such as implants, are currently used by the clinicians to sustain and prolong drug release to cure back of the eye disorders such as DR, AMD, DME, retinal vein occlusion (CRVO), and posterior uveitis. Intravitreal implants are injected or surgically implanted in the vitreous humor of the eye. Intravitreal implants can prolong the drug action up to many months and reduce the need for frequent intravitreal injection of therapeutic agents. Such frequent administration can cause retinal detachment and retinal hemorrhage, and can be painful for the patients. Such disadvantages of intravitreal injections can be minimized with the use of intravitreal implants. The following section illustrates various intravitreal ocular implants currently available in the clinic and those under clinical investigation.

Durasert Drug-Delivery Technology System. The Durasert technology system (pSivida Corp., Watertown, MA) delivers drugs at various predetermined time points depending on the implant design. The drug release ranges from days to years. Durasert consists of a drug core with surrounding polymer layers. The drug release is a function of the polymer layer permeability. Vitrasert (Bausch & Lomb) is the first intravitreal drug-delivery system loaded with an antiviral drug (ganciclovir) for the treatment of cytomegalovirus retinitis. It utilizes the Durasert technology system and releases the active drug through a small opening in the insert for a period of 6–8 months (Chang and Dunn, 2005). The Retisert...
<table>
<thead>
<tr>
<th>Drug</th>
<th>Brand Name</th>
<th>Mode of Administration</th>
<th>Excipient Controlling Release Characteristic of Drug</th>
<th>Target Indication</th>
<th>Developmental Stage</th>
<th>Clinical Trial #</th>
</tr>
</thead>
<tbody>
<tr>
<td>Azithromycin</td>
<td>AzaSite (Akorn, Inc.)</td>
<td>Eye drops</td>
<td>Polycarbophil (DuraSite)</td>
<td>Bacterial conjunctivitis</td>
<td>Launched</td>
<td>NCT00105469</td>
</tr>
<tr>
<td>Azithromycin/dexamethasone (ISV-502)</td>
<td>AzaSite Plus (Akorn Inc.)</td>
<td>Eye drops</td>
<td>Polycarbophil (DuraSite)</td>
<td>Blepharoconjunctivitis</td>
<td>Launched</td>
<td>NCT00579555</td>
</tr>
<tr>
<td>Betalanol</td>
<td>Betoptic S (Novartis pharmaceuticals)</td>
<td>Eye drops</td>
<td></td>
<td>Glaucoma</td>
<td>Launched</td>
<td>NCT00061542</td>
</tr>
<tr>
<td>Bimatoprost</td>
<td>Lumigan (Allergan)</td>
<td>Eye drops</td>
<td></td>
<td>Glaucoma</td>
<td>Launched</td>
<td>NCT01589550</td>
</tr>
<tr>
<td>Bromfenac</td>
<td>Prolensa (Bausch &amp; Lomb)</td>
<td>Eye drops</td>
<td></td>
<td>Postoperative inflammation</td>
<td>Launched</td>
<td>NCT01847658</td>
</tr>
<tr>
<td>Cyclosporine-A</td>
<td>Restasis (Allergan Inc.)</td>
<td>Eye drops</td>
<td>Cationic emulsion</td>
<td>Dry eye due to keratitis sicca</td>
<td>Launched</td>
<td>NCT02554981</td>
</tr>
<tr>
<td>Difluprednate</td>
<td>Durezol (Novartis Pharmaceuticals)</td>
<td>Eye drops</td>
<td>Emulsion</td>
<td>Anterior uveitis</td>
<td>Launched</td>
<td>NCT01201798</td>
</tr>
<tr>
<td>Timolol maleate</td>
<td>Timoptic (Bausch &amp; Lomb)</td>
<td>Eye drops</td>
<td></td>
<td>Glaucoma/intraocular hypertension</td>
<td>Launched</td>
<td></td>
</tr>
<tr>
<td>Tobramycin/dexamethasone</td>
<td>Tobradex ST (Novartis Pharmaceuticals)</td>
<td>Eye drops</td>
<td>Xanthan gum</td>
<td>Blepharitis</td>
<td>Launched</td>
<td>NCT01162244</td>
</tr>
<tr>
<td>Timolol maleate</td>
<td>Timoptic-XE (Merck &amp; Co., Inc)</td>
<td>Eye drops</td>
<td>Gellan gum</td>
<td>Glaucoma</td>
<td>Launched</td>
<td>NCT01446497</td>
</tr>
<tr>
<td>Ophthalmic emulsion</td>
<td>Cationnorm (Santen Pharmaceuticals)</td>
<td>Eye drops</td>
<td>Cationic emulsion</td>
<td>Mild dry eye</td>
<td>Launched</td>
<td>NCT03460548</td>
</tr>
<tr>
<td>Travoprost</td>
<td>iStent Inject (Glaukos Healthcare)</td>
<td>Punctum plug</td>
<td>Episceral implant</td>
<td>Open-angle glaucoma</td>
<td>Phase IV</td>
<td>NCT03624699</td>
</tr>
<tr>
<td>Cyclosporine (LX201)</td>
<td>EyeGate II (Eye Gate Pharma)</td>
<td>Iontophoresis</td>
<td>Silicone</td>
<td>Keratoconjunctivitis</td>
<td>Phase III</td>
<td>NCT00447642</td>
</tr>
<tr>
<td>Dexamethasone phosphate (EGP-437)</td>
<td>EyeGate II (Eye Gate Pharma)</td>
<td>Punctum plug</td>
<td>Hydrogel</td>
<td>Anterior uveitis</td>
<td>Phase III</td>
<td>NCT01129856</td>
</tr>
<tr>
<td>Dexamethasone (OTX-DP)</td>
<td></td>
<td></td>
<td></td>
<td>Postoperative inflammation</td>
<td>Phase II</td>
<td>NCT00650702</td>
</tr>
<tr>
<td>Latanoprost</td>
<td>Duraser (pSivida Corp.)</td>
<td>Subconjunctival insert</td>
<td>PLGA</td>
<td>Glaucoma</td>
<td>Phase II</td>
<td>NCT00024289</td>
</tr>
<tr>
<td>Loteprednol etabonate mucus-penetrating</td>
<td>Invelys (Kala Pharmaceuticals, Inc)</td>
<td>Nanoparticle</td>
<td>Mucus-penetrating particles</td>
<td>Kerato conjunctivitis sicca</td>
<td>Phase III</td>
<td>NCT03616699</td>
</tr>
<tr>
<td>particles</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Urea</td>
<td></td>
<td>Nanoparticle</td>
<td>Amphiphilic block copolymer</td>
<td>Cataract</td>
<td>Phase II</td>
<td>NCT03001466</td>
</tr>
<tr>
<td>Omega-3 fatty acids</td>
<td>Remogen Omega (TRB Chemedica)</td>
<td>Microparticle</td>
<td>Microemulsion of polyunsaturated fatty acids and hydrating polymers</td>
<td>Dry eye disease</td>
<td>Phase II</td>
<td>NCT02906282</td>
</tr>
</tbody>
</table>

TABLE 2
Currently available ocular drug-delivery systems in clinical trials for the treatment of anterior segment disorders (Kang-Mieler et al., 2014)
intravitreal implant (Bausch & Laumb, Inc.) is a steroid-eluting device implanted surgically in the vitreous humor. Retisert releases fluorocinolone acetonide up to 3 years into the vitreous humor (Jaffe et al., 2006; Kempen et al., 2011). Retisert has received a fast-track US FDA approval for treatment of posterior uveitis as an orphan drug treatment. Posterior uveitis, also called choroiditis, is the inflammation of the choroid capillaries. This can lead to damage to the optic nerve and permanent loss of vision. Retisert contains a fluorocinolone acetonide tablet encapsulated within a silicone elastomer cup containing an orifice made with a polyvinyl alcohol membrane (Hagbjou et al., 2011).

Iluvien (fluocinolone acetonide intravitreal implant, Alimera Sciences, Inc.) is the most recent US FDA-approved intravitreal injectable insert indicated for the treatment of DME. Multicenter, randomized clinical trials demonstrated that both a low dose and high dose of Iluvien resulted in a significant visual improvement with lower side effects. The onset of treatment was very rapid. Patients suffering from DME for more than 3 years had received almost twice the treatment effectiveness as compared with the control group (Campochiaro et al., 2012; Cunha-Vaz et al., 2014). Iluvien is being evaluated in phase II clinical trials for its efficacy of dry AMD (NCT00695318), wet AMD (NCT00605423), and macular edema secondary to RVO as compared with Lucentis (ranibizumab) treatment (NCT00770770).

**NOVADUR Drug-Delivery Technology.** The NOVADUR (Allergan Inc.) system consists of therapeutic agents in a polymer matrix of PLGA. PLGA is a biodegradable and biocompatible polymer which breaks down to lactic and glycolic acid when it comes in contact with the vitreous humor fluid (Hagbjou et al., 2011). Ozurdex (Allergan) is a controlled-release intravitreal dexamethasone implant approved by the US FDA for the treatment of DME, RVO, and posterior uveitis (Haller et al., 2011; Boyer et al., 2014; Sangwan et al., 2015). Ozurdex contains 0.7 mg of dexamethasone in a PLGA matrix which releases the drug up to 90 days. Mayer et al. (2012) recently investigated the effects of intravitreal bevacizumab followed by Ozurdex and Ozurdex monotherapy for the treatment of CRVO and macular edema. The research group concluded that there was no difference between the aforementioned treatment strategies for treatment of CRVO. However, for branched retinal vein occlusion, Ozurdex monotherapy resulted in a better functional outcome (Mayer et al., 2012).

Currently, a phase III clinical trial is being conducted for the possible effectiveness of intravitreal implant of Ozurdex monotherapy for the treatment of DME (NCT00168389). PLGA containing brimonidine tartrate (Allergan) is another intravitreal implant in clinical trials for dry AMD (NCT00658619) and retinitis pigmentosa (NCT00661479). Brimonidine is an α2 adrenergic agonist which releases neurotrophic factors such as ciliary neurotrophic factor and brain-derived neurotrophic factor (Kim et al., 2007). Brimonidine protects retinal cells like photoreceptor cells, RPE and ganglion cells from apoptotic cell death (Zhang et al., 2009).

**I-vation Triamcinolone Acetonide Drug-Delivery Technology.** I-vation TA (SurModics Inc.) is also an intravitreal drug-delivery implant for triamcinolone acetonide (TA). I-vation is a titanium helical coil implant coated with TA in a nonbiodegradable polymer. Preclinical experiments suggested that I-vation TA can sustain TA release in vivo up to 2 years. A phase I safety and preliminary efficacy study was conducted in 31 patients with DME after implantation of I-vation TA. The TA intravitreal implant was well tolerated by the patients as indicated by a minimal rise in IOP. The I-vation TA treatment also aided the reduction of macular thickness from baseline, indicating alleviation of DME (Dugel et al., 2009).

**Encapsulated Cell Technology.** Renexus (NT-501) is an encapsulated cell technology (ECT) for ocular implant of human RPE transfected with plasmid encoding ciliary neurotrophic factor. Renexus (NT-501) is under a phase III investigation for dry AMD, glaucoma, and retinitis pigmentosa (NCT03316300). The implant consists of a hollow tube capsule consisting of a polymeric matrix which can be loaded with genetically modified cells (Sieving et al., 2006; Emerich and Thanos, 2008). Various biocompatible polymers such as collagen and hyaluronic acid hydrogel are used for forming the ECT matrix. The implant capsule is semi-permeable, allowing diffusion of proteins across the membrane but inhibiting the entry of immune cells. The genetically modified cells in the matrix draw nutrients from the surrounding tissue after implantation. The encapsulated cell technology is implanted in the pars plana and affixed to the sclera.

ECT can be advantageous as compared to other corticosteroid implants as they can secrete biologically active molecules for a prolonged period of time, requiring less frequent implant replacement. Kontturi et al. (2015) demonstrated genetically modified RPE capable of secreting soluble VEGF receptor to suppress VEGF activity in choriocapillary neovascularization and retinal neovascularization. This proof-of-concept study indicated that the human RPE cell line remained viable with a constant secretion of soluble VEGF-1 receptor up to 50 days (Kontturi et al., 2015). Although the researchers found a modest VEGF inhibition in vivo model, this delivery technology displays promise for utilization of ECT to treat disorders such as wet AMD, DR, and DME. ECT can be considered as a versatile platform that can be used for secreting targeted therapeutic biotech drugs such as antibodies, antibody fragments, growth factors, cytokines, and prostaglandins for back of the eye disorders (Tao, 2006). Wong et al. formulated an injectable composite alginate-collagen (CAC) matrix ECT gel having human retinal pigment epithelial cells and glial cell–derived neurotrophic factor (GDNF) secreted by HEK293 cells. The GDNF-secreting HEK293 cells were transfected with lipofectamine repressor (Tet R) DNA and pro-caspase 8 transcription DNA. Tet R can be used as a biosafety switch for the ECT drug-delivery system, whereas pro-caspase 8 can trigger the inbuilt apoptotic pathway in the retinal cells. The researchers witnessed a continuous supply of bioactive glial cell–derived GDNF in vitro and effective proliferation control in rat ocular tissues. Intravitreal injections of CAC ECT in rats with retinal damage resulted in decreased apoptosis of photoreceptor and retinal function loss. Similarly, dual intravitreal injections of the ECT resulted in further reduction of photoreceptor death and gain of retinal structure and function without compromising gel viability (Fig. 3). The CAC ECT demonstrated high encapsulation efficiency of the transfected cells, high cell viability, and high mechanical stability of the implant without the use of immunosuppressant (Wong et al., 2019). Thus, ECT can be considered as a safe, effective, and well-controlled platform for the treatment of back of the eye disorders with retinal dysfunction (Baranov et al., 2017).
Fig. 3. ECT for back of the eye disorders. CAC ECT gel treatment on rats with inherited retinal degeneration. One or two units of GDNF-delivering CAC ECT gel was intravitreally injected into the eyes of dystrophic RCS/lav rats. (A) Representative H&E sections of nontreated, single, and double gel-treated rats showed different degrees of photoreceptor nuclei retention and organization in the outer nuclear layer (ONL). (B) ONL nuclei density was calculated by normalizing ONL count with retinal length. (C) Representative images showing the distribution of apoptotic cells (green) in the retina of nontreated, single, and double gel-treated animals detected by terminal deoxynucleotidyl transferase–mediated digoxigenin-deoxyuridine nick-end labeling (TUNEL) assay with 4',6-diamidino-2-phenylindole (DAPI) nuclear counterstaining. (D) Density of apoptotic cells in the ONL. (E) Representative scotopic and photopic electroretinogram wave forms showing the retinal function of dystrophic rats receiving 1 or 2 U of GDNF-secreting gel. (F) Scotopic a-wave. (G) Scotopic b-wave. (H) Photopic b-wave. *P < 0.05; **P ≤ 0.02; ***P < 0.005; ****P < 0.0005 by one-way ANOVA with Bonferroni post hoc test (Wong et al., 2019). ERG, electroretinogram; INL, inner nuclear layer.
### TABLE 3
Currently available ocular drug-delivery systems in clinical trials for the treatment of posterior segment disorders (Kang-Mieler et al., 2014)

<table>
<thead>
<tr>
<th>Drug</th>
<th>Brand Name</th>
<th>Mode of Administration</th>
<th>Excipient Controlling Release Characteristic of Drug</th>
<th>Target Indication</th>
<th>Developmental Stage</th>
<th>Clinical Trial #</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dexamethasone</td>
<td>Ozurdex (Allergan)</td>
<td>Intravitreal implant</td>
<td>PLGA (Novadur)</td>
<td>Macular edema</td>
<td>Posterior uveitis</td>
<td>Launched</td>
</tr>
<tr>
<td>Ganciclovir</td>
<td>Vitrasert (Auritec Pharmaceuticals Inc.)</td>
<td>Intravitreal implant</td>
<td>PVA/EVA</td>
<td>CMV retinitis</td>
<td>Launched</td>
<td>NCT00000135</td>
</tr>
<tr>
<td>Fluocinolone acetonide</td>
<td>Retisert (Bausch &amp; Lomb)</td>
<td>Intravitreal implant</td>
<td>PVA</td>
<td>Posterior uveitis</td>
<td>Launched</td>
<td>NCT00570830</td>
</tr>
<tr>
<td>Verteporfin</td>
<td>Visudyne (Bausch &amp; Lomb)</td>
<td>i.v. injection</td>
<td>Liposome</td>
<td>Wet AMD</td>
<td>Launched</td>
<td>NCT0024580</td>
</tr>
<tr>
<td>Dexamethasone</td>
<td>Dexuex (EyePoint Pharmaceuticals, Inc.)</td>
<td>Intravitreal implant</td>
<td>Acetyl triethyl citrate</td>
<td>Postoperative inflammation</td>
<td>Launched</td>
<td>NCT02547623</td>
</tr>
<tr>
<td>Difluprednate</td>
<td>Durezol (Novartis Pharmaceuticals)</td>
<td>Eye drops</td>
<td>Emulsion</td>
<td>DME</td>
<td>Off-label</td>
<td>NCT00429923</td>
</tr>
<tr>
<td>Betamethasone</td>
<td>Renexus (Neurotech Pharmaceuticals)</td>
<td>Intravitreal implant</td>
<td>Semipermeable hollow fiber membrane/NTC-200</td>
<td>Atrophic AMD</td>
<td>Phase II/III</td>
<td>NCT0316300</td>
</tr>
<tr>
<td>Dexamethasone</td>
<td>Iluvien (Alimera Sciences)</td>
<td>Intravitreal implant</td>
<td>Cyclohexatin microparticles</td>
<td>DME</td>
<td>Phase II/III</td>
<td>NCT01523314</td>
</tr>
<tr>
<td>Triamcinolone acetonide with ranibizumab</td>
<td></td>
<td>Intravitreal injection</td>
<td>Verisome</td>
<td>DME</td>
<td>Phase IV</td>
<td>NCT01304706</td>
</tr>
<tr>
<td>Brimonidine</td>
<td></td>
<td>Intravitreal implant</td>
<td>PLGA</td>
<td>Dry AMD</td>
<td>Phase II</td>
<td>NCT02087085</td>
</tr>
<tr>
<td>Triamcinolone acetonide (IBI-20089)</td>
<td></td>
<td>Intravitreal implant</td>
<td>Benzyl benzoate</td>
<td>Wet AMD</td>
<td>Phase II</td>
<td>NCT01175395</td>
</tr>
<tr>
<td>Triamcinolone acetonide (RETAAC)</td>
<td></td>
<td>Intravitreal implant</td>
<td>PLGA</td>
<td>DME</td>
<td>Phase III</td>
<td>NCT00407849</td>
</tr>
<tr>
<td>Dexamethasone prodrug (NOVA-63035)</td>
<td>Cortiject (Novagali Pharma SA)</td>
<td>Intravitreal implant</td>
<td>Emulsion</td>
<td>DME</td>
<td>Phase I</td>
<td>NCT00665160</td>
</tr>
<tr>
<td>Ranibizumab</td>
<td></td>
<td>Intravitreal implant</td>
<td>Drug port</td>
<td>Refillable port</td>
<td>Phase I</td>
<td>NCT03677934</td>
</tr>
<tr>
<td>VEGF-R2 (NT-503)</td>
<td></td>
<td>Intravitreal implant</td>
<td>Semipermeable hollow fiber membrane/NTC-200</td>
<td>Wet AMD</td>
<td>Phase I</td>
<td>NCT02228304</td>
</tr>
<tr>
<td>Human embryonic stem cell–derived retinal pigment epithelium (MA09-RPE) cells</td>
<td></td>
<td>Cells transplantation via subretinal injection</td>
<td>Cell suspension</td>
<td>Advanced dry AMD</td>
<td>Phase II</td>
<td>NCT01344993</td>
</tr>
<tr>
<td>AR-1105 (dexamethasone implant)</td>
<td></td>
<td>Intravitreal implant</td>
<td>Biodegradable implant</td>
<td>Macular edema due to RVO</td>
<td>Phase II</td>
<td>NCT03730593</td>
</tr>
</tbody>
</table>

CMV retinitis, cytomegalovirus retinitis; CNTF, ciliary neurotrophic factor; EVA, Ethylene-vinyl acetate copolymer; PVA, Poly(vinyl alcohol); RP, Retinitis pigmentosa.
Suprachoroidal Drug Delivery Utilizing Hollow Microneedles and Microsurgical Cannulas. Delivery of therapeutics at the suprachoroidal space has demonstrated promising potential for delivering therapeutic agents at the target tissue (retina and choroid) at a higher concentration. This can be confirmed from anatomic studies demonstrating the diffusion of therapeutic agents after drug delivery at the suprachoroidal space (Patel et al., 2012; Kadam et al., 2013; Chiang et al., 2016). Patel et al. (2012) demonstrated suprachoroidal drug delivery through the posterior pars plana of a rabbit model using a hollow microneedle. The suprachoroidal drug delivery was a minimally invasive procedure demonstrating safe delivery into the retina and choroid with no adverse effects. Gilger et al. (2013) reported the successful suppression of acute inflammation with corticosteroid delivered through the suprachoroidal route in a porcine model of noninfectious posterior uveitis. Drug delivery through the suprachoroidal route utilizing microsurgical cannulas in primate and porcine models has shown increased drug bioavailability. The researchers investigated delivery of triamcinolone acetonide and bevacizumab to evaluate the tolerability, safety, efficacy, and pharmacokinetics of suprachoroidal drug-delivery technology. Higher bioavailability of triamcinolone acetonide at the target tissue without deleterious side effects such as cataract and hypertension suggests its positive impact (Olsen et al., 2006). In contrast, bevacizumab demonstrated low bioavailability at target tissue with faster diminishing therapeutic response as compared with intravitreal injections (Olsen et al., 2011). Currently, various phase III clinical trials utilizing triamcinolone acetonide suprachoroidal injection along with various anti-VEGF agents are being investigated for the treatment of DME and posterior uveitis (NCT03203447, NCT02980874, NCT01789320). Table 3 summarizes currently available ocular drug-delivery systems in clinical trials for the treatment of posterior segment disorders.

Novel Ocular Drug-Delivery Technologies

Colloidal Nanocarriers for Anterior Segment Disorders

The chronic nature of many ocular disorders requires frequent and prolonged drug treatments. Along with this, ocular barriers reduce the bioavailability of the topically applied therapeutic agents to less than 5%. Recent developments in nanotechnology can provide opportunities to overcome drawbacks and limitations of conventional drug-delivery systems, such as low drug bioavailability and low drug permeation through ocular barriers. Nanocarriers can prolong drug action by sustained and controlled release of the drug, protect the drug from ocular enzymes, and aid in overcoming ocular barriers. This can greatly reduce the frequency of dosing and improve tissue concentrations of the drug for better pharmacological action. Colloidal nanocarriers including nanoparticles, nanomicelles, nanowafers, and microneedles are capable of encapsulating small molecules and biotech drugs for ocular delivery. The size of the nanocarriers ranges from 1 to 1000 nm. Nanoparticles greater than 10 μm can cause foreign body sensation and ocular irritation (Ali and Lehmuissaari, 2006; Liu et al., 2012). Nanocarriers can also improve the ability of drug penetration into the deeper ocular tissues, decrease drug toxicity, and reduce hyperosmolar drug loss taking place due to rapid tear turnover. Nanocarriers engineered from biodegradable and biocompatible polymers overcome ocular barriers and result in higher drug absorption in the anterior and posterior segments of the eye (Reimondez-Troitiño et al., 2015). Nanomedicine for ocular drug delivery can be highly patient compliant and have a higher tolerability than conventional eye drops for anterior segment ocular disorders (Vandervoort and Ludwig, 2007; Bachu et al., 2018; Mandal et al., 2019a).

Nanomicelles. Nanomicelles are colloidal drug-delivery systems that self-assemble in a solution and can entrap therapeutic agents at their core. Their size ranges from 10 to 200 nm, and they are made up of amphiphilic surfactants or block copolymers. Nanomicelles are formed instantaneously in a solution when the concentration of the polymers is above a specific concentration called the critical micellar concentration. Nanomicelles have the capacity to encapsulate hydrophobic drugs in the hydrophobic core of the micelles due to hydrophobic interactions. The hydrophilic corona interacts with the external aqueous fluid, increasing the solubility of a relatively lipophilic drug. This colloidal dosage form has the ability to form clear aqueous solutions which can be used as topical eye drops. Nanomicelles can be broadly classified as surfactant nanomicelles and polymeric nanomicelles. Cequa...
Sun Pharmaceuticals Inc.) is a nanomicellar formulation of 0.09% cyclosporine-A recently approved by the US FDA for dry eye disease. Cequa demonstrated improved rapid onset of action as early as 4 weeks and improvement in tear production as compared with cyclosporine-A emulsion in phase II and phase III clinical trials (Mandal et al., 2019a). The in vivo studies of the nanomicellar formulation of cyclosporine-A conducted in rabbits demonstrated enhanced bioavailability in the anterior ocular tissues as compared with cyclosporine-A emulsion with no ocular adverse effects. Here, the nanomicellar system was prepared from a polymeric mixture of two low-molecular-weight surfactants, hydrogenated castor oil-40 and octoxynol-40, which resulted in formation of a clear solution of cyclosporine-A. Mitra et al. demonstrated efficient encapsulation and enhanced ocular pharmacokinetics of hydrophobic drugs such as voclosporin, cyclosporine-A, rapamycin, triamcinolone acetonide, cidofovir prodrug, and curcumin for the treatment of various anterior and posterior ocular disorders neutrotropic. Various surfactant polymers such as vitamin E tocopheryl polyethylene glycol succinate (Vit E TPGS); hydrogenated castor oil-40, 60, 100; and octoxynol-40 were used for entrapping hydrophobic drugs in the nanomicellar core (Cholkar et al., 2015, Mandal et al., 2017b, 2019b, Trinh et al., 2017). Mandal et al. demonstrated the entrapment of hydrophobic drug and hydrophilic peptides within the core of nanomicelles for ocular drug delivery. A lipid prodrug of cyclic cidofovir (B-C12-cCDF) was encapsulated within surfactant-based nanomicelles for antiviral drug delivery for cytomegalovirus retinitis, and multilayered nanomicelles were developed for the delivery of octreotide peptide to the

Fig. 5. In vivo efficacy of PLGA fenofibrate NPs (Feno-NP) on vascular leakage and vascular permeability measured with Fundus Fluorescein Angiography (FFA). Formation of subretinal neovascularization (SRNV) and intraretinal neovascularization (IRNV) evaluated by neovascular tufts in flat-mounted choroid and retina in Vldr−/− mice 1 month after Feno-NP treatment. (A) Representative images of FFA. (B) Numbers of leakage spots in FFA. (C) Quantification of retinal vascular permeability. (D) Representative images of SRNV and IRNV in FFA. Scale bar, 1000 μm. (E) Quantification of SRNV and IRNV in flat-mounted choroid and retina. Mean ± S.E.M. (n = 8–16; one-way ANOVA followed by Bonferroni post hoc test). ***P < 0.001 vs. untreated Vldr−/− mice; ###P < 0.001 vs. blank-NP–treated Vldr−/− mice (Qiu et al., 2019).
antibody and cytokine delivery. The researchers also demonstrated that a mixed micellar structure designed from a fixed ratio of low-molecular surfactants had a lower critical micellar concentration. This indicates that the nanomicellar structure is stable over dilution in the systemic fluids and will not result in premature drug release. These highly lipophilic agents form a clear solution when encapsulated in the nanomicelles. Also, nanomicelle aid is sustained, and release of the drug to the ocular tissue is controlled (Cholkar et al., 2015; Mandal et al., 2017b, 2019b; Trinh et al., 2017).

Nanomicelles constructed from block copolymers such as PLGA, polyethylene glycol (PEG), polycaprolactone (PCL), and poly(lactide) are called polymeric nanomicelles. The polymers can be conjugated to form diblock (A-B type), triblock (A-B-A), or pentablock (A-B-C-B-A) copolymers. Block polymers have distinct hydrophilic and hydrophobic parts which impart the polymer amphiphilicity. Nanomicelles can solubilize hydrophobic drugs and improve their delivery to the ocular tissues. Methoxy poly(ethylene glycol) poly(lactides) diblock copolymer was used for constructing polymeric nanomicelles of Cyclosporine-A for efficient drug supply to the anterior ocular segment. The in vivo results demonstrated excellent ocular biocompatibility and high ocular bioavailability of the nanomicellar formulation. The results suggested that methoxy poly(ethylene glycol) poly(lactides) nanomicelles encapsulating cyclosporine-A can be used for treatment of dry eye disease, prevention of corneal graft rejection, and treatment of autoimmune uveitis (Di Tommaso et al., 2011). Polymeric micelles often offer certain advantages over surfactant micelles, such as sustained drug release and lower incidence of drug toxicity, whereas surfactant nanomicelles offer advantage of smaller nanomicellar size and rapid onset of action. Both surfactant and polymeric nanomicelles can be surface conjugated with various targeting moieties for higher drug transport through the ocular tissue (Yellepeddi and Palakurthi, 2016). Nanomicellar delivery of nucleic acids like siRNA, microRNA, plasmidDNA, and oligonucleotides is an emerging field of research. Liaw and Robinson used a nonionic copolymermic system, poly (ethylene oxide)-poly (propylene oxide) (PEO-PPO-PEO) for gene delivery (Liaw, Chang, and Hsiao, 2001). The polymeric nanomicelles encapsulated plasmid DNA with lacZ gene demonstrating efficacy in improving retinal ganglion cell survival after retinal ischemia. Rupenthal et al. and Bishat et al. evaluated connexin43 mimetic peptide PLGA NPs for retinal ischemia in zebrafish and live embryos. The study resulted in no toxicity to the ocular tissues (Chen et al., 2015; Bishat and Rupenthal, 2018). Qiu et al. (2019) developed fenofibrate PLGA nanoparticles for the management of DR and AMD. Fenofibrate is an agonist of peroxisome proliferator-activated receptor α and has efficacy against DR. The in vivo studies in diabetic rats reduced retinal vascular leakage, ameliorated retinal dysfunctions, and downregulated the overexpressed VEGF-A and ICAM-1 in 8 weeks after one intravitreal injection of fenofibrate PLGA NPs (Qiu et al., 2019). PLGA can also be used to encapsulate many well known anti-VEGFs, such as bevacizumab, ranibizumab, and aflibercept (Elsaid et al., 2016; Sousa et al., 2017; Kelly et al., 2018). However the major problem associated with the intravitreal delivery of NPs is the floating of the particles in the vitreous humor and vision obstruction (Bachu et al., 2018) (Fig. 5).

Nonsteroidal anti-inflammatory drugs such as ibuprofen, indomethacin, and flurbiprofen encapsulated in NPs can be used for the treatment of anterior segment ocular inflammation. Ibuprofen encapsulated in Eudragit RS100 (Evonik Health Care) NPs demonstrated improved drug concentrations in the aqueous humor of rabbit eyes in comparison with ibuprofen ocular solution (Pignatello et al., 2002). Eudragit RS100 was used to prepare flurbiprofen NPs for lowering anterior segment inflammation after surgical trauma. In vivo studies performed in rabbits demonstrated higher aqueous humor concentrations of flurbiprofen as compared with the control group (Pignatello et al., 2002a,b; Gupta et al., 2007; Cao et al., 2010). Biodegradable polymers such as PCL, PEG, PLGA, and poloxamer 188 were used for formulation of flurbiprofen-encapsulated nanoparticles. Topical administration of flurbiprofen nanoparticles demonstrated enhanced
anti-inflammatory efficiency and minimal toxicity, such as ocular irritation, in the rabbit eyes (Calvo et al., 1996; Valls et al., 2008). Chitosan is also a widely used anionic bio-compatible and biodegradable polymer used to prepare NPs and can improve their precorneal residence time. Cyclosporine-A is a strong immunosuppressive agent which is used to treat dry eye disease. Chitosan can be used to prepare NPs entrapment cyclosporine-A, which has shown 2-fold improved precorneal residence and higher conjunctival permeability in rabbit eyes. Chitosan polymer can also be used for the delivery of lipophilic drugs, hydrophilic drugs, and polynucleotides to the anterior ocular surface (De Campos et al., 2001; de la Fuente et al., 2010). Mitra et al. developed pentablock copolymers from polymers such as PEG, polylactide, PGA, PCL, and PLGA for making nanoparticles encapsulating hydrophilic drugs such as dexamethasone and macromolecules such as IgG, IgG(Fab), and various peptides for controlled drug delivery to the anterior as well as posterior sections of the eye (De Campos et al., 2001; de la Fuente et al., 2010). Glaucoma is the leading cause of blindness throughout the world. Li et al. (2019) created a mouse model of glaucoma demonstrating elevated intraocular pressure after the administration of dexamethasone nanoparticles composed of pentablock copolymers. This can streamline the clinical evaluation of drug candidate for glaucoma (Li et al., 2019). Current research is utilizing ligand-targeted functionalized nanoparticles for enhanced delivery of therapeutic agents as compared with nonfunctionalized nanoparticles. Targeting ligands can specifically target receptors and nutrient transporters on the conjunctiva and corneal surface. The CD44 hyaluronic acid receptor is located on the corneal and conjunctival cells. It was proven that hyaluronic acid surface–functionalized chitosan NPs encapsulating an oligomer demonstrated higher uptake in the ocular tissues as compared with NPs not surface functionalized with hyaluronic acid. Such NPs undergo active trans- portation mediated by the CD44 hyaluronic acid receptor utilizing the caveolin-dependent endocytosis pathway (Contreras-Ruiz et al., 2011). Surface-functionalized nanoparticles with targeting agents such as peptides, antibodies, vitamins (such as biotin and folic acids), and aptamers have resulted in higher uptake as compared to the nonfunctionalized nanoparticles. Kompella et al. (2006) demonstrated that transferrin-conjugated NPs had 74% higher transport across the cornea and conjunctiva in ex vivo bovine eyes as compared with nontargeted NPs. Epigallocatechin-3-gallate is a natural polyphenol compound having antioxidant, anti-inflammatory, and antiangiogenesis activity and can have efficacy against CNV. Gelatin NPs were surface functionalized with hyaluronic acid and conjugated to an RGD peptide. Encapsulated epigallocatechin-3-gallate-RGD peptide was evaluated for treatment of corneal neovascularization (CNV). Lee et al. in vivo studies in a CNV mouse model showed fewer and thinner blood vessels for mice treated with topical epigallocatechin-3-gallate-RGD peptide NPs as compared with the blank NPs (Lee et al., 2014). This result suggests a potential role of targeted nanoparticles for the treatment of CNV. Active targeting of NPs can provide efficient and rapid transport of cargo across the corneal and conjunctival epithelium.
Nanoparticles can also serve as an effective vehicle for gene delivery. Gold NPs conjugated to a 2-kD polyethylenimine were evaluated for gene delivery to rabbit cornea. The researchers observed a high uptake of the gold NPs through the rabbit stroma and a gradual clearance over time (Sharma et al., 2011).

Liposomes. Liposomes are used as ocular drug-delivery vehicles which can encapsulate hydrophilic and hydrophobic drugs. Polymers from a liposome form a lipid bilayer vesicle which separates the inner aqueous core from the exterior aqueous environment. Although liposomes have poor stability and a short half-life, they have been explored for ocular drug delivery for anterior segment disorders (Law et al., 2000). Sun et al. (2008) entrapped short-chain-conjugated ceramide and C6-ceramide in liposomes and applied to the treatment of corneal inflammation in mice. Ceramides are known for their role as an antiproliferative and proapoptotic agents in sphingolipid metabolism. The C6-ceramide liposomal formulation demonstrated significant efficacy in corneal inflammation in mice. This implies an affirmative role of ceramide-loaded liposomes for treating anterior segment ocular inflammation (Sun et al., 2008). This higher stability and superior water solubility of liposomes compared to conventional formulations make them promising for ocular drug delivery. However, liposome stability varies greatly, and factors such as the concentration of cholesterol in the liposome formulation, pH, and the presence of proteases and lipases in the ocular environment can impact liposome stability (Law et al., 2000). Furthermore, the physiochemical properties of liposomes are influenced by factors such as the composition of the lipid bilayer, the size of the vesicle, and the concentration of the drug encapsulated (Law et al., 2000).

Dendrimers. Dendrimers are polymeric nanocarriers having a branched star-shaped structure. The size and shape of the dendrimer can be controlled and customized to deliver the drug to the desired tissue. The dendrimer is composed of a core surrounded by several generations of monodispersity layers, each layer being composed of a specific functional group. This allows for targeted drug delivery, with the ability to modify the surface of the dendrimer for specific interactions. Dendrimers have been studied extensively for ocular drug delivery, with applications ranging from the treatment of dry eye disease to glaucoma (Davis et al., 2014).
and a plethora of functional groups on the surface for chemical modification. Hydrophilic and lipophilic drugs can either be conjugated to the surface of the dendrimer or be encapsulated by caging in the internal structure of the dendrimer (Kalomiraki et al., 2015; Lancina and Yang, 2017). A polyamidoamine (PAMAM) polymer having carboxylic and hydroxyl functional groups is the most commonly used dendrimer for ocular drug delivery. High branching of the PAMAM polymer can lead to primary, secondary, and tertiary generations of the dendrimer nanocarrier. Soiberman et al. (2017) designed a gel formulation of the G4-PAMAM dendrimer with cross-linked hyaluronic acid, entrapping dexamethasone intended for the treatment of corneal inflammation. Subconjunctival injection of the dendrimer formulation led to reduction in central corneal thickness and improved corneal clarity in an alkali burn rat model, which was highly clinically relevant (Soiberman et al., 2017). Another group of investigators evaluated the potential of dexamethasone-PAMAM dendrimers for the delivery to the back of the eye for the treatment of diseases such as DR and AMD. In vivo studies in rats showed that the drug-loaded dendrimers enhanced the ocular permeability of dexamethasone after subconjunctival injection as compared with the free drug (Yavuz et al., 2016). Matrix metalloproteinases-9 (MMP-9) can trigger corneal damage and result in dry eye disease. Cerofolini et al. (2017) synthesized an MMP-9 inhibitor and solubilized with PAMAM dendrimers. The synthesized inhibitor had high binding affinity to MMP-9 and can be used for the treatment of corneal inflammation and dry eye disease (Cerofolini et al., 2017). Vandamme and Brobeck (2005) entrapped tropicamide and pilocarpine nitrate in PAMAM dendrimers to study the effect of drug-release kinetics after altering the size, molecular weight, carboxylate and hydroxyl surface groups, and total number of amines present in the PAMAM dendrimer. In vivo results in New Zealand albino rabbits revealed higher drug residence time of dendrimers functionalized with carboxylic and hydroxyl functional groups (Vandamme and Brobeck, 2005).

**Microneedles.** Microneedle drug-delivery technology was originally used for overcoming the stratum corneum and was used for transdermal drug delivery (Lee et al., 2008). The effectiveness of microneedles for transdermal drug-delivery systems inspired researchers to investigate their potential to treat anterior and posterior segment ocular disorders. This minimally invasive technique can also be applied for ocular drug delivery of hydrophilic and hydrophobic drugs. Solid stainless-steel microneedles (MNs) coated with drugs such as sunitinib malate and pilocarpine resulted in higher drug bioavailability in the anterior ocular segment compared with topical drop applications in vivo (Jiang et al., 2007; Song et al., 2015). Microneedles can also be used to deliver therapeutic agents for the treatment of back of the eye disorders. Microneedle nanoparticles and microparticle suspension can be delivered to the suprachoroidal space (Patel et al., 2011). Than et al. (2018) have shown a polymeric eye patch consisting of an array of detachable and biodegradable MNs for controlled and localized ocular drug delivery. These MNs could penetrate into the corneal layers and deliver antiangiogenic monoclonal antibody (DC101) for the treatment of CNV. The MNs were double layered with DC101 to provide biphasic drug-release kinetics to enhance the therapeutic efficacy of the MNs. The DC101 MN eye patch produced approximately 90% reduction in CNV in a CNV disease mouse model as compared with a topical eye drop. The researchers also suggested that the MN patch is minimally invasive and can be self-applied by patients on their corneas (Than et al., 2018). Microneedles can greatly aid in increasing the bioavailability of a certain drug in a particular tissue by localizing the drug-delivery system. Microneedles can be a paradigm shift for the way ocular formulations are administered, but their current limitations demand further research in the field for desired clinical translation (Thakur Singh et al., 2017) (Fig. 6).

**Fig. 7.** Comparison of cyclosporine-A nanomicellar formulation (OTX-101, Cequa) and cyclosporine-A emulsion (Restasis) evaluated in New Zealand white rabbits after a single topical administration. Concentration was determined in ocular tissues such as superior bulbar conjunctiva (A), cornea (B), and sclera (C).
**Nanowafers.** Nanowafers are small, transparent, rectangular membranes or circular discs containing drug loaded into nanoreservoirs which can be smeared on the ocular surface using a fingertip. Controlled drug release from the nanowafers can increase the residence and contact time of the drug with the corneal and conjunctival surfaces. This can aid in higher drug absorption into anterior ocular tissues. The nanowafers not only enhance the drug bioavailability but also act as a protective polymer membrane to heal injured and abraded corneal surfaces commonly found in CNV and dry eye disease. This novel nanocarrier is designed from biodegradable and biocompatible polymers which can be eliminated over a period of time. Coursey et al. (2015) and Bian et al. (2016) developed a dexamethasone-loaded nanowafers (Dex-NW) for the treatment of dry eye disease. The nanowafers was fabricated using carboxymethyl cellulose polymer and consisted of an array of nano drug reservoirs filled with dexamethasone. The in vivo efficacy of Dex-NW was tested in a dry eye disease mouse model. Dex-NW was administered as once-a-day treatment on alternating days for a 5-day period of time. After the treatment duration, it was observed that Dex-NW was able to restore the corneal barrier function along with a healthy ocular surface which was similar to twice-a-day treatment of topically applied dexamethasone eye drops. Yet another interesting finding the scientists reported was that Dex-NW was effective in lowering the overexpression of inflammatory cytokines such as tumor necrosis factor-α, interferon-γ, interleukin-1β, and interleukin-6. Also, the expression of inflammatory chemokines such as CXCL-10, CCL-5, and MMP-3 and MMP-9 was lowered (Coursey et al., 2015; Bian et al., 2016). Axitinib-loaded nanowafers were developed by Yuan et al. for the treatment of CNV (Yuan et al., 2015). A murine ocular burn model was used to evaluate the in vivo efficacy of axitinib-loaded nanowafers. The laser-scanning confocal imaging and reverse-transcription polymerase chain reaction results revealed that the once-a-day axitinib-loaded nanowafers was twice as effective as compared with axitinib daily topical eye drops (Yuan et al., 2015). These findings have shown the potential of nanowafers for further evaluation in clinical trials.

**Ocular Nanocarriers Currently Approved and under Clinical Investigation.** Nanocarriers, such as nanoparticles and nanomicelles, have been widely explored for their potential to cure anterior and posterior ocular disorders. Despite the plethora of research on nanocarriers, Cequa (0.09% cyclosporine-A ophthalmic nanomicellar solution) is the only nanotechnology-derived ophthalmic approved by the US FDA for dry eye disease (Cequa, 2018). Cequa is a preservative-free, clear, and sterile nanomicellar formulation of a highly hydrophobic drug, cyclosporine-A. The phase III clinical trials of Cequa were conducted in a total of 745 patients with dry eye disease. The study showed a statistically ($P < 0.0001\%$) significant increase in the primary endpoints of the study, Schirmer’s test (measurement of tear production), and secondary endpoints. Instillation site pain (22%) and hyperemia (6%) were the adverse effects noted with the clinical trial, which are a common scenario for the drugs evaluated in this category (Sheppard et al., 2014; Tauber et al., 2015). The phase III results clearly established the safety and efficacy of Cequa (0.09% cyclosporine-A ophthalmic nanomicellar formulation) in mitigating the signs and symptoms of dry eye disease (Mandal et al., 2019a).

There are a handful of ophthalmic nanocarrier drugs currently being investigated in clinical trials to establish their safety and efficacy for the treatment of ocular disorders. A randomized, single-blind study is evaluating the efficacy of hydrating polymers and polysaturated fatty acid microemulsion for the treatment of dry eye disease (NCT02908282). In a yet another randomized, single-blind phase II clinical trial, urea-loaded nanoparticles are being evaluated as a possible treatment of cataract management (NCT03001466). A clinical study was conducted by Sun Yat-sen University to compare the efficacy of two tear substitutes, Tears Naturale Forte (Alcon Laboratories Inc.) and Liposic (Bausch & Lomb), for dry eye diseases (NCT02992392). Aston University evaluated the efficacy of liposomal spray for dry eye disease in an interventional randomized study (NCT02420834). Kala Pharmaceuticals (Waltham, MA) has developed nanoparticle-based mucus-penetrating particles of loteprednol etabonate (LE). LE is a corticosteroid, and encapsulating in mucus-penetrating particles can improve drug delivery across the ocular endothelial cells. Currently, Kala Pharmaceuticals is investigating the potential of Inveltys (KPI-121 1.0% LE) for relieving inflammation following ocular surgery (NCT02793817) and KPI-121 0.25% LE for alleviating the symptoms of dry eye disease in a phase III clinical trial (NCT03616899). The effect of KPI-121 1.0% and 0.25% LE is also being investigated for the treatment of diabetic macular edema and retinal vein occlusion (NCT02245516).

Fewer nanoformulations in clinical trials can be attributed to the limitations in the industrial development and scale-up of nanoparticles. Another major challenge involved in the clinical translation of nanoparticles is the toxicity profile of various polymers used in nanoparticles (Suresh and Sah, 2014). The majority of nanoparticles for ocular drug delivery are evaluated for their efficacy in vivo in mice, rats, and rabbits. Although rabbit ocular anatomy is similar and comparable to human ocular anatomy, rabbit ocular anatomy does not completely mimic human ocular anatomy. Rabbits have higher mucus production, higher surface sensitivity, and lower rate of blinking, which can result in better drug retention and drug penetration in comparison to human eyes (Weng et al., 2017). It is also a challenge to achieve homogeneity of particle size and particle-size distribution for a nanoparticle formulation on an industrial scale. Optimization of various formulation parameters for nanoparticle preparation is still a challenging task for many pharmaceutical scientists. Dendrimers have been shown to cause blurring of vision (Wadhwa et al., 2009). On the other hand, liposomes have limited long-term stability and lower drug-loading potential. Higher concentrations of surfactants in the nanoparticle formulation can be associated with potential ocular toxicity (Bachu et al., 2018). The recent US FDA approval of Cequa has led to an inception of the era of nanotechnology in ophthalmology. Despite limiting factors for the successful clinical translation of nanomedicine for ophthalmology, one can predict nanotechnology products being approved for ocular ailments in the near future (Fig. 7).

**Noninvasive Drug-Delivery Systems for the Posterior Disorders**

All marketed ophthalmic products used for the management of retinal disorders are invasive in nature. The intravitreal...
route is widely used for administration of biopharmaceutics to the back of the eye. This route is associated with various complications, such as intraocular inflammation, retinal detachment, glaucoma or intraocular pressure elevation, endophthalmitis, ocular hemorrhage, and cataract (Mandal et al., 2018). The following section illustrates current-state scientific research pertaining to topical delivery of potent therapeutic interventions and drugs for back of the eye diseases.

**Small Molecules.** Eye drops instilled topically are noninvasive and the most patient-compliant route of administration. Although the route is widely explored for anterior segment disorders, it remains a major challenge for delivering drugs at therapeutic concentrations at the back of the eye. Various static barriers, such as blood-retinal barrier and tear-film barrier, and dynamic barriers, such as clearance mechanisms by vitreous and aqueous humor, hinder the drug passage from the front to the back of the eye. TG100801 is a topical therapy which has demonstrated reduction in CNV in a murine model and edema reduction in rats with RVO (Doukas et al., 2008). TG100801 is a small-molecule multikinase inhibitor prodrg which is cleaved to its active form by hydrolysis in the cornea. Due to the promising results of TG100801 in the preclinical setting, it was further advanced to clinical trials. Although TG100801 was well tolerated by patients, it did not demonstrate any efficacy for alleviating the condition of AMD (NCT00509548). Pazopanib is another small-molecule multikinase inhibitor which was administered topically in a laser-induced CNV rat model (Yafai et al., 2011). Similar to TG100801, pazopanib failed to demonstrate efficacy in patients with subfoveal CNV, secondary to AMD (Singh et al., 2014). Along similar lines, acrizanib was investigated for reduction of nonvascular AMD in preclinical mouse models. Acrizanib is a VEGF receptor-2 inhibitor and demonstrated a 99% inhibitory effect for CNV, which was 3 times the daily topical application of 1% suspension in mice (Adams et al., 2018). Despite positive preclinical evaluation of acrizanib in a mouse model, topically administered acrizanib is clinically ineffective for the treatment of AMD (NCT02355028).

Although some multikinase inhibitors have failed in clinical settings, topical delivery of therapeutic agents to the back of the eye is an active area of research. A multikinase inhibitor, PAN-90806, is currently being investigated in clinical trials (phase I/II) to assess its feasibility in AMD treatment (NCT03479372). Topical application of a memantine drug (Namzaric, Actavis Plc.) was able to achieve a sufficient concentration in the retina to provide retinal neuroprotection (Hughes et al., 2005). Another small-molecular drug, dorzolamide, was administered topically to inhibit carbonic anhydrase II in a rabbit model (Inoue et al., 2004). Dexamethasone administered topically by iontophoresis showed promising results in a rabbit model. Topically administered dexamethasone by iontophoresis was further evaluated in clinical trials for macular edema. However, the clinical trial was terminated due to insufficient enrollment (NCT02485249).

**Biotech Drugs.** Biotech drugs such as antibodies or antibody fragments are high-molecular-weight charged compounds which cannot be easily absorbed by the lipid bilayer. Although topical delivery of small-molecular drugs to the back of the eye has shown some efficacy in clinical trials, the biologics face various ocular barriers to reach the posterior segment (Ambati et al., 2000a; Miao et al., 2013). Topically administered bevacizumab, an anti-VEGF IgG antibody, failed to reach the therapeutic concentration in the rabbit retina after topical dosing of 1.25 mg/0.5 ml six times daily for a week (Ambati et al., 2000a). However, topical administration of antibody against intercellular adhesion molecule 1 was able to achieve therapeutic concentrations at the retina, which resulted in successful inhibition of VEGF-induced leukostasis in the choroid (Ambati et al., 2000b). To further improve topical delivery of biologics to the back of the eye, colloidal nanoformulations such as liposomes and nanomicelles with various permeability enhancers were used. Williams et al. (2005) demonstrated that permeability enhancer sodium caprate can enhance the delivery of antibody fragment in a rabbit model. Platania et al. (2017) used annexin A5–associated liposomes for topical delivery of bevacizumab to the back of the eye. Various cell-penetrating peptides (CPPs) are increasingly

### TABLE 5

<table>
<thead>
<tr>
<th>Compound</th>
<th>Formulation</th>
<th>Preclinical Data</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>TG100801</td>
<td>Solution</td>
<td>Murine CNV model and edema in rat</td>
<td>Doukas et al., 2008</td>
</tr>
<tr>
<td>Pazopanib</td>
<td>Solution</td>
<td>Rat CNV model</td>
<td>Yafai et al., 2011,</td>
</tr>
<tr>
<td>Acrizanib</td>
<td>Suspension</td>
<td>Murine CNV</td>
<td>Adams et al., 2018</td>
</tr>
<tr>
<td>Memantine</td>
<td>Solution</td>
<td>Drug levels in rabbit retina</td>
<td>Hughes et al., 2005</td>
</tr>
<tr>
<td>Dorzolamide</td>
<td>Solution</td>
<td>Drug levels and carbic anhydrase activity</td>
<td>Inoue et al., 2004</td>
</tr>
<tr>
<td>Dexamethasone</td>
<td>Iontophoresis</td>
<td>Drug levels in retina and vitreous of rabbit</td>
<td>Ambati and Adams, 2002</td>
</tr>
<tr>
<td>Bevacizumab</td>
<td>Solution</td>
<td>Drug levels in iris/ciliary body, retina/choroid,</td>
<td>Ambati et al., 2000a</td>
</tr>
<tr>
<td>Anti-intercellular adhesion molecule-1 antibody fragment</td>
<td>Solution by osmotic pump</td>
<td>Drug levels and VEGF-induced leukostasis in the choroid and retina in rabbit</td>
<td>Ambati et al., 2000b</td>
</tr>
<tr>
<td>Bevacizumab</td>
<td>Annexin A5–based liposomes</td>
<td>Drug levels in retina of rat and rabbit</td>
<td>Williams et al., 2005</td>
</tr>
<tr>
<td>Transforming growth factor β1</td>
<td>Annexin A5–based liposomes</td>
<td>Drug levels in vitreous in rabbit</td>
<td>Davis et al., 2014</td>
</tr>
<tr>
<td>Acidic fibroblast growth factor</td>
<td>CPP (TAT)</td>
<td>Drug levels in vitreous in rabbit</td>
<td>Platania et al., 2017</td>
</tr>
<tr>
<td>Calpain inhibitory peptide</td>
<td>CPP (TAT)</td>
<td>Drug levels in retina retina</td>
<td>Wang et al., 2010</td>
</tr>
<tr>
<td>Green fluorescent protein</td>
<td>CPP (POD)</td>
<td>Drug levels in mouse cornea</td>
<td>Ozaki et al., 2015</td>
</tr>
<tr>
<td>Bevacizumab</td>
<td>CPP (R6)</td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td>Johnson et al., 2010</td>
</tr>
<tr>
<td>Bevacizumab</td>
<td>Annexin A5–based liposomes</td>
<td>Drug levels in retina/choroid, and plasma in rabbit</td>
<td>de Cogan et al., 2017</td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina of rat and rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in mouse cornea</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina/choroid, and plasma in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in mouse cornea</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina/choroid, and plasma in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in mouse cornea</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina/choroid, and plasma in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in mouse cornea</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina/choroid, and plasma in rabbit</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in retina retina</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in mouse cornea</td>
<td></td>
</tr>
<tr>
<td>Sodium caprate</td>
<td></td>
<td>Drug levels in vitreous in rat and murine CNV model</td>
<td></td>
</tr>
</tbody>
</table>

CPP (R6), cell-penetrating peptide poly-arginine-6; POD, peptide of ocular delivery; TAT, transactivator of transcription.
Parallel efforts not only in novel product development but also in product scale-up are required.

Acknowledgments
The authors thank Dr. Gerald Wyckoff, Interim Chair, Department of Pharmacology and Pharmaceutical Sciences, University of Missouri-Kansas City School of Pharmacy, and Dr. Abhirup Mandal, Postdoctoral Fellow in Bioengineering, Harvard John A. Paulson School of Engineering and Applied Sciences, Harvard University, for their guidance and support of this article.

Authorship Contributions
Wrote or contributed to the writing of the manuscript: Sicotte, Sikler, Gote, Pal.

References