Curcumin Acts as a Positive Allosteric Modulator of $\alpha_7$-Nicotinic Acetylcholine Receptors and Reverses Nociception in Mouse Models of Inflammatory Pain

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ABSTRACT

Effects of curcumin, a major ingredient of turmeric, were tested on the function of the $\alpha_7$-subunit of the human nicotinic acetylcholine ($\alpha_7$-nACh) receptor expressed in Xenopus oocytes and on nociception in mouse models of tonic and visceral pain. Curcumin caused a significant potentiation of currents induced by acetylcholine (ACh; 100 \(\mu\)M) with an EC\(_{50}\) value of 0.2 \(\mu\)M. The effect of curcumin was not dependent on the activation of G-proteins and protein kinases and did not involve Ca\(^{2+}\)-dependent Cl\(^-\) channels expressed endogenously in oocytes. Importantly, the extent of curcumin potentiation was enhanced significantly by decreasing ACh concentrations. Curcumin did not alter specific binding of \(^{[125]I}\)\(\alpha\)-bungarotoxin. In addition, curcumin attenuated nociceptive behavior in both tonic and visceral pain models without affecting motor and locomotor activity and without producing tolerance. Pharmacological and genetic approaches revealed that the antinociceptive effect of curcumin was mediated by $\alpha_7$-nACh receptors. Curcumin potentiated the antinociceptive effects of the $\alpha_7$-nACh receptor agonist \(N\)-(3R)-1-azabicyclo[2.2.2]oct-3-yl-4-chlorobenzamide (PNU282987). Collectively, our results indicate that curcumin is a positive allosteric modulator of $\alpha_7$-nACh receptor and reverses nociception in mouse models of tonic and visceral pain.

Introduction

Turmeric, the rhizome of Curcuma longa L., has been used since ancient times as a spice, coloring, flavoring, and traditional medicine. Curcumin, a polyphenolic compound isolated from turmeric, has been shown to exhibit a wide range of pharmacological activities, including anti-inflammatory, anti-cancer, antioxidant, antiatherosclerotic, antimicrobial, and wound-healing effects (Kunnakkara et al., 2017; Milani et al., 2017). Notably, in recent years, curcumin has been demonstrated to have beneficial effects in cognitive deficits and neurodegenerative disorders, such as Alzheimer and Parkinson diseases (Ji and Shen, 2014; Morales et al., 2014; Goozee et al., 2016; Spagnuolo et al., 2016). Although therapeutic effects of curcumin on a wide range of pathologic conditions have been reported, the precise mechanisms of these actions are poorly understood.

These diverse pharmacological activities of curcumin are based on its complex molecular structure and chemical features, as well as its ability to interact with multiple signaling molecules (Zhang et al., 2014). Many biologic molecules have been identified as targets of curcumin, including transcription factors, growth factors, inflammatory cytokines, protein kinases, enzymes, and ion channels (Zhou et al., 2011; Zhang et al., 2014). Many biologic molecules have been identified as targets of curcumin, including transcription factors, growth factors, inflammatory cytokines, protein kinases, enzymes, and ion channels (Zhou et al., 2011; Zhang et al., 2014). Kunnakkara et al., 2017; Milani et al., 2017).

Nicotinic acetylcholine (nACh) receptors are important members of the ligand-gated ion channel family that includes...
GABA<sub>B</sub> and 5-HT<sub>3</sub> receptors. Nicotinic receptors are divided into two groups: muscle nicotinic receptors, which are found at the skeletal muscle junction where they mediate neuromuscular transmission, and neuronal nicotinic receptors, which are found throughout the peripheral and central nervous systems where they have roles in fast synaptic transmission and in the modulation of transmitter release (Hogg et al., 2003; Albuquerque et al., 2009). The homomeric α<sub>7</sub>-nACh receptor subtype is abundantly expressed in the central nervous system and periphery (Albuquerque et al., 2009). Importantly, α<sub>7</sub>-nACh receptors, which have considerably high permeability to Ca<sup>2+</sup>, have been shown to be located on both glutamatergic and GABAergic nerve terminals, suggesting that both the excitatory and the inhibitory components of synaptic transmission can be modulated by the activity of these receptors (Hogg et al., 2003; Albuquerque et al., 2009). In fact, neuronal α<sub>7</sub>-nACh receptors are recognized targets for drug development in several preclinical models of cognitive and neurodegenerative disorders (Thomsen et al., 2010; Hone and McIntosh, 2017) and, more recently, in early human studies (Gee et al., 2017). Along with their well documented roles in cognition, activation of α<sub>7</sub>-nACh receptors produces analgesic effects in laboratory animal and human studies (Umana et al., 2013; Bagdas et al., 2017; Hone and McIntosh, 2017). Interestingly, curcumin also has been shown to attenuate pain and inflammation in various animal studies (Mittal et al., 2013; Bagdas et al., 2017; Hone and McIntosh, 2017). These findings may indirectly suggest the involvement of a common target between agonists of α<sub>7</sub>-nACh receptors and curcumin.

In the present study, we investigated the effects of curcumin on human α<sub>7</sub>-nACh receptors expressed in Xenopus oocytes and tested its effects on nociception in mouse models of tonic and visceral pain. Specifically, we provide evidence indicating that curcumin acts as a positive allosteric modulator of α<sub>7</sub>-nACh receptors in vitro and in vivo conditions.

**Materials and Methods**

**Recordings from Oocytes**

Mature female *Xenopus laevis* frogs were purchased from Xenopus Express (Haute-Loire, France), housed in dechlorinated tap water at 19–21°C with a 12/12-hour light/dark cycle, and fed food pellets supplied by Xenopus Express. The procedures followed in this study were in accordance with the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (Bethesda, MD) and approved by the Institutional Animal Care and Use Committee at the College of Medicine and Health Sciences, United Arab Emirates University. Clusters of oocytes were removed surgically under benzocaine (Sigma-Aldrich, St. Louis, MO) local anesthesia (0.03% w/v), and individual oocytes were dissected away manually in a solution containing (in millimolar) NaCl, 88; KCl, 1; NaHCO<sub>3</sub>, 2.4; MgSO<sub>4</sub>, 0.8; and HEPES, 10 (pH 7.5) as described previously (Brauneis et al., 1996; Oz et al., 2004a). Dissected oocytes were then stored for 2–7 days in modified Barth’s solution containing (in millimolar) NaCl, 88; KCl, 1; NaHCO<sub>3</sub>, 2.4; CaCl<sub>2</sub>, 2; MgSO<sub>4</sub>, 0.8; and HEPES, 10 (pH 7.5) supplemented with 2 mM sodium pyruvate, 10,000 IU/μl penicillin, 10 mg/ml streptomycin, 50 μg/ml gentamicin, and 0.5 mM theophylline. In brief, oocytes were placed in a 0.2-ml recording chamber and superfused at a rate of 3–4 ml/min. Under these conditions, solution exchange time was less than 100 ms. The bathing solution contained (in millimolar) NaCl, 96; KCl, 2; CaCl<sub>2</sub>, 1.8; MgCl<sub>2</sub>, 1, and HEPES, 5 (pH 7.5). The cells were impaled with two glass microelectrodes filled with a 3 M KCl (0.5–2 MΩ). The oocytes were routinely voltage clamped at a holding potential of −70 mV using a GeneClamp-500 amplifier (Axon Instruments Inc., Burlingame, CA), and current responses were recorded and stored digitally for further analysis.

Drugs were applied by gravity flow via a micropipette positioned about 2 mm from the oocyte. Some of the compounds were applied externally by addition to the superfusate (flow rate of 3–4 ml/min). Acetylcholine, GDPβS, methyllycaconitine (MLA), α<sub>7</sub>-bungarotoxin, pertussis toxin (PTX), and all chemicals used were obtained from Sigma-Aldrich. Procedures for the injections of NEM, 1,2-bis(oxyethylene)N,N,N',N'-tetraacetic acid (BAPTA), and γ-aminobutyric acid (GABA) were performed at 4°C.

**Radioligand Binding Studies**

Oocytes were injected with 5 ng of human α<sub>7</sub>-nACh receptor cRNA, and the functional expression of the receptors was tested by electrophysiology on day 3. Isolation of oocyte membranes was carried out by modification of a method described previously (Oz et al., 2004b; Mahgoub et al., 2013). In brief, oocytes (200–300 oocytes per assay) were suspended (approximately 20 μl/oocyte) in a homogenization buffer containing 10 mM HEPES, 1 mM EDTA, 0.1 mM phenylmethylsulfonyl fluoride (PMSF), 0.02% Na<sub>3</sub>N<sub>2</sub>, and 50 μg/ml bacitracin (pH 7.4) at 4°C on ice and homogenized using a motorized Teflon homogenizer (six strokes, 15 seconds each at high speed). The homogenate was centrifuged for 10 minutes at 800 g. The supernatant was collected, and the pellet was resuspended in homogenization buffer and centrifuged at 800 g for 10 minutes. Supernatants were then combined and centrifuged for 1 hour at 36,000 g. The membrane pellet was resuspended in homogenization buffer and used for the binding studies.

Binding assays were performed in 500 μl of 10 mM HEPES (pH 7.4) containing 50 μl of 50 μM of [³²P]α-bungarotoxin (2200 Ci/mmol; PerkinElmer, Inc., Waltham, MA). Nonspecific binding was determined using 10 μM α-bungarotoxin. Oocyte membranes were incubated with [³²P]α-bungarotoxin in the absence and presence of drugs for 1 hour at room temperature (22–24°C). The radioligand was separated by rapid filtration onto GF/C filters presoaked in 0.2% polyethyleneimine. Filters were then washed with two 5-ml washes of ice-cold HEPES buffer, and the radioactivity was determined by counting samples in a Beckman Gamma-300 γ-counter (Beckman Coulter, Inc., Indianapolis, IN).

**In Vivo Studies**

Male ICR adult (8–10 weeks of age) mice were obtained from Harlan Laboratories (Indianapolis, IN). Mice on C57BL/6J background null for the α<sub>7</sub>-subunits (B6.129S7-Chrna7tm1Bay/J; Jackson Laboratory, Bar Harbor, ME) and their wild-type (WT) littermates were housed in a 21°C animal care facility at Virginia Commonwealth University. For all experiments, mice were backcrossed for ≥9–10 generations. Mutant and WT mice were obtained by crossing homozygote mice. This breeding scheme controlled for any irregularities that might occur with crossing solely mutant animals. Mice were housed in a 21°C
humidity-controlled Association for Accreditation and A
Laboratory Animal Care—approved animal care facility. They were housed in groups of four and had free access to food and water. The rooms were on a 12-hour light/dark cycle (lights on at 7 AM). All experiments were performed during the light cycle (between 7 AM and 7 PM), and the study was approved by the Institutional Animal Care and Use Committee of Virginia Commonwealth University. All studies were carried out in accordance with the National Institutes of Health’s Guide for the Care and Use of Laboratory Animals. Animals were sacrificed via CO2 following by cervical dislocation after the experiments were finished, unless noted otherwise.

**Drugs**

MLA citrate was purchased from RBI (Natick, MA). PNU282987 [N-(3R)-1-azacyclo[2.2.2]oct-3-yl-4-chlorobenzamide] was obtained from the National Institute on Drug Abuse supply program (Bethesda, MD). [9S-(9a,10b,11b,13a)]-N-(2,3,10,11,12,13-Hexahydro-10-methoxy-9-methyl-1-oxo-9,13-epoxy-1H-9H-diindolo[1,2,3-g:h:3',2',1'-im]pyrrolo[3,4-j][1,7]benzodiazepin-11-yl)-N-methylbenzamide (PKC-412), 3-[1-(Dimethylamino)propyl]-5-methoxy-1H-indol-3-yl)-1H-pyrrole-2,5-dione (Go-6983), [9R,10S,12S]-2,3,9,10,11,12-Hexahydro-10-hydroxy-9-methyl-1-oxo-9,12-epoxy-1H-diindolo[1,2,3-fg:3',3',1'-dlpyrrolo[3,4-[1,6]benzodiazepine-10-carboxylic acid, hexyl ester (KT-5720), and 4-(2S)-2-(5-isouquinolinylsulfonyl)methylamino]-3-oxo-3-(4-phenyl-1-piperazinyl)propyl phenyl isouquinolinesulfonic acid ester (KN-62) were purchased from Tocris (Minneapolis, MN). They were dissolved in DMSO to 100 mM stock solutions. Curcumin was obtained from Santa Cruz Biotechnology, Inc. (Dallas, TX). In in vivo experiments, curcumin was dissolved in a mixture of 2:2:16 (2 volumes ethanol/2 volumes Emulphor-620 (Rhone-Poulenc, Inc., Princeton, NJ/18 volumes distilled water) and administered i.p. for systemic injections. Other drugs were dissolved in physiologic saline (0.9% sodium chloride) and injected s.c. at a total volume of 1 ml/100 g body weight, unless noted otherwise. All doses are expressed as the free base of the drug.

**Behavioral Assessments.**

**Formalin test.** The formalin test was carried out in an open Plexiglas cage (29 × 19 × 13 cm each). Mice were allowed to acclimate for 15 minutes in the test cage prior to injection. Each animal was injected intraplantarly with 20 μl of (2.5%) formalin to the right hindpaw. Mice were observed from 0 to 5 minutes (phase I) and 20–45 minutes (phase II) postformalin injection. The amount of time spent licking the injected paw was recorded with a digital stopwatch. Paw diameter (see **Measurement of Paw Edema**) was also measured before and 1 hour after formalin injection.

Curcumin (3, 10, and 30 mg/kg) or vehicle were injected in male ICR mice i.p. 45 minutes before formalin injection. For the subchronic curcumin administration study, mice were given curcumin (30 mg/kg, i.p.) or vehicle for 6 days once daily and were challenged with curcumin (30 mg/kg, i.p.) on day 7 and tested in a formalin test 45 minutes after the injection. A vehicle control group, in which mice were exposed to saline, were also included.

For the antagonist studies, α7 nicotinic antagonist MLA (10 mg/kg) or vehicle (saline) was injected s.c. 10 minutes before the curcumin (30 mg/kg, i.p.) or vehicle administration. In a separate group, curcumin (30 mg/kg, i.p.) effects in the formalin test were measured in α7 WT and knockout (KO) mice. To test possible potentiation of antinociceptive effects of curcumin on PNU282987 in the formalin test, curcumin (3 mg/kg, i.p.) was injected 45 minutes before PNU282987 (0.1 mg/kg, s.c.) administration. The formalin test was conducted 10 minutes after the last injection.

**Measurement of paw edema.** The thickness of the formalin-treated paws was measured both before and after injections at the time points indicated earlier using a digital caliper (Traceable Calipers, Friendswood, TX). Data were recorded to the nearest ±0.01 mm and expressed as change in paw thickness (ΔPD = difference in the ipsilateral paw diameter before and after injection paw thickness).

Acetic acid–induced writhing test. For the measurement of acetic acid–induced nociceptive behavior, each mouse was placed in a Plexiglas box and allowed to acclimate for 20 minutes. Then the mouse was given an i.p. injection of acetic acid (1.2%) or saline and then returned to the box. Counting of the number of typical writhing behaviors started immediately after acetic acid administration, and the number of stretches (a stretch was operationally defined as a contraction of the abdomen followed by an extension of the hind limbs) was recorded in 10-minute bins for a total of 40 minutes. Experiments were carried out by injecting the mice with either vehicle or curcumin (30 mg/kg, i.p.), and 45 minutes later, they received acetic acid and were tested as described earlier. For the antagonist studies, α7-nACh receptor (α7-nAChR) antagonist MLA (10 mg/kg) was injected s.c. 15 minutes before the curcumin (30 mg/kg, i.p.). Forty-five minutes after curcumin, mice received acetic acid or vehicle (saline) injection.

Motor coordination. The effects of drugs on motor coordination were measured using the rotarod test (IITC Inc. Life Science, Woodland Hills, CA) as previously described (Freitas et al., 2013a, b). Mice were pretreated with either i.p. vehicle or curcumin (30 mg/kg, i.p.) 45 minutes before the test. Percentage impairment was calculated as follows: % impairment = (180 – test time)/(180 × 100).

**Statistical Analysis**

In oocyte experiments, average values were calculated as the mean ± S.E.M. Statistical significance was analyzed using Student’s t test or analysis of variance (ANOVA) as indicated. Concentration-response curves were obtained by fitting the data to the logistic equation:

$$ y = \frac{E_{\text{max}}}{1 + \left(\frac{x}{EC_{50}}\right)^n} $$

where x and y are concentration and response, respectively; E_{\text{max}} is the maximal response; EC_{50} is the half-maximal concentration; and n is the slope factor (apparent Hill coefficient).

In behavioral studies, the data obtained were analyzed using GraphPad software, version 6.0 (GraphPad Software, Inc., La Jolla, CA) and expressed as the mean ± S.E.M. Statistical analysis was done using the one-way or two-way ANOVA test, followed by the post hoc Tukey’s test. Unpaired student t test was used for spontaneous activity and motor coordination. P values <0.05 were considered significant.

**Results**

Effects of Curcumin on α7-nACh Receptors. Application of 100 μM ACh for 3–4 seconds activated fast inward currents that desensitized rapidly in oocytes injected with cRNA encoding the α7-subunit of human nACh receptor (Fig. 1A). In addition, ACh-induced currents were inhibited completely with 100 nM α-bungarotoxin (Supplemental Fig. 1, A and B), indicating that these currents are mediated by the activation of α7-nACh receptors. Bath application of curcumin (100 μM) for 5 minutes did not produce detectable currents in oocytes expressing α7-nACh receptors (n = 8 oocytes).

The effect of curcumin was tested on ion currents induced with ACh (100 μM). An effect of 10-minute curcumin (1 μM) application on α7-nACh receptor—mediated currents is shown in Fig. 1A. Time courses of effects of curcumin on vehicle (0.1% DMSO) applications on the maximal amplitudes of ACh-induced currents are presented in Fig. 1B. Curcumin caused
a significant potentiation of the current, which was partially reversed during a 10–15-minute washout. In the absence of curcumin, vehicle (0.1% DMSO) alone did not alter the amplitude of the ACh-induced current, further suggesting that curcumin acts on nACh receptors (Fig. 1B, controls vs. curcumin treatment group at 10 minutes of exposure, ANOVA, n = 5–7; P < 0.05).

The potentiating effect of curcumin was significantly dependent on the application mode. For example, without preincubation, coapplication of curcumin (1 μM) and ACh (100 μM) did not alter the amplitudes of maximal currents (Fig. 1C). However, when oocytes were preincubated with curcumin, the drug was found to potentiate maximal ACh-induced currents in a time-dependent manner, reaching a maximal level within 5 minutes with a half-time (τ1/2) of 1.6 minutes (Fig. 1C). Since the magnitude of the curcumin effect was time-dependent, 10-minute curcumin application was used to equilibrate conditions. Curcumin was found to upregulate the function of α7-nACh receptor in a concentration-dependent manner with EC50 and slope values of 0.21 ± 0.14 μM and 1.6, respectively (Fig. 1D).

As shown in Fig. 1, B and C, regulation of α7-nACh receptor function by curcumin occurs gradually, reaching steady-state levels within a few minutes of curcumin application. Therefore, it is possible that activation of second messenger pathways by G-protein-coupled receptors (Pérez-Lara et al., 2011; Liu et al., 2013; Yang et al., 2015) is involved in curcumin regulation of α7-nACh receptors. Thus, we investigated the effects of pretreatments with NEM (10 mM, 50 nl, 30-minute preincubation time), a sulfhydryl-alkylating agent that blocks G-protein-effector interactions by alkylating α-subunits of PTX-sensitive GTP binding protein (Oz and Renaud, 2002), and GDPβS (10 mM, 50 nl, 30-minute preincubation time), an agent that inhibits binding of GTP to the α-subunit of G-proteins (Oz et al., 1998). Treatments with NEM and GDPβS did not alter the extent of curcumin potentiation of α7-nACh receptor (Fig. 2A).

Similarly, pretreatment with PTX (5 μg/ml, 50 nl, 30-minute preincubation time), a toxin that inhibits the α-subunit of Gi/o proteins, did not reverse the potentiating effect of curcumin (Supplemental Fig. 2). In addition, extent of curcumin-induced potentiation of currents activated by a low (30 μM) concentration of ACh was also not altered in the presence of NEM and GDPβS curcumin (Supplemental Fig. 2)

Activation of α7-nACh receptors allows sufficient Ca2+ entry to activate endogenous Ca2+-dependent Cl− channels in Xenopus oocytes (Sands et al., 1993; Uteshev, 2012). Therefore, it was important to determine whether the effect of curcumin was exerted on nACh receptor–mediated currents or on Cl− currents induced by Ca2+ entry. For this reason, we injected the Ca2+-chelator BAPTA into oocytes and replaced extracellular Ca2+ with Ba2+ since Ba2+ can pass through α7-nACh receptors but causes less activation of Ca2+-dependent Cl− channels (Sands et al., 1993). Under these conditions, we tested the effect of curcumin in a solution containing 2 mM Ba2+ in BAPTA-injected oocytes. Curcumin (1 μM) produced the same level of potentiation (195 ± 18 in controls vs. 210 ± 22 in BAPTA-injected oocytes; ANOVA, P > 0.05; n = 6 to 7) on ACh-induced currents in BAPTA-injected oocytes when currents were recorded in Ca2+-free solution containing 2 mM Ba2+ (Fig. 2B). It is important to mention that in the oocyte expression system, curcumin-induced changes in nicotinic receptor–mediated currents can be attributable to Ca2+-activated Cl− channels and concomitant alterations in the holding currents. However, in control experiments, curcumin (100 μM for 10 minutes) did not change the magnitudes of holding currents in oocytes voltage clamped at ~70 mV (n = 7), indicating that intracellular Ca2+ levels were not altered by curcumin.

We also investigated the involvement protein kinases A and C, and Ca2+-calmodulin–dependent kinase (CaM-kinase) in curcumin potentiation of α7-nACh receptors. For this purpose, the effects of curcumin were tested in oocytes pretreated with PKC-412 (nonspecific kinase inhibitor, 10 μM for 30 minutes...
pretreatment), Go-6983 (specific protein kinase C inhibitor, 10 μM for 30 minutes), KT-5720 (specific protein kinase A inhibitor, 10 μM for 30 minutes), and KN-62 (specific inhibitor of CaM-kinase II, 50 μM for 30 minutes). Curcumin continued to upregulate nicotinic receptor–mediated currents in oocytes pretreated with kinase inhibitors (Fig. 2, C and D). Similarly, potentiating effects of curcumin remained unaltered by inhibitors of protein kinases A and C and CaM-kinase at low (30 μM) ACh concentrations (Supplemental Fig. 2).

In the next series of experiments, we examined if the extent of curcumin potentiation of the α7-nACh receptor–mediated current is altered by membrane potential. As indicated in Fig. 2E, the potentiation of ACh (30 μM)-induced currents by curcumin (1 μM) does not appear to be voltage-dependent. The extent of curcumin potentiation was similar at all tested membrane potentials from −100 to +40 mV. Evaluation of the current-voltage relationship (Fig. 2F) indicates that the extent of potentiation by curcumin does not change significantly at different test potentials (P > 0.05, n = 7, ANOVA).

In the next series of experiments, we attempted to test the effects of curcumin at different ACh concentrations. Traces of low ACh (10 μM)-induced currents after 10-minute treatment with 1 μM curcumin are presented in Fig. 3A. At low ACh (10 μM) concentrations, curcumin caused an approximately 11- to 12-fold increase of ACh-induced currents with an EC50 of 58 nM (Fig. 3B). Further experiments indicated that the extent of curcumin potentiation decreased significantly with increasing concentrations of ACh (Fig. 3C). Concentration-response curves for ACh in the absence and presence of 1 μM curcumin are presented in Fig. 3D. In the presence of 1 μM curcumin, the maximal ACh response increased by 60%–70% of controls (n = 6–8). In the absence and presence of curcumin, the EC50 values for ACh were 107 ± 16 μM (open circles) and 18 ± 1.9 μM (filled circles), respectively (paired t test; n = 8; P < 0.01). These findings

**Fig. 2.** Effects of curcumin on α7-nACh receptor are not mediated by G-proteins and protein kinases, and are not dependent on intracellular Ca2+ levels and changes in membrane potential. (A) Bar presentation of the effects of 1 μM curcumin application (10 minutes) on the maximal amplitudes of ACh (100 μM)-induced currents in oocytes injected with 50 nl of distilled water, controls (n = 16), or NEM (10 mM, 50 nl, n = 8) and GDPβS (10 mM, 50 nl, n = 7) 30 minutes before recordings. (B) Bar presentation of the effects of 1 μM curcumin application (10 minutes) on the maximal amplitudes of ACh (100 μM)-induced currents in oocytes injected with 50 nl distilled water, controls (n = 5), or BAPTA (200 mM, 50 nl, n = 7). (C) Bar presentation of the effects of 1 μM curcumin on α7-nACh receptor–mediated currents in oocytes pretreated with vehicle (0.01% DMSO, n = 5) or PKC-412 (PKC; 10 μM, 30-minute pretreatment, n = 7), or Go-6983 (GO; 10 μM, 30-minute pretreatment, n = 6). (D) Bar presentation of the effects of 1 μM curcumin on α7-nACh receptor–mediated currents in oocytes pretreated with vehicle (0.01% DMSO, n = 7) or KT-5720 (KT; 10 μM, 30-minute pretreatment, n = 7) and KN-62 (KN; 50 μM, 30-minute pretreatment, n = 6). (E) Current-voltage relationships of acetylcholine-activated currents in the absence and presence of curcumin. Normalized currents activated by 30 μM ACh before (control, filled circles) and after 10-minute treatment with 1 μM curcumin (open circles). Each data point presents the normalized means and S.E.M. of seven experiments. (F) Quantitative presentation of the effect of curcumin as percentage of controls at different voltages.
suggest that curcumin caused a significant decrease of $\alpha_7$-nACh receptor desensitization. We further investigated whether curcumin can convert $\alpha_7$-nACh receptors that were already desensitized by a concentration of an agonist back to conducting state. We tested the effect of a bath application of curcumin (10 $\mu$M) on the $\alpha_7$-nACh receptors that was desensitized by 100 $\mu$M nicotine application for 25–30 seconds ($n = 6$). As illustrated in Fig. 4C, subsequent addition of curcumin resulted in activation of a sustained inward current that was reversed during washout.

$[^{125}I]$-bungarotoxin competes with ACh, an endogenous activator of $\alpha_7$-nACh receptors, by binding to the ACh binding site on the receptor (Albuquerque et al., 2009). For this reason, the effect of curcumin was investigated on the specific binding of $[^{125}I]$-bungarotoxin. Equilibrium curves for the binding of $[^{125}I]$-bungarotoxin in the presence and absence (controls) of curcumin are presented in Fig. 5A. Maximum binding activities ($B_{\text{max}}$) of $[^{125}I]$-bungarotoxin were 3.61 ± 0.37 and 3.47 ± 0.41 pM/mg (means ± S.E.M.) for controls and curcumin-treated preparations, respectively. The apparent affinities ($K_D$) of the receptor for $[^{125}I]$-bungarotoxin were 0.87 ± 0.26 and 0.67 ± 0.23 pM for controls and curcumin, respectively. There was no statistically significant difference between controls and curcumin-treated groups with respect to $K_D$ and $B_{\text{max}}$ values ($P > 0.05$, ANOVA, $n = 6–7$), suggesting that curcumin does not compete with $\alpha$-bungarotoxin at the same binding site. Curcumin up to a concentration of 100 $\mu$M did not cause a significant change on the specific binding of $[^{125}I]$-bungarotoxin (Fig. 5B). The effect of curcumin on the functional properties of other neuronal nACh receptor subtypes was also examined. Application of curcumin (10 $\mu$M for 10 minutes) did not cause alterations of ACh (100 $\mu$M)-induced currents mediated by different subtype combinations of nicotinic receptors expressed in oocytes (Fig. 5C). Similarly, curcumin (10 $\mu$M for 10 minutes) did not cause significant changes on the amplitudes of currents mediated by 5-HT$_3$A (1 $\mu$M 5-HT) subunit and glycine receptors (30 $\mu$M glycine; mediated by $\alpha_1\beta_1$, $\alpha_2\beta_1$, and $\alpha_3\beta_1$ subunit combinations) (Fig. 5D).

**Figs. 3 and 4.** (A) Records of currents activated by ACh (10 $\mu$M) in control conditions (left), after 10-minute pre-treatment with curcumin (1 $\mu$M) and coapplication of 1 $\mu$M curcumin and ACh (middle), and 10-minute washout (right). (B) Concentration-dependent effect of curcumin on $\alpha_7$-nACh receptors activated by low acetylcholine concentration. Each data point represents the mean ± S.E.M. of six to eight oocytes. The curve is the best fit of the data to the logistic equation described in the Materials and Methods section. (C) Bar presentation of the effect of curcumin at different acetylcholine concentrations. Bars represent the means ± S.E.M. of five to eight experiments. (D) Effect of curcumin on the ACh concentration-response relationship. Oocytes were voltage clamped at −70 mV, and currents were activated by applying ACh (1 $\mu$M to 3 mM). Oocytes were exposed to 10 $\mu$M curcumin for 10 minutes, and ACh was reapplied. Paired concentration-response curves were constructed and responses normalized to maximal response under control conditions. Data points obtained before (control) and after 10-minute treatment with curcumin (10 $\mu$M) are indicated by filled and open circles, respectively. Each data point presents the normalized means and S.E.M. of 7–11 experiments.

Effects of Curcumin Treatment in Formalin-Induced Pain Responses. To determine the effect of acute curcumin treatment in the formalin-induced pain model, mice were given an i.p. injection of curcumin (3, 10, and 30 mg/kg) or...
vehicle 45 minutes before intraplantar formalin injection. Ordinary two-way ANOVA revealed significant effects for phase of formalin test \( F_{\text{phase}(1,40)} = 14.54, P < 0.001 \) and dose of curcumin \( F_{\text{dose}(3,40)} = 11.04, P < 0.001 \). As seen in Fig. 6A, curcumin failed to show a significant antinociceptive effect in phase I of the test (Tukey post hoc, \( P > 0.05 \)). However, it dose-dependently attenuated nociceptive behavior in phase II (Tukey post hoc, \( P < 0.001 \)).

**Role of \( \alpha_7 \)-nACh Receptors in the Effects of Curcumin.** We explored the possible role of \( \alpha_7 \)-nAChRs in the effect of curcumin in the formalin test. We first investigated if the \( \alpha_7 \)-nAChR antagonist MLA would block curcumin’s effects in phase II of the test. Ordinary two-way ANOVA revealed significant effects for phase of formalin test \( F_{\text{phase}(1,40)} = 37.61, P < 0.001 \) and treatment \( F_{\text{dose}(3,40)} = 13.29, P < 0.001 \; \text{(Fig. 6B).} \) A post hoc Tukey test showed that while the \( \alpha_7 \)-nACh antagonist MLA (10 mg/kg, s.c.) given alone did not alter formalin-induced pain responses \( (P < 0.05) \), it completely blocked the antinociceptive effect of curcumin \( (P < 0.001) \). We then tested curcumin in the \( \alpha_7 \) WT and KO mice. Interestingly, curcumin significantly reduced the phase I nociceptive responses in \( \alpha_7 \) WT mice. Surprisingly, the antinociceptive effect of curcumin was also preserved in transgenic KO mice. On the other hand, whereas curcumin reduced formalin-induced paw licking in WT mice, the effect vanished in \( \alpha_7 \) KO mice in the phase II of the test \( F_{\text{phase}(1,44)} = 215.8, P < 0.001 \) and genotype \( F_{\text{genotype}(3,44)} = 75.87, P < 0.001 \; \text{(Fig. 6C).} \)

**Subchronic Curcumin Treatment Attenuates Formalin-Induced Pain Responses, without Producing Tolerance.** In the next experiment, we investigated whether the antinoceptive effects of curcumin in the formalin test would undergo tolerance after subchronic administration. Curcumin produced significant effects in phase I \( \text{(one-way ANOVA, Tukey post hoc,} F_{(2,15)} = 4.896, P < 0.05 \) and phase 2 \( F_{(2,15)} = 18.24, P < 0.001 \; \text{(Fig. 7A).} \) Subchronic curcumin attenuated pain responses at both phase I and phase II without producing tolerance. Moreover, curcumin also reduced the formalin-induced edema without producing tolerance \( \text{(one-way ANOVA, Tukey post hoc,} F_{(3,15)} = 26.74, P < 0.001 \) (Fig. 7B).

**Potentiation of the Antinociceptive Effects of the \( \alpha_7 \)-Nicotinic Agonist PNU282987 by Curcumin.** We determined the effects of curcumin on PNU282987-evoked antinociceptive effects in the formalin test. Data were analyzed using two-way ordinary ANOVA followed by Tukey post hoc test. Analysis revealed significant effects for phase of formalin test \( F_{\text{phase}(1,40)} = 47.55, P < 0.001 \) and treatment \( F_{\text{dose(3,40)}} = 12.51, P < 0.001 \; \text{(Fig. 8).} \) Pretreatment with a low dose of curcumin (3 mg/kg, i.p.) or PNU282987 (0.1 mg/kg, s.c.) failed to attenuate formalin-induced pain responses when tested alone \( (P > 0.05) \). However, combination of curcumin and PNU282987 significantly reversed pain behavior in phase II \( (P < 0.001) \) but not phase I of the test \( (P > 0.05) \).

**Effects of Curcumin Treatment in Acetic Acid-Induced Stretching.** Acetic acid significantly evoked stretching behavior as a nociceptive behavior outcome \( F_{\text{acetic acid}(1,56)} = 813.1, P < 0.0001 \; \text{(Fig. 9).} \) Curcumin (30 mg/kg) significantly reduced acetic acid–induced nociceptive stretching behaviors, and MLA blocked this antinociceptive effect of curcumin \( F_{(3,56)} = 20.79, P < 0.0001 \; \text{(Fig. 9).} \)

**Effects of Curcumin on Motor Activity and Coordination.** As seen in Table 1, curcumin at a dose of 30 mg/kg (i.p.), the highest active dose used in our study, did not affect 1) motor performance or 2) spontaneous activity of mice after acute injection \( (t \text{ test,} \ t = 0.2988, df = 8, P > 0.05 \) and \( t = 0.5841, df = 10, P > 0.05 \), respectively).

**Discussion**

In the present study, using electrophysiological, biochemical, and behavioral methods, we provided evidence that...
Curcumin upregulates the function of human α7-nACh receptors expressed in Xenopus oocytes and reverses nociception in mouse models of tonic and visceral pain through an α7-nACh receptor mechanism. The enhancement of α7-nACh receptor function by curcumin is reversible and occurs in a time- and concentration-dependent manner, but is independent of G-protein activation, protein kinase activity, intracellular Ca2+ levels, and membrane potential.

A relatively slow time course of curcumin effect and the results of earlier studies on curcumin modulation of various second messenger pathways and kinases (Mahmoud, 2007; Takikawa et al., 2013) suggest that activation of G-protein-coupled receptors and/or kinase-mediated phosphorylation is involved in curcumin-induced upregulation of α7-nACh receptors. However, neither treatments with established kinase inhibitors nor pharmacological disruption of G-protein activity reversed curcumin potentiation of α7-nACh receptors, suggesting that curcumin acts directly on ion channel-receptor complex.

In Xenopus oocytes, activation of α7-nACh receptors, due to their high Ca2+ permeability, allows sufficient Ca2+ entry to activate endogenous Ca2+-dependent Cl− channels (Sands et al., 1993; Hartzell et al., 2005). Therefore, the direct actions of curcumin on Ca2+-dependent Cl− channels may contribute to the observed effects of curcumin on ACh-activated currents in this expression system. In oocytes injected with BAPTA and recorded in a solution containing 2 mM Ba2+, curcumin continued to potentiate α7-nACh receptor-mediated ion currents, suggesting that Ca2+-dependent Cl− channels were not involved in curcumin potentiation of nicotinic responses. In addition, the reversal potential in solutions containing Ba2+ was not altered in the presence of curcumin, suggesting that the potentiation by curcumin is not due to alterations in the Ca2+ permeability of the α7-nACh receptor-channel complex.

Curcumin has been reported to alter Ca2+ homeostasis in various cell types (Dyer et al., 2002; Ibrahim et al., 2011; Wang et al., 2012; Moustapha et al., 2015). However, in Xenopus oocytes, Ca2+-activated Cl− channels are highly sensitive to intracellular Ca2+ levels [K0 of Ca2+-activated Cl− channels for Ca2+ is less than 1 μM; for a review, Hartzell et al., (2005)], and alterations in intracellular Ca2+ levels would be reflected by changes in the holding current under voltage- clamp conditions. In our experiments, application of curcumin (1–100 μM) did not cause alterations in baseline or holding currents, and curcumin continued to potentiate nicotinic receptors after the chelation of intracellular Ca2+ by BAPTA, suggesting that alterations in intracellular Ca2+ concentrations does not play a role in curcumin’s effect on nicotinic receptors.

Previous studies have demonstrated that curcumin acts on several integral membrane proteins, including enzymes, transporters, and ion channels (Zhang et al., 2014; Li et al., 2017). T-type Ca2+ channels in bovine adrenal cells (Enyeart et al., 2009; IC50 = 10–20 μM), L-type Ca2+ channels in hippocampal neurons (Liu et al., 2013; IC50 = 10 μM), and TREK-1 K+ channels (Enyeart et al., 2008; IC50 = 0.9 μM) were not sensitive to curcumin in bovine adrenal cells, Kv1.4 K+ channels (Liu et al., 2006) in bovine adrenal cells, Kv1.4 K+ channels (Lian et al., 2013; IC50 = 4.2 μM) in human T-lymphocytes, ERG K+ channels (Hu et al., 2012, IC50 = 5.5 μM; Choi et al., 2013, IC50 = 10.6 μM; Bandeirall et al., 2011, IC50 = 2 μM), and K+ channels in rabbit coronary arterial smooth muscle cells (Hong et al., 2013; IC50 = 1.1 μM). In addition to voltage-dependent conductances, curcumin has also been shown to act on transient-receptor potential potential receptors (Yeom et al., 2010; Zhi et al., 2013).

In this study, curcumin was applied in the concentration range of 1 nM to 100 μM, and it was found that it can enhance the effects of ACh on the function of α7-nACh receptors in a concentration-dependent manner, with EC50 values ranging from 58 nM to several micromolars. The concentration of curcumin in plasma and its ability to pass the blood-brain barrier following oral and intravenous administration have been studied previously (Anand et al., 2007).
concentration of 1.35 μg/ml (3.5 μM) was attained (Shoba et al., 1998). Since curcumin is a highly lipophilic compound with a logP (octanol–water partition coefficient) value of 3.3 (https://pubchem.ncbi.nlm.nih.gov/compound/curcumin#sec-5.Top), its membrane concentration is expected to be considerably higher than blood levels. Therefore, the functional modulation of α7-nACh receptors demonstrated in this study can be pharmacologically relevant.

As was mentioned earlier, the roles of G-proteins, kinases, and intracellular Ca\(^{2+}\) levels in curcumin actions were excluded in our functional and pharmacological studies. Radioligand binding experiments suggested that curcumin does not interact with ACh binding to receptors. Further analysis of the curcumin effect indicated that curcumin significantly (more than 3-fold) decreased desensitization of the receptor and converted the already-desensitized nACh receptor back to conducting state (Fig. 4C). Importantly, binding of α-bungarotoxin, a competitive antagonist of ACh, was not altered in the presence of curcumin, suggesting that curcumin does not interact with the ACh binding site in the receptor. It is plausible that curcumin acts as an allosteric modulator for various receptors and ion channels at the lipid membrane, accounting for some of its pharmacological actions in animal studies (Zhang et al., 2014). Allosteric modulators alter the functional properties of ligand-gated ion channels by interacting with sites that are topographically distinct from the ligand binding sites [for a review, Onaran and Costa (2009)]. Two different types of positive allosteric modulator (PAM) have been postulated (Uteshev, 2014; Chatzidaki and Millar, 2015). Whereas type I enhances agonist-induced currents without affecting macroscopic current kinetics, type II PAMs also delay desensitization and reactivate desensitized receptors. Results of our experiments indicate that curcumin significantly decreases desensitization (Fig. 4, A and B) and reactivates completely desensitized nicotinic receptors (Fig. 4C), suggesting that curcumin acts as a type II PAM.

It is likely that curcumin, a highly lipophilic agent, first dissolves into the lipid membrane and then diffuses into a nonannular lipid space to potentiate the function of the ion channel-receptor complex. Consistent with this idea, the effect of curcumin on α7-nACh receptor reached a maximal level within 5–10 minutes of application, suggesting that the binding site(s) for these allosteric modifiers is located inside the lipid membrane and requires a relatively slow (in minutes) time course to modulate the function of the receptor. It is likely that these hydrophobic agents affect the energy requirements for gating-related conformational changes in ligand-gated ion channels (Spivak et al., 2007).

Our in vitro data suggested that curcumin is a selective PAM of α7-nACh receptors. We therefore tested this possibility after systemic in vivo administration in mice. α7-nAChRs are present in supraspinal and spinal pain-transmission pathways as well as on immune and nonimmune cytokine-producing cells, such as macrophages and microglia. α7-nAChR expressed on immune cells are involved in the initiation, maintenance, and resolution of inflammation, and

Fig. 7. The antinociceptive effects of subchronic curcumin in the formalin test. (A) The effect of subchronic curcumin administration on formalin-induced pain behavior was measured. Mice were treated with curcumin (30 mg/kg, i.p.) or vehicle for 6 days once daily and were challenged with curcumin (30 mg/kg, i.p.) on day 7 and tested in a formalin (2.5%) test. A vehicle control group, in which mice were exposed to 7 days of vehicle, was also included. (B) The antiedema effect of formalin injection of curcumin, measured by the difference in the ipsilateral paw diameter before and after formalin injection (ΔPD), was assessed 1 hour after formalin injection. Data are given as the mean ± S.E.M. of six animals for each group. *P < 0.05, significantly different from its vehicle group.

Fig. 8. Antinociceptive effects of curcumin plus PNU282987 combination in the formalin test. Curcumin (3 mg/kg, i.p.) was injected 45 minutes before PNU282987 (0.1 mg/kg, s.c.) injection. The formalin test was conducted 10 minutes after PNU282987 administration in ICR mice. Data are given as the mean ± S.E.M. of six animals *P < 0.05, significantly different from vehicle (veh) group.
modulate inflammation process. In addition, α7-nAChRs expressed on microglia regulate inflammatory factors (Bagdas et al., 2017). Since α7-nACh receptor PAMs have been reported to be active in animal models of tonic and chronic pain (Munro et al., 2012; Freitas et al., 2013a,b; Bagdas et al., 2015), we evaluated the antinociceptive and anti-inflammatory effects of curcumin in the mouse formalin test, a model of tonic and persistent pain (Hunskaar and Hole, 1987). The formalin test consists of two distinct phases. The first phase (immediately after formalin injection) seems to be caused by the direct effect of formalin on sensory C-fibers. The second phase (starting later after formalin injection), known as the inflammatory phase, is associated with the development of a delayed inflammatory response and spinal dorsal horn sensitization (Abbott et al., 1995; Davidson and Carlton, 1998). In the outbred ICR mice, curcumin attenuated pain behaviors dose-dependently in the late (inflammatory) but not the early phase of the formalin test. Importantly, no changes were seen in motor locomotion or coordination with antinociceptive doses of curcumin in mice (Table 1). Using both pharmacological (i.e., the selective α7-nACh antagonist MLA) and genetic approaches (i.e., α7 KO mice), we confirmed that curcumin’s effect in the late phase of formalin is mediated by α7-nACh receptors. The effects of curcumin in the early phase of formalin is strain-dependent since, in contrast to ICR mice, curcumin at a dose of 30 mg/kg significantly attenuated pain behaviors in phase I in the α7 WT (C57BL/6J strain) mice. Interestingly, the effects of curcumin in phase I were not eliminated in the α7 KO mice, suggesting the involvement of non-α7-nACh receptor mechanisms in curcumin’s effects.

Furthermore, our results show that tolerance did not develop following subchronic exposure to the antinociceptive and anti-inflammatory effects of curcumin. PAMs are compounds that facilitate endogenous neurotransmission and/or enhance the efficacy and potency of exogenous agonists, without directly stimulating the agonist binding sites. Supporting this possibility, curcumin enhanced the effects of subactive dose of PNU282987, a full α7-nACh receptor agonist, in the formalin test.

Our vivo data extend curcumin behavioral effects reported in several animal pain models, including acetid acid–induced visceral nociception (Tajik et al., 2008) and formalin-induced orofacial pain (Mittal et al., 2009), and suggest a new mechanism for curcumin-induced antinociception. In addition, the results obtained from this study are in agreement with previous studies, which show anti-inflammatory and antinociceptive actions of PAMs for α7-nACh receptors in the tested mouse models (Freitas et al., 2013a; Bagdas et al., 2015, 2016). In conclusion, using both in vitro and in vivo approaches, this study establishes that curcumin acts as a PAM of the α7-nACh receptors and provides evidence for a new mechanism for the analgesic-like properties of curcumin.

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Authorship Contributions
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Wrote or contributed to the writing of the manuscript: El Nebrisi, Bagdas, Damaj, Oz.

TABLE 1
Effects of curcumin on motor activity and coordination of mice

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Spontaneous Activity</th>
<th>Rotarod Activity</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td># Interrupts/30 min</td>
<td>% Impairment</td>
</tr>
<tr>
<td>Vehicle</td>
<td>1018 ± 146.7</td>
<td>17.3 ± 10.3</td>
</tr>
<tr>
<td>Curcumin</td>
<td>1125 ± 111.5</td>
<td>12.5 ± 12.5</td>
</tr>
</tbody>
</table>

Fig. 9. Effects of curcumin on acetic acid-induced writhing. ICR mice were treated with i.p. vehicle or curcumin (10 or 30 mg/kg) 45 min prior to i.p. acetic acid (1.2%) injection. Animals were observed for 40 minutes for the number of typical stretching behaviors. To test blockade of the antinociceptive effect of curcumin in the writhing test, the α7 antagonist methyllycaconitine citrate (MLA, 10 mg/kg, s.c.) was given 10 min before curcumin (30 mg/kg, i.p.) or vehicle. After 45 min, acetic acid writhing test was performed. Data are given as the mean ± S.E.M. of 8 animals for each group.


Hsiao YJ, D'Souza NS, and Napolitano M (2015) Oral curcumin has anti-arthritic efficacy through somatostatin generation at ASPET Journals on November 17, 2022 jpet.aspetjournals.org Downloaded from