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Liraglutide improves renal endothelial function in obese Zucker rats on a high-salt diet

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List of non-standard abbreviations: ACh: Acetylcholine; ANG-II: Angiotensin-II; ANOVA: One-way analysis of variance; BW: Body weight; BUN: Blood urea nitrogen; CKD: Chronic kidney disease; COX: Cyclooxygenase; DBP: Diastolic blood pressure; EDHF: Endothelium derived hyperpolarizing factor; ET-1: endothelin-1; eNOS: endothelial nitric oxide synthase; FN-1: fibronectin-1; FITC: fluorescein isothiocyanate; GFR: Glomerular filtration rate; GLP-1R: Glucagon like peptide-1 receptor; GPX-1: Glutathione peroxidase-1; ID: Internal diameter; Indo: Indomethacin; LIRA: Liraglutide; NADPH: Nicotinamide adenine dinucleotide phosphate; NF- κ B: Nuclear factor kappa B; NO: Nitric oxide; Nox-1: NADPH oxidase 1; NT: Nitrotyrosine; NF- κ B: nuclear factor kappa B; PE-50: Polyethylene catheter; ROS: Reactive oxygen species; SBP: Systolic blood pressure; SNP: Sodium nitroprusside; TGF- β 1: Transforming growth factor; Vcam-1: Vascular cellular adhesion molecule-1; VEGF: Vascular endothelial growth factor

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Abstract

Metabolic syndrome is a common risk factor for chronic kidney disease. We investigated whether liraglutide (LIRA, glucagon like peptide (GLP)-1 receptor agonist) treatment improved renal vascular function and renal remodeling in male Zucker rats on a high-salt diet (6% NaCl). 8-week old Zucker lean (+/+) and Zucker obese (fa/fa) rats were treated with vehicle or LIRA (0.1mg/kg/day) for 8 weeks on high-salt diet. Glomerular filtration rate (GFR) was measured at 0 and 8 weeks using FITC-sinistrin method in conscious rats. We used X-ray microangiography to measure renal arterial vessel diameter (70-350µm) and vessel number *in vivo* in anesthetized rats. Renal protein expression levels of NT, CD-68, eNOS, VEGF, TGF-β1, COX-2 and GLP-1R were assessed by Western blotting. Renal gene expressions were determined by real time-PCR. In contrast to vehicle-treated, fa/fa-LIRA rats improved GFR, nitric oxide (NO)-mediated vasodilation in response to ACh and SNP in small arterial vessels (<200 µm diameter). LIRA treatment increased vessel responsivity to NO donors in comparison to vehicle treatment. Increases in the expressions of proinflammatory, profibrotic and oxidative stress related genes in fa/fa rats relative to +/+ were unaltered by LIRA, other than a trend for attenuation of VCAM-1 gene expression. However, LIRA treatment increased protein expressions of eNOS ($p=0.014$) and VEGF ($p=0.063$), while reducing glomerular macrophage infiltration in comparison to vehicle-treated fa/fa rats. Low-dose LIRA treatment improved renal vascular function through amelioration of vascular dysfunction and improved NO-mediated dilation of small intrarenal arteries and arterioles, and a reduction in renal inflammation.

Introduction

With the increasing prevalence of obesity in modern societies, and as a consequence of associated co-morbidities, obesity is one of the leading global causes of cardiovascular morbidity and mortality. Obesity leads to insulin resistance, vascular oxidative stress, endothelial dysfunction and consequently, reduced availability of vascular nitric oxide (NO), and impaired dilation in the microcirculation of many vascular beds, which contributes to the development of multiple cardiovascular complications including chronic kidney disease (CKD) [Sato, 2012; Rao et al., 2015]. An increasing number of the population that are obese and hypertensive develop CKD [Wu et al., 2012]. In the obese population, salt-sensitive hypertension is strongly associated with the progression of target-organ damage and renal injury associated with glomerulosclerosis, inflammation and albuminuria [Quigley et al., 2009; Wei et al., 2017]. It has also been established that a high-salt diet directly affects function of the renal vascular system in part through elevated generation of reactive oxygen species (ROS) and inflammation, independent of blood pressure changes [de Cavanagh et al., 2010]. Various experimental studies provide evidence that a high-salt intake directly impacts on the balance of production of NO and transforming growth factor (TGF)- β 1 [Sanders, 2009a; Patel et al., 2016]. Elevated endothelial TGF- β 1 production contributes directly to the development of hypertension, which in a state of endothelial dysfunction, and a pro-inflammatory and an oxidative endothelial cell environment leads to diminished NO-mediated negative regulation of TGF- β 1 effects in the vasculature, and ultimately results in glomerular fibrosis and exacerbation of vascular dysfunction [Sanders, 2009a, b; Meng et al., 2010]. Moreover, studies have shown that dietary salt intake also exacerbates insulin resistance and metabolic syndrome could play a major role in renal function decline [Garg et al., 2011; Oh et al., 2015]. The Zucker obese rat model has been shown to develop hypertension associated with impaired kidney function on a high salt diet (Ali et al., 2015;

Maheshwari et al., 2018). The combination of obesity and high-salt intake thus contributes to the acceleration of renal vascular dysfunction independent of blood pressure changes.

Glucagon-like peptide-1 (GLP-1) acts through the receptor (GLP-1R), which is a G-protein coupled receptor expressed on coronary endothelial cells, cells of the gastrointestinal tract, vascular smooth muscle, proximal tubules and glomeruli of the kidney [Ban et al., 2008]. Recent work has shown that in the streptozotocin (STZ) treated rat and in the diabetic nephropathy-prone KK/Ta-Akita mouse model treatment with a GLP-1R agonist, liraglutide (LIRA), has renoprotective effects [Hendarto et al., 2012; Fujita et al., 2014]. This was primarily attributed to an anti-oxidant effect mediated through inhibition of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, normalization of TGF- β 1 and elevation of cAMP/protein kinase A levels [Hendarto et al., 2012; Fujita et al., 2014]. Others have also reported that the protective effects of GLP-1 agonists in glomerular endothelial cells are mediated by inhibition of angiotensin (ANG)-II via AMP-dependent protein kinase mediated cRaf phosphorylation [Mima et al., 2012]. However, there are no *in vivo* studies that show the effects of sustained GLP-1 activation on functional changes in small renal arteries and arterioles, which play the most important role in regulating renal blood flow and pressure, or the effects on renal vascular remodeling. Diabetic nephropathy and CKD are often associated with arterial and glomerular arteriole hyalinosis (wall thickening with hyaline deposition) and endothelial damage [Yuzawa et al., 2008]. Moreover, no studies that we are aware of, have determined whether GLP-1 agonist can ameliorate renal dysfunction in an experimental model of obesity combined with high-salt intake.

We have successfully developed and validated effective approaches to assess segmental vascular structure and function, and blood flow distribution in the kidney in rodents using *in vivo* microangiography with synchrotron radiation [Eppel et al., 2009; Shirai et al., 2013]. Taking advantage of this lab X-ray system, we investigated herein whether

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sustained GLP-1 activation is protective against the renal vascular dysfunction and CKD in an obese rat model on a high-salt diet. Therefore, our aims were 1) to directly assess renal endothelial and smooth muscle dysfunction within the renal vascular network of male obese/lean Zucker rats maintained on a high-salt diet using *in vivo* micro-angiography; 2) to assess the direct impact of sustained GLP-1 activation on renal vascular function, glomerular filtration rate (GFR), albumin creatinine ratio and blood urea nitrogen (BUN) levels, and 3) to appraise the overall protective effect on GLP-1 agonist treatment on ROS and inflammation within the renal vascular network. Our hypothesis was that LIRA ameliorates the increased renal vasoconstriction and renal dysfunction evoked by a high-salt diet in fa/fa rats directly through increased protein expression of renal endothelial NO synthase (eNOS), enhanced endothelium-dependent NO-mediated dilation in renal small arteries and indirectly by lowering arterial blood pressure relative to vehicle treated rats.

Material and Methods

Animals and group allocation

All animal experiments were performed in accordance with the National Cerebral and Cardiovascular Centre (Osaka, Japan) animal care committee (approval number: 16086) according to National Institutes of Health guidelines. Thirty-two 8-week old male Zucker lean (+/+) and Zucker obese (fa/fa) rats were purchased from Japan SLC (Hamamatsu, Japan). Zucker +/+ rats were divided into two groups to receive either saline (Ln-Veh) or LIRA (0.1 mg/kg/day) (Ln-LIRA), and Zucker fa/fa rats were divided into two groups to receive either saline (fa/fa-Veh) or LIRA (0.1 mg/kg/day) (fa/fa-LIRA) for 8 weeks. Vehicle and LIRA were administered subcutaneously via osmotic mini-pumps (2ML-4, Alzet, CA, USA). Clinically relevant dose of LIRA was selected based on previous studies [Hendarto et al., 2012; Mima et al., 2012; Fujita et al., 2014]. All rats were provided with ad libitum access to normal drinking water and a high-salt chow diet (6% NaCl w/v, Oriental Yeast Co., Ltd,

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Tokyo, Japan). LIRA (Victoza[®]) was purchased from Japan. High-salt diet and LIRA treatment were initiated at the same-time to determine the protective beneficial effects against the high-salt induced renal dysfunction. At completion of the main study a supplementary angiography experiment was then performed to investigate the renal vascular response to ACh and SNP in Zucker +/+ rats (n=5) on a normal salt chow diet (NaCl 0.44% w/v).

Blood pressure measurements

Blood pressure was measured non-invasively in all conscious rats using the tail-cuff method (BP-98A-L, Softron Corporation, Tokyo, Japan) in 36-37°C pre-heated platform for about 10-15 minutes following training sessions prior to recording days. The mean of 4-5 systolic blood pressure (SBP) recordings were determined for each rat at the end of the 8-week treatment following recordings at the start and mid-study for accustomization.

Transcutaneous GFR measurement in conscious rats

Fluorescein isothiocyanate-(FITC)-sinistrin half-life was measured prior to high-salt diet feeding and after 8-weeks of vehicle or LIRA treatment on the high-salt diet. Animals were induced with 4% isoflurane and anesthesia maintained at 2% until the completion of device fixation and tail vein injection. For the measurement of GFR, the transcutaneous NIC-kidney device (Mannheim Pharma & Diagnostics GmbH, Mannheim, Germany) was fixed to the depilated skin on the back of rats using a double-sided adhesive patch following procedures described previously [Herrera Perez et al., 2016]. FITC-sinistrin was dissolved in physiological saline and administered through tail-vein injection (10 mg/100 g body weight, Fresenius Kabi Austria GmbH, Linz, Austria) [Ellery et al., 2014]. Animals were allowed to fully recover and move freely within a cage until transcutaneous recordings were stopped after 2 hours 15 minutes. FITC-sinistrin excretion half-life ($t_{1/2}$) was calculated using a one-compartment model (MPD Lab 1.0 Version, Mannheim Pharma & Diagnostics GmbH,

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Mannheim, Germany) conversion equation as previously described [Herrera Perez et al., 2016].

Measurement of albumin, albumin creatinine ratio and BUN

Urinary levels of albumin and albumin-creatinine ratio were measured in a single rapid colorimetric assay format using a Spectrum Diagnostic kit (Ref: 06072885-6010A, Siemens DCA 2000 Vantage analyzer, Eschborn, Germany) according to the manufacturer's protocol. BUN was measured by colorimetric qualitative analysis using commercially available Fuji Dri Chem slides (DRI-CHEM 7000, Fujifilm, Tokyo, Japan).

Measurement of non-fasted insulin and glucose levels

Insulin was determined on frozen plasma samples using a Rat Insulin enzyme-linked immunosorbent assay (ELISA) kit (AKRIN-010RU, Fujifilm Wako Pure Chemical Corporation, Tokyo, Japan) according to manufacturer recommendations. Blood glucose levels were measured using a LifeScan glucometer (Miltipias, CA, USA).

Surgical preparation of rats for microangiography

Rats were induced with 4% isoflurane and anesthesia maintained at 2.5% isoflurane throughout surgery. After anesthesia was achieved, the trachea was intubated with a angiocath sheath and connected to a small animal ventilator (Model SN-480-7, Shinano Seisakusho Co., Ltd, Tokyo, Japan). Ventilation was volume controlled at a frequency of 60 per min with a tidal volume 8-10 ml/kg of body weight (BW). Rectal temperature was monitored and regulated at 36-37°C using a combination of heating pad and or heating lamp throughout the experimental procedure. The right common carotid artery was cannulated with a polyethylene catheter (PE-50) to record mean arterial blood pressure and heart rate via pressure transducer (MLT 0670, AD instruments, Castle Hill, Australia). Then the right jugular vein was cannulated for continuous infusion of vehicle Ringer's lactate solution (4 ml/h) (Otsuka Pharmaceuticals, Chiyoda, Japan) and drug solutions used to assess vascular

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function. The right femoral artery was also cannulated retrogradely, and the tip of the catheter (BD medical Technology, UT, USA) was positioned just below the level of the left renal artery for contrast medium infusion.

X-ray microangiography system and preparation

Following the surgical preparation, the rats were placed in the microfocus X-ray system (MFX-80HK, Hitex, Osaka, Japan) for terminal microangiography experiments under isoflurane anesthesia (2%). The X-ray system consisted of an open-type 1- μ m microfocus X-ray source (L9191, Hamamatsu Photonics, Hamamatsu, Japan) and a 50/100 mm dual mode X-ray image intensifier with a CCD camera (E5877JCD1-2 N, Toshiba, Tokyo, Japan) operating at 80 kV and 40 μ A current. Micro-angiography image sequences were recorded digitally at a rate of 30 frames/s as a series of 640×480 pixel bitmap image files. Each supine rat was positioned on the sample plate above the X-ray source so that the left kidney was positioned in center of the X-ray detector field of view. Through an abdominal incision, viscera and fat pads were transected away for an uninterrupted view of the left kidney to avoid superimposition of mesenteric arteries. Vessel internal diameter (ID) determination was calibrated by placing a 50 μ m calibration tungsten wire on the surface of the exposed left kidney, which was kept moist with topical application of glycerol. After verifying that the imaging field covered the left renal arteries and most of the left kidney, heparin diluted (100U/ml) iodinated contrast medium (Iomeron 350, Eisai, Tokyo, Japan) was injected intra arterially as a bolus (250~300 μ l at a speed of 30 ml/min) with a syringe injection pump (PHD-2000, Harvard Apparatus, MA, USA). X-ray image acquisition was initiated from one second before iodine injection and continued for five seconds. Subtraction angiogram image analysis was performed as previously described [Sonobe et al., 2015]. Rats were then overdosed with 4% isoflurane followed by potassium chloride injection and the kidney was excised for further analysis.

Renal angiography protocol and rationale

Renal angiograms were recorded sequentially at the end of a 5-minute baseline of vehicle infusion (lactate Ringer's 3.0ml/h), acetylcholine (ACh) (3.0 μ g/kg/min for 5 min) and sodium nitroprusside (SNP) (3.0 μ g/kg/min for 5 min), then following combined blockade of prostaglandin production with the cyclooxygenase (COX) inhibitor indomethacin (Indo, 10 mg/kg iv bolus) and uncoupling of myoendothelial gap junctions with carbenoxolone administration (carbenox, 10 mg/kg iv bolus) following a 10 min equilibration period. We then assessed endothelium-dependent dilation after blockade with a repeat infusion of ACh (3.0 μ g/kg/min) [Fig. 5B]. Aging and insulin resistance have both been shown to impair conducted dilation, which is the transmission of hyperpolarization along vessels via endothelial cell to cell connections and smooth muscle cell to cell connections involving various potassium channels and connexins [Feher et al., 2014; Chen et al. 2016]. A reduction in NO bioavailability has been shown to impair conducted dilation in coronary arterioles by reducing NO-mediated facilitation of outward extracellular K ion efflux and subsequent smooth muscle hyperpolarization [Feher et al., 2014]. Blockade with Indo+carbenox was used in this study to inhibit local radial dilation through prostanoids and myoendothelial gap junctions to determine if reduced NO availability due to metabolic syndrome impaired conducted dilation within the renal vasculature, and whether LIRA enhanced conducted dilation. Changes in internal vessel diameter (ID) or caliber of the same vessels that were visualized in all treatment periods are considered in this study to be indicative of local (radial) dilatory function. On the other hand, a change in the number of angiographically visualized vessel segments during stimulation (i.e. not seen at baseline) is used here as an index of a change in distal or conducted dilation to the renal cortex. Therefore, we expected that any significant decrease in vessel ID or lack of change in ID during stimulation of the renal vasculature with ACh or SNP would indicate that there is underlying endothelial

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dysfunction or smooth muscle dysfunction respectively, involving impairment of NO signaling. On the other hand, a reduction in newly visualized vessels during ACh or SNP stimulation or a lack of change in visualized vessel number would indicate that there is impairment in at least one of the mechanisms responsible for conduction of distal dilation and regulation of global perfusion (NO and EDHF). Conversely, an increase in newly visualized vessels during ACh stimulation in LIRA treated rats would indicate that the distal increase in perfusion is facilitated by enhanced release of NO and or EDHF induced by LIRA treatment.

Measurement of internal vessel diameter

Image analysis was performed as previously described using Image J (version 1.48, National Institute of Health, USA) [Sonobe et al., 2015]. Quantitative analysis of vessel ID was based on measurements from the middle of the discrete vessel segments in the subtracted average images obtained from individual rats during each treatment period.

Histological examination of the kidney

Following excision, 2/3 of right kidney was immersed in 4% paraformaldehyde for more than 48 hours and then embedded either in paraffin or OCT (Tissue Tek, OCT compound, USA), whereas the remaining 1/3 was frozen into liquid nitrogen for western blotting and stored at -80°C until assays were performed. Paraffin-embedded kidney sections were stained with Azan-Mallory stain. For evaluation of collagen fiber content (staining blue), images were taken randomly under light microscopy ($\times 400$) from 20 different microscopic fields per section using a BX53 microscope (Olympus, Tokyo, Japan) equipped with a CMOS camera (DP-80, Olympus, Japan).

Immunofluorescence

Frozen sections were dried at room temperature for 10 min and rinsed 3 times in PBS for 5 min. Sections were blocked for 1 h in blocking buffer (1% BSA and 0.1% Triton X100 diluted in PBS). Sections were then incubated with anti-mouse CD-68 (1:500; ab-31630)

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overnight at 4°C. The slides were washed 3 times for 5 min in PBS and incubated for 1 h at room temperature with the conjugated secondary antibody (11004A, goat anti-mouse Alexa Fluor 568, 1/400, Life Technologies, MA, USA).

Protein expression determination by Western blotting

Right kidney tissue samples were homogenized with lysis buffer. Renal protein concentrations in these homogenized samples were measured by using bicinchoninic acid (BCA) assay method. For Western blots, proteins were separated by SDS-PAGE and the following antibodies were used to quantify the renal protein expression levels: anti-VEGF (1:1000; sc-152); anti-TGF- β 1 (1:1000; sc-146) and anti-cyclooxygenase 2 (COX-2; 1:1000, NB100-689, Novus Biologicals, USA) rabbit polyclonal antibodies; anti- β -actin (1:1000, sc-47778), anti-endothelial nitric oxide synthase (eNOS; 1:1000, sc-376751) and anti-nitrotyrosine (NT; 1:1000, sc-32757); anti-GLP-1R (1:1000, sc-390773) mouse polyclonal antibodies (Santa Cruz Biotechnology, USA). For SDS-PAGE 50 μ g of protein were loaded on 10% sodium dodecyl sulfate-polyacrylamide gels (Bio-Rad, CA, USA), and subsequently the proteins were transferred to nitrocellulose membranes. These membranes were then blocked with Blocking One solution (30953-95, Nacalai Tesque Inc., Kyoto, Japan) or 3% BSA (Sigma, St. Louis, USA) in TBS-T (20 mM Tris, pH 7.6, 137 mM NaCl, and 0.05% Tween 20) at room temperature. After incubation with the primary antibody, the bound antibody was visualized with the respective horseradish peroxidase coupled secondary antibody (1:10000; #7076S; 7074S; Cell Signaling Technology, Inc., MA, USA). Chemiluminescence signals were visualized using a chemiluminescence imaging system (Ez-Capture-II, ATTO Corporation, Tokyo, Japan) and were quantified using β -actin signal as a protein loading control.

Gene expression analysis

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RNA extracted from kidney tissues using TRIzol ® lysis reagent in conjunction with the RNA Purification kit (#74104; RNeasy Mini Kit, QIAGEN, CA, USA) with on-column DNase treatment (# 79254; RNase-Free DNase Set, QIAGEN, CA, USA). The quantity of the purified RNA was measured by using the Nano Drop One Analyzer (Thermo Fisher Scientific, CA, USA) and the RNA quality from each sample was assured by the A260/280 absorption ratio. Two µg total RNA was reverse transcribed using the Complimentary DNA synthesis kit (#11754-050; Superscript ® VILO™, Invitrogen, Thermo Fisher Scientific, CA, USA) as per the manufacturer's instructions. Real time-PCR reactions were run on a StepOne 7500 RT-PCR system (Applied Biosystems, Foster City, CA, USA) using Taqman Universal Fast PCR Master Mix and gene specific predesigned Taqman primers obtained from Applied Biosystems [GenBank Accession Nos: Rn02132634_s1 (eNOS); Rn00561129_m1 (ET-1); Rn00569575_m1 (fibronectin (FN)-1); Rn01775763_g1 (Gapdh); Rn00577994_g1 (GPX-1); Rn00586652_m1 (NADPH oxidase (Nox)-1); Rn00572010_m1 (TGF-β1); Rn00563627_m1 (VCAM-1); Rn01511602_m1 (VEGF)] (Applied Biosystems, CA, USA). Each sample was run in duplicate, and the comparative threshold cycle ($\Delta\Delta CT$) method was used to analyze the genes of interest relative to the GAPDH reference gene as described [Livak et al., 2001].

Statistical analysis

Data are presented as mean \pm SEM. The mean baseline and percentage change in vessel ID and visible vessel number from baseline of each branching order in individual rats (within-animal average) were pooled for group comparisons. To compare the means of variables among the rat treatment groups we used either a one-way analysis of variance (ANOVA) or two-way repeated measures ANOVA to compare time dependent changes between groups, such as GFR, urine albumin, and urine albumin creatinine ratio over time. $p < 0.05$ was considered statistically significant. Further, parameters obtained from the four rat groups were subjected to two-way ANOVA to evaluate significance of differences due to either

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genotype (G, fa/fa vs. fa/+) or drug treatment (T, LIRA vs. vehicle) or their interaction (I, G x T). Prism version 5.0 software by GraphPad Software (La Jolla, CA, USA) was used for statistical analysis.

Results

LIRA treatment had no impact on body weight (BW) or blood glucose but improved plasma insulin levels

BW of rats was comparable within Ln and fa/fa groups respectively throughout the duration of the study, with no significant differences observed between LIRA, saline treated groups (Fig. 1A). Blood glucose levels were not significantly altered by long-term administration of LIRA, with no significant treatment effect observed (Fig. 1B). However, saline treated fa/fa rats showed greatly elevated plasma insulin levels when compared to Ln group rats, but LIRA treatment tended to decrease insulin levels in fa/fa rats (T $p=0.089$, Fig. 1C).

LIRA suppressed blood pressure elevation in conscious fa/fa rats

Fig. 1B demonstrates that 8 weeks after high-salt diet there was a trend for SBP to be mildly elevated in fa/fa-Veh group rats compared to Ln-Veh rats (133.4 ± 5.2 vs 126.1 ± 3.8 mmHg, $p=0.063$). In contrast, LIRA treated fa/fa rats had significantly lower SBP compared with those of fa/fa-Veh group rats (122.2 ± 3.5 vs 133.4 ± 5.2 mmHg, $p=0.013$) (Fig. 1D).

LIRA effect on GFR

GFR was higher during the baseline (0 week) in fa/fa rats compared to Ln rats (genotype $p=0.001$). After 8 weeks on high-salt diet, GFR in fa/fa-Veh group rats was significantly decreased in comparison to the Ln-Veh rats (0.82 ± 0.05 vs 1.50 ± 0.05 ml/min, $p<0.01$; Fig. 1E). LIRA treated fa/fa rats maintained near normal GFR in contrast to the vehicle treated fa/fa rats (1.33 ± 0.06 vs 0.82 ± 0.05 ml/min, $p<0.001$).

LIRA partially reduced BUN and albuminuria in fa/fa rats

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Urinary albumin and albumin-creatinine ratio were elevated significantly in fa/fa-Veh group rats compared to Ln-Veh rats (Fig. 1G-H). Eight weeks of LIRA treatment in the fa/fa rats resulted in decreased BUN levels relative to fa/fa Veh rats ($p=0.032$; Fig. 1F). Further, LIRA treatment partially decreased urinary albumin (49.9 ± 7.6 vs 74.5 ± 4.7 mg/L; $p<0.01$; Fig. 1G) and albumin/creatinine ratio (254.9 ± 54.6 vs 371.9 ± 24.7 mg/L; $p<0.001$; Fig. 1H).

LIRA did not alter the profibrotic and inflammatory gene expression profile

Renal gene expression analyses revealed that the obese phenotype was associated with a significantly higher expression of ET-1, NOX1, VCAM-1 and TGF- β 1 mRNA on a high salt diet (Fig. 2A-D). However, mRNA expression of the antioxidative pathway GPX-1, fibronectin (FN-1), and genes important for vascular homeostasis eNOS and VEGF were not different between fa/fa and Ln rats (Fig. 2E-H). LIRA did not alter gene expression for any gene examined, with the notable exception of VCAM-1, which showed a strong trend to be reduced by LIRA (treatment $p=0.061$).

LIRA treatment improved eNOS and VEGF expression without appreciably changing markers of nitrosative stress, prostanoids or TGF- β 1 levels

Renal protein expression of NT and VEGF did not differ significantly between vehicle treated fa/fa rats and Ln rats (Fig. 3A, B & F). The obese phenotype was associated with a pronounced increase in macrophage marker protein CD68 (Fig. 3C; genotype $p=0.007$) and COX-2 protein (Fig. 3D, genotype $p=0.0001$), while there were similar trends for increased TGF- β 1 (Fig. 3D) and eNOS (Fig. 3E; genotype $p=0.052$) protein. Treatment with LIRA did not significantly alter NT, CD68, TGF- β 1 or COX-2 protein levels (Fig. 3B-D, G), but did restore expression of VEGF (Fig. 3F; treatment $p=0.063$) and nearly doubled mean eNOS expression (Fig. 3E; treatment $p=0.014$) in fa/fa LIRA treated rats compared to fa/fa Veh and Ln rats.

LIRA treatment attenuated GLP-1R expression

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GLP-1R protein expression in the kidney was increased in fa/fa rats relative to Ln rats. Pronounced variability in protein expression levels were observed in LIRA treated fa/fa rats compared to vehicle treated rats, but LIRA significantly reduced receptor expression protein expression (Fig. 3A, H, treatment $p=0.046$). However, immunoreactive GLP-1R expressions was not detected by immunofluorescence in the Zucker rat kidney (data not shown), even though antibody was sufficiently sensitive to enable detection of endogenous GLP-1R protein expression in kidney tissue protein extract (Fig. 3A, H).

LIRA reduced tubulointerstitial fibrosis in fa/fa rats

Collagen accumulation was evaluated to assess the extent of fibrosis. In the kidneys of the rats from both Ln groups, collagen deposition (blue color) was observed in small amounts including around the glomeruli, vessels and the borders of the tubules (Fig. 4A). Compared with Ln rats, collagen accumulation was significantly increased in fa/fa-Veh rats (Fig. 4B), even though the levels of TGF- β 1 protein upregulation did not reach statistical significance (Fig. 3A and D). In contrast, collagen accumulation was partially attenuated by LIRA treatment (Fig. 4A, C).

LIRA suppressed renal macrophage infiltration and glomerular enlargement

Fig. 4B presents representative immunofluorescence images of macrophages within kidneys. In fa/fa-Veh group rats CD-68 positive cells were observed significantly more often within the glomeruli compared with Ln rats. Macrophage accumulation was attenuated by LIRA treatment ($p<0.001$; Fig. 4B, D). However, Western blotting of renal tissue did not reveal a statically significant reduction in CD-68 marker protein (Fig. 3A and C). Although all rats were on a high salt diet, the glomeruli in fa/fa Veh rats were ~40% larger than in Ln rats (by cross sectional area and perimeter). LIRA treatment ameliorated the expansion of glomerular volume compared to fa/fa-Veh rats (Fig. 4E-F).

Baseline hemodynamics

Mean arterial pressure (MAP) during baseline was slightly elevated in vehicle-treated fa/fa rats than Ln rats (118.7 ± 9.8 vs 102.3 ± 6.2 mmHg) while mean heart rate (MHR) was comparable (323.6 ± 20.8 vs 350.9 ± 11.7 bpm) (Fig. 6A). LIRA treatment did not alter the MAP at baseline in fa/fa rats (122.9 ± 9.6 vs 118.7 ± 9.8 mmHg), but increased MHR (362.1 ± 12.1 vs 323.6 ± 20.7 bpm) compared to vehicle-treated fa/fa rats (Fig. 6A).

Vessel internal diameters and visualized vessel number at baseline

Representative X-ray angiograms of the renal vasculature are shown in Fig. 5A. Vessel ID in Ln-Veh rats were comparable to Ln-LIRA rats across 1st order, 2nd order and 3rd order arterial vessels. Veh treated fa/fa rat vessel ID was similar to LIRA treated fa/fa rat vessel ID across 1st to 3rd order branches of arterial vessels (Fig. 5C). Mean total visible vessel branching segments per animal during all treatment periods in the field of view, was 10 to 12 vessels in Ln rats, as compared to 6-8 in fa/fa-Veh group rats and 9-12 in LIRA treated fa/fa rats. More 3rd order vessel branching segments were observed in LIRA treated fa/fa rats at baseline than vehicle-treated fa/fa rats (Fig. 5A, D).

LIRA treatment improved renal dilation to endothelium-dependent and independent stimulations

Fig. 7A shows the mean change in vessel caliber across 1st to 3rd order branching during ACh infusion. Caliber increases in fa/fa-Veh rats were smaller than Ln rats across 2nd and 3rd order vessels. LIRA treatment significantly increased dilation in the 3rd order vessels and there was a similar trend in the 2nd order, but there were no significant differences in the large arteries of the first branching order. Further there was tendency for the number of visible 2nd order vessels to increase with ACh stimulation (Fig. 7A; $p=0.07$). There was a trend for a larger decrease in MAP in fa/fa rats in response to ACh than Ln rats (-12.8 ± 7.6 vs -1.1 ± 3.3 mmHg, genotype $p=0.078$, Fig. 6C). However, LIRA treated rats did not differ significantly in the size of the MAP change from vehicle-treated rats. Hence, the larger dilator response to

ACh in LIRA treated rats was not due to differences in arterial pressure. Ln-Veh group rats essentially showed no response to SNP across any branching order, and fa/fa-Veh rats showed slight constriction in the smallest vessels (Fig. 7B). On the other hand, SNP evoked potent dilation in LIRA treated rats that was greatest in the 3rd order vessels in Ln rats along with a small increase in visible 3rd order vessels. MAP changes during SNP infusion were similar across all groups, suggesting that the potent dilation only in LIRA rat groups was not due to differences in arterial pressure (Fig. 6C). However, there was a non-significant modest depression of MHR during SNP in the fa/fa-LIRA rats. Overall, compared to the vehicle-treated rats, the LIRA treated rats exhibited a significant enhancement of the vasodilator responses within the renal vasculature that was not directly related to arterial pressure changes at the time of imaging.

In light of the absence of calibre changes to SNP in the Ln-Veh treated rats we investigated whether this was due to strain influences or the effect of the high salt diet in +/- rats on a normal chow diet. We found that naïve +/- rats had slightly smaller calibres, but not significantly so, in both 2nd and 3rd order vessels compared to the Ln-Veh rats (Fig. 8B). In general, renal vessels in the naïve +/- rats showed a 20-30% increase in calibre with a significant increase in the number of perfused 3rd order vessel during ACh stimulation (Fig. 8D), whereas calibre change during SNP stimulation was ~10% in the renal vessels (Fig. 8E). The latter finding suggests that the sensitivity to NO donors in the renal vascular bed is low in naïve Zucker rats and mildly exacerbated by exposure to a high-salt diet (main study).

LIRA treatment enhanced residual local dilation following blockade of prostaglandins and myoendothelial gap junctions

MAP decreased significantly in fa/fa rats during blockade, while in Ln-Veh rats MAP increased by ~18 mm Hg (genotype $p=0.011$). Vessel caliber of vehicle-treated fa/fa rats showed no change from baseline in 1st and 2nd order vessels, but a tendency to be constricted

in the 3rd order following blockade (Fig. 7C). In contrast, LIRA treatment resulted in a significantly larger caliber in 3rd order vessels, ~20% greater than at baseline ($p=0.023$), which did not differ between genotypes. Furthermore, LIRA-treated rats maintained similar visible vessel numbers in the 2nd and 3rd branching order during blockade as baseline, while vehicle-treated rats showed reduction in 2nd order vessels ($p=0.089$). These findings suggest that dilator prostaglandins most likely contribute to the maintenance of basal renal perfusion in small to medium sized arteries in the Veh treated rats irrespective of obesity. However, the small calibre constriction in 3rd order vessels in fa/fa-Veh rats is likely to be in part due to the small arterial pressure reduction during blockade (Fig. 6C). ACh stimulation during blockade of COX and uncoupling of myoendothelial gap junctions evoked a similar degree of increase in vessel ID as that seen during ACh stimulation prior to blockade (Fig. 7D) with only a moderate decrease in MAP observed in Ln rats (Fig. 6B, interaction $p=0.016$). While local radial dilation was observed in all rats during the final ACh stimulation, we observed that there was a trend for greater dilation 1st order large arteries in LIRA treated rats, and moreover, a significantly larger dilation in the 3rd order resistance vessels in LIRA treated rats. There was only a weak trend for an increase in the number of visible 2nd order arteries during ACh stimulation post blockade in LIRA treated rats. Thus, these findings taken together suggest that LIRA enhanced the low sensitivity of renal smooth muscle that we observed in all Veh-treated rats on a high-salt diet and significantly enhanced endothelium-dependent dilation to ACh to a similar extent in Ln and fa/fa rats, but NO-mediated dilation was generally greater in the Ln rats. On the other hand, there was little evidence that LIRA modified the extent of conducted dilation within the kidney of Zucker rats.

Discussion

In the present study, we investigated whether chronic LIRA treatment is able to improve renal vascular function in a rat model of metabolic syndrome and hypertension in part through enhancement of endothelium-dependent dilation in small intrarenal resistance arteries and arterioles. It is well established that the increased risk for cardiovascular disease in pre-diabetes is multifactorial, with etiologies including insulin resistance, dyslipidemia, hypertension, systemic inflammation and oxidative stress, as well as renal dysfunction. Recently, it was shown that diabetic nephropathy is improved following treatment with LIRA [Mann et al., 2017]. Herein, we showed that low dose LIRA treatment first and foremost, maintained NO-mediated dilation in the kidney independent of blood pressure changes, and normalized blood pressure in conscious rats to partially restore GFR in the obese Zucker rat model. While blood pressure reduction in the conscious rats is likely to contribute to this improvement in renal filtration, the microvessel dilation observed in LIRA rats during ACh and SNP stimulations is not attributable to this blood pressure lowering effect as basal MAP did not differ between rat treatment groups during anesthesia and the minor non-significant MAP changes during stimulations were similar in Veh-treated rats. Furthermore, LIRA treatment partially prevented glomerular macrophage infiltration, resulting in a significant reduction in the development of tubulointerstitial fibrosis but only a modest inhibitory effect on glomerular expansion. Taken together, these findings suggest that LIRA treatment ameliorated the progression of CKD in this study, in part by lowering blood pressure and thereby increasing the capacity for dilation of small intrarenal arteries and arterioles rather than by altering renal oxidative stress per se.

It has already been established that in Zucker fa/fa rats monocyte-macrophage infiltration, glomerular hypertrophy, interstitial fibrosis, accumulation of extracellular matrix (ECM) proteins and a decline in GFR lead to renal dysfunction and end-organ damage [Qian

et al., 2008; Reiniger et al., 2010; Patel et al., 2016]. TGF- β 1 plays a major role in the pathogenesis of interstitial fibrosis and glomerulosclerosis. Other studies have also revealed that, ET-1 plays an important role in mediating fibrosis in various disease pathologies including hypertension and diabetes [Widyantoro et al., 2010; Wang et al., 2015], and that augmented ET-1 induced an increase in ECM protein FN-1 production in the kidney in diabetes [Xu et al., 2008]. Indeed, in this study we found that fa/fa rats on a high-salt diet had increased expression of mRNA for TGF- β 1, ET-1, Nox-1 and Vcam-1 relative to lean fa/+ rats (Fig. 2). Previously it was shown that treatment with a GLP-1R agonist inhibited the upregulation of TGF- β 1 in human mesangial cells in vitro [Li et al., 2012]. However, treatment of Zucker rats with LIRA in this study did not result in significant reductions in renal mRNA expression of TGF- β 1, ET-1, Nox-1 or FN-1, and protein expression of TGF- β 1 was not significantly altered (Fig. 2 and Fig. 3D). On the other hand, endothelial function in the kidney was improved by LIRA, as indicated by the aforementioned improved vasodilator responses and substantially increased expression of eNOS protein (Fig. 3E) and a strong trend for reduced endothelial activation, as indicated by Vcam-1 mRNA expression. The later findings are in line with a previous study, which revealed that LIRA treatment improved renal injury by eNOS activation via inhibition of the nuclear factor kappa B (NF- κ B) pathway in STZ-induced diabetic rats [Zhou et al., 2014].

Based on the lack of effect of LIRA on TGF- β 1 and ET-1 signaling in this study, the modest amelioration of renal dysfunction and end-organ damage in the fa/fa rats on the high salt diet by LIRA treatment suggests that this partial amelioration is attributable to improvements in renal microvessel endothelial function and normalization of renal vasomotor tone (hypothesised mechanism shown in Fig. 9). Partial reductions in nephrosclerosis, BUN and urinary albumin elevation were associated with reduced macrophage infiltration of the nephrons and CD68-positive protein expression (Fig. 3A, C and 4A-D). Unlike a previous

study [Fujita et al., 2014], we did not observe a large reduction by LIRA in urinary albumin, albumin/creatinine ratio and body weight as reported for obese diabetic mice on a normal diet (Fig. 1D-E). This difference between studies might be in part due to the lower dose of LIRA administered in this study and the extra renal challenge presented by the chronic exposure to a high-salt diet.

Sanders [2009a, b] has shown that in salt-sensitivity, a high salt intake stimulates the production of vasoconstrictors TGF- β 1 and ET-1, which evokes ROS generation (Fig. 9). Further, it is well established that increased ROS generation in endothelial and vascular smooth muscle cells, contributes to a state of oxidative stress in hypertension and diabetes [Li et al., 2003; Viridis et al., 2011; Nagasu et al., 2016]. NADPH oxidase activation is triggered by the upregulation of vasoconstrictors including ANG-II, norepinephrine and ET-1 [Zweier et al., 2011]. Other studies implicate Nox1 in vascular inflammation, as oxidase activity and Nox1 expression are increased in smooth muscle cells and aortas of mice and rats with experimentally induced diabetes [Wendt et al., 2005; San Martin et al., 2007a, b]. In addition, it has been reported that a high-salt diet caused accumulation of oxidative and nitrosative injury in renal tissues of Zucker obese rats [Patel et al., 2016]. Even though it has been suggested that LIRA inhibits renal NADPH oxidase in diabetes models [Fujita et al., 2014; Herrera et al., 2016], it is somewhat surprising that we found no effect of LIRA on the expression of genes associated with elevation of oxidative stress and that renal NT protein did not differ significantly between rat groups (Fig. 2B, E and 3A-B). Therefore, we suggest here that direct inhibition of oxidative stress signaling is unlikely to be the main renoprotective action of LIRA in metabolic syndrome.

In this present study, we used a 1- μ m microfocus X-ray system to visualize the renal vasculature and investigate endothelial function in anesthetized Zucker rats. *In vivo* studies, including our own have demonstrated that ACh induced endothelium-dependent vasodilation

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is impaired in diabetes and high-salt diet animal models [DuPont et al., 2013; Pearson et al., 2013; Sonobe et al., 2015]. Microangiography revealed that the impairment in small artery-arteriole vasodilation was prevented in the LIRA treated rats, and more microvessels were perfused at baseline in fa/fa-LIRA group rats than in fa/fa-Veh rats (Fig. 5). Endothelium-dependent vasodilatory responses were most impaired in the small arteries and large arterioles in fa/fa-Veh group rats. Notably, using direct visualization of the renal vasculature we showed that smooth muscle dysfunction was also evident in the smaller vessels of fa/fa-Veh group rats, most likely due to the upregulation of TGF- β 1, oxidative stress and downregulation of soluble guanylyl cyclase in the renal vasculature [Du et al., 2019] and this was prevented by concurrent treatment with LIRA. NO and or EDHF-mediated dilation in both conduit and resistance vessels was maintained by chronic LIRA treatment in the fa/fa rats. Only in the vehicle treated fa/fa rats did the renal vessels become more constricted following blockade of COX and uncoupling of myoendothelial gap junctions. Gap junctions including connexins and various potassium channels are more important for longitudinal or conducted dilation [Feher et al., 2014], which increases the perfusion of side branches and more distal segments, rather than radial transmission of the hyperpolarization from the endothelium to the smooth muscle wall through myoendothelial gap junctions. Therefore, we assume that the observed constriction of vessels that were visibly perfused throughout the imaging protocol, which was evoked by blockade in this study is due to the inhibition of COX in the vasculature rather than inhibition of the myoendothelial gap junctions. This would imply that dilator prostanoids contribute significantly to the maintenance of renal perfusion in the Zucker fa/fa rats exposed to high salt intake, possibly in compensation for reduced NO bioavailability, impaired NO-mediated signaling and or increased prostanoids. Our findings suggest that the upregulation of inducible isoform COX-2 in the fa/fa-Veh rats is associated with production of dilator prostanoids on the basis of the changes in perfusion

following non-selective COX inhibition. However, both COX-1 and COX-2 produce dilator and constrictor prostanoids. Upregulation and overactivation of COX-2 associated with vascular aging changes ACh-mediated relaxation to constriction [Guo et al., 2018]. Furthermore, it has been demonstrated that in the Zucker fa/fa rat that COX-2 derived prostanoids contribute to coronary constrictor responses mediated by oxidative stress [Santiago et al., 2016]. Since ACh did not evoke constriction, and LIRA somewhat reduced COX-2 expression, there is no direct evidence at present to suggest COX-2 mediated renal constriction in our hypertensive Zucker fa/fa rats at this age. Previous studies have shown that, EDHF mediated dilation is increased in the Dahl salt-sensitive model rat and following chronic NOS inhibition [Goto et al., 2012; Simonet et al., 2012]. However, in direct contrast to these findings, in the early stages of diabetes localized deficits in coronary EDHF-mediated dilation develop while NO-mediated dilation is still well maintained [Jenkins et al., 2012]. A limitation of this study is that we did not attempt to establish if any of the known EDHF pathways are also impaired by a high-salt intake and or metabolic syndrome, or indeed, if LIRA alters EDHF-mediated dilation contributions in the kidney. In this study, LIRA did not appear to significantly modify conducted dilation within the kidney, which is largely mediated by various EDHF. Recently, it has been demonstrated that GLP-1 agonists evoke endothelium-dependent dilation in isolated resistance arteries through both NO and EDHF (Ca^{2+} -activated K channels) signaling pathways [Bayram et al., 2014]. We speculate that the upregulation of renal GLP-1R protein in the fa/fa-Veh rats might be a compensatory mechanism to maintain renal perfusion in the face of diminished NO signaling as protein expression was normalized in association with NO-mediated dilation to SNP stimulation in the fa/fa-LIRA rats. Importantly, such vasodilatory actions of LIRA provide further support for the hypothesised renoprotective actions of LIRA in this study (Fig. 9).

Conclusion and Perspectives

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Taken together our findings and the current literature suggests that high-salt diet induced renal dysfunction and injury occurs through the upregulation of oxidative stress and inflammation, which subsequently leads to exacerbation of renal dysfunction in the presence of obesity (Fig. 9). *In vivo* renal angiograms revealed that LIRA treatment improved endothelium dependent relaxation and restored renal blood flow in Zucker rat renal arteries most likely through increased NO production rather than through an effect on TGF- β 1 and ET-1 or oxidative stress and inflammation per se. Improvements in renal endothelial function and normalized blood pressure following, LIRA treatment attenuated the progression of renal fibrosis and glomerular hypertrophy. However, further studies are warranted to determine if greater benefits in this salt-sensitive model of hypertension can be obtained with higher doses of LIRA.

Authors contributions

Participated in research design: Sukumaran, Shirai and Pearson

Conducted experiments: Sukumaran, Tsuchimochi and Sonobe

Performed data analysis: Sukumaran

Wrote or contributed to the writing of the manuscript: Sukumaran and Pearson

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Footnote page

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Figure legends

Fig. 1. Effects of LIRA on A) Body weight, B) Blood glucose, C) plasma insulin levels, D) systolic blood pressure (SBP), and F) Blood urea nitrogen levels after 8 weeks high-salt diet in Zucker rats. The effects of LIRA treatment on E) glomerular filtration rate (GFR), G) urinary albumin and H) albumin creatinine ratio during baseline and after 8 weeks high-salt diet in Zucker rats. The data are shown as the mean \pm SEM; N=4-7 rats per group. The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 2. The effects of LIRA treatment on renal mRNA expression of ET-1, Nox-1, VCAM-1, TGF- β 1, GPX-1, FN-1, eNOS and VEGF (A-H). GAPDH gene $\Delta\Delta$ CT value was used as an internal control. The data are shown as the mean \pm SEM. N=8 rats per group. The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 3. The effects of LIRA treatment on protein expression of NT, CD-68, TGF- β 1, eNOS, VEGF, COX-2 and GLP-1R in the kidney. A) Protein expression levels of NT, CD-68, TGF- β 1, eNOS, VEGF, COX-2 and GLP-1R detected by Western blotting. β -actin was used as an internal control. Bands are representative of seven separate experiments. B-H) Relative density values of NT, CD-68, TGF- β 1, eNOS, VEGF, COX-2 and GLP-1R. The data are shown as the mean \pm SEM. N=5-6 rats per group. The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

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Fig. 4. Effects of LIRA on collagen accumulation and macrophage number in Zucker rat kidneys. A) Azan-Mallory staining of collagen. Azan-Mallory-stained kidney sections from a Ln-Veh and Ln-LIRA rats showing low levels of collagen deposition (blue fibrils); large patches of strong blue fibrotic tissue reactivity in a vehicle-treated fa/fa rats (blue fibrils); LIRA-treated fa/fa rats showing moderate blue fibrous tissue reactivity around glomerulus. Scale bar is 20µm. B) Immunolocalization of macrophage-specific antigen CD-68 in the glomeruli. The merged CD-68/DAPI image is presented. C) Quantification of tubulointerstitial fibrosis (%). D) Quantitative measurement of CD-68 positive cell numbers/20 fields. Original magnification x400. E-F) bar diagram shows that mean glomerular area and perimeter (N=4). The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 5. Representative X-ray angiograms of the renal vasculature in Zucker rats after 8 weeks on a high-salt diet. (A) Images during baseline, infusions of acetylcholine (ACh), sodium nitroprusside (SNP), ACh stimulation during blockade of prostaglandin production and uncoupling of gap junctions. B) Acute drug infusion protocol during renal vasculature imaging. C) Vessel internal diameter categorized by branching order during baseline Ringer's lactate infusion. D) Visualized vessel number during baseline. Calibration wire (white line) = 50 µm. The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 6. Mean arterial pressure (MAP) and mean heart rate (MHR) and their change during the acute infusion. A) MAP and MHR during baseline and B-C) infusions of ACh, SNP, indomethacin + carbenoxolone (blockade) and post-blockade + ACh relative to vehicle Ringer's lactate solution (baseline). Mean ± SEM. N=5-6 rats per group. The significance of

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group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 7. LIRA treatment improved the capacity to increase perfused segments by NO-mediated dilation in Zucker fa/fa and Ln rats. (A-D) Percentage change in 1st to 3rd order vessel caliber and visible vessel number during infusions of ACh, SNP and ACh stimulation during the blockade of prostaglandin production and uncoupling of gap junctions from baseline. Mean \pm SEM. N=7 rats per group. The significance of group differences was determined by 2-way ANOVA for factors of genotype (G) and drug treatment (T) and their interaction (I).

Fig. 8. Representative X-ray angiograms of the renal vasculature in 9 weeks old Zucker rats on a normal-salt diet. (A) Images during baseline, infusions of ACh and SNP. B) Vessel internal diameter and number categorized by branching order during baseline Ringer's lactate infusion. C) MAP and MHR during baseline. (D-E) Percentage change in 1st to 3rd order vessel caliber, visible vessel number, MAP and MHR during infusions of ACh, SNP from baseline. Mean \pm SEM. N=5 rats per branching order. The significance was determined by column statistics.

Fig. 9. Hypothesized protective effects of low dose-LIRA in renal dysfunction associated with the additive effects of obesity, insulin resistance and a high-salt diet. High-salt diet induced renal dysfunction is characterized by upregulation of COX-2 and associated increase in renal vasoconstrictor tone and ROS-production, endothelial dysfunction and reduced eNOS, leading to elevated blood pressure and reduced GFR. LIRA treatment improved endothelial function and reduced renal vasomotor tone through the upregulation of eNOS and endothelium-dependent vasodilation (see discussion for detailed explanation).

Fig. 1

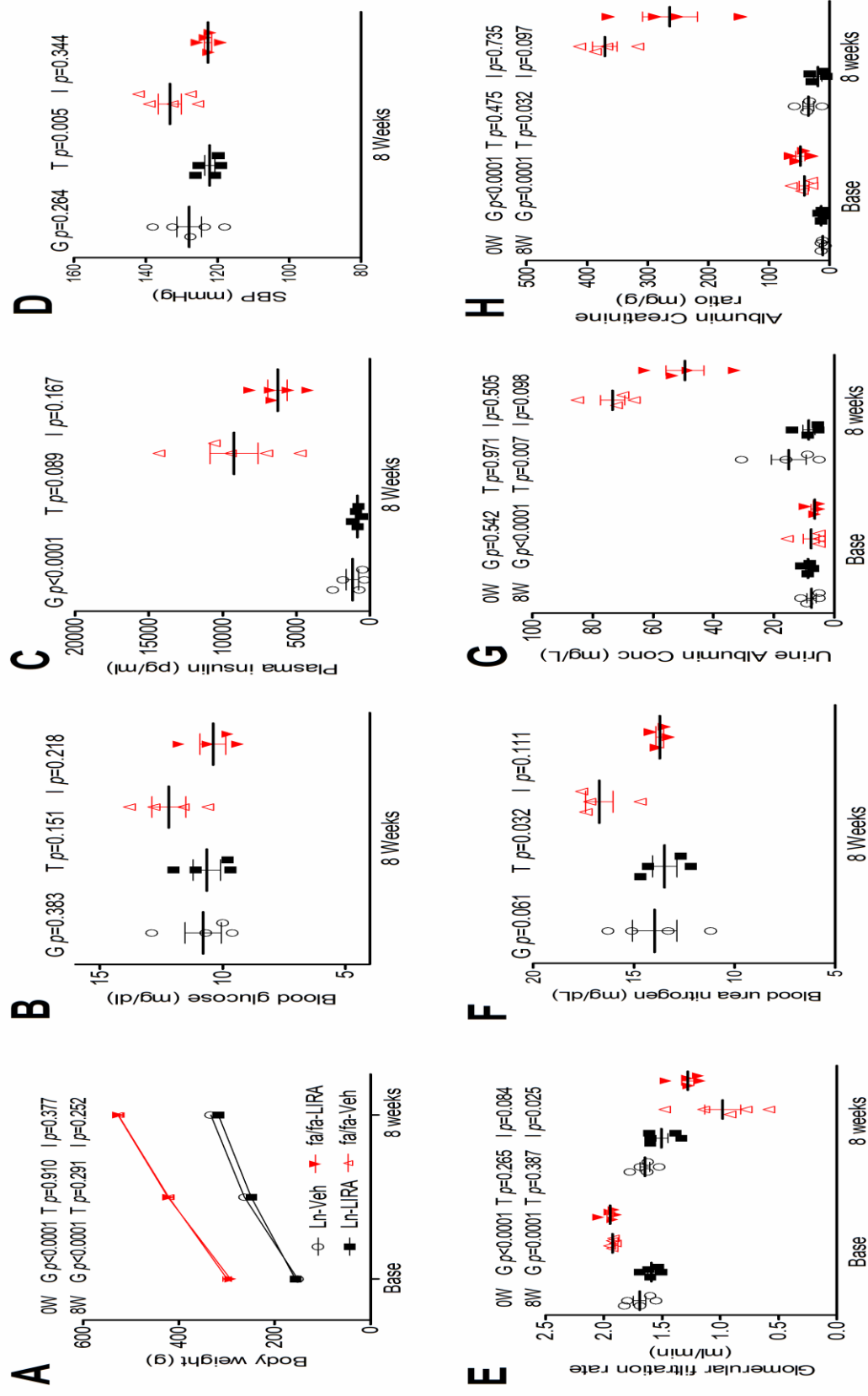


Fig. 2

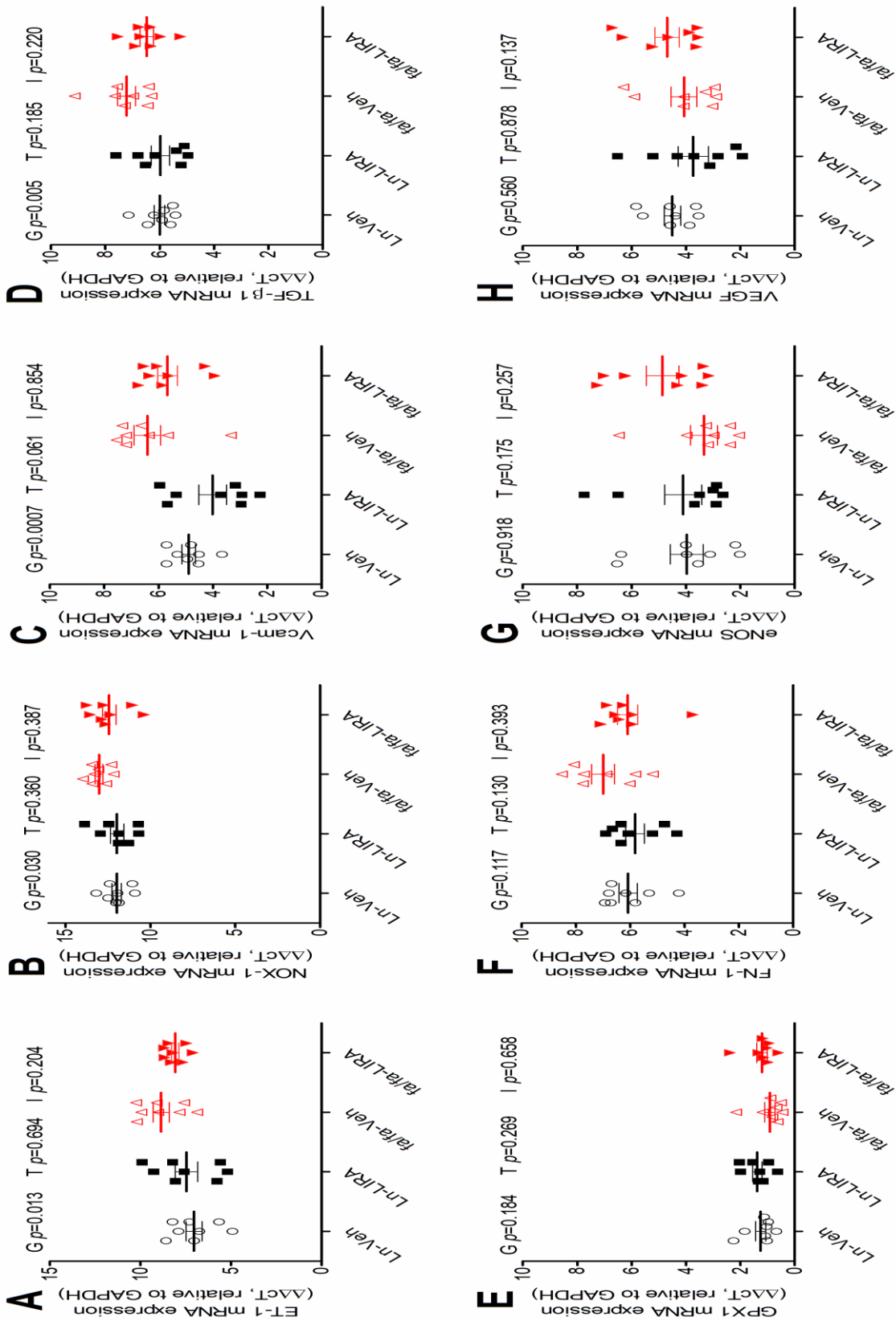


Fig. 3

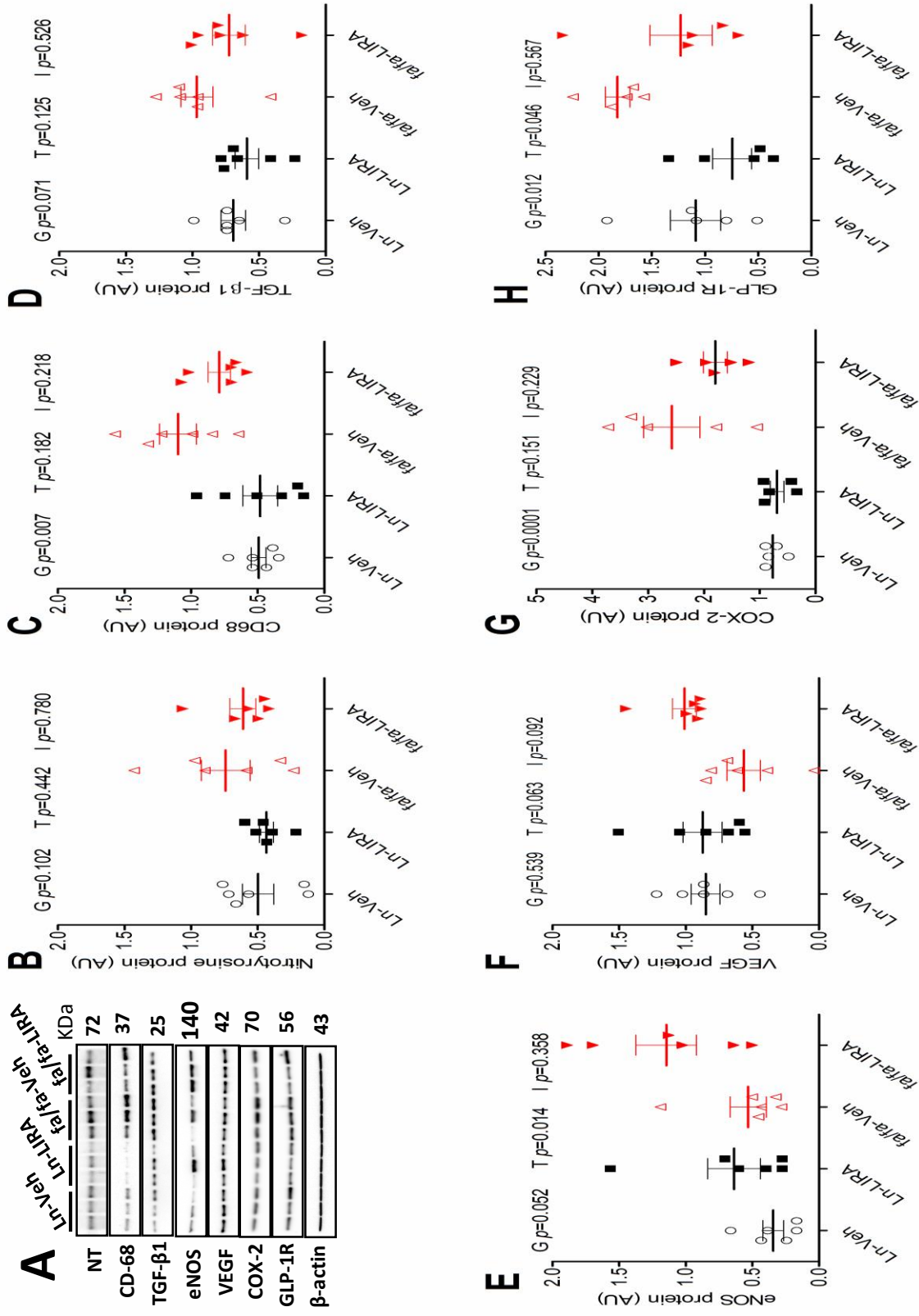


Fig. 4

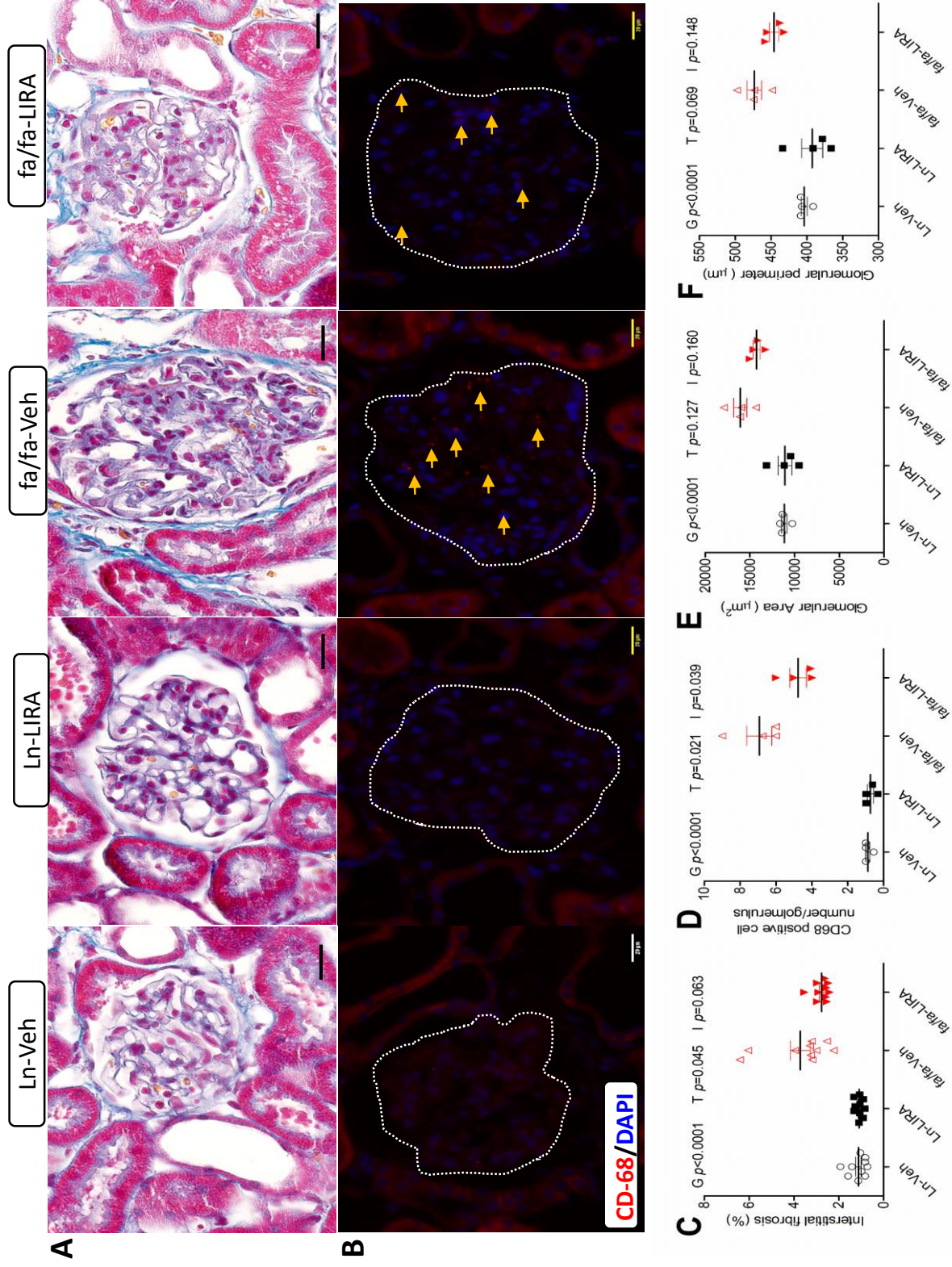


Fig. 5

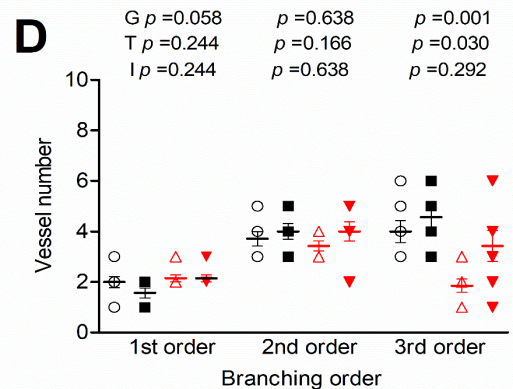
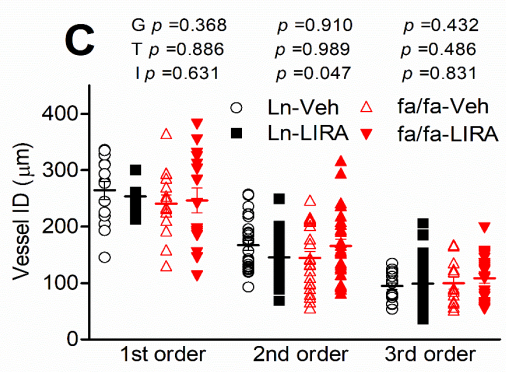
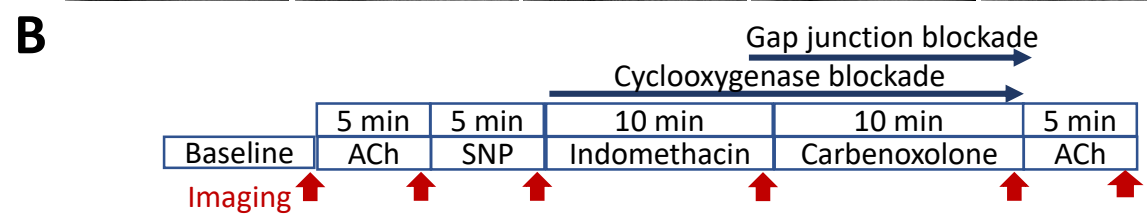
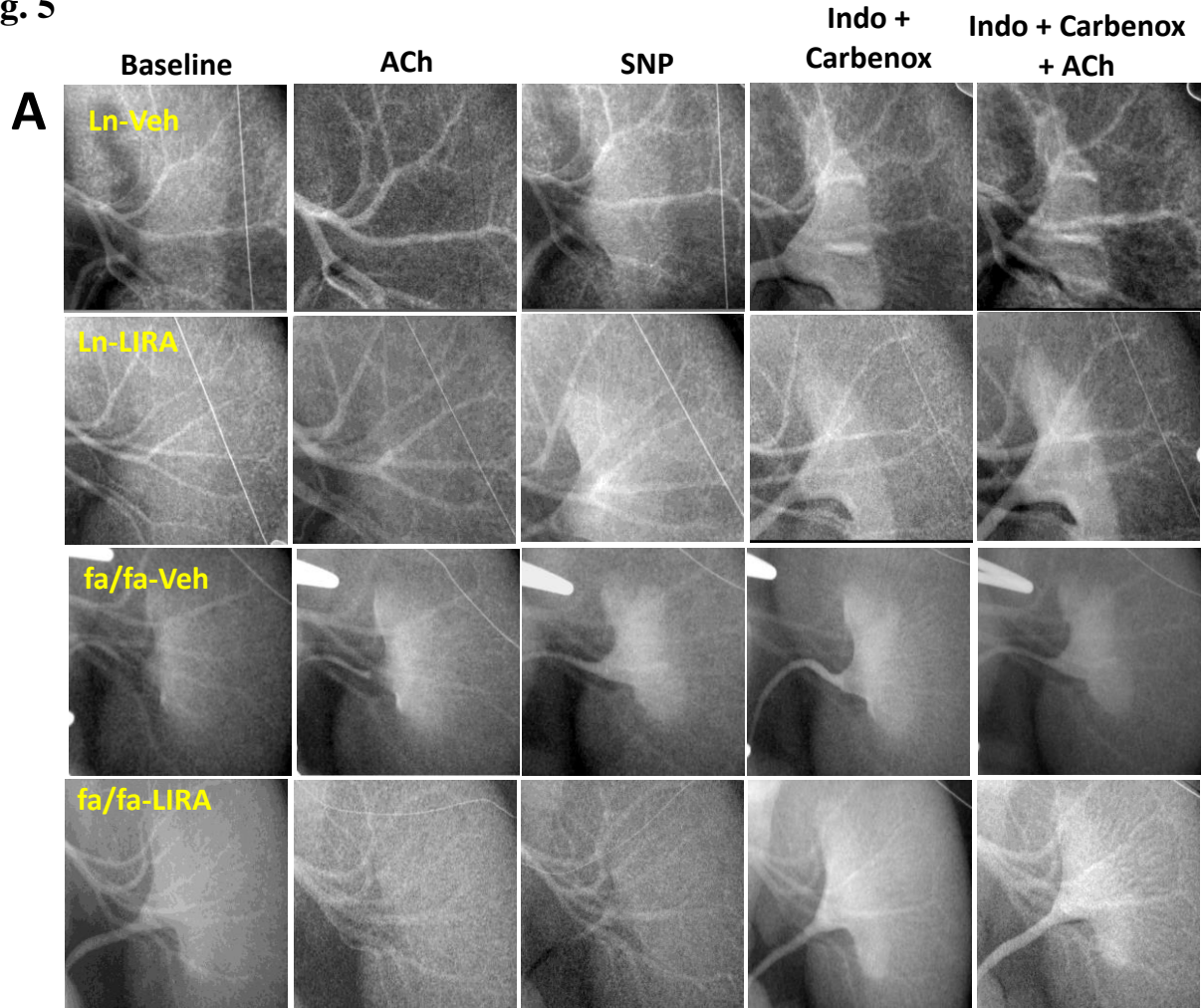


Fig. 6

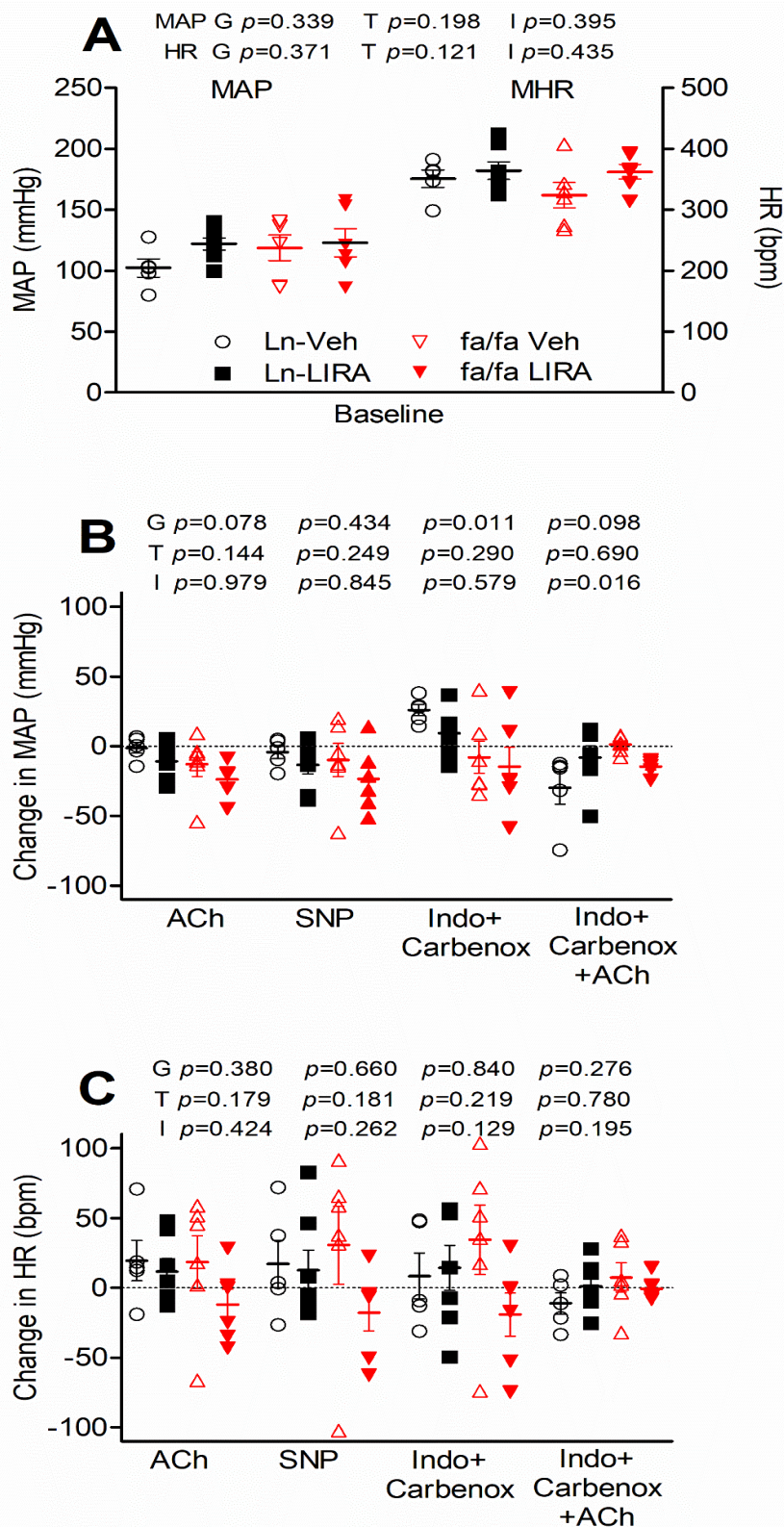


Fig. 7

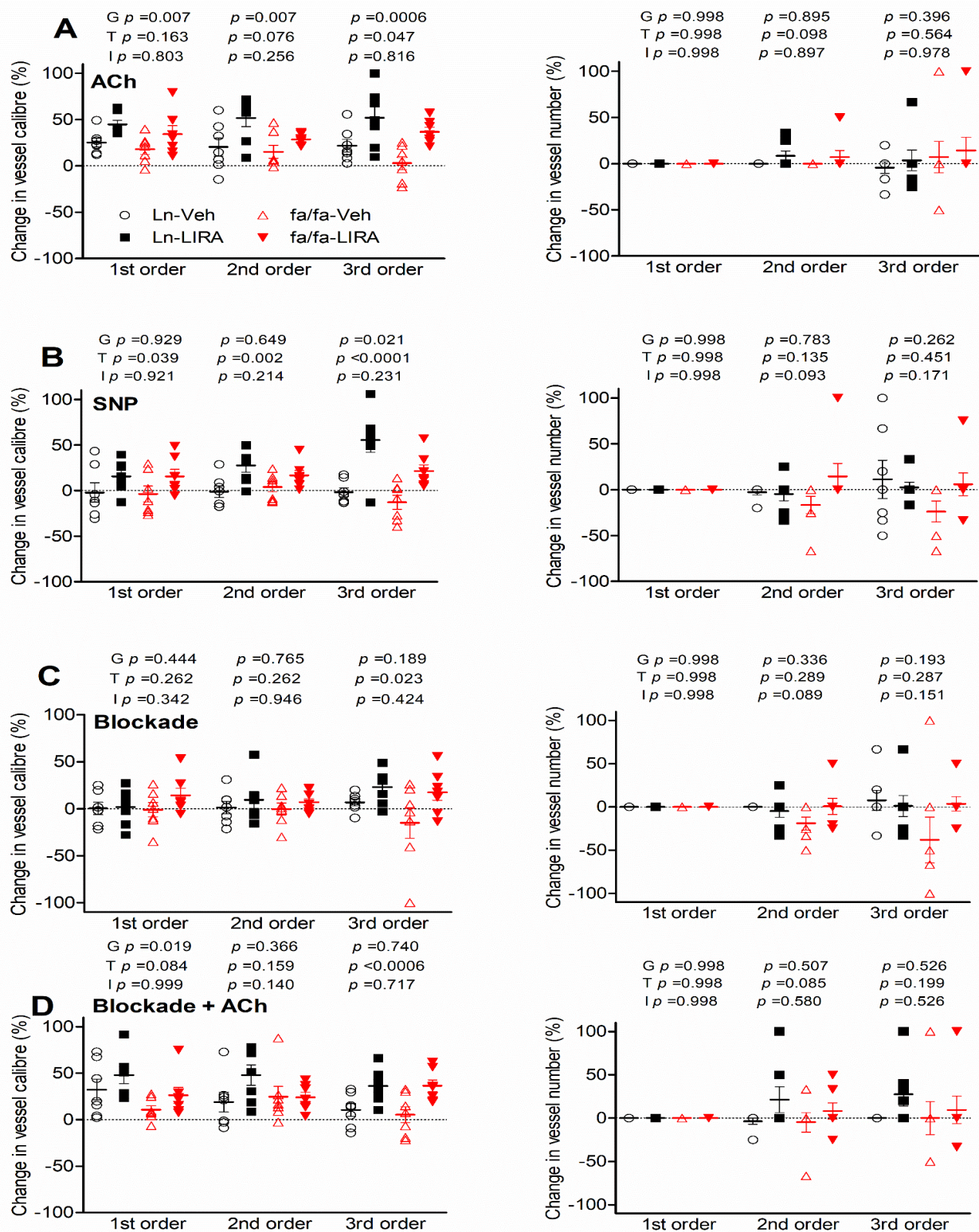


Fig 8

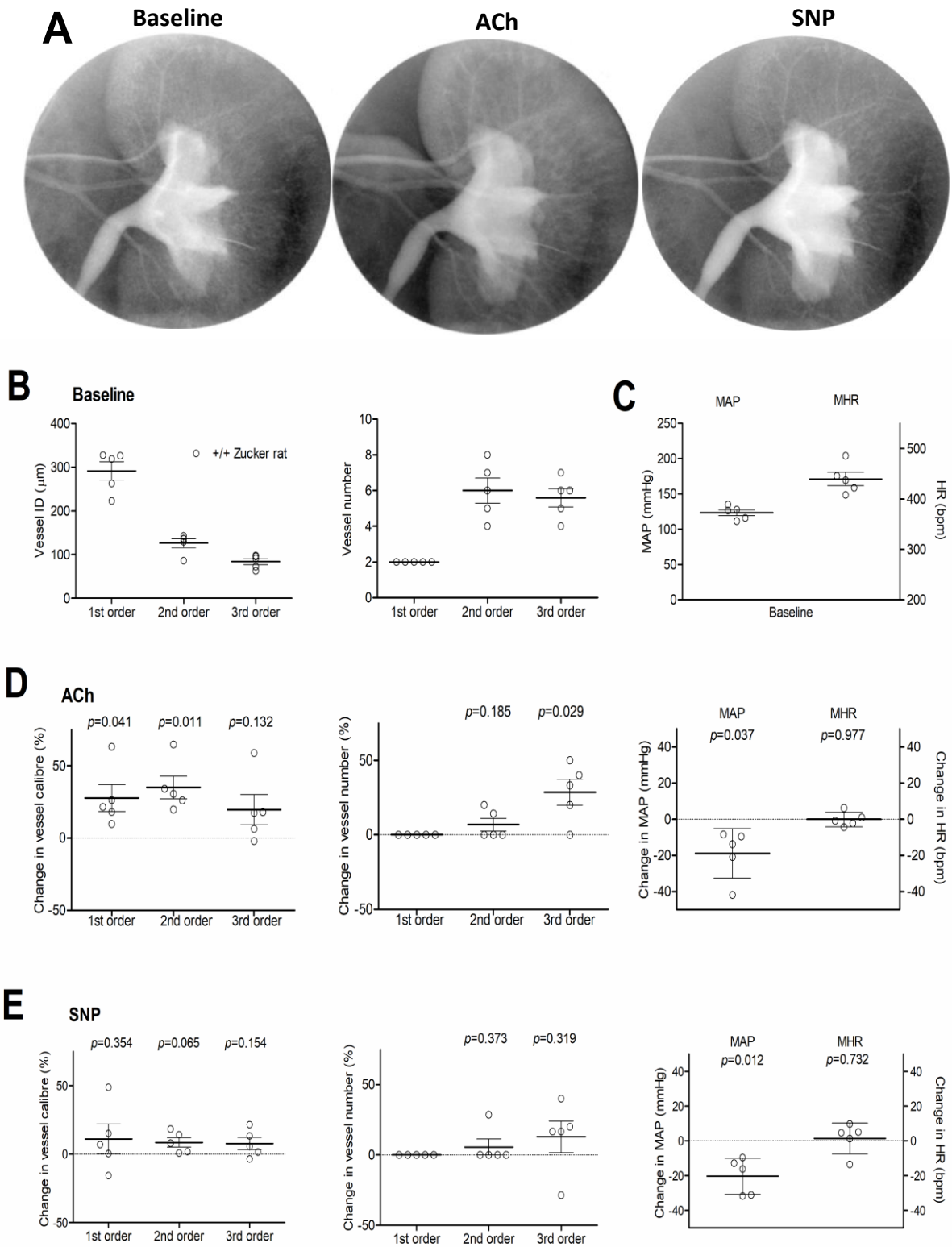


Fig. 9

