

JPET #246165

Reduction in secretion of very low density lipoprotein-triacylglycerol by a matrix
metalloproteinase inhibitor in a rat model of diet-induced hypertriglyceridemia

Yoichi Kawashima, Yoshihiro Eguchi, Tohru Yamazaki, Minako Karahashi, Hiroshi Kawai and

Naomi Kudo

Faculty of Pharmacy and Pharmaceutical Sciences, Josai University, 1-1 Keyakidai, Sakado,

Saitama 350-0295, Japan

JPET #246165

a) Running title: A matrix metalloproteinase inhibitor reduces VLDL secretion

b) Corresponding author:

Name: Yoichi Kawashima

Address: Faculty of Pharmacy and Pharmaceutical Sciences, Josai University, 1-1 Keyakidai,

Sakado, Saitama 350-0295, Japan

Tel.: +81-49-271-7676

Fax: +81-49-271-7984.

E-mail address: ykawash@josai.ac.jp

c) Number of text pages: 45

Number of tables: 2

Number of figures: 7

Number of references: 60

Number of words in the Abstract: 250

Number of words in the Introduction: 712

Number of words in the Discussion: 1498

JPET #246165

d) Abbreviations: ACC1, acetyl-CoA carboxylase 1; ACLY, ATP-citrate lyase; ACOT1, acyl-CoA thioesterase 1; ACSL, long-chain acyl-CoA synthetase; apo, apolipoprotein; ATGL, adipose triglyceride lipase; BSA, bovine serum albumin; CD36, fatty acid translocase; CGI-58, comparative gene identification-58; ChREBP, carbohydrate response element-binding protein; CPT1a, carnitine palmitoyltransferase 1a; CVD, cardiovascular disease; DGAT, acyl-CoA:diacylglycerol acyltransferase; FA, fatty acid; FAS, fatty acid synthase; FATP, fatty acid transport protein; GPAT, glycerol-3-phosphate acyltransferase; G6PD, glucose-6-phosphate dehydrogenase; HSD, high sucrose diet; IRS, insulin receptor substrate; LCAD, long-chain acyl-CoA dehydrogenase; LDL, low density lipoprotein; LPK, L-type pyruvate kinase; LPL, lipoprotein lipase; MCAD, medium-chain acyl-CoA dehydrogenase; ME1, malic enzyme 1; MMPI, matrix metalloproteinase inhibitor; MTP, microsomal triglyceride transfer protein; ND, normal diet; NEFA, non-esterified fatty acids; PPAR α , peroxisome proliferator-activated receptor α ; SCD1, stearoyl-CoA desaturase 1; SREBP-1c, sterol regulatory element-binding protein-1c; TAG, triacylglycerol (s); TRL, triacylglycerol-rich lipoprotein; UCP2, uncoupling protein 2; VLDL, very low density lipoprotein.

e) Recommendation section: Drug Discovery and Translational Medicine

JPET #246165

ABSTRACT

Matrix metalloproteinase inhibitors (MMPi) reduced serum triacylglycerol (TAG) levels in streptozotocin-induced diabetic rats and Zucker *fa/fa* rats in our previous study. However, the mechanisms underlying TAG reduction by MMPi remain unclear. The present study aimed to elucidate the mechanism by which F81-1144b, a MMPi, lowers serum TAG levels in an animal model of high sucrose diet (HSD)-induced hypertriglyceridemia. F81-1144b was repeatedly administered to rats fed HSD, and its effects were evaluated on TAG levels in serum and the liver, very low density lipoprotein (VLDL) secretion, de novo fatty acid (FA) synthesis in the liver, and the expression of genes regulating the metabolism of FA, TAG, and VLDL in the liver and serum. F81-1144b lowered TAG levels in serum and the liver, VLDL-TAG secretion, de novo FA synthesis in the liver, and serum levels of insulin and glucose. F81-1144b suppressed the expression of genes related to the de novo synthesis of FA and TAG, key proteins (lipin 1 and apolipoprotein CIII) responsible for VLDL metabolism, and sterol regulatory element-binding protein-1c and carbohydrate response element-binding protein. F81-1144b little affected the expression of genes related directly to the degradation of TAG or FA, while it up-regulated that of gene for uncoupling protein 2 in the liver. These results suggest that MMPi are a novel type of therapeutic agent for the treatment of hypertriglyceridemia because the metabolic effects of F81-1144b expected from changes in the expression of genes regulating lipid metabolism would

JPET #246165

alter metabolism differently from those induced by fibrates, niacin, or *n*-3 FAs.

Introduction

Low density lipoprotein (LDL) has been firmly established as the primary atherogenic lipoprotein. However, even at low plasma LDL levels among patients treated with statins, the high residual risk of cardiovascular disease (CVD) is not abrogated (Fruchart et al., 2008; Kearney et al., 2008). The independent relationship between plasma triacylglycerol (TAG) and the risk of future CVD events has remained controversial; however, recent studies demonstrated that elevated plasma TAG levels are strongly and independently associated with the risk of CVD (Harchaoui et al., 2009; Neil et al., 2010; Sarwar et al., 2010; Langsted et al., 2011; Nordestgaard and Varbo, 2014; Nordestgaard, 2016). A number of metabolic conditions are frequently associated with hypertriglyceridemia. The factors associated with increases in plasma TAG levels are a genetic predisposition (such as familial hypertriglyceridemia and familial combined hyperlipidemia), diseases (such as insulin resistance, metabolic syndrome, type 2 diabetes, type 1 diabetes, and renal diseases), lifestyle factors (such as obesity, alcohol consumption, and a diet enriched with simple carbohydrates), and medications (such as estrogens) (Boullart et al., 2012). Very low density lipoprotein (VLDL) overproduction appears to be central to the pathophysiology of hypertriglyceridemia in an insulin-resistant state (Taskinen, 2003; Adiels et al., 2008). Hypertriglyceridemic subjects with more visceral fat storage have been reported to have larger TAG-enriched VLDL particles (Wang et al., 2011). In patients with hepatic steatosis, VLDL

JPET #246165

overproduction results from compensatory mechanisms for hepatic lipid overload, which ultimately increases the risk of developing hypertriglyceridemia and atherosclerosis (Chatrath et al., 2012).

The aberrant metabolism of TAG-rich lipoproteins (TRL) (VLDL and chylomicrons) results in increased plasma TRL remnants and elevated small dense LDL, lipoprotein particles that are strongly associated with an increased risk of CVD (Miller et al., 2011). Under conditions of the overproduction and/or delayed clearance of TRL, TRL and/or TRL remnants formed during lipolysis are more likely to enter artery walls, causing foam cell formation and activating vascular cell inflammation (Nordestgaard et al., 1995; Schwartz and Reaven, 2012). Moreover, VLDL particles are larger and LDL particles are smaller in insulin-resistant states; LDL size is inversely associated with plasma TAG levels (Berneis and Krauss, 2002). Small dense LDL particles are retained for longer period of time in the circulation, are glycosylated and oxidized, and are prone to cross the endothelial surface, contributing to plaque formation. Thus, increased TRL remnants play a pivotal role in the pathophysiology of atherosclerosis, and thereby contribute to CVD (Chapman et al., 2011; Nordestgaard, 2016). In this context, the prevalence of elevated TRL is a major concern (Miller et al., 2011).

Fibrates, niacin, and *n*-3 fatty acids (FAs) are currently the first choice treatments for hypertriglyceridemia (Ito, 2015). The mechanistic actions of these agents markedly differ from each other. Considering the diversity in the pathogenesis of hypertriglyceridemia, the wide variety of agents available for selection in order to reduce plasma TAG is therapeutic benefit. We

JPET #246165

previously demonstrated that among various metabolic conditions that are known to be associated with increases in serum TAG, the administration of MMPIs, the structures of which are similar to that of F81-1144b, reduced the serum levels of TAG, VLDL-TAG, and LDL-cholesterol, but not those of high density lipoprotein-cholesterol in streptozotocin-induced diabetic rats (a model of type 1 diabetes) and genetically obese Zucker *fa/fa* rats (a model of genetic obesity due to a dysfunction in the actions of leptin) (Morikawa et al., 2007). In comparison to a population with hypertriglyceridemia attributed to these two metabolic disorders, however, a population with hypertriglyceridemia that is due to changes in dietary habits, a marked increase in dietary intake of fructose from sucrose and fructose-enriched sweeteners is considered to be much greater. Epidemiological studies have shown that the consumption of sucrose or a mixture of glucose and fructose is associated with the occurrence of metabolic disorders including hypertriglyceridemia (Tappy and Lê, 2010; Ouyang et al., 2008). Nevertheless, no information is currently available on whether MMPIs reduce serum TAG in animal models with diet-induced hypertriglyceridemia, but not diabetes or obesity. Moreover, the mechanisms underlying MMPI-induced reductions in serum TAG remain unclear. In this context, the present study aimed (i) to investigate whether MMPIs suppress serum TAG levels in animals with high sucrose diet (HSD)-induced hypertriglyceridemia and, if this is the case, (ii) to elucidate the metabolic mechanism by which MMPIs reduce serum TAG levels.

Materials and Methods

Chemicals. The following materials were obtained from the indicated commercial sources: [$1\text{-}^{14}\text{C}$] acetic acid (2.035 TBq/mol), [$\text{U-}^{14}\text{C}$]glycerol-3-phosphate (5.55 TBq/mol), and [$1,3\text{-}^3\text{H}$]glycerol (1,480 TBq/mol) (American Radiolabeled Chemicals, Inc., St. Louis, MO); acetyl-CoA, palmitoyl-CoA, glycerol-3-phosphate, Triton WR-1339 and bovine serum albumin (BSA) (Sigma-Aldrich, St. Louis, MO). F81-1144b, the structure of which is shown in Fig. 1, was kindly provided by Daiichi Fine Chemical Co., Ltd. (Takaoka, Japan); F81-1144b is a subcutaneously active hydroxamic acid MMP inhibitor that exhibits potent activity against a number of MMPs, including MMP-1 ($\text{IC}_{50} = 5 \text{ nM}$) and MMP-3 ($\text{IC}_{50} = 29 \text{ nM}$) (Fujisawa *et al.*, 2001).

Animals. All animal procedures were approved by the Institutional Animal Care Committee of Josai University in accordance with the Guidelines for the Proper Conduct of Animal Experiments (Science Council of Japan). Six-week-old male Wistar rats were purchased from SLC Inc. (Hamamatsu, Japan). Rats were acclimatized for 1 week on a standard rodent chow (CE-2; Clea Japan Inc., Tokyo, Japan).

Study design. After acclimatization for 1 week, 7-week-old rats were divided into four groups. Group 1 was fed the standard diet for an additional 21 days (normal diet (ND)-fed rats).

JPET #246165

Group 2 was fed HSD for 21 days. HSD (ICN high carbohydrate diet) was purchased from Oriental Yeast Co. (Tokyo, Japan) and contained (by weight): 68% sucrose, 8% cottonseed oil, 18% milk casein, 2% beer yeast, and the required vitamins and minerals. Group 3 was maintained on HSD for 21 days and subcutaneously received F81-1144b at a dose of 50 mg/kg once a day for 7 days before being sacrificed. Group 4 was fed HSD for 21 days and subcutaneously received F81-1144b at a dose of 100 mg/kg once a day for 7 days before being sacrificed. F81-1144b was suspended in saline at a concentration of 50 mg/ml. At the age of 10 weeks, rats were sacrificed in the fed state. Rats were anesthetized with diethyl ether, and blood was withdrawn from the inferior vena cava. The liver was rapidly removed, washed with ice-cold saline, and weighed. Two portions of the liver were frozen in liquid nitrogen and stored at -80°C until the analysis of mRNA and acetyl-CoA. The remainder of the liver was used for a lipid analysis and the preparation of homogenates and microsomes. The parts of the liver that were used to prepare homogenates and microsomes were perfused with ice-cold saline.

Serum clinical chemistry. Serum was obtained from blood by centrifugation at 1,200g for 15 minutes. Serum TAG, total cholesterol, non-esterified fatty acids (NEFA), and glucose were measured using colorimetric enzymatic assay kits from Wako Pure Chemicals (Osaka, Japan). Serum insulin was measured using a rat RIA kit from Millipore Corp. (Billerica, MA).

Hepatic lipid analysis. After the addition of a known amount of triheptadecanoin as an internal standard, total lipids were extracted from liver homogenates by the reported method (Bligh

JPET #246165

and Dyer, 1959). Cholesteryl ester, TAG, diacylglycerol, free FA, and phospholipid were separated by thin-layer chromatography on silica gel G plates, which were developed with *n*-hexane/diethyl ether/acetic acid (80:30:1, v/v/v). After visualization by spraying 0.001% (w/v) primuline in 80% acetone, each region that corresponded to TAG and phospholipids on the plates was scraped off and transferred to tubes. A known amount of methyl heptadecanoate was added to the tubes containing phospholipids as an internal standard. TAG and phospholipids were extracted from silica gel as described previously (Imai et al., 1999). FA methyl esters were prepared using sodium methoxide/methanol. FA methyl esters were measured by gas-liquid chromatography (Shimadzu GC-2014; Shimadzu, Kyoto, Japan) as described previously (Yamazaki et al., 2014).

Acetyl-CoA measurement in the liver. Acetyl-CoA was extracted from one portion of the liver and measured enzymatically using a reported method (Decker, 1985).

Measurement of VLDL-TAG secretion in HSD-fed rats in the fed state. Rats that were maintained on HSD for 21 days were subcutaneously administered F81-1144b at a dose of 100 mg/kg once a day for 7 days before being killed at 10 weeks. Rats were anesthetized with urethane at a dose of 0.9 g/kg and underwent surgical catheter implantation via the jugular vein (SP-31) and portal vein (SP-10). Rats were administered [1, 3-³H]glycerol solution (24.7 MBq/0.37 ml saline/kg) via the portal vein, and Triton WR1339 solution (600 mg/3 ml saline/kg) was injected via the tail vein 15 minutes later. Triton WR1339 was utilized to inhibit lipoprotein lipase (LPL) (Millar et al., 2005). Blood samples were collected into heparinized tubes via the jugular vein 1

JPET #246165

minute before and 30, 60, 90, and 120 minutes after the Triton WR1339 injection. Plasma samples were prepared by centrifugation at 3,000g for 5 minutes. Total lipids were extracted from plasma by the method of Bligh and Dyer (1959). TAG was separated by thin-layer chromatography on silica gel G plates and then extracted from silica as described above. Radioactivity was measured using a liquid scintillation counter (Aloka LSC6100, Hitachi Ltd., Tokyo, Japan).

Measurement of in vivo synthesis of FA in the liver. Hepatic FA synthesis was estimated by measuring the in vivo incorporation of [$1\text{-}^{14}\text{C}$]acetic acid into FA in the liver according to the previously reported method (Sakurai et al., 1978) with some modifications. In brief, [^{14}C]acetic acid was dissolved in saline (370 kBq/ml). Under anesthesia with diethyl ether, HSD-fed control rats and HSD-fed rats that had been treated daily with F81-1144b at a dose of 50 or 100 mg/kg for 7 days were intraperitoneally injected with [^{14}C]acetic acid at a dose of 1.48 MBq/10 μmol /4 ml saline/kg body weight. Ten minutes after the injection, livers were immediately isolated and snap-frozen in liquid nitrogen. Lipids were extracted from the liver (Bligh and Dyer, 1959). Extracted lipids were saponified with 10% methanolic KOH at 80°C for 60 minutes under a nitrogen atmosphere. After being diluted with water, unsaponifiable matter was extracted with *n*-hexane three times. The aqueous phase was acidified with 6 M HCl, and FAs were extracted with *n*-hexane three times. FAs were mixed with scintillation fluid after drying down, and radioactivity was measured using a liquid scintillation counter.

Real-time quantitative PCR. Total RNA was isolated from liver tissues using the

JPET #246165

QIAzol reagent and RNeasy kit (QIAGEN, Hilden, Germany). cDNA was synthesized from 500 ng of total RNA with avian myeloblastosis virus reverse transcriptase (Takara Bio Inc., Otsu, Japan). PCR amplification was conducted using SYBR Premix EX Taq (Takara). Amplification and detection were performed with the Step One Plus real-time PCR system (Life Technologies Corp., Carlsbad, CA). The thermal cycling program was as follows: 10-second denaturation steps at 95°C, followed by 50 cycles of 5-second denaturation steps at 95°C, and 34-second annealing steps at 60°C. After the reaction, dissociation curve analyses were performed in order to confirm the amplification of a single PCR product. Changes in gene expression were calculated using the comparative threshold cycle (Ct) method. Ct values were first normalized by subtracting the Ct value obtained from β -actin (control). The sequences of primers used in the present study are listed in Table 1.

Assay for glycerol-3-phosphate acyltransferase (GPAT). One portion of the perfused liver was homogenized in 4 volumes of 0.25 M sucrose/1 mM EDTA/10 mM Tris-HCl buffer (pH 7.4). Hepatic microsomes were prepared by sequential centrifugation as previously described (Karahashi et al., 2013b), and protein was assessed using a previously reported method (Lowry et al., 1951) with BSA as the standard. The activities of GPAT in microsomes were assessed by the reported method (Yamada and Okuyama, 1978). In brief, the assay mixture consisted of 2 mM (9.26 kBq/ml) [U-¹⁴C]glycerol-3-phosphate, 40 μ M palmitoyl-CoA, 0.5 mg/ml of BSA (essentially FA free), 0.1 M Tris-HCl (pH 7.5), and 200 μ g/ml of microsomal proteins. Incubations were

JPET #246165

performed at 25°C for 2 minutes. Lipids were extracted with chloroform/methanol after acidification of the incubation mixture with 0.1 M HCl. Extracted lipids were mixed with scintillation fluid after drying down, and radioactivity was measured.

Statistical analysis. Data are presented as the mean \pm standard deviation (S.D.). The homogeneity of variance was established using a one-way ANOVA. When a difference was significant ($P < 0.05$), Scheffé's multiple range test was used as a post-hoc test. Levels of significance were set at $P < 0.05$. The significance of differences between two groups was analyzed using the Student's *t*-test.

Results

Effects on serum TAG levels in HSD-fed rats. It was initially confirmed that feeding HSD causes hypertriglyceridemia. When rats were fed HSD, serum TAG levels gradually increased by as much as 4-fold on day 21 (Fig. 2A). In order to estimate the effects of F81-1144b on serum TAG levels, rats that were maintained on HSD for 21 days were subcutaneously administered F81-1144b once a day for 7 days at doses of 50 or 100 mg/kg before being killed, and biochemical and physiological parameters were measured. Elevations in serum TAG levels by the feeding of HSD significantly decreased in a dose-dependent manner following the treatment of rats with F81-1144b, and TAG levels in the F81-1144b dosing group at a dose of 100 mg/kg were similar to those in ND-fed control rats (Fig. 2B). The feeding of HSD did not affect body weight, while the treatment with F81-1144b at a dose of 100 mg/kg insignificantly reduced body weight (Table 2). No significant changes were noted in the overall health of rats following the administration of F81-1144b (100 mg/kg). The feeding of HSD slightly increased liver weights and relative liver weights. Serum levels of cholesterol were markedly elevated by the feeding of HSD, and these increases were reduced by F81-1144b in a dose-dependent manner. Serum levels of glucose slightly increased upon the feeding of HSD; these levels returned to normal values following the treatment of rats with F81-1144b. The treatment with F81-1144b at a dose of 50 or 100 mg/kg significantly suppressed the serum levels of TAG, but not glucose, in rats that were maintained on

JPET #246165

the normal diet (ND) (Supplemental Fig. 1A, B).

Effects on the expression of genes related to VLDL formation and degradation.

Regarding VLDL degradation, HSD feeding tended to down-regulate the expression of gene for lipoprotein lipase (LPL) (Student's *t*-test, $P = 0.0075$, ND versus HSD-fed control), and significantly up-regulated that for apolipoprotein (apo) CIII (Fig. 3A, B). The administration of F81-1144b to rats maintained on HSD significantly increased the expression of *Lpl* in a dose-dependent manner, and markedly reduced the elevated levels of mRNA for *Apoc3* in a dose-dependent manner to the levels of ND-fed control rats (Fig. 3A, B). When ND-fed rats were treated with F81-1144b, the expression of *Lpl* was up-regulated and that of *Apoc3* was down-regulated (Supplemental Fig. 1C, D). Regarding the formation of VLDL, HSD feeding markedly up-regulated the expression of gene encoding lipin 1, and significantly elevated the levels of mRNAs for microsomal triglyceride transfer protein (MTP) and Apo-CIII (Fig. 3B, C, D). F81-1144b dosing little affected the expression of *Mtp*, but markedly lowered that of *Apoc3* and *Lipin1* in a dose-dependent manner to the levels of ND-fed control rats (Fig. 3B, C, D). The expression of gene for Apo-AI, the main component of high density lipoprotein, was little affected by HSD feeding, and markedly elevated by F81-1144b dosing (Fig. 3E).

Effects on VLDL-TAG secretion in HSD-fed rats in the fed state. In order to

estimate the effects of F81-1144b on the secretion of VLDL-TAG that formed in the liver, [³H]glycerol was injected into the portal vein and Triton WR-1339 was injected into the tail vein of

JPET #246165

HSD-fed control rats and in HSD-fed rats treated with F81-1144b. [³H]TAG that appeared in serum was then measured (Fig. 4). It is important to note that this experiment was performed in the fed state of rats in order to avoid the influence of fasting. No marked differences were observed in diet intake between control and F81-1144b-treated rats (Supplemental Fig. 2A). [³H]TAG that appeared in the circulation was markedly less in F81-1144b-treated rats than in HSD-fed control rats (Fig. 4A). The [³H]TAG secretion rate, which was calculated between 0 and 60 minutes, was reduced by 45% with F81-1144b dosing (Fig. 4B).

Effects on TAG metabolism in the liver of HSD-fed rats. The administration of F81-1144b to HSD-fed rats significantly reduced hepatic TAG levels without affecting hepatic phospholipid contents (Fig. 5A, B). F81-1144b dosing markedly changed the FA profile (mol %) of TAG and phospholipids in the liver (Supplemental Table 1). F81-1144b at a dose of 100 mg/kg reduced the proportions of palmitic, palmitoleic, stearic, and octadecenoic acids, but increased those of linoleic, 8, 11, 14-eicosatrienoic, and arachidonic acids in TAG. However, the treatment with F81-1144b reduced the proportions of octadecenoic, α -linolenic, and 5, 8, 11- eicosatrienoic acids in phospholipids. When these results were compared by differences in the content ($\mu\text{mol/g}$ liver) of a particular FA between HSD-fed control rats and HSD-fed rats receiving 100 mg/kg of F81-1144b (Fig. 5C, D), the differences in the masses of de novo synthesized FAs (palmitic, octadecenoic, palmitoleic, and stearic acids) in TAG were significantly decreased (Fig. 5C); changes in the differences in the masses of these FAs in phospholipids were less prominent than

JPET #246165

those in TAG (Fig. 5D). The changes caused by 50 mg/kg of F81-1144b in profiles (mol %) and differences in the masses of de novo synthesized FAs were less than those induced by 100 mg/kg of F81-1144b.

In order to gain insights into the molecular basis for understanding the reduction in hepatic TAG by F81-1144b, the mRNA levels of key enzymes and proteins involved in the synthesis of FA and TAG were measured (Fig. 6, Supplemental Table 2). The expression of gene for fatty acid synthase (FAS) was markedly up-regulated by the feeding of HSD, and the elevated expression was suppressed by F81-1144b dosing in a dose-dependent manner (Fig. 6A). The treatment of ND-fed rats with F81-1144b tended to suppress hepatic expression of *Fas* (Scheffé's multiple range test, $P = 0.0551$, control versus group receiving 100 mg/kg) (Supplemental Fig. 1E). The levels of mRNA of *Fas* in HSD-fed rats receiving 100 mg/kg of F81-1144b were similar to those in ND-fed control rats receiving F81-1144b at a dose of 100 mg/kg. Regarding the other enzymes related to de novo FA synthesis, the expression of genes encoding acetyl-CoA carboxylase 1 (ACC1), glucose-6-phosphate dehydrogenase (G6PD), ATP-citrate lyase (ACLY), malic enzyme 1 (ME1), and stearoyl-CoA desaturase 1 (SCD1) were all significantly down-regulated by the treatment of HSD-fed rats with F81-1144b (Supplemental Table 2). The effects of F81-1144b on de novo FA synthesis in the liver were estimated in vivo by measuring the incorporation of [^{14}C]acetate, which was intraperitoneally injected, into FAs; the incorporation of [^{14}C]acetate into hepatic FA was significantly suppressed by the treatment with F81-1144b at a dose of 100 mg/kg (Fig. 7A).

JPET #246165

Hepatic concentrations of acetyl-CoA in rats treated with F81-1144b were slightly greater than those in HSD-fed control rats; however, the extent of the increase was only 1.27-fold (Fig. 7B). Therefore, the decrease observed in the formation of [¹⁴C]-labeled FAs by F81-1144b did not appear to be due to the isotopic dilution of acetyl-CoA in the liver. F81-1144b dosing markedly affected the expression of genes encoding enzymes related to glycerolipid synthesis (Fig. 6B, Fig. 6C, Supplemental Table 2). The expression of gene for GPAT4 was significantly up-regulated by HSD feeding; the elevated levels were suppressed by F81-1144b dosing (Fig. 6C). The activity of microsomal GPAT was increased by the feeding of HSD, and the elevated activity was significantly decreased by the treatment of rats with F81-1144b (Fig. 7C). The expression of *Gpat1* was little affected by HSD feeding, whereas that in HSD-fed rats was significantly down-regulated by F81-1144b dosing (Fig. 6B). The expression of genes encoding diacylglycerol acyltransferase (DGAT) 2 was suppressed by F81-1144b dosing, while the levels of mRNA for DGAT1 were unchanged (Supplemental Table 2). Among FA transporters in the liver, the expression of gene encoding fatty acid translocase (CD36) tended to be down-regulated by the feeding of HSD (Student's *t*-test, *P* = 0.0234, ND versus HSD-fed control), and that was markedly up-regulated by the treatment of HSD-fed rats with F81-1144b in a dose-dependent manner (Fig. 6D). Fatty acid transport protein (FATP) 2 was also significantly increased by the administration of F81-1144b (Supplemental Table 2). F81-1144b dosing did not change the serum levels of NEFA (Fig. 7D), and lowered the relative weight of epididymal fat (Supplemental Fig. 2B). The treatment of rats with

JPET #246165

F81-1144b significantly decreased the levels of mRNAs for long-chain acyl-CoA synthetase (ACSL) 1 and ACSL5 (Supplemental Table 2).

Regarding the effects of F81-1144b dosing on TAG catabolism in the liver, HSD feeding did not affect the expression of genes for adipose triglyceride lipase (ATGL) or carnitine palmitoyltransferase 1a (CPT1a) (Fig. 6E, F), and the administration of F81-1144b did not up-regulate the expression of genes encoding the key enzymes and proteins related to either TAG degradation [ATGL and comparative gene identification-58 (CGI-58)] or FA catabolism [CPT1a, medium-chain acyl-CoA dehydrogenase (MCAD), and long-chain acyl-CoA dehydrogenase (LCAD)] (Fig. 6E, Fig. 6F, Supplemental Table 2). The levels of mRNA encoding uncoupling protein 2 (UCP2) in the liver were not affected by the feeding of HSD, and those were markedly augmented by the treatment of HSD-fed rats with F81-1144b (Fig. 6G).

The effects of F81-1144b on the expression of genes encoding transcription factors regulating the metabolism of FA and TAG in the liver of HSD-fed rats were estimated (Fig. 6H, Fig. 6I, Supplemental Table 2). The expression of gene encoding sterol regulatory element-binding protein-1c (SREBP-1c) was significantly up-regulated by the feeding of HSD, and F81-1144b dosing markedly down-regulated it (Fig. 6H). Similarly, the expression of gene for carbohydrate response element-binding protein (ChREBP) was significantly up-regulated by HSD feeding, and the elevated levels were markedly decreased by F81-1144b dosing (Fig. 6I); the mRNA levels for L-type pyruvate kinase (LPK), a typical ChREBP target gene (Thompson and Towel, 1991), were

JPET #246165

reduced by F81-1144b dosing (Supplemental Table 2). The administration of F81-1144b to ND-fed rats markedly down-regulated the expression of genes for these two transcription factors (Supplemental Fig. 1F, G). The expression of the gene for peroxisome proliferator-activated receptor α (PPAR α) was suppressed by the treatment of HSD-fed rats with F81-1144b (Supplemental Table 2); nevertheless, the levels of mRNAs for acyl-CoA thioesterase 1 (ACOT1), CPT1a, and ATGL, typical PPAR α target genes (Rakhshandehroo et al., 2010; Karahashi et al., 2013a), were unchanged (Supplemental Table 2). The feeding of HSD significantly increased serum concentrations of insulin, and the elevated levels were markedly reduced by the treatment of HSD-fed rats with F81-1144b (Fig. 7E). However, the administration of F81-1144b to streptozotocin-induced diabetic rats significantly lowered the serum levels of TAG without changing glucose concentrations (Supplemental Fig. 3).

Discussion

Overfeeding of sucrose or fructose leads to several adverse metabolic effects, including hypertriglyceridemia, hepatic steatosis, and insulin resistance (Tappy and Lê, 2010). Fructose increases the expression of lipogenic enzymes by inducing SREBP-1c and activating ChREBP, and suppresses FA oxidation by reducing PPAR α expression (Nagai et al., 2002; Koo et al., 2008; Softic et al., 2016), thereby favoring FA esterification and VLDL formation in the liver. Moreover, fructose deteriorates VLDL-TAG clearance (Hirano *et al.*, 1989). Collectively, these metabolic changes cause hepatic steatosis and hypertriglyceridemia. The present study showed that F81-1144b reduced serum TAG levels in rats with HSD-induced hypertriglyceridemia. Two reasons have been proposed for this reduction. F81-1144b enhanced TAG degradation by LPL in plasma or suppressed VLDL-TAG secretion. Regarding VLDL-TAG metabolism in plasma, F81-1144b up-regulated *Lpl* expression and down-regulated *Apoc3* expression in the liver. Since apo-CIII is attached to VLDL and delays VLDL breakdown by inhibiting LPL (Jong et al., 1999), the increase in *Lpl* expression and decrease in *Apoc3* expression by F81-1144b appears to result in LPL activation, thereby reducing VLDL-TAG. Besides its potential effects on VLDL-TAG degradation suggested by gene expression data, the present study showed that F81-1144b suppressed VLDL-TAG secretion from the liver to the circulation in rats in the HSD-fed state. The secretion rate is considered to depend not only on hepatic TAG availability, but also on the capacity for

JPET #246165

VLDL assembly. Regarding factors related to VLDL assembly, the present study showed that the expression of *Lipin1* and *Apoc3* in the liver was markedly down-regulated by F81-1144b, while that of *Mtp* was not. *Lipin1* positively affects the secretion pathway of TAG and apoB (Khalil et al., 2010), and apo-CIII is involved in the TAG loading of VLDL (Qin et al., 2011). Moreover, the present study revealed that intrahepatic TAG was reduced by F81-1144b. Therefore, the decreased expression of *Lipin1* and *Apoc3* in concert with the reduced TAG supply within hepatocytes by F81-1144b may have reduced VLDL formation and secretion.

We then focused on elucidating the metabolic mechanisms underlying reductions in the hepatic TAG supply by F81-1144b. Changes in hepatic TAG concentrations occur due to an imbalance between lipid acquisition (de novo lipogenesis and the uptake of circulating FA) and disposal (TAG degradation and TAG export as a component of VLDL). Regarding its effects on lipogenesis, F81-1144b significantly down-regulated the expression of genes encoding key enzymes (*Fas*, *Acc1*, *G6pd*, *Acly*, and *Me1*) related to de novo FA synthesis, strongly suggesting the suppression of de novo FA synthesis by F81-1144b. As expected, the in vivo incorporation of [¹⁴C]acetate into hepatic FAs was decreased by F81-1144b. Moreover, F81-1144b reduced the expression of *Gpat1*, *Gpat4*, *Dgat2*, and *Scd1*, but not *Dgat1*, all of which are directly or indirectly involved in TAG synthesis. GPAT 1 and 4 are rate limiting enzymes of TAG synthesis (Coleman and Mashek, 2011). The decrease observed in hepatic TAG levels by F81-1144b appears to be due to the suppressed expression of *Gpat1* and/or *Gpat4*. GPAT1 and DGAT2 are responsible for

JPET #246165

incorporating de novo synthesized FAs into TAG (Qi et al., 2012; Wendel et al., 2013). DGAT2 appears to be linked with the esterification of monounsaturated FAs endogenously formed by SCD1 to produce TAG (Man et al., 2006). Therefore, the decreased supply of de novo formed FAs (particularly palmitic, octadecenoic and palmitoleic acids) by FAS and SCD1 in concert with the reduced expression of *Gpat1* and *Dgat2* by F81-1144b appears to reduce TAG formation in the liver. Regarding FAs from the circulation, since F81-1144b markedly up-regulated *Cd36* expression, a transporter that mediates the uptake of circulating FAs (Glatz et al., 2010), and did not change serum NEFA levels, the supply of exogenous FAs potentially available for TAG synthesis and/or β -oxidation within hepatocytes may be increased by F81-1144b. GPAT4 is prone to utilize exogenous FAs for TAG formation (Wendel et al., 2013), and resides on the endoplasmic reticulum (Beigneux et al., 2006). Consistent with reduced *Gpat4* expression by F81-1144b, it significantly suppressed microsomal GPAT activity, potentially suppressing TAG formation from circulating FAs. DGAT1 plays a greater role in esterifying exogenous FAs (Qi et al., 2012). F81-1144b markedly increased *Ucp2* expression in the liver. This may ultimately help to limit TAG accumulation within hepatocytes because TAG synthesis is an ATP-dependent process and the up-regulation of *Ucp2* provides hepatocytes with a mechanism to constrain ATP production (Cortez-Pinto et al., 1999). Regarding lipid degradation, ACSL5 and GPAT1 are considered to mediate FA channeling between anabolic and catabolic pathways. Namely, the knockdown of *Acs15* decreased hepatic TAG secretion and increased FA oxidation (Bu and Mashek, 2010).

JPET #246165

GPAT1 is localized on outer mitochondrial membranes and channels FAs toward glycerolipids such as TAG, thereby diverting them away from CPT1a-mediated entry into mitochondria (Wendel et al., 2013). Therefore, the present result that F81-1144b significantly down-regulated the expression of *Acs15* and *Gpat1* suggests that F81-1144b enhanced FA degradation by elevating the supply of FAs available for β -oxidation, even though F81-1144b did not up-regulate the expression of genes for the enzymes related directly to TAG degradation (*Atgl*) and FA β -oxidation (*Cpt1a* and *Acs11*) (Coleman and Mashek, 2011; Lee et al., 2011).

The changes induced in the hepatic expression of key regulatory genes related to gluconeogenesis and de novo lipogenesis by fructose overfeeding are largely similar to those induced by a leptin deficiency (Shimomura et al., 2000; Koo et al., 2008). This state produces a vicious cycle of co-existing insulin resistance (overproduction of glucose due to resistance to the normal actions of insulin through the down-regulation of insulin receptor substrate (IRS)-2) and insulin sensitivity (elevated lipogenesis due to the continuous stimulation of SREBP-1c production by insulin signals through IRS-1). Since HSD causes hepatic insulin resistance (Tappy and L e, 2010) and the elevated expression of many lipogenic genes (Softic et al., 2016), HSD-fed rats may be in a state of co-existing insulin resistance and insulin sensitivity, as reported in leptin-deficient mice. F81-1144b dosing to HSD-fed rats decreased the hepatic expression of *Srebp1c* and serum insulin levels; these changes subsequently suppressed the expression of genes (*Fas*, *Acc1*, *G6pd*, *Acly*, *Me1*, *SCD1*, *Gpat1*, *Acs11*, and *Acs15*) related to FA and TAG metabolism because these

JPET #246165

genes are known to be regulated by insulin (Ning et al., 2011; Xu et al, 2013; Grevengoed et al., 2014). These results imply that F81-1144b ameliorates dysfunctions in responses to insulin in HSD-fed rats. It is noteworthy that F81-1144b decreased serum TAG levels without changing serum glucose levels in streptozotocin-induced diabetic rats. Moreover, SREBP-1c induction and subsequent elevations in the expression of genes for key lipogenic enzymes in the liver were shown to be independent of insulin in streptozotocin-treated mice (Matsuzaka et al., 2004). Therefore, the effects of F81-1144b on TAG metabolism in the liver and plasma in HSD-fed rats may be secondary, at least in part, to its effects on insulin, and another mechanism, which is independent of insulin, may operate for the metabolic changes induced in lipids by F81-1144b.

Fibrates, niacin, and *n*-3 FAs are typical tools in interventions to reduce serum TAG (Ito, 2015). Fibrates facilitate lipid degradation (increased expression of *Atgl* and *Cpt1a*) in the liver by stimulating PPAR α (Staels et al., 1998; Rakhshandehroo et al., 2010; Karahashi et al., 2013a), decreasing VLDL secretion; fibrates also promote VLDL lipolysis by elevating *Lpl* expression and suppressing *Apoc3* expression (Chapman et al., 2011). Niacin inhibits FA mobilization from adipose tissue and DGAT2 activities in the liver, decreasing VLDL secretion (Karpe and Frayn, 2004; Kamanna and Kashyap, 2008). *n*-3 FAs activate PPAR α and suppress SREBP-1c, resulting in the stimulation of FA β -oxidation and inhibition of lipogenesis, and also block the accumulation of apo-CIII on VLDL, potentially increasing lipolysis by LPL (Chapman et al., 2011; Shearer et al., 2012). Consequently, the mechanisms underlying the effects of F81-1144b may markedly differ

JPET #246165

from those of fibrates, niacin, and *n*-3 FAs.

In conclusion, the present study revealed that F81-1144b up-regulated *Lpl* expression, down-regulated *Apoc3* expression, and suppressed VLDL-TAG secretion, which may reduce serum TAG. The reductions observed in VLDL-TAG secretion may be due to the down-regulated expression of *Lipin1* and *Apoc3*, key factors required in VLDL assembly, and/or reduced hepatic TAG levels. Decreases in intrahepatic TAG appear to be due to the reduced de novo synthesis of FAs and TAG through the suppressed expression of SREBP-1c and ChREBP in the liver. Reductions of serum insulin by F81-1144b are at least partly responsible for the observed suppression of *Srebp1c* expression; other mechanism underlying the metabolic alterations induced by F81-1144b yet remains to be elucidated. Although F81-1144b did not affect the expression of genes (*Atgl* and *Cpt1a*) related to the degradation of TAG and FAs, but up-regulated *Ucp2* expression, it significantly decreased the expression of *Gpat1* and *Acs15* and markedly increased *Cd36* expression, changes that may consequently augment FA availability for β -oxidation. These potential metabolic changes induced by F81-1144b would markedly differ from those induced by fibrates, niacin, or *n*-3 FAs. These results suggest that F81-1144b and its analogues reported in our previous study (Morikawa et al., 2007) are a novel type of therapeutic agent for the treatment of hypertriglyceridemia.

JPET #246165

Acknowledgments

The authors acknowledge Dr. Tadanori Morikawa and Ishinori Shimizu (Daiichi Fine Chemical Co., Ltd.) for helpful suggestions in the study design and continuing encouragement. We are grateful to Dr. Takashi Tanikawa and Shingo Yamada (Josai University) for their technical assistance.

Authorship Contributions

Participated in research design: Kudo, Kawai, and Kawashima.

Conducted experiments: Eguchi, Yamazaki, and Karahashi.

Performed data analysis: Kudo, Karahashi, and Kawashima.

Wrote or contributed to the writing of the manuscript: Kudo and Kawashima.

JPET #246165

References

- Adiels M, Olofsson SO, Taskinen MR and Boren J (2008) Overproduction of very low-density lipoproteins is the hallmark of the dyslipidemia in the metabolic syndrome. *Arterioscler Thromb Vasc Biol* **28**:1225-1236.
- Beigneux AP, Vergnes L, Qiao X, Quatela S, Davis R, Watkins SM, Coleman RA, Walzem RL, Philips M, Reue K and Young SG (2006) *Agpat6*—a novel lipid biosynthetic gene required for triacylglycerol production in mammary epithelium. *J Lipid Res* **47**:734-744
- Berneis KK and Krauss RM (2002) Metabolic origins and clinical significance of LDL heterogeneity. *J Lipid Res* **43**:1363-1379.
- Bligh EG and Dyer WJ (1959) A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* **37**:911-917.
- Boullart ACI, de Graaf J and Stalenhoef AF (2012) Serum triglycerides and risk of cardiovascular disease. *Biochim Biophys Acta* **1821**:867-875.
- Bu SY and Mashek DG (2010) Hepatic long-chain acyl-CoA synthetase 5 mediates fatty acid channeling between anabolic and catabolic pathways. *J Lipid Res* **51**:3270-3280.
- Chapman MJ, Ginsberg HN, Amarenco P, Andreotti F, Boren J, Catapano AL, Descamps OS, Fisher E, Kovanen PT, Kuivenhoven JA, Lesnik P, Masana L, Nordestgaard BG, Ray KK, Reiner Z, Taskinen MR, Tokgozoglul L, Tybjaerg-Hansen A and Watts GF (2011) Triglyceride-rich lipoproteins and high-density lipoprotein cholesterol in patients at high

JPET #246165

risk of cardiovascular disease: evidence and guidance for management. *Eur Heart J*
32:1345-1361.

Chatrath H, Vuppalanchi R and Chalasani N (2012) Dyslipidemia in patients with nonalcoholic
fatty liver disease. *Semin Liver Dis* **32**:22-29.

Coleman RA and Mashek DG (2011) Mammalian triacylglycerol metabolism: synthesis, lipolysis,
and signaling. *Chem Rev* **111**:6359-6386.

Cortez-Pinto H, Zhi Lin H, Qi Yang S, Odwin Da Costa S and Diehl AM (1999) Lipids up-regulate
uncoupling protein 2 expression in rat hepatocytes. *Gastroenterology* **116**:1184-1193.

Decker K (1985) Acetyl-coenzyme A. In: Bremer J. (ed), *Methods of enzymatic analysis, vol. 7*,
Wiley-Blackwell, New York .

Fruchart JC, Sacks F, Hermans MP, Assmann G, Brown WV, Ceska R, Chapman MJ, Dodson PM,
Fioretto P, Ginsberg HN, Kadowaki T, Lablanche JM, Marx N, Plutzky J, Reiner Z,
Rosenson RS, Staels B, Stock JK, Sy R, Wanner C, Zambon A and Zimmet P (2008) The
Residual Risk Reduction Initiative: a call to action to reduce residual vascular risk in
patients with dyslipidemia. *Am J Cardiol* **102**:1k-34k.

Fujisawa T, Kotake S, Hongo K, Ito H, Otani M, Yasuda J, Morikawa T (2001) Metalloproteinase
inhibitors containing hydroxamic acids. Jpn Kokai Tokkyo Koho (Japanese Unexamined
Patent Application Publication), JP 2001055327 A.

Glatz JFC, Luiken JJFP and Bonen A (2010) Membrane fatty acid transporters as regulators of

JPET #246165

lipid metabolism: implications for metabolic disease. *Physiol Rev* **90**:367-417.

Grevengoed TJ, Klett EL and Coleman RA (2014) Acyl-CoA metabolism and partitioning. *Annu Rev Nutr* **34**:1-30.

Harchaoui KE, Visser ME, Kastelein JJ, Stroes ES and Dallinga-Thie GM (2009) Triglycerides and cardiovascular risk. *Curr Cardiol Rev* **5**:216-222.

Hirano T, Mamo JC, Poapst ME, Kuksis A and Steiner G (1989) Impaired very low-density lipoprotein-triglyceride catabolism in acute and chronic fructose-fed rats. *Am J Physiol Endocrinol Metab* **256**:E559-E565.

Imai K, Koyama M, Kudo N, Shirahata A and Kawashima Y (1999) Increase in hepatic content of oleic acid induced by dehydroepiandrosterone in the rat. *Biochem Pharmacol* **58**:925-933.

Ito MK (2015) Long-chain omega-3 fatty acids, fibrates and niacin as therapeutic options in the treatment of hypertriglyceridemia: a review of the literature. *Atherosclerosis* **242**:647-656.

Jong MC, Hofker MH and Havekes LM (1999) Role of ApoCs in lipoprotein metabolism: functional differences between ApoC1, ApoC2, and ApoC3. *Arterioscler Thromb Vasc Biol* **19**:472-484.

Kamanna VS and Kashyap ML (2008) Mechanism of action of niacin. *Am J Cardiol* **101**:20B-26B.

Karahashi M, Hoshina M, Yamazaki T, Sakamoto T, Mitsumoto A, Kawashima Y and Kudo N (2013a) Fibrates reduce triacylglycerol content by upregulating adipose triglyceride lipase in the liver of rats. *J Pharmacol Sci* **123**:356-370.

JPET #246165

Karahashi M, Ishii F, Yamazaki T, Imai K, Mitsumoto A, Kawashima Y and Kudo N (2013b)

Up-regulation of stearoyl-CoA desaturase 1 increases liver MUFA content in obese Zucker but not Goto-Kakizaki rats. *Lipids* **48**:457-467.

Karpe F and Frayn KN (2004) The nicotinic acid receptor--a new mechanism for an old drug.

Lancet **363**:1892-1894.

Kearney PM, Blackwell L, Collins R, Keech A, Simes J, Peto R, Armitage J and Baigent C (2008)

Efficacy of cholesterol-lowering therapy in 18,686 people with diabetes in 14 randomised trials of statins: a meta-analysis. *Lancet* **371**:117-125.

Khalil MB, Blais A, Figeys D and Yao Z (2010) Lipin — The bridge between hepatic glycerolipid

biosynthesis and lipoprotein metabolism. *Biochim Biophys Acta* **1801**:1249-1259.

Koo H-Y, Wallig MA, Chung BH, Nara TY, Cho BHS and Nakamura MT (2008) Dietary fructose

induces a wide range of genes with distinct shift in carbohydrate and lipid metabolism in fed and fasted rat liver. *Biochim Biophys Acta* **1782**:341-348.

Langsted A, Freiberg JJ, Tybjaerg-Hansen A, Schnohr P, Jensen GB and Nordestgaard BG (2011)

Nonfasting cholesterol and triglycerides and association with risk of myocardial infarction and total mortality: the Copenhagen City Heart Study with 31 years of follow-up. *J Intern Med* **270**:65-75.

Lee K, Kerner J and Hoppel CL (2011) Mitochondrial carnitine palmitoyltransferase 1a (CPT1a) is

part of an outer membrane fatty acid transfer complex. *J Biol Chem* **286**:25655-25662.

JPET #246165

Lowry OH, Rosebrough NJ, Farr AL and Randall RJ (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* **193**:265-275.

Man WC, Miyazaki M, Chu K and Ntambi J (2006) Colocalization of SCD1 and DGAT2: implying preference for endogenous monounsaturated fatty acids in triglyceride synthesis. *J Lipid Res* **47**:1928-1939.

Matsuzaka T, Shimano H, Yahagi N, Amemiya-Kudo M, Okazaki H, Tamura Y, Iizuka Y, Ohashi K, Tomita S, Sekiya M, Hasty A, Nakagawa Y, Sone H, Toyoshima H, Ishibashi S, Osuga J-i and Yamada N (2004) Insulin-independent induction of sterol regulatory element-binding protein-1c expression in the livers of streptozotocin-treated mice. *Diabetes* **53**:560-569.

Millar JS, Cromley DA, McCoy MG, Rader DJ and Billheimer JT (2005) Determining hepatic triglyceride production in mice: Comparison of poloxamer 407 with Triton WR-1339. *J Lipid Res* **46**:2023-2028.

Miller M, Stone NJ, Ballantyne C, Bittner V, Criqui MH, Ginsberg HN, Goldberg AC, Howard WJ, Jacobson MS, Kris-Etherton PM, Lennie TA, Levi M, Mazzone T and Pennathur S (2011) Triglycerides and cardiovascular disease: a scientific statement from the American Heart Association. *Circulation* **123**:2292-2333.

Morikawa T, Toyama T, Kudo N and Kawashima Y (2007) Reducing effect of matrix metalloproteinase inhibitors on serum triacylglycerol in streptozotocin-induced diabetic

JPET #246165

rats and Zucker fa/fa rats. *Biol Pharm Bull* **30**:1461-1467.

Nagai Y, Nishio Y, Nakamura T, Maegawa H, Kikkawa R and Kashiwagi A (2002) Amelioration of high fructose-induced metabolic derangements by activation of PPAR α . *Am J Physiol Endocrinol Metab* **282**:E1180-E1190.

Neil HA, Cooper J, Betteridge DJ, Capps N, McDowell IF, Durrington PN, Seed M, Mann JI and Humphries SE (2010) All-cause and cardiovascular mortality in treated patients with severe hypertriglyceridaemia: A long-term prospective registry study. *Atherosclerosis* **211**:618-623.

Ning J, Hong T, Yang X, Mei S, Liu Z, Liu H-Y and Cao W (2011) Insulin and insulin signaling play a critical role in fat induction of insulin resistance in mouse. *Am J Physiol Endocrinol Metab* **301**:E391-E401.

Nordestgaard BG (2016) Triglyceride-rich lipoproteins and atherosclerotic cardiovascular disease: new insights from epidemiology, genetics, and biology. *Circ Res* **118**:547-563.

Nordestgaard BG and Varbo A (2014) Triglycerides and cardiovascular disease. *Lancet* **384**:626-635.

Nordestgaard BG, Wootton R and Lewis B (1995) Selective retention of VLDL, IDL, and LDL in the arterial intima of genetically hyperlipidemic rabbits in vivo. Molecular size as a determinant of fractional loss from the intima–inner media. *Arterioscler Thromb Vasc Biol* **15**:534-542.

JPET #246165

Ouyang X, Cirillo P, Sautin Y, McCall S, Bruchette JL, Diehl AM, Johnson RJ and Abdelmalek

MF (2008) Fructose consumption as a risk factor for non-alcoholic fatty liver disease. *J*

Hepatol **48**:993-999.

Qi J, Lang W, Geisler JG, Wang P, Petrounia I, Mai S, Smith C, Askari H, Struble GT, Williams R,

Bhanot S, Monia BP, Bayoumy S, Grant E, Caldwell GW, Todd MJ, Liang Y, Gaul MD,

Demarest KT and Connelly MA (2012) The use of stable isotope-labeled glycerol and oleic

acid to differentiate the hepatic functions of DGAT1 and -2. *J Lipid Res* **53**:1106-1116.

Qin W, Sundaram M, Wang Y, Zhou H, Zhong S, Chang C-C, Manhas S, Yao EF, Parks RJ,

McFie PJ, Stone SJ, Jiang ZG, Wang C, Figeys D, Jia W and Yao Z (2011) Missense

mutation in APOC3 within the C-terminal lipid binding domain of human ApoC-III results

in impaired assembly and secretion of triacylglycerol-rich very low density lipoproteins:

Evidence that ApoC-III plays a major role in the formation of lipid precursors within the

microsomal lumen. *J Biol Chem* **286**:27769-27780.

Rakhshandehroo M, Knoch B, Muller M and Kersten S (2010) Peroxisome proliferator-activated

receptor alpha target genes. *PPAR Res* **2010**.

Sakurai T, Miyazawa S and Hashimoto T (1978) Effects of di-(2-ethylhexyl)phthalate

administration on carbohydrate and fatty acid metabolism in rat liver. *J Biochem*

83:313-320.

Sarwar N, Sandhu MS, Ricketts SL, Butterworth AS, Di Angelantonio E, Boekholdt SM,

JPET #246165

Ouwehand W, Watkins H, Samani NJ, Saleheen D, Lawlor D, Reilly MP, Hingorani AD,

Talmud PJ and Danesh J (2010) Triglyceride-mediated pathways and coronary disease:

collaborative analysis of 101 studies. *Lancet* **375**:1634-1639.

Schwartz EA and Reaven PD (2012) Lipolysis of triglyceride-rich lipoproteins, vascular

inflammation, and atherosclerosis. *Biochim Biophys Acta* **1821**:858-866.

Shearer GC, Savinova OV and Harris WS (2012) Fish oil — How does it reduce plasma

triglycerides? *Biochim Biophys Acta* **1821**:843-851.

Shimomura I, Matsuda M, Hammer RE, Bashmakov Y, Brown MS and Goldstein JL (2000)

Decreased IRS-2 and increased SREBP-1c lead to mixed insulin resistance and sensitivity

in livers of lipodystrophic and *ob/ob* mice. *Mol Cell* **6**:77-86.

Softic S, Cohen, DE and Kahn, CR (2016) Role of dietary fructose and hepatic de novo lipogenesis

in fatty liver disease. *Dig Dis Sci* **61**:1282-1293.

Staels B, Dallongeville J, Auwerx J, Schoonjans K, Leitersdorf E and Fruchart J-C (1998)

Mechanism of action of fibrates on lipid and lipoprotein metabolism. *Circulation*

98:2088-2093.

Tappy L and Lê K-A (2010) Metabolic effects of fructose and the worldwide increase in obesity.

Physiol Rev **90**:23-46.

Taskinen MR (2003) Diabetic dyslipidaemia: from basic research to clinical practice. *Diabetologia*

46:733-749.

JPET #246165

Thompson KS and Towle HC (1991) Localization of the carbohydrate response element of the rat

L-type pyruvate kinase gene. *J Biol Chem* **266**:8679-8682

Wang YI, Schulze J, Raymond N, Tomita T, Tam K, Simon SI and Passerini AG (2011)

Endothelial inflammation correlates with subject triglycerides and waist size after a high-fat meal. *Am J Physiol Heart Circ Physiol* **300**:H784-791.

Wendel AA, Cooper DE, Ilkayeva OR, Muoio DM and Coleman RA (2013) Glycerol-3-phosphate

acyltransferase (GPAT)-1, but not GPAT4, incorporates newly synthesized fatty acids into triacylglycerol and diminishes fatty acid oxidation. *J Biol Chem* **288**:27299-27306.

Xu X, So JS, Park JG and Lee AH (2013) Transcriptional control of hepatic lipid metabolism by

SREBP and ChREBP. *Semin Liver Dis* **33**:301-311.

Yamada K and Okuyama H (1978) Selectivity of diacylglycerophosphate synthesis in subcellular

fractions of rat liver. *Arch Biochem Biophys* **190**:409-420.

Yamazaki T, Kadokura M, Mutoh Y, Sakamoto T, Okazaki M, Mitsumoto A, Kawashima Y and

Kudo N (2014) Inducing effect of clofibric acid on stearyl-CoA desaturase in intestinal mucosa of rats. *Lipids* **49**:1203-1214.

JPET #246165

Footnotes

Address correspondence to: Dr. Yoichi Kawashima, Faculty of Pharmacy and Pharmaceutical

Sciences, Josai University, 1-1 Keyakidai, Sakado, Saitama 350-0295, and Japan. E-mail:

ykawash@josai.ac.jp

JPET #246165

Figure Legends

Fig. 1. Molecular structure of F81-1144b.

Fig. 2. Effects of HSD on serum TAG levels (A) and effects of F81-1144b on serum TAG

levels in HSD-fed rats (B). (A) Rats were maintained on HSD for the indicated periods of time. (B)

Rats were maintained on ND or HSD for 21 days. HSD-fed rats subcutaneously received

F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed. Data are

presented as the mean \pm S.D. ($n = 8/\text{group}$). ^{a, b, c} Differences in the mean without a common

superscript are significant ($P < 0.05$). HSD, high sucrose diet; ND, normal diet; TAG,

triacylglycerol.

Fig. 3. Effects of F81-1144b on the expression of genes related to VLDL formation and

degradation. Rats were maintained on ND or HSD for 21 days. HSD-fed rats subcutaneously

received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed. (A)

Lpl (lipoprotein lipase); (B) *Apoc3* (apolipoprotein CIII); (C) *Lipin1* (lipin 1); (D) *Mtp* (microsomal

triglyceride transfer protein); (E) *Apoa1* (apolipoprotein AI). The levels of mRNA in the liver of

HSD-fed rats relative to those in the liver of ND-fed control rats are presented as the mean \pm S.D.

($n = 6/\text{group}$). ^{a, b, c} Differences in the mean without a common superscript are significant ($P < 0.05$).

HSD, high sucrose diet; ND, normal diet; VLDL, very low density protein.

JPET #246165

Fig. 4. Effects of F81-1144b on VLDL-TAG secretion in HSD-fed rats in the fed state. Rats that were maintained on HSD for 21 days were subcutaneously administered F81-1144b at a dose of 0 or 100 mg/kg once a day for 7 days before measuring VLDL-TAG secretion. Rats were administered [³H]glycerol via the portal vein, and Triton WR1339 was injected via the tail vein 15 minutes later. Blood samples were collected 1 minute before and 30, 60, and 120 minutes after the Triton WR1339 injection. The incorporation of radioactivity into plasma TAG was measured. (A) Time-course of the appearance of [³H]TAG in plasma. (B) The rate of [³H]TAG secretion between 0 and 60 minutes. Values are presented as the mean ± S.D. (*n* = 4 or 5/group). * Significantly different from HSD-fed control (**P* < 0.05). HSD, high sucrose diet; TAG, triacylglycerol; VLDL, very low density protein.

Fig. 5. Effects of F81-1144b on contents and fatty acid composition of TAG and phospholipid in the liver of HSD-fed rats. Rats were maintained on ND or HSD for 21 days. HSD-fed rats subcutaneously received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed. (A) Effects of F81-1144b on TAG contents in the liver of HSD-fed rats (*n* = 8/group). (B) Effects of F81-1144b on phospholipid contents in the liver of HSD-fed rats (*n* = 8/group). (C) Differences in contents (μmol/g liver) of fatty acids in hepatic TAG between HSD-fed rats receiving F81-1144b at a dose of 50 mg/kg and HSD-fed control rats and between HSD-fed rats

JPET #246165

receiving F81-1144b at a dose of 100 mg/kg and HSD-fed control rats. Regarding each fatty acid, differences in contents between the two groups were calculated from the data in Supplemental Table 1 ($n = 8/\text{group}$). (D) Differences in contents ($\mu\text{mol/g}$ liver) of fatty acids in hepatic phospholipids between HSD-fed rats receiving F81-1144b at a dose of 50 mg/kg and HSD-fed control rats and between HSD-fed rats receiving F81-1144b at a dose of 100 mg/kg and HSD-fed control rats ($n = 8/\text{group}$). Regarding each fatty acid, differences in contents between the two groups were calculated from the data in Supplemental Table 1. Values are presented as the mean \pm S.D. ^{a, b, c} Differences in the mean without a common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$). HSD, high sucrose diet; ND, normal diet; TAG, triacylglycerol.

Fatty acids are designated by the numbers of carbon atoms and double bonds; palmitic acid, 16:0; palmitoleic acid, 16:1n-7; stearic acid, 18:0; octadecenoic acid, 18:1; linoleic acid, 18:2n-6; α -linolenic acid, 18:3n-3; 5,8,11-eicosatrienoic acid, 20:3n-9; 8,11,14-eicosatrienoic acid, 20:3n-6; arachidonic acid, 20:4n-6; 5,8,11,14,17-eicosapentaenoic acid, 20:5n-3; 7,10,13,16,19-docosapentaenoic acid, 22:5n-3; 4,7,10,13,16,19-docosahexaenoic acid, 22:6n-3.

Fig. 6. Effects of F81-1144b on the expression of genes related to TAG formation and degradation in the liver of HSD-fed rats. Rats were maintained on ND or HSD for 21 days.

HSD-fed rats subcutaneously received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7

JPET #246165

days before being killed. (A) *Fas* (fatty acid synthase); (B) *Gpat1* (glycerol-3-phosphate acyltransferase 1); (C) *Gpat4* (glycerol-3-phosphate acyltransferase 4); (D) *Cd36* (fatty acid translocase); (E) *Atgl* (adipose triglyceride lipase); (F) *Cpt1a* (carnitine palmitoyltransferase 1a); (G) *Ucp2* (uncoupling protein 2); (H) *Srebp1c* (sterol regulatory element-binding protein-1c); (I) *Chrebp* (carbohydrate response element-binding protein). The levels of mRNA in the liver of HSD-fed rats relative to those in the liver of ND-fed control rats are presented as the mean \pm S.D. ($n = 5$ or 6 /group). ^{a, b, c} Differences in the mean without a common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$). HSD, high sucrose diet; ND, normal diet.

Fig. 7. Effects of F81-1144b on TAG formation in the liver of HSD-fed rats. Rats were maintained on ND or HSD for 21 days. HSD-fed rats subcutaneously received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed. (A) Effects of F81-1144b on the in vivo formation of fatty acids from [¹⁴C]acetate in the liver of HSD-fed rats ($n = 8$ /group). (B) Effects of F81-1144b on acetyl-CoA contents in the liver of HSD-fed rats ($n = 11$ /group). (C) Effects of F81-1144b on microsomal activity of GPAT in the liver of HSD-fed rats ($n = 4$ /group). (D) Effects of F81-1144b on NEFA in the serum of HSD-fed rats ($n = 8$ /group). (E) Effects of F81-1144b on insulin levels in the serum of HSD-fed rats (ND-fed control, $n = 6$; HSD-fed rats, $n = 12$ /group). Values are presented as the mean \pm S.D. ^{a, b, c} Differences in the mean without a

JPET #246165

common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$). GPAT, glycerol-3-phosphate acyltransferase; HSD, high sucrose diet; ND, normal diet; NEFA, non-esterified fatty acids; TAG, triacylglycerol.

TABLE 1

Sequences of primers used for real-time PCR

Gene	Forward primer (5'-3')	Reverse primer (5'-3')	Accession No.
<i>Lpl</i>	AGCGCTCCATCCATCTCTTCAT	TCAGGCAGAGCCCTTTCTCAA	NM_012598
<i>Apoc3</i>	GACAATCGCTTCAAATCCCT	CGGCTCAAGAGTTGGTGTG	NM_012501
<i>Mtp</i>	ACGTGGTATTCCCGCCTCA	CGTCAAAGCATTTCGTTCTCG	BC012686
<i>Lipin1</i>	AGGGAGGGAGATGGTGGTTT	CTCTCCGGTATTGTGGCCCTT	NM_001012111
<i>Apoa1</i>	GGCACCGAGCTTCACAAAAA	TGATCGCTGTAGAGCCCAAAC	NM_012738.1
<i>Fas</i>	CGCCGACCAGTATAAACCCA	GTTGTAATCGGCACCCAAGTC	M76767
<i>Acc1</i>	AACGCCTTCACACCACCTTG	AGTCGCAGAAGCAGCCCAT	J03808
<i>G6pd</i>	CTTTGGACCCATCTGGAATCG	TCAAATAGCCCCACGACC	NM_017006
<i>Acly</i>	AAACTGTATCGCCCAGGCAGT	GTAACGCAGCACGTGATCCAT	J05210
<i>Me1</i>	ACAATACAGTTTGGCATTCCG	AGGATTCGCTCTCCATCAGTCA	NM_012600
<i>Scd1</i>	TCACCTTGAGAGAAGAATTAGCA	TTCCATTCCCTTCACTCTGA	J02585
<i>Gpat1</i>	AGACACAGGCAGGGAATCCAC	AATCCCGGAGAAGCCCAG	AF021348
<i>Gpat4</i>	TTGGAGTCCTGGAATTTGCTGA	GGCTAATCCCTGTGAATGCCA	NM_001047849
<i>Dgat1</i>	CCGTGGTATCCTGAATTGGT	GGCGTTCTCAATCTGAAAT	NM_053437
<i>Dgat2</i>	ATCTTCTCTGTACCTGGCT	ACCTTTCTTGGGCGTGTTC	NM_001012345
<i>Cd36</i>	CGAAGGCTTGAATCCTACCG	TGTTGACCTGCAGTCGTTT	NM_031561
<i>Fatp2</i>	TTCAACAGTGGCGATCTCCTG	ACCGGAAGGTGTCTCCAAC	NM_031736
<i>Acs11</i>	TCAGAGCAGTTCATCGGCATC	GTCGTTCCAAGCGTGTGATA	NM_012820
<i>Acs15</i>	CAAACATGGCTGCTTTCCTCA	ACCCTGGACAAGCCTCTCAA	NM_053607
<i>Atgl</i>	TCACCAACACCAGCATCCAA	TCCATCTCGGTAGCCCTGTTT	NM_001108509
<i>Cpt1a</i>	AAGGCAGCGTTCTTCGTGA	GTCAAAGCATCTTCCATGC	NM_031559
<i>Mead</i>	CTTTGCCTCTATTGCGAAGGC	TCCGAAAATCTGCACAGCATC	J02791
<i>Lcad</i>	TGTATTGGTGCCATAGCCATGA	CCCAGACCTTTTGGCATTGT	L11276
<i>UCP2</i>	CAAGACCATTGCACGAGAGGA	CAGTTGACAATGGCATTTCGG	NM_019354.2
<i>Srebp1c</i>	GGAGCCATGGATTGCACATT	AGGAAGGCTTCCAGAGAGGA	AF286469
<i>Chrebp</i>	AATAGAGGAGCTCAATGCT	CCCAGAACTTCCAGTTGTGC	AB074517
<i>Ppara</i>	AATGCCCTCGAACTGGATGAC	CACAATCCCCTCCTGCAACTT	NM_013196
<i>Lpk</i>	TGTGTACCACCGCCAGTTGTT	AGCACTTGAAGGAAGCCTCCA	M17685
<i>Acot1</i>	ACTACGACGACCTCCCCAAGA	TGGCCACGCAGGTAGTTCA	NM_031315.1
<i>β-Actin</i>	TGCAGAAGGAGATTACTGCC	CGCAGCTCAGTAACAGTCC	V01217

TABLE 2

Effects of F81-1144b on physiological and serum measurements of HSD-fed rats

Values are represented as the mean \pm S.D. ($n = 10$ /group)

F81-1144b (mg/kg)	ND		HSD		
	–		0	50	100
Body weight (g)	266.2 \pm 8.8 ^a		274.5 \pm 10.6 ^a	261.7 \pm 8.0 ^b	253.0 \pm 7.3 ^b
Liver weight (g)	10.79 \pm 0.48 ^a		13.35 \pm 1.25 ^b	13.99 \pm 1.2 ^b	14.13 \pm 0.97 ^b
Relative liver weight (% body weight)	4.05 \pm 0.11 ^a		4.86 \pm 0.35 ^b	5.35 \pm 0.4 ^c	5.58 \pm 0.32 ^c
Serum					
Cholesterol (mg/dl)	47.39 \pm 2.66 ^a		75.16 \pm 12.42 ^b	67.98 \pm 8.5 ^b	49.89 \pm 7.94 ^a
Glucose (mg/dl)	143.6 \pm 8.2 ^a		170.9 \pm 29.1 ^b	142.7 \pm 4.5 ^b	146.3 \pm 6.0 ^a

Rats were maintained on the ND or HSD for 21 days. HSD-fed rats subcutaneously received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed.

^{a, b, c} Differences in the mean without a common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$).

HSD, high sucrose diet; ND, normal diet.

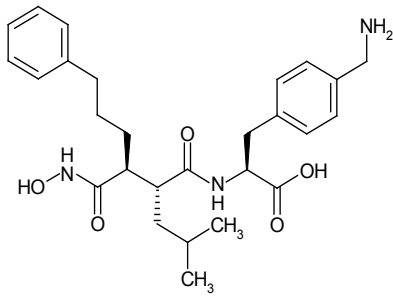


Fig. 1

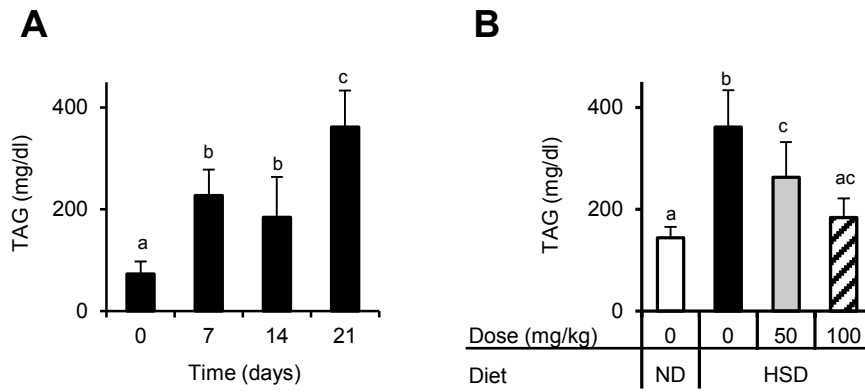


Fig. 2

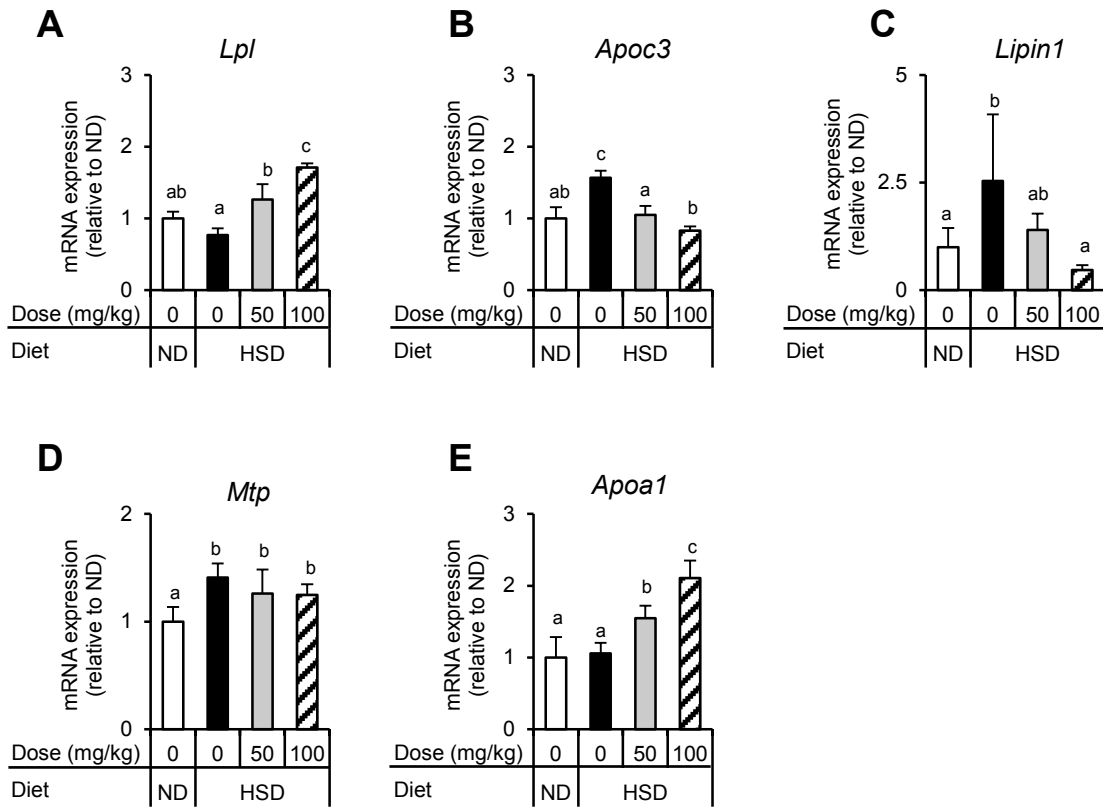


Fig. 3

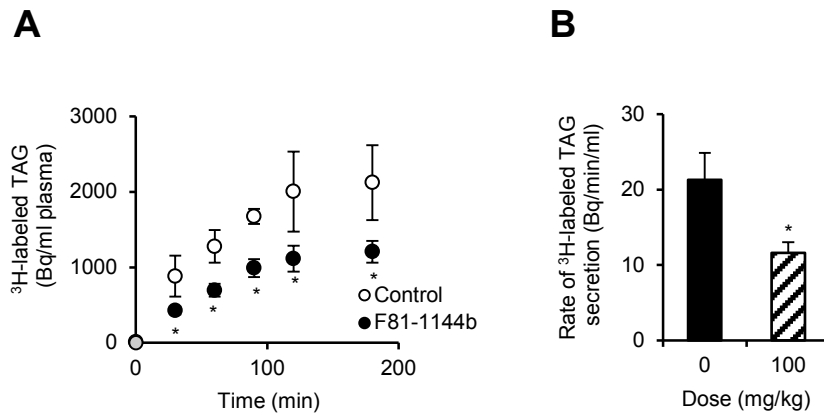


Fig. 4

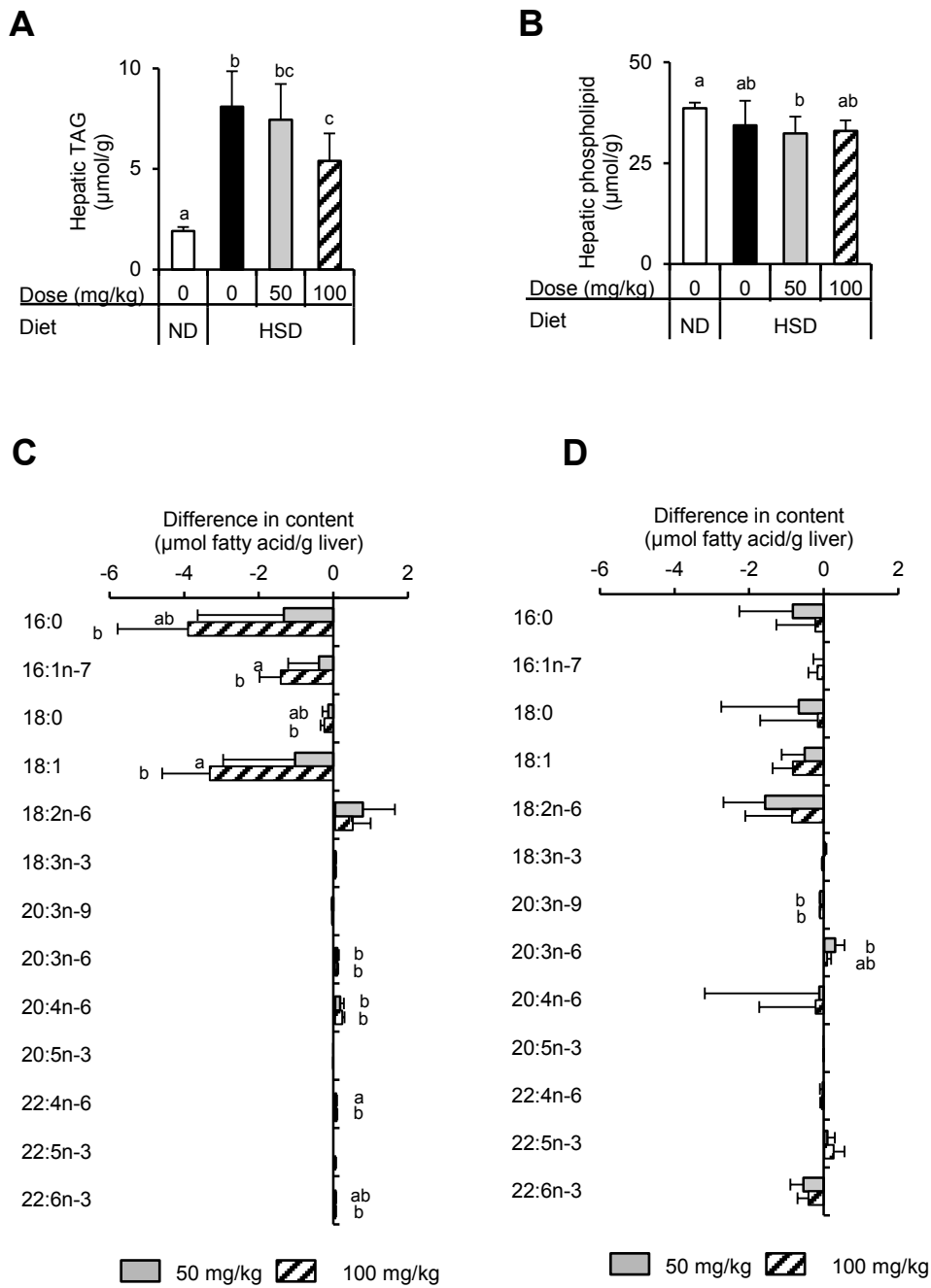


Fig. 5

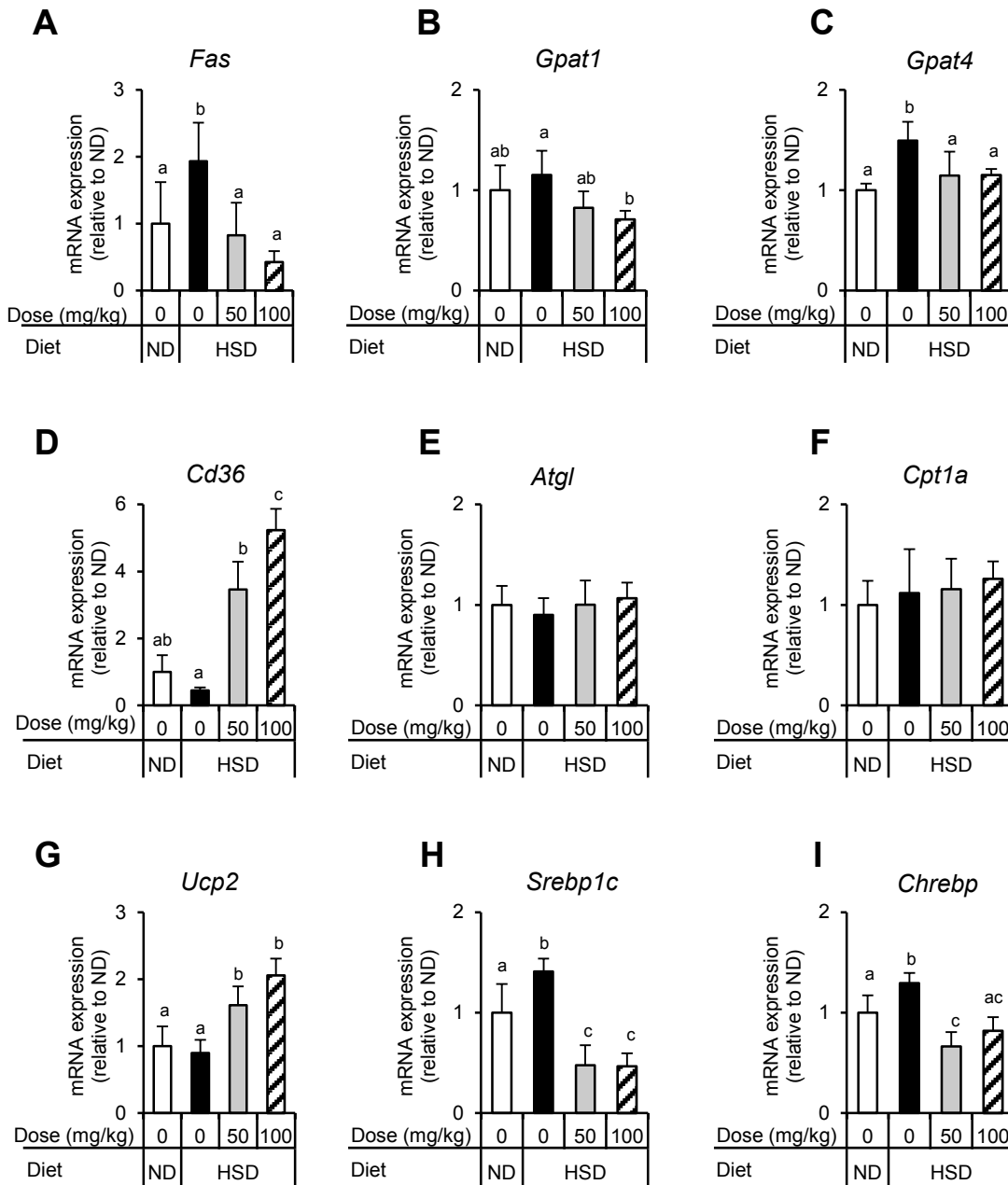


Fig. 6

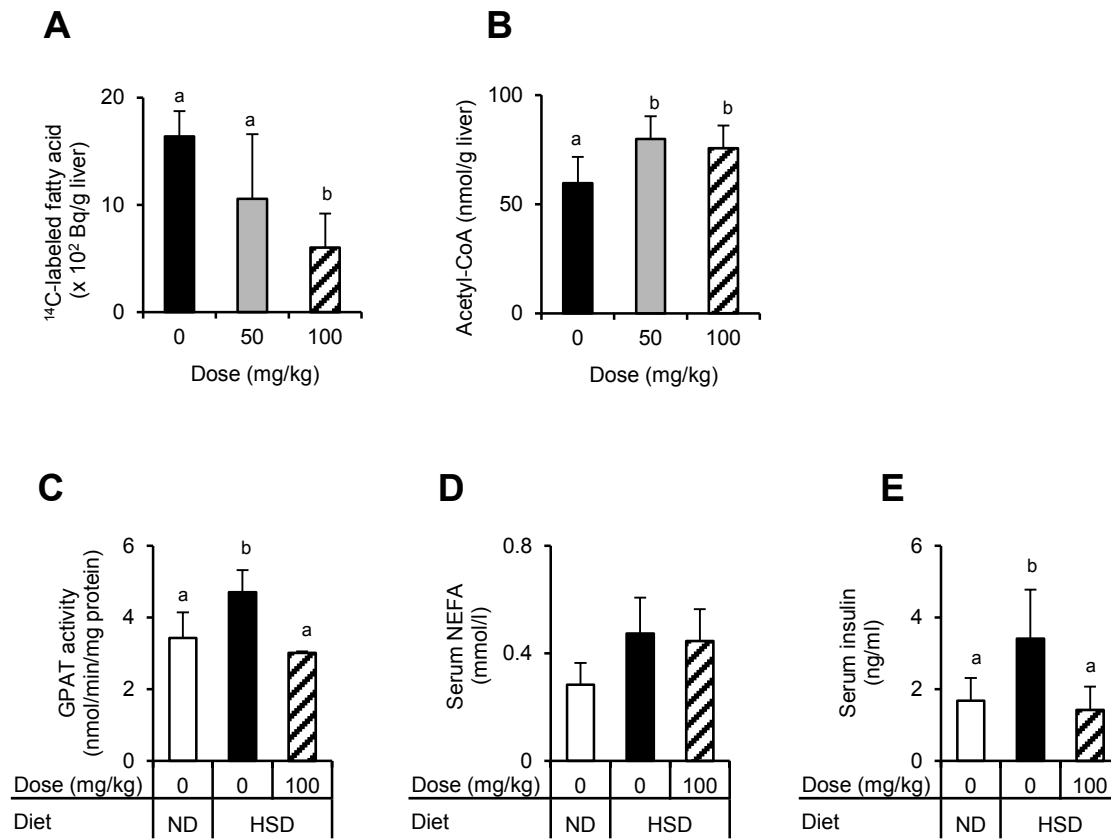


Fig. 7

JPET #246165

Supplemental Data

Reduction in secretion of very low density lipoprotein-triacylglycerol by a matrix

metalloproteinase inhibitor in a rat model of diet-induced hypertriglyceridemia

Authors:

Yoichi Kawashima, Yoshihiro Eguchi, Tohru Yamazaki, Minako Karahashi, Hiroshi Kawai and

Naomi Kudo

Journal of Pharmacology and Experimental Therapeutics

Supplemental TABLE 1

Effects of F81-1144b on fatty acid profiles of TAG and phospholipid in the liver of HSD-fed rats

Values are represented as the mean \pm S.D. (n = 4/group).

F81-1144b	ND	HFD		
	—	0 mg/kg	50 mg/kg	100 mg/kg
Fatty acids				
(mol %)				
TAG				
16:0	33.68 \pm 1.82 ^a	40.25 \pm 1.34 ^b	37.74 \pm 2.09 ^{bc}	36.01 \pm 2.83 ^{ac}
16:1 _{n-7}	4.79 \pm 0.84 ^a	10.87 \pm 0.51 ^b	10.13 \pm 2.23 ^b	7.35 \pm 2.01 ^c
18:0	1.58 \pm 0.14 ^a	2.34 \pm 0.35 ^b	1.88 \pm 0.25 ^a	1.93 \pm 0.24 ^a
18:1	23.61 \pm 0.58 ^a	33.58 \pm 0.79 ^b	31.83 \pm 1.89 ^{bc}	30.00 \pm 2.62 ^c
18:2 _{n-6}	24.57 \pm 2.04 ^a	11.56 \pm 1.82 ^b	16.12 \pm 3.66 ^c	20.79 \pm 4.61 ^{ac}
18:3 _{n-3}	1.40 \pm 0.21 ^a	0.10 \pm 0.04 ^b	0.16 \pm 0.06 ^b	0.16 \pm 0.09 ^b
20:3 _{n-9}	0.11 \pm 0.11	0.20 \pm 0.48	0.09 \pm 0.02	0.14 \pm 0.02
20:3 _{n-6}	0.50 \pm 0.58 ^{ab}	0.13 \pm 0.08 ^b	0.40 \pm 0.14 ^{ab}	0.61 \pm 0.18 ^a
20:4 _{n-6}	1.82 \pm 0.27 ^{ab}	0.65 \pm 0.16 ^c	1.23 \pm 0.36 ^{bc}	2.2 \pm 0.71 ^a
20:5 _{n-3}	1.54 \pm 0.48 ^a	0.10 \pm 0.13 ^b	0.07 \pm 0.06 ^b	0.06 \pm 0.03 ^b
22:5 _{n-3}	2.45 \pm 0.49 ^a	0.03 \pm 0.04 ^b	0.04 \pm 0.03 ^b	0.11 \pm 0.07 ^b
22:6 _{n-3}	3.20 \pm 0.34 ^a	0.05 \pm 0.02 ^b	0.07 \pm 0.05 ^b	0.17 \pm 0.11 ^b
Total (μ mol/g liver)	5.73 \pm 0.61 ^a	24.30 \pm 5.26 ^b	22.36 \pm 5.34 ^b	16.18 \pm 4.10 ^c
(mol %)				
Phospholipid				
16:0	22.99 \pm 0.71 ^a	19.53 \pm 0.5 ^b	19.48 \pm 1.13 ^b	19.98 \pm 0.51 ^b
16:1 _{n-7}	1.55 \pm 0.27 ^a	2.29 \pm 0.2 ^b	2.43 \pm 0.48 ^b	2.12 \pm 0.38 ^b
18:0	20.26 \pm 0.54 ^a	23.08 \pm 0.55 ^b	23.45 \pm 0.87 ^b	23.79 \pm 0.87 ^b
18:1	7.49 \pm 0.24 ^a	8.31 \pm 0.27 ^b	8.02 \pm 0.43 ^b	7.38 \pm 0.65 ^a
18:2 _{n-6}	16.22 \pm 0.47 ^a	12.51 \pm 0.97 ^b	10.88 \pm 1.20 ^c	11.70 \pm 1.20 ^{bc}
18:3 _{n-3}	0.17 \pm 0.06 ^a	0.08 \pm 0.05 ^b	0.12 \pm 0.09 ^a	0.03 \pm 0.02 ^c
20:3 _{n-9}	0.28 \pm 0.03 ^a	0.21 \pm 0.03 ^b	0.07 \pm 0.04 ^c	0.06 \pm 0.01 ^c
20:3 _{n-6}	0.88 \pm 0.08 ^a	1.13 \pm 0.08 ^{ab}	1.65 \pm 0.33 ^c	1.28 \pm 0.12 ^b
20:4 _{n-6}	21.17 \pm 0.75 ^a	28.08 \pm 0.54 ^b	29.59 \pm 1.26 ^c	29.00 \pm 0.53 ^{bc}

20:5n-3	0.92 ± 0.05 ^a	0.04 ± 0.02 ^b	0.05 ± 0.02 ^b	0.04 ± 0.01 ^b
22:5n-3	1.62 ± 0.13 ^a	0.41 ± 0.15 ^b	0.57 ± 0.33 ^b	0.81 ± 0.49 ^b
22:6n-3	6.28 ± 0.36 ^a	3.74 ± 0.40 ^b	3.14 ± 0.25 ^c	3.30 ± 0.40 ^{bc}
Total (μ mol/g liver)	77.31 ± 2.67 ^a	68.72 ± 12.18 ^{ab}	64.72 ± 8.31 ^b	65.94 ± 5.34 ^{ab}

Rats were maintained on ND or HSD for 21 days. HSD-fed rats subcutaneously received F81-1144b at a dose of 0, 50, or 100 mg/kg once a day for 7 days before being killed.

HSD, high sucrose diet, ND, normal diet.

Fatty acids are designated by the numbers of carbon atoms and double bonds; palmitic acid, 16:0; palmitoleic acid, 16:1n-7; stearic acid, 18:0; octadecenoic acid, 18:1; linoleic acid, 18:2n-6; α -linolenic acid, 18:3n-3; 5,8,11-eicosatrienoic acid, 20:3n-9; 8,11,14-eicosatrienoic acid, 20:3n-6; arachidonic acid, 20:4n-6; 5,8,11,14,17-eicosapentaenoic acid, 20:5n-3; 7,10,13,16,19-docosapentaenoic acid, 22:5n-3; 4,7,10,13,16,19-docosahexaenoic acid, 22:6n-3.

^{a, b, c} Significantly different without a common superscript ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$).

Supplemental TABLE 2

Effects of F81-1144b on gene expression in the liver of HSD-fed rats

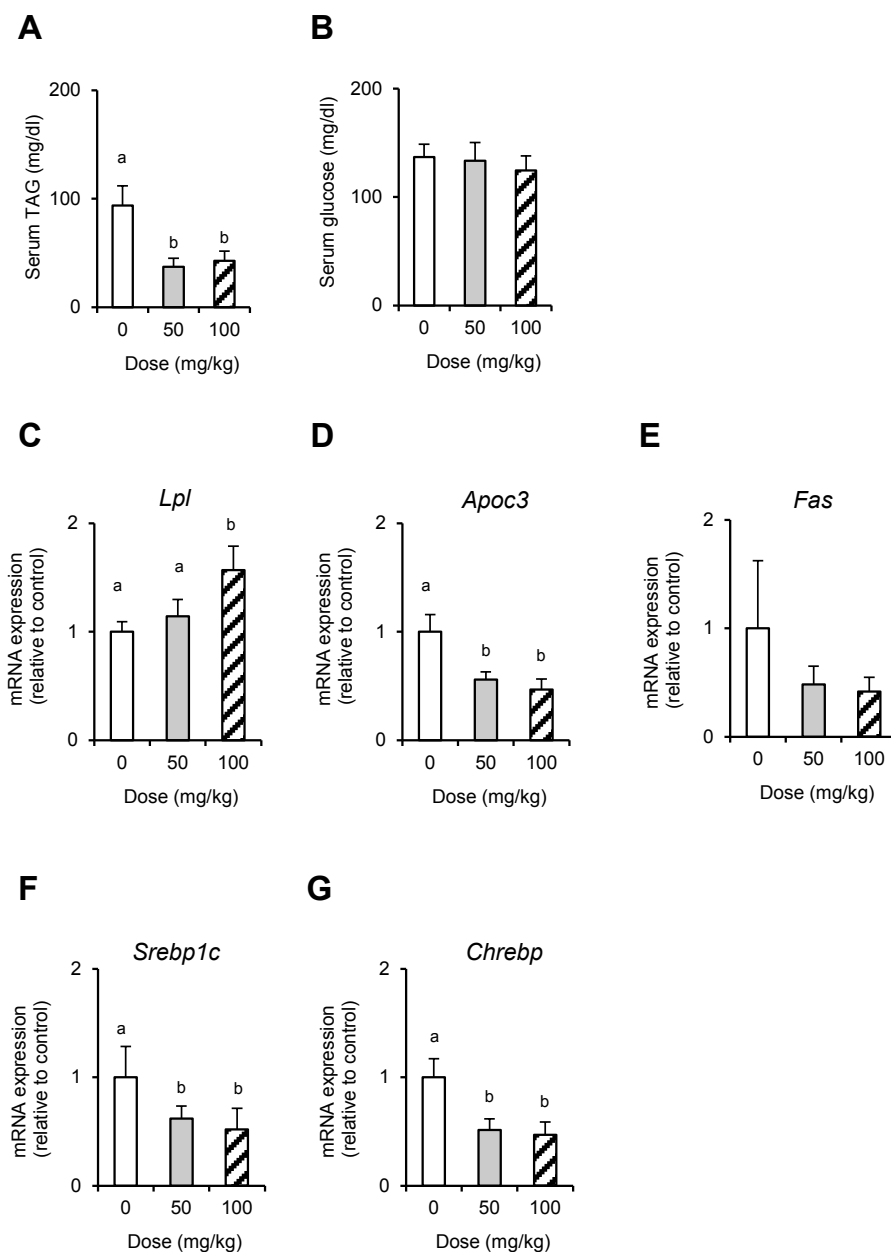
Values are represented as the mean \pm S.D. (n = 10/group)

Gene	Gene product	HSD-fed control	F81-1144b (100 mg/kg)
Lipogenesis			
<i>Fas</i>	Fatty acid synthase	1.00 \pm 0.25	0.29 \pm 0.18 ^{***}
<i>Acc1</i>	Acetyl-CoA carboxylase	1.00 \pm 0.14	0.54 \pm 0.20 ^{***}
<i>G6pd</i>	Glucose-6-phosphate dehydrogenase	1.00 \pm 0.16	0.55 \pm 0.24 ^{**}
<i>Acly</i>	ATP-citrate lyase	1.00 \pm 0.14	0.31 \pm 0.14 ^{***}
<i>Me1</i>	Malic enzyme 1	1.00 \pm 0.08	0.40 \pm 0.20 ^{***}
<i>Scd1</i>	Stearoyl-CoA desaturase 1	1.00 \pm 0.11	0.45 \pm 0.28 ^{***}
<i>Gpat1</i>	Glycerol-3-phosphate acyltransferase 1	1.00 \pm 0.16	0.60 \pm 0.23 ^{**}
<i>Gpat4</i>	Glycerol-3-phosphate acyltransferase 4	1.00 \pm 0.22	0.77 \pm 0.13 ^{**}
<i>Dgat1</i>	Acyl-CoA:diacylglycerol acyltransferase 1	1.00 \pm 0.33	1.05 \pm 0.20
<i>Dgat2</i>	Acyl-CoA:diacylglycerol acyltransferase 2	1.00 \pm 0.28	0.46 \pm 0.13 ^{***}
Fatty acid trafficking			
<i>Cd36</i>	Fatty acid translocase	1.00 \pm 0.35	11.14 \pm 3.13 ^{***}
<i>Fatp2</i>	Fatty acid transport protein 2	1.00 \pm 0.11	1.31 \pm 0.13 ^{***}
<i>Acs11</i>	Long-chain acyl-CoA synthetase 1	1.00 \pm 0.24	0.65 \pm 0.08 ^{***}
<i>Acs15</i>	Long-chain acyl-CoA synthetase 5	1.00 \pm 0.08	0.75 \pm 0.18 ^{***}
Lipid degradation			
<i>Atgl</i>	Adipose triglyceride lipase	1.00 \pm 0.32	1.16 \pm 0.23
<i>Cgi58</i>	Comparative gene identification-58	1.00 \pm 0.36	1.22 \pm 0.44
<i>Cpt1a</i>	Carnitine palmitoyltransferase 1a	1.00 \pm 0.35	1.09 \pm 0.32
<i>Mcad</i>	Medium-chain acyl-CoA dehydrogenase	1.00 \pm 0.12	0.73 \pm 0.17 ^{**}
<i>Lcad</i>	Long-chain acyl-CoA dehydrogenase	1.00 \pm 0.19	1.02 \pm 0.21
<i>Ucp2</i>	Uncoupling protein 2	1.00 \pm 0.28	2.30 \pm 0.48 ^{***}
Transcription factors and related proteins			
<i>Srebp1c</i>	Sterol regulatory element-binding protein-1c	1.00 \pm 0.12	0.50 \pm 0.26 ^{***}
<i>Chrebp</i>	Carbohydrate response element-binding protein	1.00 \pm 0.19	0.66 \pm 0.17 ^{***}
<i>Ppara</i>	Peroxisome proliferator-activated receptor α	1.00 \pm 0.16	0.56 \pm 0.15 ^{***}
<i>Lpk</i>	L-type pyruvate kinase	1.00 \pm 0.13	0.48 \pm 0.19 ^{***}
<i>Acot1</i>	Acyl-CoA thioesterase 1	1.00 \pm 0.21	0.94 \pm 0.10

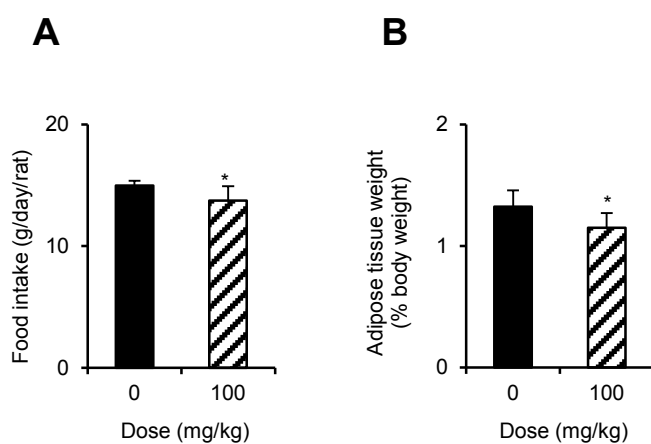
Rats were maintained on HSD for 21 days. HSD-fed rats subcutaneously received F81-1144b at a dose of 0 or 100 mg/kg once a day for 7 days before being killed.

^{*}, ^{**}, ^{***} Significantly different from the HSD-fed control (^{*} $P < 0.05$; ^{**} $P < 0.01$; ^{***} $P < 0.001$).

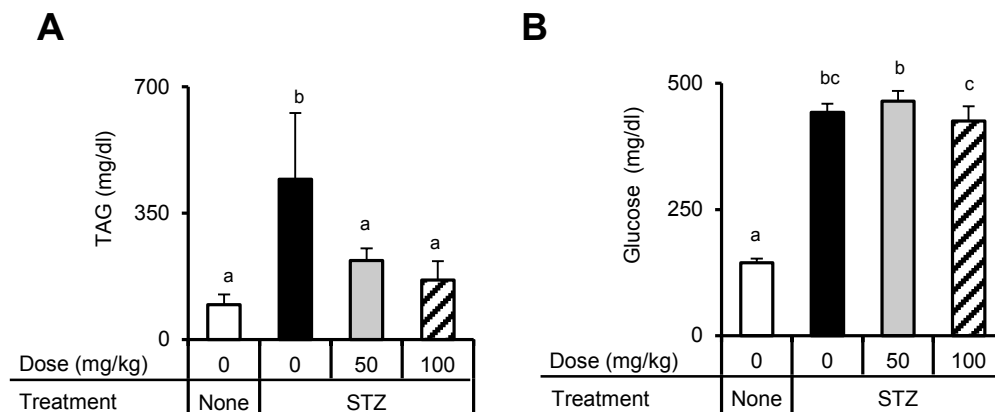
HSD, high sucrose diet.



Supplemental Figure 1. Effects of F81-1144b on serum TAG, serum glucose, and hepatic expression of genes encoding enzymes and proteins related to TAG formation and degradation in ND-fed rats. Rats were maintained on the ND throughout the experiments. Rats subcutaneously received F81-1144b at a dose of 0, 50 or 100 mg/kg once a day for 7 days before being killed. (A) TAG in serum (n = 8/group); (B) Glucose in serum (n = 8/group); (C) *Lpl* (lipoprotein lipase); (D) *Apoc3* (apolipoprotein CIII); (E) *Srebp1c* (sterol regulatory element-binding protein-1c); (F) *Chrebp* (carbohydrate response element-binding protein). The levels of mRNA in the liver of rats receiving F81-1144b relative to those in the liver of ND-fed control rats are presented (n = 5 or 6/group). Data are presented as means \pm S.D. ^{a, b} Differences in the mean without a common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$). ND, normal diet; TAG, triacylglycerol.



Supplemental Figure 2. Effects of F81-1144b on food intake (A) and the relative weight of epididymal fat (B) in HSD-fed rats. Rats were maintained on HSD for 21 days. HSD-fed rats subcutaneously received F81-114b at a dose of 0 or 100 mg/kg once a day for 7 days before being killed. Data are presented as means \pm S.D. (n = 8/group). * Significantly different from the HSD-fed control (* P < 0.05). HSD, high sucrose diet.



Supplemental Figure 3. Effects of F81-1144b on TAG (A) and glucose (B) in serum of streptozotocin-induced diabetic rats. Diabetes was induced in rats aged 6 weeks by an intravenous injection of streptozotocin (dissolved in 50 mM citrate buffer, pH 4.5) at a dose of 40 mg/kg, and rats with a blood glucose concentration greater than 500 mg/dl 21 days after the administration of streptozotocin were considered to be diabetic. Twenty-one days after the diabetic induction, rats subcutaneously received F81-1144b at a dose of 0, 50 or 100 mg/kg once a day for 7 days before being killed. Rats were maintained on the normal diet throughout the experiments. Data are presented as means \pm S.D. ($n = 6/\text{group}$). ^{a, b} Differences in the mean without a common superscript are significant ($P < 0.05$). In the absence of a superscript, the difference in the means is not significant ($P > 0.05$). TAG, triacylglycerol.