NS19504: A Novel BK Channel Activator with Relaxing Effect on Bladder Smooth Muscle Spontaneous Phasic Contractions

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ABSTRACT

Large-conductance Ca2+-activated K+ channels (BK, K\textsubscript{Ca}1.1, MaxiK) are important regulators of urinary bladder function and may be an attractive therapeutic target in bladder disorders. In this study, we established a high-throughput fluorometric imaging plate reader–based screening assay for BK channel activators and identified a small-molecule positive modulator, NS19504 (5-[4-bromophenyl]methyl]-1,3-thiazol-2-amine), which activated the BK channel with an EC\textsubscript{50} value of 11.0 ± 1.4 μM. Hit validation was performed using high-throughput electrophysiology (QPatch), and further characterization was achieved in manual whole-cell and inside-out patch-clamp studies in human embryonic kidney 293 cells expressing hBK channels: NS19504 caused distinct activation from a concentration of 0.3 and 10 μM NS19504 left-shifted the voltage activation curve by 60 mV. Furthermore, whole-cell recording showed that NS19504 activated BK channels in native smooth muscle cells from guinea pig urinary bladder. In guinea pig urinary bladder strips, NS19504 (1 μM) reduced spontaneous phasic contractions, an effect that was significantly inhibited by the specific BK channel blocker iberiotoxin. In contrast, NS19504 (1 μM) only modestly inhibited nerve-evoked contractions and had no effect on contractions induced by a high K+ concentration consistent with a K+ channel–mediated action. Collectively, these results show that NS19504 is a positive modulator of BK channels and provide support for the role of BK channels in urinary bladder function. The pharmacologic profile of NS19504 indicates that this compound may have the potential to reduce nonvoiding contractions associated with spontaneous bladder overactivity while having a minimal effect on normal voiding.

Introduction

Ca2+-activated large conductance K+ channels (BK, K\textsubscript{Ca}1.1, MaxiK, Slo1), encoded by KCNMA, are unique in the family of K+-selective ion channels by showing dual activation by both Ca2+ and membrane depolarization. The ability to respond strongly to increases in intracellular Ca2+ resulting from action potential activity makes this channel an important negative-feedback mechanism for Ca2+ entry in many excitable cell types. In addition to direct regulation by intracellular Ca2+ and voltage, BK channel activity is also modulated by other factors, such as phosphorylation state, pH, and the presence of regulatory β-subunits. Alternative splice variants of the KCNMA1 α-subunit with distinct basic physiologic and pharmacological properties have been described (Salkoff

ABBREVIATIONS: ANOVA, analysis of variance; Apa, apamin; BMS-204352, (3S)-3-(5-chloro-2-methoxyphenyl)-3-fluoro-6-(trifluoromethyl)-2-indoline; BMS-223131, 4-(5-chloro-2-hydroxyphenyl)-3-(2-hydroxyethyl)-6-(trifluoromethyl)-3,4-dihydro-1H-quinoxalin-2-one; BTC-AM, benzothia-zole coumarin acetoxymethyl ester; DMSO, dimethylsulfoxide; DRG, dorsal root ganglia; EFS, electrical field stimulation; FLIPR, fluorometric imaging plate reader; HEK, human embryonic kidney; HTS, high-throughput screening; IbTx, iberiotoxin; MPO, multiple parameter optimization; NS11021, 5-[2-[[3,5-bis(trifluoromethyl)phenyl]thiourea]-5-bromophenyl]-1H-1,2,3,4-tetrazole; NS1608, 4-chloro-2-[3-[3-( trifluoromethyl)phenyl]ureido]phenol; NS1619, 1-[2-hydroxy-5-(trifluoromethyl)phenyl]-5-(trifluoromethyl)-1,3-diaza-1,3-dihydroinden-2-one; NS19504, 5-[4-bromophenyl]methyl]-1,3-thiazol-2-amine; OAB, overactive bladder; PSS, physiologic saline solution; SPC, spontaneous phasic contraction; UBSM, urinary bladder smooth muscle.

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et al., 2006). BK channels are expressed in both excitatory and nonexcitable cells and are involved in many cellular functions, including regulation of the tone of vascular, uterine, gastrointestinal, airway and bladder smooth muscle, neuronal excitability, as well as neurotransmitter and hormone release (Ghata et al., 2006; Lu et al., 2006). As a consequence of their physiologic roles, BK channels have long been regarded as attractive therapeutic targets for a number of diseases, including urinary bladder dysfunction disorders.

In urinary bladder smooth muscle (UBSM), BK channels have been shown to be key regulators of excitability and contractility. Bladder emptying is mediated by the activation of parasympathetic nerves, which release ATP and acetylcholine to increase UBSM excitability (action potentials) and thereby contractility (Heppner et al., 2005; Petkov et al., 2005; Naush et al., 2010). BK pore-forming α and accessory β1- and β3-subunits are expressed in UBSM cells (Ohya et al., 2000; Ohl et al., 2001a,b; Petkov et al., 2001; Werner et al., 2007; Chen and Petkov, 2009; Hristov et al., 2011), and the BK channel is believed to play an important modulatory role in urinary bladder function through action potential shortening and membrane potential hyperpolarization (Heppner et al., 1997). Consistent with this role, inhibiting BK channel activity with the specific BK blocker iberiotoxin (IbTx) has been shown to increase spontaneous phasic, agonist-induced, and nerve-evoked contractions of detrusor strips from humans (Darblade et al., 2006), pigs (Buckner et al., 2002), guinea pigs (Heppner et al., 1997; Kobayashi et al., 2000; Mora and Suarez-Kurtz, 2005), rats (Uchida et al., 2005), and mice (Herrera et al., 2005).

In bladder strips from mice in which the BK pore-forming α-subunit has been deleted (slo−/− mice), contractility is markedly increased in response to both cholinergic and purinergic agonists, as well as neurotransmitter and hormone release (Ghatta et al., 2004; Thorneloe et al., 2005, 2006). BK channels have long been regarded as attractive therapeutic targets, the known activators of BK channels suffer to a varying degree from limited selectivity (Holland et al., 1996; Bentzen et al., 2009) as well as undesirable physiochemical and pharmacodynamic properties that compromise their usefulness in validating the BK channel as a viable target or their value as therapeutic agents in their own right. Accordingly, there is a continued interest in identifying new positive BK channel modulators with improved properties.

In the present study, we introduce NS19504 (5-[(4-bromophenyl) methyl]-1,3-thiazol-2-amine) identified in a high-throughput fluorescence imaging plate reader (FLIPR) screening campaign as a new BK channel activator. The compound was validated and characterized further in automated and manual patch-clamp experiments, in which it was shown to activate heterologously expressed and native BK channels in a concentration-dependent manner. A favorable selectivity profile was revealed in a screen of 68 receptors and by functional tests on Na+, Ca2+, SK, and IK channels. NS19504 potently inhibits urinary bladder spontaneous phasic contractions (SPCs) while having only a modest effect on contractions evoked by electrical field stimulation (EFS) and no effect on high K+-induced contractions. These findings suggest that NS19504 may have the potential to alleviate bladder overactivity without compromising the capacity for voluntary bladder emptying.

**Materials and Methods**

NS19504 was synthesized from readily available starting materials, as described by Obushak et al. (2004). The compound is also commercially available and can be purchased from Enamine (Kiev, Ukraine). IbTx was purchased from Tocris Bioscience (Bristol, UK) and dissolved in water.

**Cells.** Establishment of human embryonic kidney (HEK) 293 cells stably expressing hBK channels (α-subunit) has previously been described (Ahbring et al., 1997). The cells were grown in Dulbecco's minimum essential medium supplemented with 10% fetal calf serum and selection antibiotics (G418; Invitrogen, Taastrup, Denmark) and maintained at 37°C in a 5% CO2 atmosphere.

Smooth muscle cells from guinea pig urinary bladder were isolated, as described before (Layne et al., 2010). Briefly, pieces (2 × 2 mm) of guinea pig detrusor were incubated for 20 minutes at 37°C in dissection solution (80 mM monosodium glutamate, 55 mM NaCl, 6 mM KCl, 10 mM glucose, 10 mM HEPES, and 2 mM MgCl2, adjusted to pH 7.3) containing 1 mg/ml papain (Warthington Biochemical, Freehold, NJ) and 1 mg/ml dithioerythritol. Tissue pieces were then transferred to dissection solution containing 1 mg/ml collagenase (type II; Sigma-Aldrich, Brøndby, Denmark) and 100 μM CaCl2, incubated for 6 minutes at 37°C. After rinses in ice-cold dissection solution, tissue pieces were gently triturated with fire-polished Pasteur pipette to release smooth muscle cells. Smooth muscle cells were used on the next day.

For culturing of dorsal root ganglia (DRG) cells, see Supplemental Methods.

**High-Throughput Screening.** To identify new BK channel modulators, we established a T1+ influx assay, exploiting the fact that T1+ can constitute a surrogate for K+ influx. The screening campaign was performed using HEK293 cells stably expressing hBK channels and loaded with the T1+-sensitive fluorescent dye benzothiazole coumarin acetoxyethyl ester (BTC-AM). A high-throughput screening (HTS) campaign using the FLIPR methodology was launched to identify potential modulators of hBK channels in a compound collection of 174,879 compounds, of which 11,261 were proprietary, and the remainder was comprised of commercially available screening libraries.
HEK293 cells stably expressing hBK channels were seeded on 384-well, clear-bottom, black-walled Optiplates (Corning, Corning, NY) coated with poly-D-lysine (10 μg/ml) at a density of ~3 × 10^4 cells/ml in 20 μl Dulbecco’s minimum essential medium containing 10% fetal calf serum. After incubation overnight at 37°C in a humidified 5% CO2 environment, cells were washed once in Cl−-free assay buffer (in mM: 140 NaCl, 2.5 KCl, 6 CaCl2, 1 MgCl2, and 5 glucose, 10 HEPES, pH 7.3) and then loaded with the fluorescent dye BCT-AM (Invitrogen) for 1 hour in loading buffer ( assay buffer containing an additional 2 mM BCT-AM, 2 mM amaranth (Sigma-Aldrich), and 1 mM tartrazine (Sigma-Aldrich)). Assay plates were tested in a FLIPR (Molecular Devices, Sunnyvale, CA) using the 488-nm line of an argon laser for excitation and a 540 ± 30 nm bandpass filter for emission in a two-addition protocol. Following collection of a baseline signal, test compounds from the compound library were added in Cl−-free assay buffer to give a final test concentration of 30 μM [final dimethylsulfoxide (DMSO) concentration <0.1%]. Thereafter, stimulus buffer (Cl−-free assay buffer supplemented with 6 mM A23187, 18.9 mM Tl2SO4, 12.5 mM K2SO4, 2 mM amaranth, and 1 mM tartrazine) was added to the assay plates to yield final concentrations of Ti2+ and A23187 of 9.45 mM and 1.5 μM, respectively. Test compound responses were normalized to the stimulus buffer response. Screening hits were retested in quadruplicate for final concentrations of Tl2+ by Zhang et al. (1999). A Z' coefficient Z' was subsequently subjected to concentration-response fitting to Boltzmann equations of the following form:

$$I_{\text{tail}}(V) = I_{\text{tail}}(\infty) \frac{1 + \exp\left(-\frac{V - V_1/2}{S}\right)}{1 + \exp\left(-\frac{V - V_2/2}{S}\right)}$$

where $V_{1/2}$ is the potential at which half-maximal activation is obtained and $S$ is the slope factor. When results from several experiments were pooled, data were normalized with respect to fitted $I_{\text{tail}}(\infty)$ from the individual experiments.

To test for selectivity toward other classes of ion channels, recordings of whole-cell voltage-clamp experiments were conducted using rat dorsal root ganglion neurons and HEK293 cell lines expressing hIK, hSK3, hSK2, and nNa1.2 channels, respectively. hIK, hSK2, and hSK3 currents were activated by using a pipette solution with free [Ca2+]1 buffered at 0.4 μM, and currents were measured upon application of voltage ramps from −120 mV to +30 mV every 5 seconds. nNa1.2 channels in HEK293 as well as Na+ and Ca2+ channels endogenously expressed in DRG neurons were activated by 15-millisecond voltage steps to 0 mV (see Supplemental Material).

**Patch-Clamp Electrophysiology of Single Freshly Isolated Guinea Pig USBM Cells.** Whole-cell BK channel currents in response to 200-millisecond voltage ramps (~100 to +50 mV; holding potential, −50 mV) were recorded using the conventional configuration of the patch-clamp technique (Axopatch 200B amplifier and Clampex software; Molecular Devices) at room temperature. Currents were filtered at 2 kHz and digitized at 20 kHz. The bathing solution contained (in mM): 134 NaCl, 6 KCl, 1 MgCl2, 2 CaCl2, 10 glucose, and 10 HEPES, pH 7.4. The patch pipettes had tip resistance of ~3 MOhm. The pipette solution contained in mM: KCl 128.6, KOH 11.4, NaOH 10, CaCl2 3.2, ETG 5.0, Mg 1.091, pH 7.2. The free Ca2+ concentration was calculated to be 300 nM (WEBMAXC Standard, http://www.stanford.edu/~cpatton/webmaxc5.htm; Chris Paton, Stanford University).

**Guinea Pig Bladder Myography.** Juvenile guinea pigs of either sex were euthanized by isoflurane anesthesia, followed by exsanguination, according to a protocol approved by the Institutional Animal Care and Use Committee of the University of Vermont. The urinary bladder was removed and kept in ice-cold Ca2+-free HEPES-buffered solution (Ca-free HEPES; in mM: 55 NaCl, 5.6 KCl, 2 MgCl2, 80 sodium glutamate, 10 HEPES, 10 glucose, pH adjusted to 7.3 with NaOH). The caudal part of the urinary bladder (proximal urethra, trigone, distal ureters) was removed, and the remaining tissue was cut open with a longitudinal incision. Residual urine was rinsed off the tissue with ice-cold Ca-free HEPES. After removal of the urethromus, the detrusor was cut into eight strips (~6 × 1.5 mm), and loops made from silk suture were attached to either end of the tissue strips.

To myography, the strips were transferred to the organ bath of a MyoMED myograph system (Med Associates, Georgia, VT) and, by means of the loops, attached to a force transducer. The strips were bathed in physiologic saline solution (PSS; in mM: 119 NaCl, 4.7 KCl, 24 NaHCO3, 1.2 KH2PO4, 2.5 CaCl2, 1.2 MgSO4, 0.023 EDTA, 11 glucose). PSS was warmed to 37°C and bubbled with 95% O2 and 5% CO2 to provide oxygenation and maintain physiologic pH. Initially, a tension of 10 mN was applied and the tissue was allowed to equilibrate for 1 hour. During equilibration, the PSS was changed every 20 minutes. Most strips developed SPCs during equilibration. The effect of 1 μM NS19504 was tested on SPCs. After equilibration, linear voltage ramp from −120 to +80 mV every 5 seconds from a holding potential of −90 mV or a voltage step protocol (from a holding potential of −90 mV, 20 mV steps, 50-millisecond duration, were applied from −120 to +120 mV, followed by a 60-millisecond step to −120 mV). Sample rate was 10 kHz to allow for proper resolution of peak tail currents. Serial resistance (∝R) compensation (80%) was performed in the whole-cell experiments. Data are reported in the absence of leak subtraction.

For the calculation of steady-state activation curves, peak tail currents were measured at −120 mV. Steady-state activation curves were fitted to Boltzmann equations of the following form:
15 minutes after application of NS19504 or DMSO. Vehicle (DMSO) controls were run in parallel. For concentration-response curves, stock solution of NS19504 in DMSO was added in an additive fashion directly to the bath to yield 0.1, 0.3, 1, 3, 10, and 30 µM NS19504. Contractile force was analyzed before adding NS19504 and then 20 minutes after each concentration increase.

Nerve-evoked contractions were induced by EFS. A PHM-152V stimulator was used to generate square pulses of 20 V amplitude and 0.2-millisecond duration. Bursts of pulses were delivered via platinum electrodes parallel to the tissue strip for 2 seconds at 20 Hz every 2 minutes. After 30 minutes of stimulation, 1 µM NS19504 or 0.1% DMSO (vehicle) was added to the bath. Contractile force was analyzed before and 20 minutes after application of NS19504 or DMSO and 20 minutes thereafter.

For K⁺-induced depolarization of strips, the PSS in the organ bath was replaced with PSS containing 60 mM K⁺ (K⁺ PSS, in mM: 64.9 NaCl, 58.8 KCl, 24 NaHCO₃, 1.2 KH2PO₄, 2.5 CaCl₂, 1.2 MgSO₄, 0.023 EDTA, 11 glucose). After 15 minutes, 1 µM NS19504 or 0.1% DMSO (vehicle) was added to the bath. Contractile force was analyzed before application of NS19504 or DMSO and 20 minutes thereafter.

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For myography data analysis, we analyzed amplitude, force integral (area under the curve) and frequency of SPCs, amplitude and force evoked by 60 mM KCl (K⁺ PSS). Data were collected before and after application of NS19504 (1 µM) and compared using Student’s t tests. To compare different treatments, we expressed the value after treatment as percentage of the value before treatment (% of control) and performed Student’s t tests; one- or two-way analyses of variance (ANOVA) were used to compare two or more groups. Half-maximal effective concentrations (EC₅₀) were obtained from fits of concentration-response curve data to a sigmoidal concentration-response function (GraphPad Prism; GraphPad Software, La Jolla, CA). The number of experiments (n) refers to the number of tissue strips (from at least four guinea pigs).

All statistical comparisons were performed using GraphPad Prism software; P values <0.05 were considered statistically significant. All data are presented as mean ± S.E.M.

Results

Fluorescence-Based Tl⁺ Influx Assay for the Identification of hBK Modulators. A HTS campaign was launched to test for potential modulators of hBK channels in a compound collection of 174,879 compounds. As described in Materials and Methods, the screening campaign was performed as a FLIPR-based Tl⁺ influx assay. Figure 1A demonstrates that an increased fluorescence signal was readily detected upon addition of the Ca²⁺ ionophore A23187 (second addition) in the presence of a BK-positive modulator, in this two-addition Tl⁺ influx assay. The screening assay was performed in 384-well format, as demonstrated in Fig. 1B, in which columns 23 and 24 served as controls. As part of the assay validation procedure, the

![Fig. 1. FLIPR-based hBK assay.](image)
ability of known BK channel openers to potentiate the basic stimulus response was confirmed. These BK channel openers included, among others, NS1619 and NS11021 (data not shown). The screening campaign delivered 345 confirmed hits, which correspond to a hit rate of 0.2%. The confirmed FLIPR hits were subsequently validated in an automated patch-clamp assay using QPatch. Supplemental Figure 1 shows an example of a test of the compound NS19504 in the QPatch system. The resulting 231 validated hits were clustered in 30 putative chemical series, including a particularly interesting group of benzothiazoleamine compounds. The prototype compound of this group, NS19504, activated BK channels concentration-dependently in the Tl⁺ assay (Fig. 1C) with an estimated EC₅₀ value of 11.0 ± 1.4 μM (n = 4). To rule out any potential nonspecific fluorescent effects of NS19504, the compound was also tested in the presence of the BK channel antagonist paxilline. In this study, 5 μM paxilline completely abolished the modulatory response of NS19504 (data not shown). NS19504 has a structure markedly different from previously reported BK activators (Fig. 1D) as well as a low molecular mass of 269.2 Da and good calculated physicochemical properties (ACDlabs Software version 12), showing a cLogD of 2.5 at pH 7.4, a calculated intrinsic solubility of 0.097 mg/ml [measured solubilities of 0.02 mg/ml (water), 9.0 mg/ml (15% methyl-β-cyclodextrin), 0.43 mg/ml (15% hydroxypropyl-β-cyclodextrin), and 3.6 mg/ml (10% Cremophor), respectively], and a polar surface area of 67.2 Å². In addition, NS19504 is devoid of an acidic group in contrast to most known small-molecule BK channel activators. NS19504 was therefore selected for further characterization as a promising chemical lead.

**NS19504 Activation of BK Channel Currents.** NS19504 was further characterized in whole-cell patch-clamp experiments on the same hBK/HEK293 cell line as used in the FLIPR and QPatch assays. Figure 2A shows individual current responses to an applied voltage ramp in the absence and presence of NS19504 in concentrations from 0.32 to 10 μM. Figure 2B shows the time course of the entire experiment as the current measured at a ramp potential of 25 mV (indicated by the arrow in Fig. 2A). Figure 2C is a summary bar graph showing the percentage increase in BK current at the various concentrations tested. Clearly, the BK channel is reversibly modulated in a concentration-dependent manner starting at submicromolar concentrations of NS19504. Because the BK channel is expressed at high levels in the cell line used, we were not able to apply higher concentrations of NS19504 (voltage clamp not possible).

![Figure 2](https://example.com/figure2.png)

**Fig. 2.** Activation of hBK channels measured in whole-cell patch-clamp experiments. (A) hBK-mediated currents recorded from voltage ramps (top) before (Saline) and in the presence of NS19504 in the concentrations indicated to the right of the traces. Currents were measured at a ramp potential of 25 mV as indicated by arrow. (B) The currents at +25 mV were measured and plotted as a function of time. At the 0.03 μM free Ca²⁺ used in this experiment, activation was prominent from a concentration of 1 μM NS19504. Due to the high numbers of channels in the HEK293 cell line, the currents were too large to test higher concentrations of the compound. (C) Summary bar graph showing percentage BK current (control = 100%) at various concentrations of NS19504. The average values ± S.E.M. are 0.1 μM: 100% (n = 1); 0.32 μM: 121 ± 5.7% (n = 6); 1 μM: 205 ± 34% (n = 6); 3.2 μM: 530 ± 74% (n = 6); and 10 μM: 1834 ± 917 (n = 4).
yielded a potential of the subsequent step to the previous step potential. Figure 3C shows the current in response to steps to potentials from $-120$ to $+120$ mV. Tail currents were analyzed once a steady current level was obtained. The peak tail current reflects the relative fraction of open channels at the time course record in Fig. 3B). The peak tail current was measured at $+80$ mV. Application of NS19504 ($10 \, \mu M$) resulted in an increase in current at potentials positive to $0$ mV, indicating that less membrane depolarization is needed for opening of BK channels. Figure 3B shows the time course of an experiment in which current was measured at $+80$ mV. Application of NS19504 ($10 \, \mu M$) induced a fast increase in current that reached a new steady level within seconds and completely reversed to control levels upon compound washout. To determine the effect of NS19504 on the steady-state voltage activation curve, we measured tail currents at $-120$ mV after 50-millisecond steps to potentials from $-120$ to $+120$ mV. Tail currents were analyzed once a steady current level was obtained. The peak tail current reflects the relative fraction of open channels at the previous step potential. Figure 3C shows the current in response to steps to $-60$, $0$, $+60$, and $+120$ mV from a holding potential of $-90$ mV and the inward tail current in response to the subsequent step to $-120$ mV. Plotting tail currents as a function of the step potential yielded the activation curves depicted in Fig. 3D. Under control conditions, the threshold for activation was approximately $+40$ mV, and the fitted curve yielded a $V_{1/2}$ of approximately $+120$ mV. In the presence of $10 \, \mu M$ NS19504, the voltage activation curve was left-shifted with a threshold for activation of $0$ mV and $V_{1/2}$ of $+60$ mV, indicating more current at a given potential.

**Selectivity Testing.** The functional effect of NS19504 on the IK and SK Ca$^{2+}$-activated K$^+$ channels was tested in whole-cell experiments (see Supplemental Material). The pipette saline with a free Ca$^{2+}$ concentration of $0.4 \, \mu M$ was used in these experiments, and using this saline in experiments with hBK. 0.3 $\mu M$ NS19504 increased the current relatively to the baseline current to $164 \pm 26\%$ ($n = 4$), whereas $1 \, \mu M$ augmented it to $569 \pm 96\%$ ($n = 3$). On hIK a modest increase in current was observed at $1 \, \mu M$ NS19504 ($119 \pm 5\%$ S.E.M., $n = 5$), whereas clear activation was observed at $10 \, \mu M$ ($234 \pm 29\%$, $n = 5$). At hSK3 and hSK2 channels, the effects of $10 \, \mu M$ NS19504 were $113 \pm 5\%$, $n = 7$ and $290 \pm 80\%$, $n = 3$, respectively.

Furthermore, NS19504 ($10 \, \mu M$) inhibited negligibly $3 \pm 3\%$ of the peak current mediated by Na$_{a,1.2}$ channels expressed in HEK293 cells ($n = 6$) (see Supplemental Material; Supplemental Fig. 2) and $18\%$ ($n = 2$) of the Na$_{a}$ current in DRG neurons. Finally, NS19504 at $10 \, \mu M$ had no effect on high-threshold Ca$_{a}$ channels in DRG neurons ($n = 3$) (for experimental details, see Supplemental Material).

To further address receptor selectivity, we tested binding of NS19504 ($10 \, \mu M$) at 68 receptors in radioligand receptor-binding assays (Ricerca, LeadProfilingScreen, Taipei, Taiwan). Effects (defined as >50% inhibition) were observed against the following: norepinephrine transporter (SLC6A2; 74%), dopamine transporter (SLC6A3; 75%), and sigma nonopioid intracellular receptor $1$ ($\sigma_1$R; 59%). No effect was observed against the other 65 channels and receptors tested, including L- and N-type Ca$^{2+}$ channels; Na$_{a}$, K$_{ATP}$, and hERG channels; muscarinic M$_{1}$, M$_{2}$, and M$_{3}$ receptors; neuropeptide Y$_1$ and Y$_2$ receptors; NK1 receptor; and purinergic P2X and P2Y receptors (Supplemental Table 1). These observations provide experimental support for the selectivity profile of NS19504 for BK channels.

**NS19504 Activation of BK Channels in Bladder Smooth Muscle Cells.** To test the effect of NS19504 on native BK channel currents in urinary bladder, smooth muscle cells were isolated from guinea pig bladder and whole-cell

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**Fig. 3.** Electrophysiological characterization and activation of BK channels by NS19504. Current was measured in inside-out patches obtained from HEK293 cells stably expressing BK channels. Experiments were conducted at a physiologic K$^+$ gradient (4/154 mM K$^+$) and at a free intracellular Ca$^{2+}$ concentration of 0.3 $\mu M$. (A) Current-voltage (I-V) relationships measured in the absence (Control) or presence of 10 $\mu M$ NS19504 in the intracellular/bath solution. Currents were elicited by applying linear voltage ramps from $-120$ to $+80$ mV from a holding potential of $-90$ mV. (B) BK current measured at $+80$ mV depicted as a function of time. NS19504 ($10 \, \mu M$) was applied to the inside of the patch, indicated by the bar. Breaks in the recording indicate periods where voltage step protocols were applied. Data are from a single experiment representative of four independent experiments. (C) BK current activated by membrane potential steps, as illustrated in the drawing to the right. (D) Normalized tail currents depicted as a function of step potential. Data are mean ± S.E.M. of four independent experiments. Peak tail currents were measured by stepping to $-120$ mV after obtaining steady current activation at depolarized potentials either in the absence or presence of $10 \, \mu M$ NS19504.
responses to voltage ramps were investigated (Fig. 4). NS19504 increased whole-cell currents induced by voltage ramps of 200 milliseconds (−100 to +50 mV, holding potential −50 mV, 300 nM free Ca$^{2+}$) in a concentration-dependent manner. At a concentration of 0.32 μM, NS19504 increased the response to 127 ± 7% of control (n = 7). When 1 μM NS19504 was applied, a response to 194 ± 16% (n = 8) was observed; at a concentration of 3.2 μM, NS19504 increased the response to 258 ± 24% (n = 8); and at a concentration of 10 μM, NS19504 increased the response to 561 ± 114% (n = 7; data not shown). The augmented response was abolished by the BK channel inhibitor paxilline (1 μM), supporting the BK selectivity of NS19504 in guinea pig urinary bladder cells.

**Effect of NS19504 on SPCs in Guinea Pig Bladder Strips.** We studied the effect of NS19504 in urothelium-denuded bladder strips from guinea pigs, as this preparation is highly suitable for analyzing SPCs of myogenic origin (Fig. 5). SPCs developed in most strips during equilibration. We measured amplitude, force integral, and frequency of SPCs. In controls, the amplitude was 0.89 ± 0.14 mN, the force integral was 2.28 ± 0.44 mN s, and the frequency was 3.72 ± 0.32 minute$^{-1}$. A representative recording is depicted in Fig. 5A.

Application of 1 μM NS19504 significantly reduced the amplitude of SPCs to 56.3% ± 4.4% of controls (from 1.14 ± 0.26 to 0.62 ± 0.13 mN, P = 0.0035, n = 10) and force integral to 53.7% ± 5.6% of control (from 3.02 ± 0.72 to 1.56 ± 0.39 mN s, P = 0.0028, n = 10). The frequency of SPCs was also reduced by NS19504 to 76.5% ± 5.9% of control (from 4.6 ± 0.5 to 3.4 ± 0.3 minute$^{-1}$, P = 0.0064, n = 10). A representative recording is shown in Fig. 5B.

The response to NS19504 was further investigated using two different toxin blockers of Ca$^{2+}$-activated K$^+$ channels, as follows: IbTx, which selectively blocks BK channels, and Apa, which selectively blocks SK channels. After pretreating with (or without) BK or SK channel blocker for 30 minutes to allow strips to equilibrate, we applied NS19504 (1 μM) or vehicle (0.1% DMSO) and then measured amplitude, force integral, and frequency of SPCs. The specific BK channel blocker IbTx (100 nM) alone (prior to application of NS19504 or vehicle) increased amplitude, force integral, and frequency of SPCs to 4.8 ± 0.8 mN, 9.0 ± 1.6 mN s, and 8.5 ± 0.7 minutes$^{-1}$, respectively. NS19504 (1 μM) had no effect on SPCs in IbTx-pretreated strips compared with DMSO time controls (P > 0.05, one-way ANOVA; representative recording in Fig. 5C), suggesting that NS19504 acts through BK channels. To confirm this, we tested the effect of NS19504 on strips pretreated with the SK channel blocker Apa (300 nM). As was the case with inhibition of BK channels, block of SK channels increased the amplitude and force integral of SPCs to 2.0 ± 0.3 mN and 6.1 ± 0.9 mN s, respectively, but did not increase SPC frequency (2.6 ± 0.4 minutes$^{-1}$ after Apa). In contrast to pretreatment with IbTx, Apa pretreatment did not inhibit the effects of subsequently added NS19504 (1 μM).

In the presence of Apa, NS19504 reduced amplitude to 56.6% ± 4.0% of control, force integral to 57.3% ± 5.5% of control, and frequency to 77.1% ± 7.7% of control, effects that were not significantly different from those in strips treated with NS19504 alone (P > 0.05, one-way ANOVA; representative recording in Fig. 5D). Collectively, these data indicate that NS19504 (1 μM) reduces the amplitude, force integral, and frequency of the SPC by activating BK channels (Fig. 5E).

To investigate the ability of NS19504 to affect bladder strips in greater detail, we determined the relationship between NS19504 concentration and phasic contractions (Fig. 6). Stock solutions of NS19504 in DMSO were added directly to the bath. NS19504 decreased the force integral of SPCs in a concentration-dependent manner, producing a 24% decrease at 100 nM and completely eliminating SPCs at 10 μM (Fig. 6A). A fit of the data to a sigmoidal dose-response function yielded an EC$_{50}$ for NS19504 of 640 nM (log EC$_{50}$ = −0.19 ± 0.11 where EC$_{50}$ is in μM units). To confirm that the effects of NS19504 were due to activation of BK channels, we also performed concentration-response experiments with detrusor strips pretreated with IbTx (100 nM). Inhibition of BK channels shifted the concentration-response curve to the right (Fig. 6B). In the presence of IbTx, the EC$_{50}$ for NS19504 was approximately 4.3 μM, whereas the effect of NS19504 at concentrations ≤1 μM was effectively eliminated by IbTx.

In addition to spontaneous contractions, nerve-evoked contractions elicited by EFS, which causes release of the

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**Fig. 4.** NS19504 increases BK currents in freshly isolated single USBM cells. (A) Representative whole-cell currents in response to 200-millisecond voltage ramps from −100 to +50 mV (insert in top) before (saline) and in the presence of NS19504 in the concentrations indicated to the right of the trace. BK channel inhibitor paxilline (1 μM, Pax) added at the end of experiment eliminates the effect of NS19504 and decreases the currents below the control (saline) amplitude. (B) Summary of relative to control percent increase of whole-cell BK currents by 0.32, 1.0, and 3.2 μM NS19504 measured at +50 mV. BK currents were obtained by subtraction of the currents in the presence of paxilline (1 μM) from control currents and those in the presence of NS19504. Data are means ± S.E. (n = 7–8; *P < 0.05, one-way ANOVA).
neurotransmitters ATP and acetylcholine from parasympathetic nerves, were also measured (Fig. 7). The tissue was stimulated every 2 minutes with a 2-second burst of 40 pulses of 20 V amplitude and 0.2-millisecond duration. Amplitude and force integral of EFS-induced contractions were measured before and 20 minutes after the application of vehicle (0.1% DMSO) or NS19504 (1 μM). Application of vehicle did not significantly affect amplitude (31.8 ± 6.4 mN before and 30.6 ± 3.9 mN after treatment; \( P = 0.246, n = 6 \)) or force integral (85.9 ± 15.4 mN/s before and 84.8 ± 13.8 mN/s after treatment; \( P = 0.745, n = 6 \)). NS19504 (1 μM) modestly decreased force amplitude (34.3 ± 3.1 mN before and 30.1 ± 2.9 mN after treatment; \( P < 0.001, n = 8 \)) and force integral (86.2 ± 7.9 mN/s before and 75.4 ± 7.1 mN/s after treatment; \( P < 0.001, n = 8 \); Fig. 7). Expressed as a percentage, NS19504 (1 μM) reduced amplitude to 87.6 ± 2.6% (\( P = 0.003 \)) and force integral to 87.3 ± 0.9% (\( P = 0.004 \)) of DMSO controls. Thus, NS19504 (1 μM) reduces nerve-evoked contraction, but the effect is small (13% reduction).

Finally, contractions were evoked by inducing depolarization with a high concentration of extracellular K+ (60 mM; K+ PSS) to activate voltage-gated Ca2+ channels (Fig. 8), and contractile force was measured before and 15 minutes after application of NS19504 (1 μM) or vehicle (0.1% DMSO). Depolarization of urinary bladder smooth muscles by application of K+ PSS caused a biphasic contraction, with an initial peak followed by a plateau phase (Fig. 8A). Force stabilized at a value of 4.7 ± 0.4 mN (\( n = 12 \)) about 15 minutes after adding K+ PSS. Vehicle had no effect on steady-state contractions induced by K+ PSS (5.5 ± 0.6 mN before and 5.4 ± 0.7 mN after DMSO; \( P = 0.256, n = 5 \)). Treatment with NS19504 (1 μM, 15 minutes) also failed to reduce contractile force (4.0 ± 0.5 mN in control and 3.7 ± 0.5 mN with NS19504; \( P = 0.200, n = 6 \); Fig. 8B). Collectively, these results demonstrate that NS19504 (1 μM) does not exert its function by inhibiting voltage-gated Ca2+ channels.

Discussion

The BK channel has long been pursued as a therapeutic target, and several studies using BK subtype-deficient mice and selective channel blockers have suggested that activation of BK channels may be beneficial in overactive bladder (OAB) disorders. Moreover, genetic transfer of DNA encoding the
The pore-forming BK subunit has been reported to be effective in animal models of OAB (Christ et al., 2001), strengthening support for the BK channel as a valid target in the treatment of OAB. Different classes of BK channel activators have been reported and shown to be effective in studies in vitro and in animal disease models. Two promising small-molecule BK channel activators, NS-8 and TA-1702, entered clinical trials for OAB, but none of them appear to be in active development. Therefore, there is a continued search for new small-molecule BK channel activators with more favorable properties (Nardi and Olesen, 2008).

In this study, we introduce NS19504, which represents a novel chemotype among BK activators. The structure of NS19504 is markedly different from that of well known BK activators, such as the benzimidazolidin-2-one NS1619, the quinolinone BMS-223131 [4-(5-chloro-2-hydroxyphenyl)-3-(2-hydroxyethyl)-6-(trifluoromethyl)-3,4-dihydro-1H-quinolin-2-one], the oxindole BMS-204352, the anthraquinone analog BK activators of the GoSlo-SR family (Roy et al., 2012, 2014), and the bisaryl thiourea NS11021 (Bentzen et al., 2007). NS19504 is characterized by a low mol. wt. and generally very good calculated physicochemical properties, including a low cLogD value and low polar surface area as well as the absence of an acidic function, which otherwise characterize most of the known small-molecule BK activators. Such properties may be considered promising with respect to the prospects of achieving good absorption, distribution, metabolism, and excretion and safety properties. In addition, the structure and low mol. wt. appear to provide a good starting point for generation of analogs. We have adopted the multiple parameter optimization (MPO) value introduced by Wager et al. (2010), who, based on investigations of approved central nervous system active drugs, developed the MPO approach to predict drug-like properties of small molecules and expressed this as the MPO score on a scale from 0 to 6, a score of 4 being desirable. The MPO value for NS19504 is 5.24 and close to the ideal of 6. Thus, NS19504 stands out from earlier reported compounds because of its low mol. wt. and absence of an acidic function, and therefore represents an interesting lead in the search for new BK channel modulators.

NS19504 was identified using a HTS approach employing a new FLIPR-based Ti⁺-flux assay for BK channel activators as a primary screen and an automated QPatch assay as a secondary screen. Our goal was to establish an assay for the detection of positive modulators of the hBK channel of sufficient quality to permit us to conduct a HTS campaign. It has previously been reported that Ti⁺ influx can be monitored in FLIPR assays to detect activation of K⁺ channels such as Kᵥ7.2 and KᵥCa2.3 (Weaver et al., 2004). In addition, our earlier studies demonstrated the capacity to detect activators of another Ca²⁺-activated K⁺ channel
Moreover, bladder contractions evoked by 60 mM K^+ channels in an exogenous expression system; effects on IK which was highly efficacious at recombinant concentration, NS19504 had a relatively small effect (\( \mu \)M). This suggests that some other factor, such as BK channel blockers greatly increase the amplitude, force integral, and frequency of SPCs. Therefore, the lack of effect of NS19504 in the presence of IbTx was not due to an increase in force. In contrast to its inhibitory effect on SPCs, NS19504 (1 \( \mu \)M) had only a modest effect on nerve-evoked contractions (13% decrease in force integral) in guinea pig bladder strip preparations, although loss of BK channel function has been shown to result in a substantial increase of nerve-evoked contractions. Nerve-evoked contractions, which are dependent on both activation of UBSM purinergic and muscarinic receptors, trigger a burst of action potentials and concurrent calcium influx (Nausch et al., 2010) to contract urinary bladder smooth muscle. This presumably leads to a strong activation of BK channels, and therefore it is conceivable that the addition of an exogenous BK channel opener may have only a small additional effect.

Spontaneous nonvoiding contractions are not only a hallmark sign of overactive bladder, but are also believed to contribute to its pathogenesis (Brading, 1997; Andersson, 2010). Therefore, a compound capable of inhibiting nonvoiding contractions without affecting normal voiding elicited by nerve-evoked contractions may be considered to have a desired profile as a candidate to treat bladder overactivity. The current observations that NS19504 potently inhibits SPCs in guinea pig bladder strips while only modestly affecting EFS-induced contractions suggest a promising profile for treating overactive detrusor. A similar profile was previously reported for the BK channel blocker Apa, supporting that NS19504 (1 \( \mu \)M) acts on urinary bladder smooth muscle via BK channels and not SK channels. It should be noted that both SK and BK channel blockers greatly increase the amplitude, force integral, and frequency of SPCs. Therefore, the lack of effect of NS19504 in the presence of IbTx was not due to an increase in force. In contrast to its inhibitory effect on SPCs, NS19504 (1 \( \mu \)M) had only a modest effect on nerve-evoked contractions (13% decrease in force integral) in guinea pig bladder strip preparations, although loss of BK channel function has been shown to result in a substantial increase of nerve-evoked contractions. Nerve-evoked contractions, which are dependent on both activation of UBSM purinergic and muscarinic receptors, trigger a burst of action potentials and concurrent calcium influx (Nausch et al., 2010) to contract urinary bladder smooth muscle. This presumably leads to a strong activation of BK channels, and therefore it is conceivable that the addition of an exogenous BK channel opener may have only a small additional effect.

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