

JPET #257972

## **Preclinical Pharmacokinetics of Foscicliprox, a Novel Treatment for Urothelial Cancers in Rats and Dogs**

Scott J. Weir, Robyn Wood, Karl Schorno, Amanda E. Brinker, Prabhu Ramamoorthy, Kathy Heppert, Lian Rajewski, Mehmet Tanol, Tammy Ham, Michael J. McKenna, William McCulloch, Michael Dalton, Gregory A. Reed, Roy A. Jensen, Michael J. Baltezor, Shrikant Anant, John A. Taylor III

Department of Pharmacology, Toxicology and Therapeutics, University of Kansas Cancer Center (S.J.W., R.W., A.E.B., G.A.R., R.A.J., M.J.B., S.A., J.A.T.); Institute for Advancing Medical Innovation (S.J.W., R.W., A.E.B., M.J.B.), and Department of Cancer Biology (S.J.W., P.R., S.A.) University of Kansas Medical Center, Kansas City, KS; Biotechnology Innovation and Optimization Center, University of Kansas, Lawrence, KS (K.S., K.H., L.R., M.T., M.J.B.); School of Pharmacy, Istanbul Kemerburgaz University, Istanbul, Turkey (M.T.); CicloMed LLC, Kansas City, MO (T.H.); Navigator LSA, Wilmington, NC (M.J.M.); Alba BioPharm Advisors Inc., Durham, NC (W.M.); The Gnomon Group, Carrboro, NC (M.D.); Department of Pathology (R.A.J) and Department of Urology (J.A.T.) University of Kansas Medical Center, Kansas City, KS.

**Key Words:** Foscicliprox, ciclopirox, ciclopirox glucuronide, pharmacokinetics, drug metabolism, excretion, bladder cancer, allometric scaling

JPET #257972

**Running title:** Fosciclopirox Pharmacokinetics in Rats and Dogs

**Address correspondence and reprints to:**

Scott J. Weir, Pharm.D., Ph.D.

The Institute for Advancing Medical Innovation

University of Kansas Medical Center

2009 Wahl Hall West, Mailstop 1027

3901 Rainbow Boulevard

Kansas City, KS 66160

T: 913-588-3729

F: 913-588-4701

Email: sweir@kumc.edu

Current text pages: 17

Current number of tables: 9

Current number of figures: 5

Current number of references: 19

Abstract current word count: 233 <<**no more than 250 words**>>

Introduction current word count: 548 <<**no more than 750 words**>>

Discussion current word count: 430 <<**no more than 1500 words**>>

JPET #257972

### **List of Nonstandard Abbreviations**

AUC<sub>0-n</sub> = area under the plasma CPX concentration versus time curve from time zero to the last measurable time point; AUC<sub>0-∞</sub> = area under the plasma CPX concentration versus time curve from time zero to infinity; AUMC<sub>0-n</sub> = CPX area under the first moment versus time curve from time zero to the last measurable time point; AUMC<sub>0-∞</sub> = CPX area under the first moment versus time curve from time zero to infinity; C<sub>0</sub> = extrapolated plasma CPX concentration at time zero following intravenous bolus administration; Cl = CPX systemic drug clearance; Cl<sub>r</sub> = CPX renal clearance; CPX = ciclopirox; CPX-O = ciclopirox olamine; CPX-POM = fosciclopirox; F = CPX absolute bioavailability; IV = intravenous administration; Kel = apparent first-order elimination rate constant; MRT = CPX mean residence time; SC = subcutaneous administration; V<sub>d</sub> = CPX apparent volume of distribution; V<sub>ss</sub> = CPX apparent steady state volume of distribution.

### **Section Assignment**

Drug Discovery and Translational Medicine

JPET #257972

## Abstract

Pharmacokinetic studies in rats and dogs were performed to characterize the *in vivo* performance of a novel prodrug, fosciclopirox. Ciclopirox olamine (CPX-O) is a marketed topical antifungal agent with demonstrated *in vitro* and *in vivo* preclinical anticancer activity in several solid tumor and hematologic malignancies. The oral route of administration for CPX-O is not feasible due to low bioavailability and dose-limiting gastrointestinal toxicities. To enable parenteral administration, the phosphoryl-oxymethyl ester of ciclopirox (CPX), fosciclopirox (CPX-POM), was synthesized and formulated as an injectable drug product. In rats and dogs, intravenous (IV) CPX-POM is rapidly and completely metabolized to its active metabolite, CPX. The bioavailability of the active metabolite is complete following CPX-POM administration. CPX and its inactive metabolite, ciclopirox glucuronide (CPX-G) are excreted in urine resulting in delivery of drug to the entire urinary tract. The absolute bioavailability of CPX following subcutaneous (SC) administration of CPX-POM is excellent in rats and dogs, demonstrating the feasibility of this route of administration. These studies confirmed the oral bioavailability of CPX-O is quite low in rats and dogs compared to IV CPX-POM. Given its broad-spectrum anticancer activity in several solid tumor and hematologic cancers and renal elimination, CPX-POM is being developed for the treatment of urothelial cancer. The safety, dose tolerance, pharmacokinetics and pharmacodynamics of IV CPX-POM are currently being characterized in a United States multi-center first-in-human Phase 1 clinical trial in patients with advanced solid tumors (NCT03348514).

JPET #257972

## Introduction

Ciclopirox (CPX) is a broad spectrum antifungal agent (Gupta, 2001). CPX is believed to act as a fungicidal agent by chelating polyvalent metal cations such as  $\text{Fe}^{3+}$  and  $\text{Al}^{3+}$ , resulting in the inhibition of peroxide degradation (Gupta and Plott, 2004) and mitochondrial electron transport processes and energy production (Gupta et al., 1994; Sakurai et al., 1978). Several topical drug products containing CPX free acid or CPX olamine (CPX-O), including a cream, lotion, gel, nail lacquer, and shampoo, are currently marketed in the US and other countries. Consistent with its iron chelation properties, researchers at the Ontario Cancer Institute (OCI) demonstrated that CPX inhibited ribonucleotide reductase *in vitro* in leukemia, myeloma, and a number solid tumor cancer cells, resulting in decreased cell growth and viability at concentrations associated with cell death (Eberhard et al., 2009). Oral administration of CPX-O decreased tumor weight and volume in mouse xenograft models of leukemia and lymphoma and also prevented engraftment of primary acute myeloid leukemia (AML) cells in mice (Eberhard et al., 2009). Song and colleagues subsequently showed that CPX inhibited Wnt signaling through its iron chelation effects (Song et al., 2011), suggesting a possible mechanism of action for CPX's anti-cancer activity.

We collaborated with OCI and The Leukemia and Lymphoma Society to translate these findings to patients with advanced hematologic malignancies. The safety, pharmacokinetics and pharmacodynamics of CPX-O were characterized in patients with relapsed or refractory hematologic malignancy (Minden et al., 2014), representing the first oral, multiple dose CPX-O trial in humans. In relapsed and refractory leukemia and lymphoma patients, circulating CPX and CPX-G appeared to increase proportionately with increasing oral dose across a dose range of 5 to 80 mg/m<sup>2</sup> once daily (Minden et al., 2014). Plasma CPX concentrations were quite low, however, while the major metabolite, CPX-G, circulated in plasma at concentrations at least 10-fold higher than those of CPX (Minden et al., 2014). Although repression of survivin expression

JPET #257972

was observed in two patients, administration of 80 mg/m<sup>2</sup> CPX-O four times daily to patients was associated with grade 3 dose-limiting gastrointestinal toxicities (Minden et al., 2014).

These data, combined with results from an oral [<sup>14</sup>C]-CPX-O human metabolism and mass balance study demonstrating complete absorption (Kellner et al., 1981), led us to conclude the drug undergoes significant presystemic metabolism following oral administration. Low oral bioavailability, coupled with the dose-limiting gastrointestinal toxicities observed in patients, resulted in discontinuation of efforts to repurpose oral CPX-O for the treatment of hematologic malignancies.

Our CPX drug repurposing research efforts turned to developing a drug product that could be administered parenterally to cancer patients. Unfortunately, CPX and CPX-O are sparingly soluble in water (sanofi-aventis, 2006) creating challenges in developing a suitable injectable formulation for this drug. We were successful in employing a prodrug strategy, synthesizing the phosphoryl-oxymethyl (POM) derivative (Stella, 1996; Tanol and Weir, 2013, 2015, 2016) of CPX. Fosciclopirox (CPX-POM) possesses outstanding aqueous solubility and is readily formulated into an injectable drug product. Herein, we have described the plasma and urine pharmacokinetics of CPX-POM, its active metabolite, CPX, and major inactive metabolite, ciclopirox glucuronide (CPX-G), in rats and dogs following intravenous (IV) and subcutaneous (SC) administration. A United States multi-center first-in-human Phase 1 clinical trial is ongoing to characterize the safety, dose tolerance, pharmacokinetics and pharmacodynamics of IV CPX-POM in patients with advanced solid tumors (NCT03348514).

JPET #257972

## Materials and Methods

### *Test Articles*

Disodium ((6-cyclohexyl-4-methyl-2-oxopyridin-1(2H)-yl) oxy) methyl phosphate heptahydrate (CPX-POM), the structure of which is illustrated in **Figure 1**, has a proposed International Nonproprietary Name (INN) of **fosciclopirox**. Fosciclopirox has a molecular formula of  $C_{13}H_{32}NNa_2O_{13}P$  and molecular weight of 487.35 g/mol. CPX-POM exists as a white solid, is water soluble, and possesses solution stability for parenteral administration. CPX-POM and CPX-O were formulated as sterile injectable solutions in 25 mM phosphate buffer, pH 7, with 50 mM Captisol® and stored under refrigerated conditions at 2°C to 8°C. CPX-O was administered orally in Orasweet SF and Water. All test article nominal dosing parameters for following studies are summarized in Table 1.

### *Animal Care and Use Statement*

All studies in animals have been carried out at Xenometrics LLC, Stilwell, KS, a US-based Good Laboratory Practice (GLP) compliant, United States Department of Agriculture registered, Association for Assessment and Accreditation of Laboratory Animal Care accredited nonclinical contract laboratory, in accordance with the Guide for the Care and Use of Laboratory Animals as adopted and promulgated by the U.S. National Institutes of Health and were approved by the Institution's Animal Care and Use Committee.

### *Absolute Bioavailability of CPX Following IV CPX-POM in Rats*

Plasma and urine pharmacokinetics of CPX were characterized following IV administration of CPX-O and CPX-POM to 16 fasted male Sprague-Dawley rats. In this study, CPX-O served as the reference treatment to characterize the absolute bioavailability of CPX following IV CPX-POM. A single IV dose of CPX-O was administered to six rats employing the IV formulation

JPET #257972

described above. In a second group of six male rats, a single IV dose of CPX-POM was administered. IV doses of CPX-O and CPX-POM were administered directly into the tail vein using a syringe and needle at a volume of 2 mL/kg. IV doses of 13.0 mg/kg CPX-O and 17.5 mg/kg CPX-POM doses were administered using IV formulation concentrations of 6.47 mg/mL and 8.72 mg/mL, respectively. Dose volumes were adjusted for the weight of the animal on the morning of drug administration. Serial blood (plasma) samples were collected using a syringe via jugular vein catheter prior to and 0.083, 0.167, 0.25, 0.5, 0.75, 1, 1.5, 2, 3 and 4 hours post-dose. In a third group of four male rats, complete urine was collected at pre-dose and at 0-8 hour and 8-24 hour collection intervals post-dose following IV CPX-O and CPX-POM. The same four animals were dosed with both test articles in a nonrandomized, complete crossover fashion with 72 hours separating treatments. Only urine was collected in this treatment group. Each treatment was administered under fasting conditions.

#### *Absolute Bioavailability of CPX Following IV CPX-POM in Dogs*

Plasma and urine pharmacokinetics of CPX were characterized following IV administration of CPX-O and CPX-POM to four fasted male beagle dogs. In this study, CPX-O served as the reference treatment to characterize the absolute bioavailability of CPX following IV CPX-POM. Single IV doses of 4 mg/kg CPX-O and 5.4 mg/kg CPX-POM were administered under fasting conditions on separate occasions in a nonrandomized, complete crossover fashion with 72 hours separating treatments. IV doses were administered through an indwelling catheter in the cephalic vein at a volume of 2 mL/kg. CPX-O and CPX-POM were formulated at strengths of 1.94 mg/mL and 2.61 mg/mL, respectively. Dose volumes were adjusted for the weight of the animal on the morning of drug administration. Each treatment was administered under fasting conditions. Serial blood (plasma) samples were collected using a syringe via jugular vein prior to and 0.083, 0.167, 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6 and 8 hours post-dose. Complete urine was



JPET #257972

also collected at pre-dose and at 0-8 hour and 8-24 hour collection intervals post-dose following IV CPX-O and CPX-POM.

*Absolute Bioavailability of CPX Following SC CPX-POM and Oral CPX-O in Rats*

Plasma and urine pharmacokinetics of CPX were characterized following subcutaneous (SC) CPX-POM, oral CPX-O, and IV CPX-POM to 22 male Sprague-Dawley rats. In this study, IV CPX-POM served as the reference treatment. The feasibility of SC CPX-POM was determined by comparing the plasma pharmacokinetics of the active metabolite, CPX, following SC and IV CPX-POM. The absolute bioavailability of CPX following oral CPX-O (compared to IV CPX-POM) was determined to bridge previously reported GLP toxicology data generated in the rat (Kellner et al., 1981; Weir et al., 2018). A single IV dose of CPX-POM was administered to six rats employing the IV formulation described above. In a second group of six male rats, a single SC dose of CPX-POM was administered. IV doses of CPX-POM were administered directly into the tail vein using a syringe and needle. SC doses of CPX-POM were administered in two different locations in the scapular region. IV and SC CPX-POM doses of 47 mg/kg and 70.6 mg/kg, respectively, were administered to two separate groups of six rats each, of using IV formulation concentrations of 23.53 mg/mL. In a third group of six male rats, a single oral dose of 38.8 mg/kg CPX-O was administered using the oral formulation described above at a concentration of 29.39 mg/mL. All dose volumes were adjusted for the weight of the animal on the morning of drug administration. Each treatment was administered under fasting conditions. Serial blood (plasma) samples were collected using a syringe via jugular vein catheter prior to and 0.083, 0.167, 0.25, 0.5, 0.75, 1, 1.5, 2, 3 and 4 hours post-dose. In a fourth group of four male Sprague-Dawley rats, complete urine was collected at CPX-POM, SC CPX-POM, and oral CPX-O. The same four animals received all three treatments in a nonrandomized, complete crossover fashion with 72 hours separating treatments. Only urine was collected in this treatment group. Each treatment was administered under fasting conditions.

JPET #257972

*Absolute Bioavailability of CPX Following SC CPX-POM and Oral CPX-O in Dogs*

Plasma and urine pharmacokinetics of CPX were characterized following single SC CPX-POM, oral CPX-O, and IV CPX-POM dose administration under fasting conditions to four male beagle dogs. In this study, IV CPX-POM served as the reference treatment. The feasibility of SC CPX-POM was determined by comparing the plasma pharmacokinetics of the active metabolite, CPX, following SC and IV CPX-POM. The absolute bioavailability of CPX following oral CPX-O (compared to IV CPX-POM) was determined to bridge previously reported GLP toxicology data generated in the dog (Kellner et al., 1981; Weir et al., 2018). IV and SC doses of CPX-POM were administered to dogs employing the IV formulation described above at a concentration of 3.5 mg/mL. IV doses of CPX-POM were administered via an indwelling catheter into the cephalic vein at a volume of 2 mL/kg resulting in a dose of 7.3 mg/kg. SC CPX-POM was administered to the back between the scapula at a volume of 6 mL/kg resulting in a dose of 21.8 mg/kg. CPX-O was administered directly into the stomach via gavage tube. Immediately following oral dose administration, the gavage tube was flushed with approximately 10 mL of water to assure complete delivery of the intended dose. Oral doses were administered at a volume of 6 mL/kg, resulting in an oral CPX-O dose of 12.2 mg/kg. All dose volumes were adjusted for the weight of the animal on the morning of drug administration. Each treatment was administered under fasting conditions. Serial blood (plasma) samples were collected using a syringe via jugular vein prior to and 0.083, 0.167, 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6 and 8 hours post-dose. Complete urine was also collected at pre-dose and at 0-8 hour and 8-24 hour collection intervals post-dose following IV CPX-POM, SC CPX-POM and oral CPX-O.

JPET #257972

### *Bioanalytical Sample Collection, Storage, and Shipment*

Blood samples were transferred to blood collection tubes containing K<sub>2</sub>EDTA, inverted several times to ensure adequate mixing of the blood with the anticoagulant, and placed on ice prior to processing. Blood samples were centrifuged at 3200 revolutions per minute for 10 minutes at approximately 5°C. Plasma samples were transferred into 1.1 mL 96-well plate tubes then stored at -20 ± 5°C until bioanalysis. Urine samples were collected frozen on dry ice into glass collection jars, thawed, transferred to tared plastic jars, and weighed. All urine was stored at -20 ± 5°C until bioanalysis. Plasma and urine samples were transferred from the animal facility to the bioanalytical laboratory on dry ice by same day courier.

### *Bioanalytical Methods in Rat and Dog Plasma and Urine*

Rat and dog plasma and urine bioanalytical methods were developed and validated under non-GLP conditions to quantitate concentrations of CPX-POM, CPX and CPX-G. CPX-POM and CPX-G do not appear to be active based on *in vitro* studies in human high-grade urothelial cancer cell lines (unpublished results).

CPX-POM concentrations were determined in 20 µL sample volumes. Samples were placed in polypropylene centrifuge tubes followed by the addition of another phosphoryl-oxymethyl ester, SN38 prodrug, as the internal standard. An additional 50 µL of acetonitrile was added to each sample, as well as 60 µL of 0.1% formic acid. Samples were then vortexed and extracted with ethyl acetate. Samples were frozen and organic phase transferred into clean polypropylene tubes and evaporated to dryness. Plasma samples were reconstituted in 50 µL of 30% methanol in water containing 0.1% formic acid and 15 µL of each sample injected for analysis by liquid chromatography-mass spectrometry and tandem mass spectrometry (LC-MS/MS) for CPX-POM. The chromatographic separation of CPX-POM and internal standard was achieved on a Phenomenex Kinetex C18 column (100 mm x 2.1 mm, 2.6 µm) maintained at 40°C. The

JPET #257972

mobile phase gradient program with solvent A (95:5:0.1 deionized water/acetonitrile/formic acid) and solvent B (95:5:0.1 acetonitrile/deionized water/formic acid) was run at a flow rate of 0.25 mL/min. The gradient started at 25% solvent B and increased linearly to 95% over 6.5 minutes, then reduced linearly to the initial conditions within 0.1 minutes. Total run time was 10 minutes for each sample. An ABSciex 3200 Linear Ion TRAP quadrupole mass spectrometer was operated in the negative electrospray ionization (ESI) mode for mass detection and analysis of CPX-POM. Multiple reaction monitoring (MRM) was used to monitor the precursor/product ion transitions of 316.07/79.1 (CPX-POM) and 500.79/373.0 (SN38 prodrug, internal standard). All mass spectrometer settings were standardized to optimize detection and analysis of CPX-POM and internal standard. The lower limit of quantitation (LLOQ) for CPX-POM was 100 ng/mL and the standard curve was linear between 100 ng/mL and 5000 ng/mL, with a correlation coefficient of 0.998.

CPX concentrations were determined in 50  $\mu$ L sample volumes. Samples were placed in polypropylene centrifuge tubes followed by the addition of internal standard (d3-methoxycyclopirox). An equal volume of 0.2 M sodium hydroxide was added to each tube and the sample vortexed. Samples were chilled until ice cold and then derivatized with the addition of 25  $\mu$ L dimethyl sulfate. Samples were then heated in an oven at 37°C for 30 minutes. After the derivatization reaction, 25  $\mu$ L of triethylamine was added to each sample to neutralize the reaction. Plasma samples were then extracted using methyl tert-butyl ether and frozen over into clean polypropylene tubes for evaporation and sample concentration. Plasma samples were reconstituted in 100  $\mu$ L of 40% acetonitrile in water. Five microliters of each prepared sample were injected onto the LC-MS/MS for determination of CPX concentration. The chromatographic separation of CPX and internal standard was achieved on a Waters Sunfire C18 column (50 mm x 2 mm, 3.5  $\mu$ m) that was maintained at 40°C. The mobile phase gradient program with solvent A (95:5:0.1 deionized water/acetonitrile/formic acid) and solvent B (95:5:0.1

JPET #257972

acetonitrile/deionized water/formic acid) was run at a flow rate of 0.25 mL/min. The gradient started at 25% B and increased linearly to 95% over 6.5 minutes, then reduced linearly to the initial conditions within 0.1 minutes. Total run time for each sample was 10 minutes. An ABSciex 3200 Linear Ion TRAP quadrupole mass spectrometer was operated in the positive ESI mode for mass detection and analysis of CPX. MRM was used to monitor the precursor/product ion transitions of 222.2/136.0 (CPX) and 225.2/136.0 (d3 methoxy-ciclopirox, internal standard). All mass spectrometer settings were standardized to optimize detection and analysis of CPX and internal standard. The LLOQ for CPX was 25 ng/mL and the standard curve was linear between 25 ng/mL and 5000 ng/mL, with a correlation coefficient of 0.998.

CPX-G concentrations were determined in 20  $\mu$ L sample volumes. Samples were placed in polypropylene centrifuge tubes followed by the addition of ciclopirox-D11- $\beta$ -glucuronide as the internal standard in 20  $\mu$ L of 1:1 acetonitrile:deionized water. Samples were vortexed followed by transfer of the supernatant to clean polypropylene tubes. Fifteen milliliter aliquots were injected for analysis by LC-MS/MS for CPX-G. For urine samples, an aliquot of urine was diluted 1:10 with 1% formic acid, internal standard added, and samples mixed well prior to injection onto the LC-MS/MS for CPX-G determination. The chromatographic separation of CPX-G and internal standard was achieved on a Zorbax Agilent column (50 mm x 2.1 mm, 2.6  $\mu$ m) that was maintained at 15°C. The mobile phase gradient program with solvent A (95:5:0.1 deionized water/acetonitrile/formic acid) and solvent B (95:5:0.1 acetonitrile/deionized water/formic acid) was run at a flow rate of 0.40 mL/min. The gradient started at 15% solvent B and increased linearly to 95% over 1.0 minutes beginning at 2.0 minutes, then reduced linearly at 5.1 minutes to the initial conditions at 6.3 minutes. Total run time for each sample was 10 minutes. An ABSciex 3200 Linear Ion TRAP quadrupole mass spectrometer was operated in the negative ESI mode for mass detection and analysis of CPX-G. MRM was used to monitor the precursor/product ion transitions of 384.1 (CPX-G) and 395.1 (ciclopirox-D11- $\beta$ -

JPET #257972

glucuronide), internal standard. All mass spectrometer settings were standardized to optimize detection and analysis of CPX-G and internal standard. The LLOQ for CPX-G was 250 ng/mL and the standard curve was linear between 100 ng/mL and 5000 ng/mL, with a correlation coefficient of 0.998.

### *Pharmacokinetic Analysis*

Non-parametric plasma and urine pharmacokinetic parameters for CPX-POM, CPX and CPX-G were generated from resultant concentration-time data using Phoenix WinNonlin (Certara™, Version 6.3) software. For plasma data, actual blood sample collection times were used. Pre-dose plasma concentration values reported as below the LLOQ were designated as below the quantifiable limit (BQL) and treated as zero in calculation. BQL values observed in the terminal phase of the plasma concentration-time profiles were treated as missing values in determination of the apparent first-order elimination rate constant and area under the plasma concentration-time curve. Given the absence of quantifiable concentrations of CPX-POM in plasma and urine for all studies in rats and dogs, pharmacokinetic data analysis was not possible for parent drug.

### *Statistical Analysis*

Descriptive statistics (mean ± standard deviation) were reported for the resultant plasma and urine pharmacokinetic parameters. Given the objective of the studies conducted, inferential statistical analysis was not performed.

### *Data Management*

The study protocol, in-life report including dosing and sample collection data, Excel bioanalytical data, pharmacokinetic data analysis, data tabulation, and graphics files are stored at CicloMed LLC, Kansas City, MO.

JPET #257972

## Results

The rationale for developing the injectable formulation of CPX-POM was based on pH, solubility, and the desired concentration of active pharmaceutical ingredient in the injectable formulation. The desired concentration of the injectable CPX-POM formulation for study in early phase clinical trials, based on the anticipated dose range being evaluated, was 76.8 mg/mL as CPX-POM disodium heptahydrate (which is equivalent to concentrations of 50 mg/mL and 32.6 mg/mL CPX-POM and CPX, respectively). Formulating an injectable product containing CPX at this concentration required approximately 300 mg/mL Captisol® and adjustment of the pH of the solution to between 9.5 and 10.0. In contrast, CPX-POM for injection is supplied as a sterile solution containing sterile water for injection, anhydrous dibasic sodium phosphate as a buffer, and sodium hydroxide and hydrochloric acid for pH adjustment. The much more desirable pH of the CPX-POM injectable formulation is between 7.5 and 8.0.

Treatments administered to rats and dogs in the four studies, including test article, formulation description, route of administration, dose volume administered, test article dose as well as CPX equivalent dose are summarized in Table 1. CPX-POM IV and SC doses chosen for these studies were selected as well tolerated doses determined in non-GLP toxicology dose range finding studies conducted in rats and dogs. Oral and IV CPX-O doses were selected based on published data as well as available Freedom-of-Information data released following FDA approval of topical CPX-containing antifungal drug products. Derived CPX plasma and urine pharmacokinetic parameters (e.g., clearance, absolute bioavailability) were calculated based on CPX equivalent doses. CPX has a pKa of approximately 6.8 for the phenolic hydrogen. The anionic form of CPX has excellent solubility, exceeding 500 mg/mL. The neutral protonated form of CPX, in contrast, is poorly soluble at approximately 0.3 mg/mL. Captisol®, at 50 mM (108.15 mg/mL), was added to the CPX formulations used in the studies described herein to avoid potential precipitation of the neutral protonated phenolic form of CPX. The CPX-O intravenous

JPET #257972

dosing solution for the rat was formulated at pH 7.0 at a concentration of 6.47 mg/mL (5.0 mg/mL CPX). The IV CPX-O dosing solution for the dog was more dilute, 1.94 mg/mL CPX-O (1.5 mg/mL CPX). Although not required to solubilize CPX-POM, Captisol® was utilized in the prodrug injectable formulation for rat and dog studies to control for any potential effects of this excipient on the pharmacokinetics of CPX.

The plasma and urine pharmacokinetics of CPX were characterized in the rat following IV administration of CPX-O and CPX-POM doses of 13.0 mg/kg and 17.5 mg/kg, respectively. IV CPX-O was used as the reference in characterizing the bioavailability of CPX following IV CPX-POM in this rodent toxicology species. Metabolism of CPX-POM was rapid and complete following IV administration of CPX-POM. Only a few blood samples, collected within minutes of IV bolus administration, contained plasma CPX-POM concentrations exceeding the assay LLOQ. Plasma CPX concentrations declined rapidly in a bi-exponential fashion following IV administration of both test articles as shown in **Figure 2**. Resultant plasma CPX pharmacokinetic parameters are summarized in **Table 2**. Following IV administration of CPX-POM, the systemic availability of CPX was 97% compared to IV administration of CPX-O. The apparent elimination half-life and mean residence time values for the active metabolite were less than one hour and were similar for both treatments. The systemic clearance of CPX was 3,326 mL/hr/kg following IV CPX-POM. The apparent and steady-state volume of distribution values for CPX following IV CPX-POM were 2,664 mL /kg and 1,853 mL/kg, respectively. These values were comparable to those observed following IV CPX-O administration. CPX-POM was not detected in any urine samples collected. Urine CPX pharmacokinetic parameters obtained in rats following IV CPX-O and CPX-POM are presented in **Table 3**. Following single IV doses of CPX-POM and CPX-O, less than 1% of the dose was excreted as CPX while a significant portion of the administered dose, approximately 18-34%, was eliminated as the glucuronide metabolite of CPX following both treatments.



JPET #257972

The plasma and urine pharmacokinetics of CPX were characterized in the dog following IV administration of 4 mg/kg CPX-O and 5.4 mg/kg CPX-POM. IV CPX-O was used as the reference in characterizing the bioavailability of CPX following IV CPX-POM in this non-rodent toxicology specie. Metabolism of CPX-POM was rapid and complete following IV administration of CPX-POM. Only a few blood samples, collected within minutes of IV bolus administration, contained plasma CPX-POM concentrations exceeding the LLOQ. Like the rat, plasma CPX concentrations declined bi-exponentially in the dog following IV administration of both test articles as illustrated in **Figure 3**. As summarized in **Table 4**, the systemic availability of CPX was 105% following IV CPX-POM compared to IV administration of CPX-O. The apparent elimination half-life and mean residence time values for the active metabolite ranged from 2 to 4 hours following both treatments. The systemic clearance of CPX was 762 mL/hr/kg following IV CPX-POM. The apparent and steady-state volume of distribution values for CPX were 4,133 mL/kg and 3,686 mL/kg, respectively, following IV CPX-POM. These values were comparable to those observed following IV CPX-O administration. Urine CPX pharmacokinetic parameters obtained in dogs following IV CPX-O and CPX-POM are summarized in **Table 5**. CPX-POM was not detected in any urine samples collected. Following single IV doses of CPX-POM and CPX-O, less than 1% of the dose was excreted as CPX. Approximately 63% of test article doses were recovered in urine as the glucuronide metabolite.

The pharmacokinetics of CPX were characterized following IV administration of 47 mg/kg CPX-POM and SC administration of 70.6 mg/kg CPX-POM, as well as oral administration of 38.8 mg/kg of CPX-O in fasted rats. In this study, IV CPX-POM was used as the reference treatment to characterize the absolute bioavailability of SC CPX-POM as well as to establish a pharmacokinetic bridge between IV CPX-POM and oral CPX-O. Resultant mean plasma CPX concentration-time profiles following administration of IV and SC CPX-POM as well as orally

JPET #257972

administered CPX-O are illustrated in **Figure 4**. Plasma CPX concentrations declined rapidly following IV administration of CPX-POM. Plasma concentrations of CPX following SC CPX-POM were consistently greater than those observed following IV CPX-POM over the terminal portion of the profile and clearly demonstrated an absorption phase. Comparatively, plasma CPX concentrations following oral administration of CPX-O were consistently quite low. Resultant plasma CPX pharmacokinetic parameters following administration of the three treatments are summarized in **Table 6**. Systemic clearance, apparent, and steady-state volume of distribution values for CPX following IV CPX-POM were 3,696 mL/hr/kg, 3,160 mL/kg, and 1,106 mL/kg, respectively. These values were comparable to those observed following IV CPX-POM administration to rats described above. The absolute bioavailability of CPX following SC CPX-POM was 80% with a relatively rapid absorption rate ( $T_{max} = 0.26$  hour). In contrast, the bioavailability of CPX following oral administration of CPX-O was 4.6% compared to IV CPX-POM. CPX-POM was not detected in any blood or urine sample collected in this study. Urine CPX pharmacokinetic parameters obtained in rats are summarized in **Table 7**. Less than 1% of the administered dose was recovered in the urine following SC CPX-POM and CPX-O administration as CPX. Approximately 3.5% and 9% of the dose was recovered in urine as CPX and CPX-G, respectively, following IV CPX-POM administration.

The pharmacokinetics of CPX were characterized following IV administration of 7.3 mg/kg CPX-POM and SC administration of 21.8 mg/kg CPX-POM, as well as the administration of 12.2 mg/kg CPX-O to four fasted Beagle dogs. In this study, IV CPX-POM was used as the reference treatment to characterize the absolute bioavailability of SC CPX-POM as well as to establish a pharmacokinetic bridge between IV CPX-POM and oral CPX-O. Resultant plasma CPX pharmacokinetic parameters are summarized in **Table 8**. Plasma CPX concentrations declined rapidly following IV administration of CPX-POM as illustrated in **Figure 5**. Plasma concentrations of CPX following SC CPX-POM were consistently greater than those observed

JPET #257972

following IV CPX-POM over the terminal portion of the profile while plasma CPX concentrations following oral administration of CPX-O were comparatively quite low. The absolute bioavailability of CPX following SC CPX-POM was complete, 106%, in the dog based on area-under-the-plasma-CPX-concentration-time curve to the last sampling time (8 hours post-dose). The rate of absorption from the injection site was dramatically slower,  $T_{max} = 5.5$  hours, compared to the rat. The absolute bioavailability of CPX following oral CPX-O was approximately 17%. CPX-POM was not detected in any blood or urine sample collected. Systemic clearance, apparent and steady-state volume of distribution values for CPX following IV CPX-POM were 615 mL/hr/kg, 4,486 mL/kg, and 3,776 mL/kg, respectively. These values were comparable to those observed following IV CPX-POM administration to dogs described above. Urine CPX pharmacokinetic parameters obtained in dogs are summarized in **Table 9**. Less than 1% of the dose was excreted as CPX following each the three treatments while 37-49% of the dose was eliminated as the glucuronide conjugate.

## Discussion

Incorporation of the phosphoryl-oxymethyl (POM) prodrug moiety has been an effective approach to improving aqueous solubility and oxidative stability of successful commercial products such as fosphenytoin and fospropofol (Dhareshwar and Stella, 2010; Stella, 1996; Stella and Nti-Addae, 2007). *In vivo* conversion of POM prodrugs to their active metabolites is mediated by phosphatases (Dhareshwar and Stella, 2008). Following single IV and SC doses of CPX-POM to rats and dogs, rarely was the prodrug detected in plasma and never in urine. Further, we demonstrated complete bioavailability of the active metabolite, CPX, following IV CPX-POM. This rapid and complete metabolism of the POM prodrug of CPX observed in rats and dogs is consistent with circulating phosphatases.

JPET #257972

The rodent and non-rodent toxicology species evaluated in oral GLP toxicology studies conducted to support registration of CPX and CPX-O containing topical drug products were rats and dogs, respectively (FDA, 2003). To support development of CPX-POM, these same species were selected for the studies described herein. Systemic clearance values for CPX determined in rats and dogs were quite comparable to reported hepatic blood flow rates (Davies and Morris, 1993) suggesting the active metabolite is a high clearance drug. Despite rapid clearance of the active metabolite, CPX-POM administered IV to rats and dogs at doses utilized in these studies, resulted in plasma CPX concentrations exceeding *in vitro* IC50 values in human leukemia and lymphoma cell lines several-fold (Eberhard et al., 2009; Song et al., 2011). CPX has been shown to be extensively metabolized via glucuronidation followed by renal excretion in rats, dogs, and humans (Kellner et al., 1981). The relatively low urinary recoveries of CPX observed in rats and dogs, along with renal clearance values of the active metabolite, are consistent with extensive hepatic metabolism. Despite reported protein binding of 96% (Kellner et al., 1981), CPX distributes beyond the central (blood) compartment in rats and dogs based on steady-state and apparent volume of distribution values observed in these studies.

The low absolute bioavailability of CPX following oral administration of CPX-O to rats and dogs, compared to IV CPX-POM, is consistent with the drug or active metabolite undergoing significant presystemic metabolism. These data established a pharmacokinetic bridge between CPX-POM and the olamine salt of CPX contained in marketed antifungal agents and was useful in designing IV CPX-POM GLP toxicology studies subsequently conducted in these species. Excellent bioavailability of CPX was observed following SC administration demonstrating feasibility of this route of administration should CPX-POM treatment prove appropriate in an outpatient or ambulatory cancer treatment setting. CPX-POM is being developed for the treatment of non-muscle invasive and muscle invasive bladder cancer.

JPET #257972

### **Acknowledgments**

We would like to thank KUCC's Lead Development and Optimization Shared Resource for their screening and formulation expertise as well as Nancy Bella, PharmD, of The Gnomon Group, LLC for her editorial support.

JPET #257972

**Authorship Contributions.**

*Participated in research design:* SJW, RW, MJM, WM, MD, RAJ, MJB, SA, JAT

*Conducted experiments:* SJW, RW, KS, MJM

*Contributed new reagents or analytic tools:* PR, KH, LR, MT, GAR, MJB, SA, JAT

*Performed data analysis:* SJW, RW, AEB

*Wrote or contributed to the writing of the manuscript:* SJW, RW, AEB, TH, GAR, RAJ, MJB, SA,

JAT

## References

- Davies, B., and Morris, T. (1993) Physiological parameters in laboratory animals and humans. *Pharmaceutical research* 10: 1093-1095.
- Dhareshwar, S.S., and Stella, V.J. (2008) Your prodrug releases formaldehyde: should you be concerned? No! *Journal of pharmaceutical sciences* 97: 4184-4193.
- Dhareshwar, S.S., and Stella, V.J. (2010) A novel prodrug strategy for beta-dicarbonyl carbon acids: syntheses and evaluation of the physicochemical characteristics of C-phosphoryloxymethyl (POM) and phosphoryloxymethyl (POMOM) prodrug derivatives. *Journal of pharmaceutical sciences* 99: 2711-2723.
- Eberhard, Y., McDermott, S.P., Wang, X., Gronda, M., Venugopal, A., Wood, T.E., Hurren, R., Datti, A., Batey, R.A., Wrana, J., *et al.* (2009) Chelation of intracellular iron with the antifungal agent ciclopirox olamine induces cell death in leukemia and myeloma cells. *Blood* 114: 3064-3073.
- FDA, U.S. (2003) Loprox (Ciclopirox) Shampoo Drug Approval Package. In IND 21-129, H.H. Services, ed. (U.S. Food & Drug Administration Drug Approvals Database: Drugs@FDA.gov).
- Gupta, A.K. (2001) Ciclopirox: an overview. *International journal of dermatology* 40: 305-310.
- Gupta, A.K., and Plott, T. (2004) Ciclopirox: a broad-spectrum antifungal with antibacterial and anti-inflammatory properties. *International journal of dermatology* 43 *Suppl 1*: 3-8.
- Gupta, A.K., Sauder, D.N., and Shear, N.H. (1994) Antifungal agents: an overview. Part I. *Journal of the American Academy of Dermatology* 30: 677-698; quiz 698-700.
- Kellner, H.M., Arnold, C., Christ, O.E., Eckert, H.G., Herok, J., Hornke, I., and Rupp, W. (1981) [Pharmacokinetics and biotransformation of the antimycotic drug ciclopiroxolamine in animals and man after topical and systemic administration]. *Arzneimittel-Forschung* 31: 1337-1353.
- Minden, M.D., Hogge, D.E., Weir, S.J., Kasper, J., Webster, D.A., Patton, L., Jitkova, Y., Hurren, R., Gronda, M., Goard, C.A., *et al.* (2014) Oral ciclopirox olamine displays biological activity in a phase I study in patients with advanced hematologic malignancies. *American journal of hematology* 89: 363-368.
- Sakurai, K., Sakaguchi, T., Yamaguchi, H., and Iwata, K. (1978) Mode of action of 6-cyclohexyl-1-hydroxy-4-methyl-2(1H)-pyridone ethanolamine salt (Hoe 296). *Chemotherapy* 24: 68-76.
- sanofi-aventis (2006) PENLAC (Ciclopirox Topical Solution, 8% w/w) Nail Lacquer. In Product Monograph, S.-A.C. Inc, ed.
- Song, S., Christova, T., Perusini, S., Alizadeh, S., Bao, R.Y., Miller, B.W., Hurren, R., Jitkova, Y., Gronda, M., Isaac, M., *et al.* (2011) Wnt inhibitor screen reveals iron dependence of beta-catenin signaling in cancers. *Cancer research* 71: 7628-7639.
- Stella, V. (1996) A case for prodrugs: Fosphenytoin. *Advanced Drug Delivery Reviews* 19: 311 -

JPET #257972

330.

Stella, V.J., and Nti-Addae, K.W. (2007) Prodrug strategies to overcome poor water solubility. *Adv Drug Deliv Rev* 59: 677-694.

Tanol, M., and Weir, S. (2013) Issued Patent US 8609637 *Prodrugs of 6-cyclohexyl-1-hydroxy-4-4-methylyridin-2-(1H)-one and derivatives thereof.*, USPTO, ed. (US).

Tanol, M., and Weir, S. (2015) Issued Certificate of Japanese Patent No 5853028 *Prodrugs of 6-cyclohexyl-1-hydroxy-4-methylyridin-2-(1H)-one and derivatives thereof.*, JPO, ed. (Japan).

Tanol, M., and Weir, S. (2016) Issued Patent PCT 2646035. *Prodrugs of 6-cyclohexyl-1-hydroxy-4-4-methylyridin-2-(1H)-one and derivatives thereof.*, WIPO, ed. (Austria, Belgium, Croatia, Czech Republic, Denmark, Finland, France, Germany (Patent No. 602011023919.8), Greece, Hungary, Italy, Ireland, Luxembourg, Netherlands, Norway, Poland, Portugal, Serbia, Spain, Sweden, Switzerland, Turkey and United Kingdom).

Weir, S.J., Wood, R., Ham, T., Ranjarajan, P., Ramamoorthy, P., Rajewski, L., Heppert, K., Haslam, J., Schorno, K., Dalton, M., *et al.* (2018) Preclinical development of ciclopirox prodrug for the treatment of non-muscle invasive and muscle invasive bladder cancer. *Journal of Clinical Oncology* 36: e14576-e14576.



JPET #257972

### **Footnotes**

Research reported in this presentation was supported by the National Cancer Institute Cancer Center Support Grant [Grant P30 CA168524], and by a public-private partnership between The Institute for Advancing Medical Innovation at the University of Kansas Medical Center and CicloMed LLC, Kansas City, MO.

JPET #257972

## Figure Legends

**Figure 1: Chemical Structure of Fosciclopirox ((6-cyclohexyl-4-methyl-2-oxopyridin-1(2H)-yl) oxy) methyl phosphate heptahydrate (CPX-POM).**

**Figure 2: Mean  $\pm$  SD Plasma CPX Concentration-Time Profiles Following Single Dose IV Administration of 17.5 mg/kg CPX-POM and 13 mg/kg CPX-O to Male Sprague-Dawley Rats**

Data are presented on rectilinear (A) and semilogarithmic (B) plots as mean  $\pm$  SD, n = 6 per treatment group.

CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; hr, hour; IV, intravenous; SD, standard deviation.

**Figure 3: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following Single Dose IV Administration of 5.4 mg/kg CPX-POM and 4.0 mg/kg CPX-O to Male Beagle Dogs**

Plasma CPX concentration-time profiles following single dose administration of 5.4 mg/kg IV CPX-POM and 4.0 mg/kg CPX-O to four male beagle dogs in a complete crossover fashion.

Data are presented on rectilinear (A) and semilogarithmic (B) plots as mean  $\pm$  SD.

CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; hr, hour; IV, intravenous; SD, standard deviation.

JPET #257972

**Figure 4: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following Administration of 47 mg/kg IV CPX-POM, 70.6 mg/kg SC CPX-POM, and 38.8 mg/kg CPX-O to Six Rats per Treatment Group**

Plasma CPX concentration-time profiles following single dose administration of 47mg/kg IV CPX-POM, 71 mg/kg SC CPX-POM, and 39 mg/kg CPX-O to male Sprague-Dawley rats. Data are presented on rectilinear (A) and semilogarithmic (B) plots as mean  $\pm$  SD, n = 6 per treatment group.

CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; hr, hour; IV, intravenous; SC, subcutaneous; SD, standard deviation.

**Figure 5: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following IV Administration of 7.3 mg/kg CPX-POM, SC Administration of 21.8 mg/kg SC CPX-POM, and 12.2 mg/kg Oral CPX-O to Four Male Beagle Dogs**

Plasma CPX concentration-time profiles following single dose administration of 7.3 mg/kg IV CPX-POM, 21.8 mg/kg SC CPX-POM, and 12.2 mg/kg oral CPX-O to four male beagle dogs in a complete crossover fashion. Data are presented on rectilinear (A) and semilogarithmic (B) plots as mean  $\pm$  SD.

CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; hr, hour; IV, intravenous; SC, subcutaneous; SD, standard deviation.

JPET #257972

**Tables**

**Table 1: Treatments administered to rats and dogs including test article, test article strength, dose volume, formulation description, test article dose and ciclopirox equivalent dose.**

Study	Species	Route	Test Article	Test Article Strength (mg/mL)	Dose Volume (mL/kg)	Formulation Description	Test Article Dose (mg/kg)	Ciclopirox Equivalent Dose (mg/kg)
1	Rat	IV	CPX-O	6.47	2.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	13.0	10.0
		IV	CPX-POM	8.72	2.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	17.5	7.4
2	Dog	IV	CPX-O	1.94	2.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	4.0	3.1
		IV	CPX-POM	2.61	2.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	5.4	2.3
3	Rat	IV	CPX-POM	23.53	2.1	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	47.0	20.0
		SC	CPX-POM	23.53	3.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	70.6	30.0
		Oral	CPX-O	29.39	1.3	Orasweet SF, Water	38.8	30.0
4	Dog	IV	CPX-POM	3.53	2.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	7.3	3.1
		SC	CPX-POM	3.53	6.0	25mM Phosphate Buffer, pH 7, 50 mM Captisol®	21.8	9.3
		Oral	CPX-O	1.94	6.0	Orasweet SF, Water	12.2	9.3

JPET #257972

**Table 2: Mean ± SD Plasma CPX Pharmacokinetic Parameters in Male Sprague-Dawley Rats following Single IV Doses of CPX-POM and CPX-O**

Parameter	Treatment <sup>a</sup>	
	CPX-POM	CPX-O
Body weight (kg)	0.26 ± 0.01	0.27 ± 0.01
CPX dose (mg/kg) <sup>b</sup>	7.47 ± 0.12	10.02 ± 0.04
Test article dose (mg/kg) <sup>c</sup>	17.56 ± 0.28	12.96 ± 0.05
C <sub>0</sub> (ng/mL)	9,310 ± 5,251	25,883 ± 13,321
K <sub>el</sub> (hr <sup>-1</sup> )	1.580 ± 0.286	1.453 ± 0.634
T <sub>½</sub> (hr)	0.452 ± 0.087	0.615 ± 0.404
AUC <sub>0</sub> <sup>n</sup> (ng•hr/mL)	2,121 ± 501	3,077 ± 427
AUC <sub>0</sub> <sup>∞</sup> (ng•hr/mL)	2,161 ± 477	3,141 ± 349
AUMC <sub>0</sub> <sup>n</sup> (ng•hr <sup>2</sup> /mL)	827 ± 275	947 ± 608
AUMC <sub>0</sub> <sup>∞</sup> (ng•hr <sup>2</sup> /mL)	950 ± 229	1,291 ± 1,206
MRT (hr)	0.455 ± 477	0.446 ± 0.474
Cl (mL/hr/kg)	3,326 ± 296	3,225 ± 342
V <sub>d</sub> <sub>z</sub> (mL/kg)	2,664 ± 670	2,983 ± 2,319
V <sub>ss</sub> (mL/kg)	1,853 ± 899	1,563 ± 1,853
F (%)	96.7	

JPET #257972

AUC<sub>0-n</sub>, area under the plasma CPX concentration versus time curve from time zero to the last measurable time point; AUC<sub>0-∞</sub>, area under the plasma CPX concentration versus time curve from time zero to infinity; AUMC<sub>0-n</sub>, CPX area under the first moment versus time curve from time zero to the last measurable time point; AUMC<sub>0-∞</sub>, CPX area under the first moment versus time curve from time zero to infinity; C<sub>0</sub>, extrapolated plasma CPX concentration at time zero following intravenous bolus administration; Cl, CPX systemic drug clearance; CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; F, CPX bioavailability; Kel, apparent first-order elimination rate constant; MRT, CPX mean residence time; T<sub>½</sub>, apparent elimination half-life, V<sub>d</sub>, CPX apparent volume of distribution; V<sub>ss</sub>, CPX apparent steady state volume of distribution.

- a N = 6 rats per treatment.
- b Ciclopirox free acid dose.
- c CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 8.72 mg/mL (3.71 mg/mL ciclopirox free acid), CPX-O was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 6.47 mg/mL (5.0 mg/mL ciclopirox free acid).

JPET #257972

**Table 3: Mean ± SD Urine CPX and CPX-G Pharmacokinetic Parameters in Male Sprague-Dawley Rats following Single Doses of IV CPX-POM and CPX-O**

Parameter	Treatment <sup>a</sup>	
	CPX-POM	CPX-O
Body weight (kg)	0.252 ± 0.006	0.238 ± 0.006
CPX dose (mg/kg) <sup>b</sup>	7.38 ± 0.01	9.98 ± 0.06
Test article dose (mg/kg) <sup>c</sup>	17.35 ± 0.03	12.91 ± 0.08
U <sub>CPX</sub> 0-8 hr (ng/mL)	1,182 ± 498	921 ± 112
U <sub>CPX</sub> 8-24 hr (ng/mL)	198 ± 172	118 ± 45
U <sub>CPX</sub> 0-8 hr (µM)	4.45 ± 0.54	8.79 ± 2.41
U <sub>CPX</sub> 8-24 hr (µM)	0.57 ± 0.22	0.96 ± 0.83
dXe/dt <sub>CPX</sub> 0-8hr (µg/hr)	0.23 ± 0.09	0.76 ± 0.26
dXe/dt <sub>CPX</sub> 8-24hr (µg/hr)	0.07 ± 0.01	0.13 ± 0.04
Xe <sub>CPX</sub> (µg)	2.88 ± 0.78	8.18 ± 2.90
Fe <sub>CPX</sub> (%)	0.15 ± 0.04	0.34 ± 0.12
Fe <sub>CPX-G</sub> (%)	34.07 ± 12.30	17.58 ± 2.46

CPX, ciclopirox; CPX-G, ciclopirox glucuronide metabolite; Fe<sub>CPX</sub>, fraction of the administered dose excreted as CPX; dXe/dt<sub>CPX</sub>, mass of CPX excreted per hour over x hours following drug administration; Fe<sub>CPX-G</sub>, fraction of the administered dose excreted as CPX-G; U<sub>CPX</sub>, urine CPX concentration; Xe<sub>CPX</sub>, mass of CPX excreted in urine.

<sup>a</sup> Four rats received both treatments in a complete, nonrandomized, crossover fashion.

<sup>b</sup> Ciclopirox free acid dose.

JPET #257972

- c CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 8.72 mg/mL (3.71 mg/mL ciclopirox free acid), CPX-O was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 6.47 mg/mL (5.0 mg/mL ciclopirox free acid).



JPET #257972

**Table 4: Mean ± SD Plasma CPX Pharmacokinetic Parameters in Male Beagle Dogs following Single IV Doses of CPX-POM and CPX-O**

Parameter	Treatment <sup>a</sup>	
	CPX-POM	CPX-O
Body weight (kg)	9.65 ± 0.38	9.64 ± 0.34
CPX dose (mg/kg) <sup>b</sup>	2.28 ± 0.01	3.10 ± 0.02
Test article dose (mg/kg) <sup>c</sup>	5.40 ± 0.02	4.01 ± 0.02
C <sub>0</sub> (ng/mL)	2,970 ± 969	5,996 ± 1,387
K <sub>el</sub> (hr <sup>-1</sup> )	0.242 ± 0.030	0.185 ± 0.019
T <sub>½</sub> (hr)	2.90 ± 0.329	3.77 ± 0.40
AUC <sub>0</sub> <sup>n</sup> (ng•hr/mL)	2,571 ± 436	3,314 ± 655
AUC <sub>0</sub> <sup>∞</sup> (ng•hr/mL)	3,075 ± 591	4,009 ± 802
AUMC <sub>0</sub> <sup>n</sup> (ng•hr <sup>2</sup> /mL)	5,485 ± 1,293	7,293 ± 1,520
AUMC <sub>0</sub> <sup>∞</sup> (ng•hr <sup>2</sup> /mL)	11,680 ± 3,682	16,665 ± 3,971
MRT (hr)	3.74 ± 0.58	2.20 ± 0.09
Cl (mL/hr/kg)	762 ± 158	804 ± 200
V <sub>d<sub>z</sub></sub> (mL/kg)	4,133 ± 363	4,253 ± 1,266
V <sub>ss</sub> (mL/kg)	3,686 ± 431	3,218 ± 810
F (%)	105.4 ± 11.9	

AUC<sub>0</sub><sup>n</sup>, area under the plasma CPX concentration versus time curve from time zero to the last measurable time point; AUC<sub>0</sub><sup>∞</sup>, area under the plasma CPX concentration versus time curve

JPET #257972

from time zero to infinity;  $AUMC_0^n$ , CPX area under the first moment versus time curve from time zero to the last measurable time point;  $AUMC_0^\infty$ , CPX area under the first moment versus time curve from time zero to infinity;  $C_0$ , extrapolated plasma CPX concentration at time zero following intravenous bolus administration;  $Cl$ , CPX systemic drug clearance; CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox;  $F$ , CPX bioavailability;  $IV$ , intravenous;  $K_{el}$ , apparent first-order elimination rate constant;  $MRT$ , CPX mean residence time;  $SD$ , standard deviation;  $T_{1/2}$ , apparent elimination half-life;  $V_{dz}$ , CPX apparent volume of distribution;  $V_{ss}$ , CPX apparent steady state volume of distribution.

- <sup>a</sup> N = 4 dogs receiving both treatments in a complete, nonrandomized, cross-over study design.
- <sup>b</sup> Ciclopirox free acid dose.
- <sup>c</sup> CPX-POM disodium heptahydrate was formulated in 25 mM phosphate pH 7 with 50 mM Captisol® at a strength of 2.61 mg/mL (1.11 mg/mL ciclopirox free acid), CPX-O was formulated in 25 mM phosphate pH 7 with 50 mM Captisol® at a strength of 1.94 mg/mL (1.50 mg/mL ciclopirox free acid).

JPET #257972

**Table 5: Mean ± SD Urine CPX Pharmacokinetic Parameters in Male Beagle Dogs following Single IV Doses of CPX-POM and CPX-O**

Parameter	Treatment <sup>a</sup>	
	CPX-POM	CPX-O
Body weight (kg)	9.65 ± 0.38	9.64 ± 0.34
CPX dose mg/kg) <sup>b</sup>	2.28 ± 0.01	3.10 ± 0.02
Test article dose (mg/kg) <sup>c</sup>	5.40 ± 0.02	4.01 ± 0.02
U <sub>CPX</sub> 0-8 hr (ng/mL)	838 ± 149	1,006 ± 370
U <sub>CPX</sub> 8-24 hr (ng/mL)	112.0 ± 56.1	72.5 ± 54.5
U <sub>CPX</sub> 0-8 hr (µM)	4.04 ± 0.72	4.86 ± 1.79
U <sub>CPX</sub> 8-24 hr (µM)	0.541 ± 0.271	0.350 ± 0.263
dXe/dt <sub>CPX</sub> 0-8hr (µg/hr)	9.88 ± 0.3.89	12.58 ± 5.61
dXe/dt <sub>CPX</sub> 8-24hr (µg/hr)	0.97 ± 0.19	0.43 ± 0.42
Xe <sub>CPX</sub> (µg)	94.5 ± 32.6	107.5 ± 39.9
Fe <sub>CPX</sub> (%)	0.428 ± 0.138	0.358 ± 0.123
Clr (mL/hr/kg) <sup>d</sup>	3.38 ± 1.80	2.82 ± 0.90
Fe <sub>CPX-G</sub> (%)	63.46 ± 6.68	62.67 ± 12.15

CPX, ciclopirox; CPX-G, ciclopirox glucuronide metabolite; Fe<sub>CPX</sub>, fraction of the administered dose excreted as CPX; dXe/dt<sub>CPX</sub>, mass of CPX excreted per hour over x hours following drug administration; FE<sub>CPX-G</sub>, fraction of the administered dose excreted as CPX-G; IV, intravenous; SD, standard deviation; U<sub>CPX</sub>, urine CPX concentration; Xe<sub>CPX</sub>, mass of CPX excreted in urine.

JPET #257972

- a N = 4 dogs receiving both treatments in a complete, nonrandomized, cross-over study design.
- b Ciclopirox free acid dose.
- c CPX-POM disodium heptahydrate was formulated in 25 mM phosphate pH 7 with 50 mM Captisol® at a strength of 2.61 mg/mL (1.11 mg/mL ciclopirox free acid), CPX-O was formulated in 25mM phosphate pH 7 with 50 mM Captisol® at a strength of 1.94 mg/mL (1.50 mg/mL ciclopirox free acid).
- d Renal clearance calculated per kg body weight as  $Cl_r = X_{e_{cpx}} / AUC_0^{\infty}$ .

JPET #257972

**Table 6: Mean ± SD Plasma CPX Pharmacokinetic Parameters in Male Sprague-Dawley Rats following Single Doses of IV CPX-POM, SC CPX-POM, and Oral CPX-O**

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POM SC	CPX-O Oral
Body weight (kg)	0.314 ± 0.012	0.316 ± 0.004	0.314 ± 0.013
CPX dose (mg/kg) <sup>b</sup>	19.98 ± 0.10	30.04 ± 0.06	29.78 ± 0.16
Test article dose (mg/kg) <sup>c</sup>	47.01 ± 0.24	70.68 ± 0.15	38.55 ± 0.21
C <sub>0</sub> (ng/mL)	33,569 ± 13,124		
C <sub>max</sub> (ng/mL)		5,005 ± 808	373 ± 158
T <sub>max</sub> (hr)		0.264 ± 0.123	0.250 ± 0.129
K <sub>el</sub> (hr <sup>-1</sup> )	1.529 ± 0.680	1.039 ± 0.253	1.234 ± 0.742
T <sub>½</sub> (hr)	0.537 ± 0.238	0.700 ± 0.163	0.801 ± 0.558
AUC <sub>0</sub> <sup>n</sup> (ng•hr/mL)	5,449 ± 756	6,548 ± 1007	211.2 ± 84.5
AUC <sub>0</sub> <sup>∞</sup> (ng•hr/mL)	5,496 ± 756	6,633 ± 1,019	378.0 ± 140.4
AUMC <sub>0</sub> <sup>n</sup> (ng•hr <sup>2</sup> /mL)	1,534 ± 457		
AUMC <sub>0</sub> <sup>∞</sup> (ng•hr <sup>2</sup> /mL)	1,687 ± 478		
MRT (hr)	0.285 ± 0.092		
Cl (mL/hr/kg)	3,696 ± 531		
V <sub>d</sub> (mL/kg)	3,160 ± 1696		

JPET #257972

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POM SC	CPX-O Oral
V <sub>ss</sub> (mL/kg)	1,106 ± 394		
F (%)		80.3	4.61

AUC<sub>0-n</sub>, area under the plasma CPX concentration versus time curve from time zero to the last measurable time point; AUC<sub>0-∞</sub>, area under the plasma CPX concentration versus time curve from time zero to infinity; AUMC<sub>0-n</sub>, CPX area under the first moment versus time curve from time zero to the last measurable time point; AUMC<sub>0-∞</sub>, CPX area under the first moment versus time curve from time zero to infinity; C<sub>0</sub>, extrapolated plasma CPX concentration at time zero following intravenous bolus administration; Cl, CPX systemic drug clearance; CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciciclopirox; F, CPX bioavailability; IV, intravenous; Kel, apparent first-order elimination rate constant; MRT, CPX mean residence time; SC, subcutaneous; SD, standard deviation; T<sub>1/2</sub>, apparent elimination half-life; V<sub>d</sub>, CPX apparent volume of distribution; V<sub>ss</sub>, CPX apparent steady state volume of distribution.

<sup>a</sup> N = 6 rats per treatment.

<sup>b</sup> CPX free acid dose.

<sup>c</sup> CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 23.53 mg/mL (10.0 mg/mL ciclopirox free acid); ciclopirox olamine was formulated in Orasweet® SF and Water at a strength of 29.39 mg/mL (22.7 mg/mL ciclopirox free acid).

JPET #257972

**Table: 7 Mean ± SD Urine CPX and CPX-G Pharmacokinetic Parameters in Male Sprague-Dawley Rats following Single Doses of IV CPX-POM, SC CPX-POM, and Oral CPX-O**

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POP SC	CPX-O Oral
Body weight (kg)	0.276 ± 0.008	0.293 ± 0.007	0.289 ± 0.010
CPX dose (mg/kg) <sup>b</sup>	20.02 ± 0.07	29.97 ± 0.18	38.89 ± 0.23
Test article dose (mg/kg) <sup>c</sup>	47.10 ± 0.16	70.53 ± 0.08	30.44 ± 0.18
U <sub>CPX</sub> (ng/mL) 0-24 hr	20,623 ± 11,948	5,931 ± 5689	1,711 ± 1,427
U <sub>CPX</sub> (µM) 0-24 hr	99.63 ± 57.72	28.65 ± 27.48	8.27 ± 6.89
Xe <sub>CPX</sub> (mcg) 0-24 hr	190.5 ± 103.0	85.44 ± 65.89	22.30 ± 16.65
Fe <sub>CPX</sub> (%)	3.46 ± 1.90	0.98 ± 0.76	0.25 ± 0.18
dXe/dt <sub>CPX</sub> (µg/hr) 0-8 hr	18.36 ± 11.6	6.22 ± 6.50	0.63 ± 0.16
dXe/dt <sub>CPX</sub> (µg/hr) 8-24 hr	2.76 ± 1.79	2.22 ± 2.76	1.08 ± 1.11
Cl <sub>r</sub> (mL/hr)	34.6		
Cl <sub>r</sub> (mL/hr/kg)	126		
Fe <sub>CPX-G</sub> (%)	9.35 ± 9.05	13.73 ± 8.83	9.13 ± 4.28

Cl<sub>r</sub>, renal clearance of the drug from plasma; CPX, ciclopirox; CPX-G, ciclopirox glucuronide metabolite; Fe<sub>CPX</sub>, fraction of the administered dose excreted as CPX; dXe/dt<sub>CPX</sub>, mass of CPX excreted per hour over x hours following drug administration; FE<sub>CPX</sub>, fraction of the administered dose excreted as CPX; IV, intravenous; SC, subcutaneous; SD, standard deviation; U<sub>CPX</sub>, urine CPX concentration; Xe<sub>CPX</sub>, mass of CPX excreted in urine.

JPET #257972

- a N = 4 rats who received all three treatments in a complete, nonrandomized, crossover fashion.
- b CPX free acid dose.
- c CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 23.53 mg/mL (10.0 mg/mL ciclopirox free acid).



JPET #257972

**Table 8: Mean ± SD Plasma CPX Pharmacokinetic Parameters in Male Beagle Dogs following Single Doses of IV CPX-POM, SC CPX- POM, and Oral CPX-O**

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POM SC	CPX-O Oral
Body weight (kg)	9.58 ± 0.49	9.91 ± 0.81	9.48 ± 0.64
CPX dose (mg/kg) <sup>b</sup>	3.10 ± 0.02	9.28 ± 0.09	9.39 ± 0.041
Test article dose (mg/kg) <sup>c</sup>	7.30 ± 0.04	21.88 ± 0.14	12.14 ± 0.05
C <sub>0</sub> (ng/mL)	3,770 ± 1280 <sup>d</sup>		
C <sub>max</sub> (ng/mL)		1,988 ± 374	1,017 ± 271
T <sub>max</sub> (hr)		0.264 ± 0.122	0.250 ± 0.129
K <sub>el</sub> (hr <sup>-1</sup> )	0.153 ± 0.072	0.137 ± 0.031	0.126 ± 0.009
T <sub>½</sub> (hr)	5.54 ± 2.92	5.27 ± 1.18	5.52 ± 0.39
AUC <sub>0</sub> <sup>n</sup> (ng•hr/mL)	3,622 ± 255	11,518 ± 2,632	1,901 ± 650
AUC <sub>0</sub> <sup>∞</sup> (ng•hr/mL)	5,262 ± 1243	19,614 ± 6161	2,697 ± 1013
AUMC <sub>0</sub> <sup>n</sup> (ng•hr <sup>2</sup> /mL)	8,836 ± 1,186		
AUMC <sub>0</sub> <sup>∞</sup> (ng•hr <sup>2</sup> /mL)	38,965 ± 29,492		
MRT (hr)	6.76 ± 3.76		
Cl (mL/hr/kg)	615 ± 145		
Vd <sub>z</sub> (mL/kg)	4,486 ± 1366		
V <sub>ss</sub> (mL/kg)	3,776 ± 1,266		

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POM SC	CPX-O Oral
F (%)		106 ± 22.3 <sup>e</sup>	17.4 ± 5.65

AUC<sub>0-n</sub>, area under the plasma CPX concentration versus time curve from time zero to the last measurable time point; AUC<sub>0-∞</sub>, area under the plasma CPX concentration versus time curve from time zero to infinity; AUMC<sub>0-n</sub>, CPX area under the first moment versus time curve from time zero to the last measurable time point; AUMC<sub>0-∞</sub>, CPX area under the first moment versus time curve from time zero to infinity; C<sub>0</sub>, extrapolated plasma CPX concentration at time zero following intravenous bolus administration; Cl, CPX systemic drug clearance; C<sub>max</sub>, maximum plasma CPX concentration following single dose administration; CPX, ciclopirox; CPX-O, ciclopirox olamine; CPX-POM, fosciclopirox; F, CPX bioavailability; IV, intravenous; Kel, apparent first-order elimination rate constant; MRT, CPX mean residence time; SC, subcutaneous; SD, standard deviation; T<sub>1/2</sub>, apparent elimination half-life; T<sub>max</sub>, time to maximal plasma CPX concentration; V<sub>d</sub>, CPX apparent volume of distribution; V<sub>ss</sub>, CPX apparent steady state volume of distribution.

- <sup>a</sup> N = 4 dogs who received all three treatments in a nonrandomized, complete cross-over fashion.
- <sup>b</sup> Ciclopirox free acid dose.
- <sup>c</sup> CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 3.53 mg/mL (1.50 mg/mL ciclopirox free acid), CPX-O was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 1.94 mg/mL (1.50 mg/mL ciclopirox free acid).
- <sup>d</sup> C<sub>0</sub> corresponds to y-intercept value derived from non-parametric pharmacokinetic data analysis.
- <sup>e</sup> Absolute bioavailability for SC CPX-POM was determined using AUC<sub>0-n</sub><sup>f</sup> given 39% of AUC<sub>0-∞</sub><sup>f</sup>

JPET #257972

was comprised of extrapolated AUC.

JPET #257972

**Table 9: Mean  $\pm$  SD Urine CPX and CPX-G Pharmacokinetic Parameters in Male Beagle Dogs following Single Doses of IV CPX-POM, SC CPX-POM, and Oral CPX-O**

Parameter	Treatment <sup>a</sup>		
	CPX-POM IV	CPX-POP SC	CPX-O Oral
Body weight (kg)	9.58 $\pm$ 0.49	9.91 $\pm$ 0.81	9.48 $\pm$ 0.64
CPX dose (mg/kg) <sup>b</sup>	3.10 $\pm$ 0.02	9.28 $\pm$ 0.09	9.39 $\pm$ 0.04
test article dose (mg/kg) <sup>c</sup>	7.30 $\pm$ 0.04	21.88 $\pm$ 0.14	12.14 $\pm$ 0.05
U <sub>CPX</sub> (ng/mL) 0-24 hr	957 $\pm$ 508	2,530 $\pm$ 958	1,602 $\pm$ 750
U <sub>CPX</sub> ( $\mu$ M) 0-24 hr	4.62 $\pm$ 2.45	12.2 $\pm$ 4.6	7.74 $\pm$ 3.62
Xe <sub>CPX</sub> (mcg) 0-24 hr	208 $\pm$ 117	339 $\pm$ 206	299 $\pm$ 208
Fe <sub>CPX</sub> (%)	0.70 $\pm$ 0.38	0.38 $\pm$ 0.24	0.32 $\pm$ 0.21
Xe/dt <sub>CPX</sub> ( $\mu$ g/hr) 0-8 hr	19.8 $\pm$ 15.2	23.0 $\pm$ 16.6	31.6 $\pm$ 26.2
Xe/dt <sub>CPX</sub> ( $\mu$ g/hr) 8-24 hr	3.10 $\pm$ 3.33	9.67 $\pm$ 9.49	2.90 $\pm$ 1.70
Cl <sub>r</sub> (mL/hr/kg) <sup>d</sup>	4.47 $\pm$ 3.35		
Fe <sub>CPX-G</sub> (%)	49.2 $\pm$ 34.47	43.0 $\pm$ 19.7	37.0 $\pm$ 18.7

CPX, ciclopirox; CPX-G, ciclopirox glucuronide metabolite; CPX-O, ciclopirox olamine;

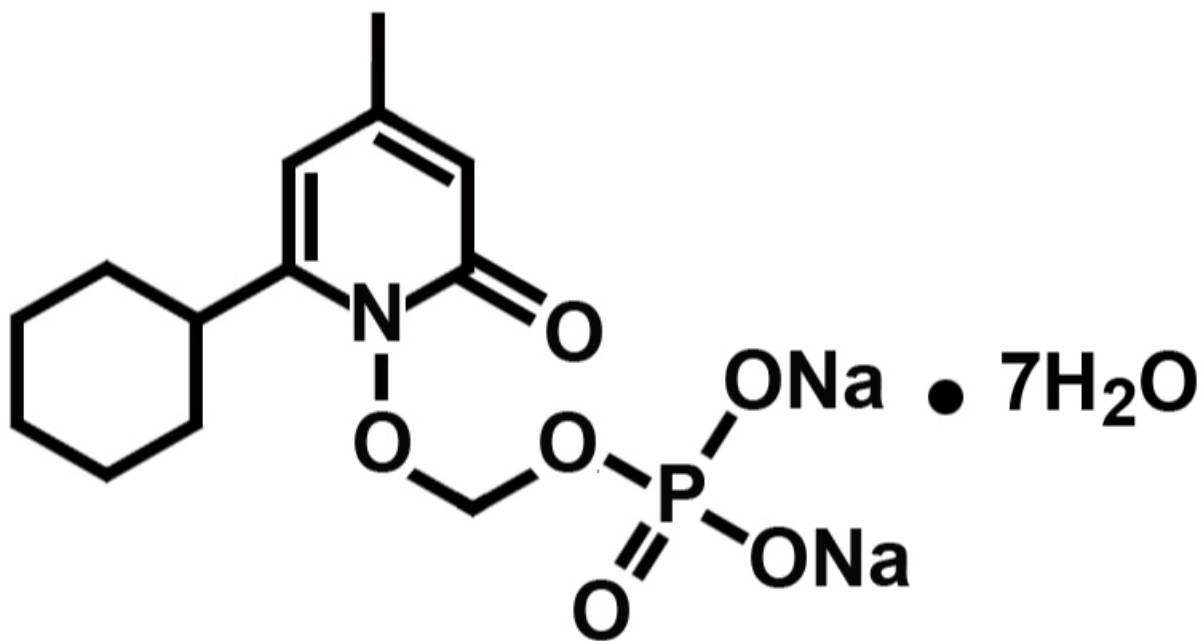
CPX-POM, fosciclopirox; Fe<sub>CPX</sub>, fraction of the administered dose excreted as CPX; dXe/dt<sub>CPX</sub>, mass of CPX excreted per hour over x hours following drug administration; FE<sub>CPX-G</sub>, fraction of the administered dose excreted as CPX-G; SC, subcutaneous; IV, intravenous; SD, standard deviation; U<sub>CPX</sub>, urine CPX concentration; XE<sub>CPX</sub>, mass of CPX excreted in urine.

<sup>a</sup> N = 4 dogs who received all three treatments in a nonrandomized, complete crossover fashion.

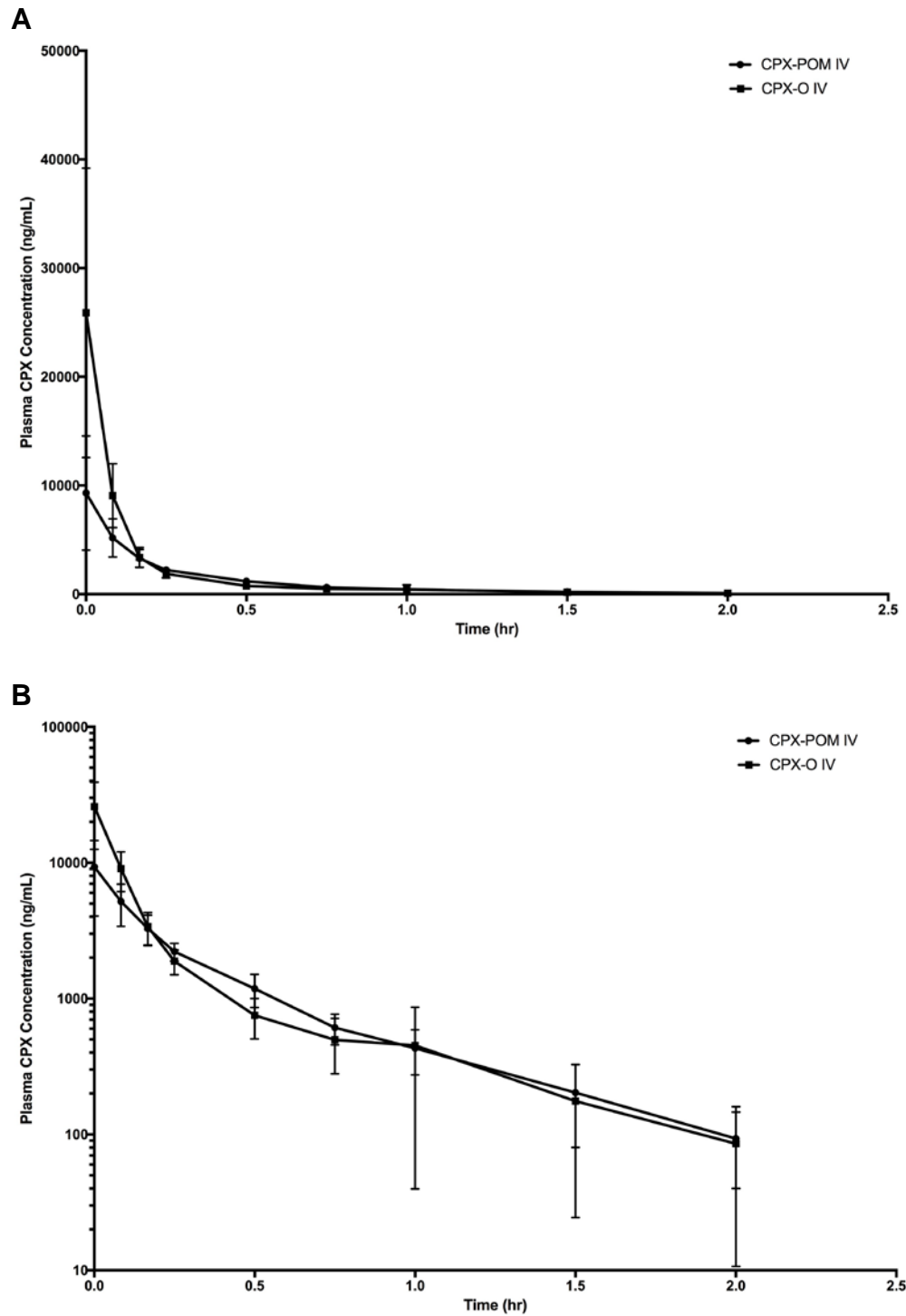
JPET #257972

- b Ciclopirox free acid dose.
- c CPX-POM disodium heptahydrate was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 3.53 mg/mL (1.50 mg/mL ciclopirox free acid), CPX-O was formulated in 25 mM phosphate buffer pH 7 with 50 mM Captisol® at a strength of 1.94 mg/mL (1.50 mg/mL ciclopirox free acid).
- d Renal clearance calculated per kg body weight as  $Cl_r = X_{e_{cpx}} / AUC_0^{\infty}$ .

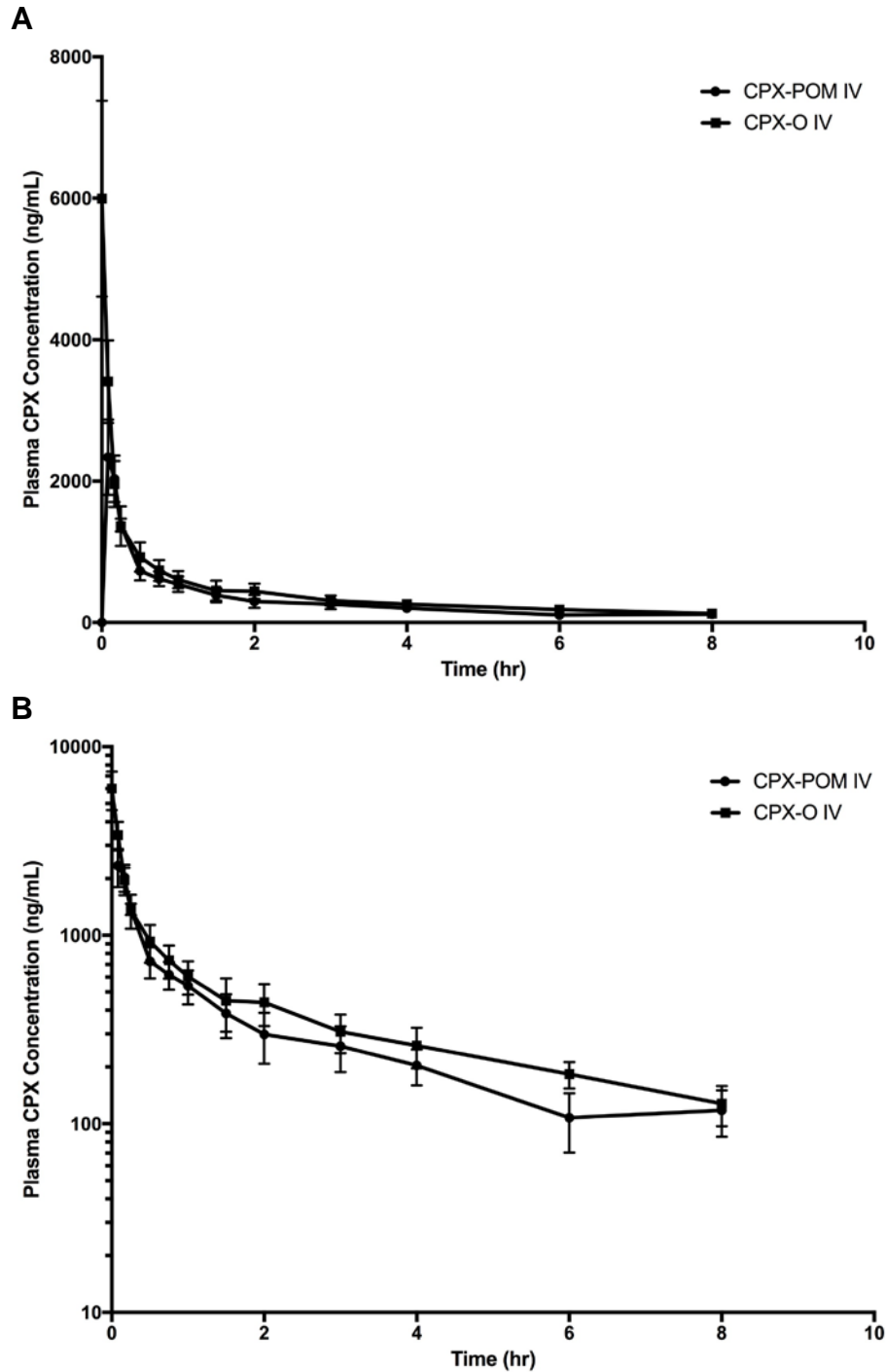
**Figure 1: Chemical Structure of Fosciclopirox ((6-cyclohexyl-4-methyl-2-oxopyridin-1(2H)-yl) oxy) methyl phosphate heptahydrate (CPX-POM).**



**Figure 2: Mean  $\pm$  SD Plasma CPX Concentration-Time Profiles Following Single Dose IV Administration of 17.5 mg/kg CPX-POM and 13 mg/kg CPX-O to Male Sprague-Dawley Rats**

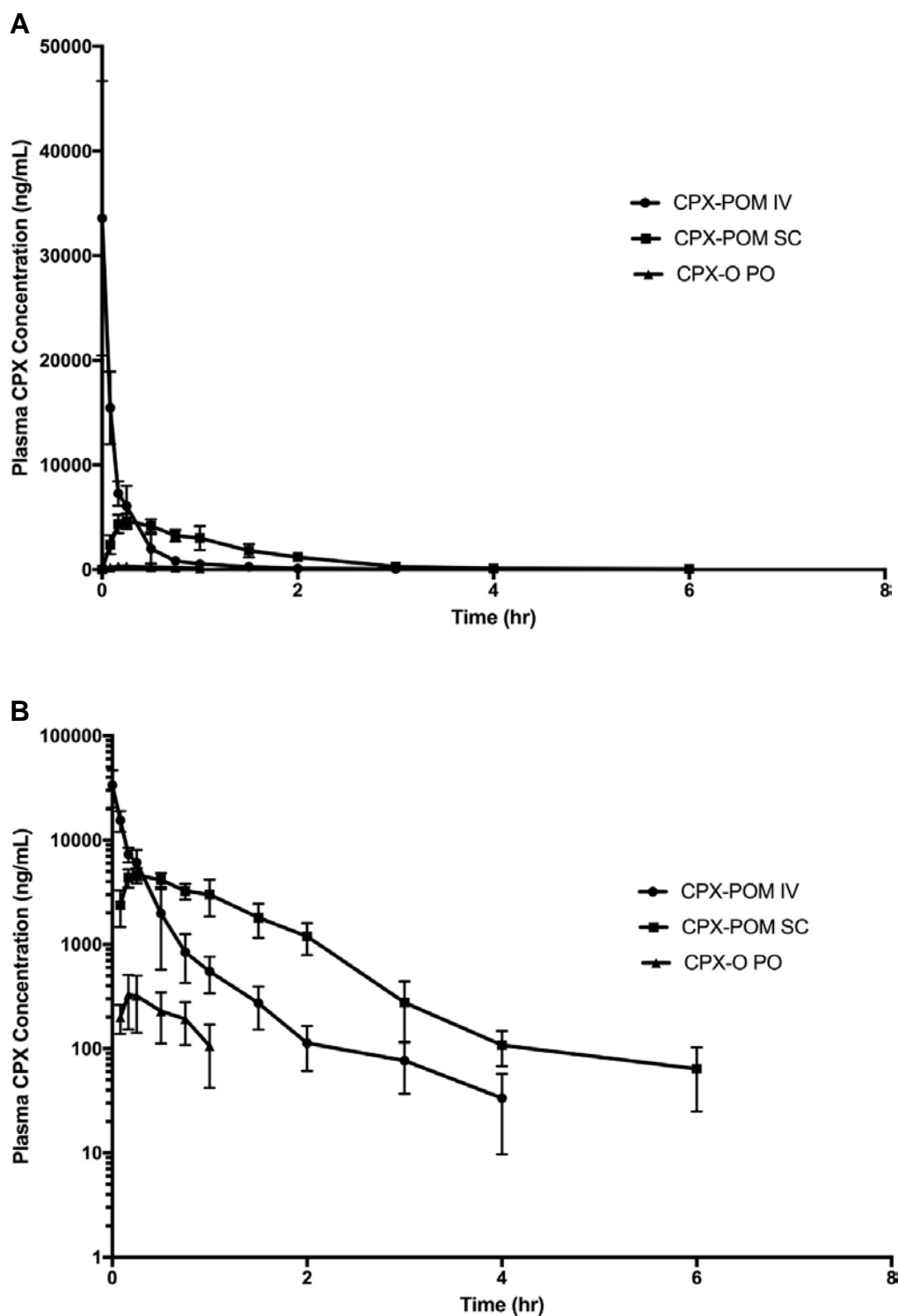


**Figure 3: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following Single Dose IV Administration of 5.4 mg/kg CPX-POM and 4.0 mg/kg CPX-O to Male Beagle Dogs**





**Figure 4: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following Administration of 47 mg/kg IV CPX-POM, 70.6 mg/kg SC CPX-POM, and 38.8 mg/kg CPX-O to Six Rats per Treatment Group**



**Figure 5: Mean  $\pm$  SD Plasma CPX Concentration-Time Rectilinear Profiles following IV Administration of 7.3 mg/kg CPX-POM, SC Administration of 21.8 mg/kg SC CPX-POM, and 12.2 mg/kg Oral CPX-O to Four Male Beagle Dogs**

