Implication of Kv7 Channels in the Spinal Antinociceptive Actions of Celecoxib

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ABSTRACT

Celecoxib is a nonsteroidal anti-inflammatory drug (NSAID) commonly used to treat pain conditions in humans. In addition to its blocking activity on cyclooxygenase (COX) enzymes, several other targets could contribute to its analgesic activity. Here we explore the spinal antinociceptive actions of celecoxib and the potential implication of Kv7 channels in mediating its effects. Spinal cord in vitro preparations from hind paw-inflamed animals were used to assess the segmental sensory-motor and the early sensory processing of nociceptive information. Electrophysiological recordings of ventral roots and dorsal horn neurones were obtained, and the effects of celecoxib and Kv7 modulators on responses to repetitive dorsal root stimulation at C-fiber intensity were assessed. Celecoxib applied at clinically relevant concentrations produced depressant effects on responses to dorsal root stimulation recorded from both ventral roots and individual dorsal horn neurones; by contrast, the non-nociceptive monosynaptic reflex was unaffected. The NSAID indomethacin had no effect on spinal reflexes, but further coapplication of celecoxib still produced depressant effects. The depressant actions of celecoxib were abolished after Kv7 channel blockade and mimicked by its structural analog dimethyl-celecoxib, which lacks COX-blocking activity. The present results identify Kv7 channels as novel central targets for celecoxib, which may be relevant to its analgesic effect. This finding contributes to better understand the pharmacology of celecoxib and reinforces both the role of Kv7 channels in modulating the excitability of central pain pathways and its validity as target for the design of analgesics.

Introduction

Celecoxib, a nonsteroidal anti-inflammatory drug (NSAID) with selective activity on cyclooxygenase 2 (COX-2) isoforms, is used for the treatment of rheumatoid arthritis and osteoarthritis in humans (Clemett and Goa, 2000). Celecoxib applied systemically is effective in reducing nociceptive behaviors in various animal models of pain, including the formalin test and the carrageenan model of paw inflammation (Penning et al., 1997; Zhang et al., 1997; Smith et al., 1998; Inoue et al., 2009; Sun et al., 2013). In addition, the application of celecoxib and other NSAIDs by the intrathecal route also produces antinociceptive effects in the same models (Malmberg and Yaksh, 1992; Dirig et al., 1998; Nishiyama, 2006; Lee and Seo, 2008). These reports suggest that, in addition to peripheral actions, the spinal cord may be a relevant site for NSAID-induced analgesia. In addition to its COX-blocking actions, celecoxib has been reported to interact with opioid and cannabinoid systems as well as a variety of ionic channels (Park et al., 2007; Brueggenmann et al., 2009; Du et al., 2011; Rezende et al., 2012; Frolov and Singh, 2014).

Of special interest for nociceptive processing, celecoxib affects the activity of voltage-dependent potassium channels of the K7 family (Du et al., 2011). These channels are expressed in peripheral and central elements of the nociceptive system (Rivera-Arconada et al., 2009), and their modulation alters the excitability of primary afferents and spinal neurones due to its fundamental role regulating membrane resting potential and repetitive firing (Jentsch, 2000; Passmore et al., 2003; Riveria-Arconada and Lopez-Garcia, 2005). The K7 channel opener retigabine when applied systemically shows analgesic-like effects on animal models of inflammatory, visceral, neuropathic, and cancer pain (Blackburn-Munro and Jensen, 2003; Passmore et al., 2003; Hirano et al., 2007; Zheng et al., 2013). Application of celecoxib to dorsal root ganglion neurones in vitro produces an enhancement of K7-mediated currents, an effect that may be relevant to its analgesic actions (Mi et al., 2013).

Here we explore the potential contribution of K7 channels, as expressed in spinal neurones, to the analgesic effects of celecoxib. To this end, we used electrophysiological recordings of spinal reflexes and dorsal horn neurones in an in vitro preparation of the mouse spinal cord obtained after the induction of inflammation. The effect of celecoxib on spinal function was assessed and the contribution of K7 channels was tested using a variety of pharmacologic tools. Our results are...
in agreement with the hypothesis that modulation of \( K_{v7} \) channels is relevant to the spinal antinociceptive actions of celecoxib.

**Materials and Methods**

All experimental protocols were performed following the European Union and Spanish Government regulations for animal handling, were approved by the local ethics committee and the Government of the Community of Madrid (Ref. PROEX 018/16) and comply with the ARRIVE guidelines. Experiments were performed on 6- to 11-day-old CD1 mice of either sex weighing between 4 and 9.7 g. The animals were bred in house, kept together with their mothers, and were maintained under a 12-hour light/dark cycle with 55%–65% humidity. The number of animals used was kept to the minimum that allowed a clear interpretation of main results. All efforts were made to minimize animal suffering.

**Induction of Peripheral Inflammation and Behavioral Testing.** Peripheral inflammation was induced by intraplantar injection of carrageenan (3% in saline; 30 \( \mu l \)) in both hind paws. This procedure ensured maximum inflammation and allowed the use of either side of the cord in electrophysiologic experiments. Paw diameters and mechanical withdrawal thresholds were measured before and 20 hours after carrageenan injection. The withdrawal threshold was defined as the minimum mechanical force applied with von Frey filaments (from 0.008 to 2 g) that elicited a withdrawal response in at least three out of five trials. For statistical analyses, the threshold and paw diameter of the hind paw ipsilateral to the hemi cord used for electrophysiologic recordings were considered.

**In Vitro Spinal Cord Preparations and Dorsal Root Stimulation.** Mice were anesthetized with urethane (2 mg kg\(^{-1}\), i.p.), and their spinal cords were extracted after a dorsal laminectomy following a procedure described in full elsewhere (Lopez-Garcia and Laird, 1998). The whole cord was used for recording spinal reflexes. The spinal cord was hemisected to make multielectrode recordings from deep dorsal horn neurones. To record neurones located in superficial laminae, the lumbar enlargement of the whole spinal cord was sliced in a vibratome to obtain a single slice of \( \approx 400 \mu m \) containing the dorsal part of the cord (laminae from I to V) together with the attached dorsal roots.

In the recording chamber, the cord was pinned down to the Sylgard base of the chamber with the medial or sectioned side upward to facilitate electrode penetration. Cords were continuously superfused (4–6 ml min\(^{-1}\)) with oxygenated (95% O\(_2\); 5% CO\(_2\)) artificial cerebrospinal fluid (aCSF) at room temperature (22 ± 1°C). The composition of the aCSF was (in millimolars): NaCl (127), KCl (1.9), KH\(_2\)PO\(_4\) (1.5), MgSO\(_4\) (1.3), CaCl\(_2\) (2), NaHCO\(_3\) (22), and glucose (10); pH 7.4.

Electrical stimuli were applied to the L4 dorsal root via tight-fitting glass suction electrodes. Electrical stimulation consisted of a train of 15 consecutive C-fiber intensity stimuli (200 \( \mu A\), 200 microseconds) applied at 1 Hz to produce wind-up responses. Trains of stimuli were applied at 30-minute intervals.

**Ventral Root Reflex Recordings.** The L4 ventral root was placed in a tight-fitting glass suction electrode. Signals were obtained using a Multiclamp 7A amplifier (Molecular Devices,
Digital band-pass filtering of the original DC signal between 200 and 1200 Hz allowed us to generate an AC channel to collect fast spike-like events produced by the firing of action potentials in motor neurones (Rivera-Arconada et al., 2016). Events crossing an amplitude threshold were counted in a time window between 100 and 950 milliseconds from stimulus artifact and summed to obtain the total spike counts to each train of stimuli. Stable baseline responses were usually obtained after two to four repetitions.

Multielectrode Recordings from Dorsal Horn Neurones and Estimation of Recording Sites. Multielectrode arrays (MEAs; A4 × 4 3 mm-50/100-125–177-A16 or Buzsaki32; NeuroNexus Technologies, Ann Arbor, MI) had four shanks with four to eight iridium electrodes with a resistivity of 100–125 mΩ. The MEA tips were placed into the tissue by means of a micromanipulator, and the distance of the electrode tip to the dorsal border of the spinal cord was measured with a micromanipulator. To estimate the position of the deep dorsal horn neurones recorded, extracellular signals were preamplified and isolated from each experiment. The compounds were prepared in DMSO as concentrated stock solutions (10 mM–1 M) and stored at −20°C. All compounds were diluted to the final concentration in aCSF just before use and were superfused to the entire preparation for at least 25-minute periods to ensure a complete tissue equilibration.

To study the effect of the compounds, each preparation received applications of only one of the compounds at a maximum of three increasing concentrations. In experiments designed to assess the implication of COX enzymes, 10 μM indomethacin was applied during 60 minutes and then 10 μM celecoxib was added to the perfusion medium. In other series of experiments, the K_+ channel blockers XE-991 or linopirdine were superfused for 60–90 minutes before the application of celecoxib or DMC.

Statistical Analysis. Statistical analysis was performed with GraphPad Prism (GraphPad Software, San Diego, CA). All data are presented as mean ± S.E.M. unless otherwise stated. Analyses of data from behavioral assays were made using Wilcoxon matched pairs test. The effects of celecoxib on spinal reflexes were first compared with control experiments with DMSO applications using the same three concentrations schedule to avoid a misinterpretation of drug actions due to the long-lasting protocol employed. These comparisons were done using a two-way ANOVA test with data normalized to predrug (or presolvent) values. Concentration-dependent effects of the drugs from predrug values were analyzed in raw data using Bonferroni post hoc test after repeated measures one-way ANOVA. For electrophysiological studies with a single concentration of drug, the comparisons to predrug values were performed using paired t test analysis.

Results

Celecoxib Actions on Spinal Reflexes Recorded from Paw-Inflamed Mice. Intraplantar injections of carrageenan induced an inflammatory process in mice pups characterized by paw swelling and allodynia. Twenty hours after carrageenan injection, paw diameter increased from 2.18 ± 0.02 to 3.32 ± 0.15 mm.

![Figure 2](https://example.com/fig2.png)

Fig. 2. Celecoxib still depressed spinal reflexes after indomethacin application. (A) Original recordings showing the lack of effect of indomethacin (P = 0.16 for cumulative depolarization, and P = 0.33 for spike wind-up; paired t test) and the subsequent actions of celecoxib. In the presence of indomethacin, celecoxib still produces a profound depression of nociceptive reflexes (P = 0.01; P = 0.04 for cumulative depolarization and spike wind-up; paired t test). (B) Graph summarizing the results obtained in the six experiments made. *P < 0.05; **P < 0.01 stand for statistically significant differences between indomethacin and celecoxib values as obtained with paired t test analysis.
coapplied during 30 minutes. Indomethacin alone showed a 60% reduction of mechanical withdrawal thresholds assayed with von Frey filaments were reduced from a median force of 1 g to a median force of 0.04 g (Fig. 2). However, further application of celecoxib produced a significant depression of both parameters compared with values obtained under indomethacin (Fig. 2).

Implication of K\textsubscript{7} Channels in the Actions of Celecoxib on Spinal Reflexes. The previous results suggest that depressant effects of celecoxib on spinal nociceptive reflexes are mediated by a COX-independent mechanism. To test the possible contribution of K\textsubscript{7} channels, we studied the effect of K\textsubscript{7} channel blockade on the depressant action of celecoxib. Superfusion of the K\textsubscript{7} channel blocker XE-991 at 10 \mu M on its own produced a strong increase in spike wind-up, but not on the underlying cumulative depolarization (n = 7; Fig. 3, A and B). The depressant effects of celecoxib were fully abolished after K\textsubscript{7} blockade with XE-991 (Fig. 3, A and B). Similar results were obtained using linopirdine, another blocker of K\textsubscript{7} channels (n = 4; Fig. 3C).

To further confirm the implication of K\textsubscript{7} channels on celecoxib effects, we superfused the cord with DMC, an analog of celecoxib devoid of activity on COX enzymes. DMC applied at 10 \mu M on its own did not produce significant changes on spinal reflexes (P = 0.069 for cumulative depolarization and P = 0.075 for spike wind-up; paired t test). No depressant effects of celecoxib were observed either in cumulative depolarization (P = 0.47; paired t test) or spike wind-up (P = 0.61; paired t test). **Significant differences (P < 0.001) between predrug and XE-991 values.

Effects of Celecoxib on the Responses of Dorsal Horn Neurones. A first series of experiments was conducted to address the potential effects of celecoxib on the spinal sensory neurones that contribute to the initial steps of sensorimotor processing. Extracellular recordings with MEAs were obtained from deep dorsal horn neurones using a hemisection spinal cord preparation that allowed us to gain access directly to the gray matter. Twenty-four neurones were successfully isolated from 18 hemisection cords. Recording sites recovered after DiI labeling, and histologic processing showed that electrode tracks were located in lamina IV to VI, between 223 and 436 \mu m from the medial side (Fig. 5A).
Celecoxib was applied at 3 and 10 μM to 13 neurones. Repetitive dorsal root stimulation elicited wind-up responses in three neurones, whereas another eight neurones were able to maintain spike firing during the train of stimuli and the remaining two neurones showed a spike wind-down. In nine of them, action potential firing in response to C-fiber stimulation, was depressed by ≥20% (Fig. 5, B and D). Pooling together data from all neurones we obtained statistically significant differences at both concentrations. The four neurones insensitive to celecoxib showed a pattern of maintained firing in response to repetitive dorsal root stimulation.

Occlusion experiments with XE-991 were performed to explore the involvement of Kv7 channels in the effects of celecoxib. Responses to repetitive stimulation in predrug conditions for the five neurones tested included wind-up responses (two), neurones maintaining spike firing (two), and one neurone showing spike wind-down. A 60-minute application of XE-991 at 10 μM produced a mild but significant change in neuronal responses to repetitive stimuli (n = 5). After the superfusion of XE-991, applications of celecoxib at 3 and 10 μM were completely devoid of effects in the five neurones tested (Fig. 5, C and E).

In an additional set of experiments, we wanted to explore the effects of celecoxib on neurones more specifically related to nociceptive processing. Using a slice containing the laminae from I to V of the dorsal horn, we obtained MEAs recordings of the activity of superficial dorsal horn neurones in response to repetitive dorsal root stimulation at C-fiber intensity. Twenty-one superficial dorsal horn neurones were isolated from eight spinal cord slices. Electrode tips were situated at 20–60 μm from the dorsal border of the spinal cord, allowing all the sensors to be located within laminae I–III (see Fig. 6A).

Celecoxib applied at 10 μM depressed the responses to dorsal root stimulation in 10 out of 12 neurones tested (Fig. 6, B and D). Neurones inhibited by celecoxib showed wind-up (two), wind-down (three), or maintained (five) responses to repetitive stimulation. Two more neurones insensitive to celecoxib had wind-up responses.

Occlusion experiments with XE-991 were made in nine additional neurones showing wind-up (five), wind-down (two), or maintained firing (two). XE-991, applied at 10 μM during 60 minutes, produced slightly increased responses in seven neurones, but this trend did not reach statistical significance. The posterior coapplication of 10 μM celecoxib was devoid of effects in the nine neurones tested (Fig. 6, C and E).

Discussion

Applications of celecoxib to the in vitro mouse spinal cord produced a concentration-dependent depression of nociceptive reflexes, manifested as a reduction of action potential wind-up and the underlying cumulative depolarization, a model widely used to study nociceptive processing in the spinal cord (Herrero et al., 2000). On the other hand, the monosynaptic reflex, a purely non-nociceptive response, was unaffected by celecoxib. This profile of action on spinal reflexes is very similar to that of other analgesics, including morphine (Mazo et al., 2015). In addition, celecoxib reduced the firing of dorsal horn neurones in response to C-fiber stimulation, indicating a specific action of the compound on sensory neurones. These results are in line with previous reports showing that celecoxib applied by intrathecal route reduces nociceptive behaviors evoked by formalin injection in the hind paws (Nishiyama, 2006; Lee and Seo, 2008).

The present results show significant depressant effects of celecoxib at concentrations ≥3 μM. These are in range with plasmatic levels of celecoxib achieved in humans after treatment (100–400 mg, 1–4 μM) (Davies et al., 2000), which suggests that the spinal actions of celecoxib may be relevant to its analgesic effects under clinical conditions.

In this work, carrageenan-inflamed mice were used in an attempt to maximize COX activity. Both COX isozymes are constitutively present in the spinal cord, and their expression appears to up-regulate under neuropathic and inflammatory processes (Hay and de Belleroche, 1997; Zhu and Eisenach, 2003). However, some reports suggest that spinal COX activity is not relevant to acute pain transmission nor to the maintenance of hyperalgesia in inflammatory models (Yamamoto and Nozaki-Taguchi, 1996; Dirig et al., 1998; Vardeh et al., 2009). This view is reinforced by the lack of effect of indomethacin on spinal reflexes and the large, likely unspecific concentrations of other NSAIDs required to cause depression (Lopez-Garcia and Laird, 1998; Lizarraga et al., 2006).

Our carrageenan-inflamed mice showed signs of edema and allodynia. Previous reports using this in vitro model have shown larger spinal reflexes in treated rats than in controls (Hedo et al., 1999). In our study, the application of indomethacin at 10 μM, a concentration above its therapeutic range and IC50 values for COX inhibition (Kurumbail et al., 1996;
Kalgutkar et al., 2000; Lucas, 2016), was devoid of effect on spinal reflexes, whereas subsequent coapplication of celecoxib produced clear depressant actions. These observations suggest a minor role for COX blockade in the actions of celecoxib on spinal transmission and a likely contribution of other unspecific targets for this compound.

Celecoxib applied at concentrations similar to those used in the present experiments has been shown to exert direct actions on heterologously expressed Kv7 channels and on native M-currents expressed in dorsal root ganglia neurones (Brueggemann et al., 2009; Du et al., 2011; Mi et al., 2013). A recent report has shown that Kv7 subunits 2, 3, and 5 are present in the spinal cord and are expressed in both inhibitory and excitatory dorsal horn neurones (Häring et al., 2018). Interestingly, they found that the Kv7.5 subunit is expressed in two populations of glutamatergic neurones (glut 11 and 12), which became activated by noxious thermal stimulation. In this line, other work employing a similar approach (Sathya-murthy et al., 2018) also reported the expression of Kv7.5 subunits in a population of excitatory neurones (DE-7) that express the Fos gene after formalin application.

Celecoxib can interact with several homo- and heteromeric conformations of Kv7 channels showing EC50 values between 2 and 5 μM. Celecoxib shifts the activation curve of Kv7-mediated currents to the left and produces an approximately 2-fold increase in conductance (Du et al., 2011). In previous studies, we showed that retigabine, the standard Kv7 channel opener, has depressant actions on spinal reflexes comparable to those obtained here for celecoxib (Rivera-Arconada et al., 2004; Vicente-Baz et al., 2016). Like celecoxib, retigabine has similar affinity for different Kv7 channel subunits and EC50 values in the low micromolar range (Tatulian et al., 2001; Wickenden et al., 2001).

The major proof obtained here in favor of the implication of Kv7 channels in mediating the spinal effects of celecoxib comes from the experiments with Kv7 channel blockers. Under the present conditions, XE-991 showed clear potentiating actions on the firing of motor neurones while the effects on dorsal horn neurones were smaller. A minor effect of XE-991 on dorsal horn neurones has been observed in intracellular recordings obtained in rats (Rivera-Arconada and Lopez-Garcia, 2005). The lack of effect of XE-991 on dorsal horn neurones may be explained by the little impact that this compound has on dorsal horn neurones and, consequently, on their synaptic input to motor neurones.

The concentration of XE-991 used here was sufficient to produce complete blockade of the effects of the specific

Fig. 5. Effects of celecoxib on the activity of deep dorsal horn neurones. (A) Drawing of a transverse section of the hemisected spinal cord (dorsal side up) with a reconstruction of its borders and laminar organization. The approximate location of the multielectrode shanks used for recording were recovered after DiI labeling. Dotted lines indicate the trajectory of the electrodes inside the spinal cord recovered from the seven experiments in which labeling with DiI was obtained. (B) Original recordings showing celecoxib (3 and 10 μM) effects on action potential firing in response to repetitive high-intensity stimulation of the dorsal root as obtained from a deep dorsal horn neurone. (C) Illustration of the lack of effect of celecoxib after Kv7 channel blockade with XE-991. Celecoxib at 3 and 10 μM was coapplied with 10 μM XE-991, but only the higher concentration is shown. Insets show the average of all action potentials in each condition. Calibration bars for all recordings: 0.2 mV and 2 seconds (0.2 mV and 1 millisecond for insets). (D) Graph showing the effects of celecoxib on action potential firing in response to repetitive dorsal root stimulation in the 13 neurones tested. Celecoxib had depressant effects on neuronal firing (F(2,24) = 15.28; \( P < 0.001 \); one-way ANOVA). The vertical axis was broken to improve visualization. (E) Graph summarizing the effects of celecoxib at 3 and 10 μM after Kv7 channel blockade with XE-991 in the five neurones tested. XE-991 increased the firing of deep dorsal horn neurones (\( P = 0.026 \); paired \( t \) test). In the presence of XE-991, the coapplication of celecoxib was devoid of effect (F(2,8) = 1.58; \( P = 0.26 \); one-way ANOVA). For both graphs, the values for individual neurones are shown in gray; mean ± S.E.M. values for all neurones included in the graph are indicated in black. Asterisks (**\( P < 0.01 \) and ***\( P < 0.001 \)) and plus signs (\( + P < 0.05 \)) indicate statistically significant differences from predrug values as obtained using the Bonferroni post hoc test after one-way ANOVA or paired \( t \) test, respectively.
Kv7 openers N-(2,4,6-trimethylphenyl)-bicyclo[2.2.1]heptane-2-carboxamide (ML213) and N-(2-chloro-5-pyrimidinyl)-3,4-difluorobenzamide (ICA-0696 73) in previous experiments (Vicente-Baz et al., 2016). We have also proof to our satisfaction that XE-991 does not prevent the effects of 1 mMDiazepam, an allosteric modulator of GABA-A receptors under identical experimental conditions (not shown). Thus, we believe that the effects of XE-991 should be considered effective and selective for Kv7 channels. Under these conditions of specific Kv7 channel blockade, celecoxib had no effect on spinal reflexes. Also relevant is the finding that XE-991 prevented the depressant effects of celecoxib on the responses of dorsal horn neurones to C-fiber activation. This shows that early sensory processing of nociceptive information is depressed by celecoxib using a mechanism that likely involves Kv7 channels as well.

Further evidence supporting the implication of Kv7 channels on celecoxib depression of spinal nociceptive transmission was obtained in experiments with DMC, a structural analog of celecoxib that lacks COX blocking activity (Kardosh et al., 2005). This compound produced depressant effects on spinal reflexes similar to those of celecoxib, which were also abolished by XE-991 application.

All these observations support a role for Kv7 channels on celecoxib action although a minor contribution of other targets might be possible. Celecoxib can interact with other targets like sodium and calcium channels which may also contribute to its observed effects (Park et al., 2007; Brueggemann et al., 2009). Although experiments with XE-991 may suggest direct actions on Kv7 channels, Kv7 channel blockade may depolarize resting potential originating an inactivation of sodium and calcium channels, also preventing celecoxib effects mediated by these other channels. We have shown previously that spinal neurones can be depolarized by the application of 10 μM XE-991 (Rivera-Arconada and Lopez-Garcia, 2005). Depolarizations ranged from 1 to 5 mV, with higher values found in motor neurones, which also showed more negative potentials. These small depolarizations are unlikely to substantially affect high-threshold calcium channels or sodium channels to avoid subsequent actions of celecoxib. However, further studies will be necessary to definitively rule out this possibility.

In summary, the present results prove the concept that Kv7 channels are relevant potential mediators of the antinociceptive actions of celecoxib acting on the spinal cord. Although the results have to be interpreted cautiously due to the limitations of the present experimental approach, it is clear that celecoxib produces depression of the responses of sensory neurones and spinal nociceptive reflexes activating nonspecific targets. Although several nonspecific targets have been identified, the present results are consistent with an involvement of Kv7 channels in these actions of celecoxib. In conclusion, we suggest that celecoxib may combine its main peripheral actions as a blocker of COX enzymes, with activity at Kv7 channels located in spinal nociceptive circuits to achieve analgesia.

Authorship Contributions

Participated in research design: Lopez-Garcia, Rivera-Arconada.
Conducted experiments: Vicente-Baz.
Performed data analysis: Vicente-Baz, Rivera-Arconada.
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