Activation of Protein Kinase C (PKC)\(\alpha\) or PKC\(\varepsilon\) as an Approach to Increase Morphine Tolerance in Respiratory Depression and Lethal Overdose\(^S\)

Hong-You Lin, Ping-Yee Law, and Horace H. Loh

Department of Pharmacology, University of Minnesota, Minneapolis, Minnesota

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ABSTRACT

Long-term use of opioids is hindered by respiratory depression and the possibility for fatal overdose in drug abusers. This is attributed to higher levels of tolerance that develops against antinociception than to respiratory depression. Identifying important mechanisms that would increase morphine respiratory depression and overdose tolerance could lead to the safer use of opioids. Because protein kinase C (PKC) activity mediates the development and maintenance of morphine antinociceptive tolerance, we hypothesized that activating PKC\(\alpha\) or PKC\(\varepsilon\) at the pre-Bötzer complex (preBo\(\ddot{t}\)C) can increase morphine tolerance in respiration and overdose. Laser microdissection and quantitative reverse transcriptase-polymerase chain reaction were used to compare the relative mRNA abundances of PKC\(\alpha\), \(\gamma\), and \(\varepsilon\) between ventrolateral periaqueductal gray (vlPAG) and preBo\(\ddot{t}\)C. To test whether PKC\(\alpha\) or \(\varepsilon\) could enhance morphine tolerance in respiratory depression and overdose, lentivirus carrying the wild type, constitutively activated mutants, and small interference RNA against PKC\(\alpha\) or \(\varepsilon\) was stereotaxically injected into the preBo\(\ddot{t}\)C. Expression of constitutively active PKC (CAPKC)\(\alpha\) or \(\varepsilon\), but not wild-type PKC (WTPKC)\(\alpha\) or \(\varepsilon\), at the preBo\(\ddot{t}\)C allowed rats to develop tolerance to morphine respiratory depression. In terms of lethality, expression of WTPKC\(\alpha\), CAPKC\(\alpha\), or CAPKC\(\varepsilon\) at preBo\(\ddot{t}\)C increased morphine tolerance to lethal overdose. CAPKC\(\varepsilon\)-expressing rats developed the highest level of respiratory depression tolerance. Furthermore, when CAPKC\(\varepsilon\)-lentivirus was injected into the vlPAG, rats were able to develop significant antinociceptive tolerance at low doses of morphine that normally do not cause tolerance. The approach of increasing morphine respiratory depression and lethality tolerance by increasing PKC\(\alpha\) or \(\varepsilon\) activity at preBo\(\ddot{t}\)C could be used to make opioids safer for long-term use.

Introduction

Opioids are the most effective analgesics for the treatment of moderate to severe pain, and morphine is the prototypical opiate. The long-term use of morphine is hampered both by the development of analgesic tolerance and the relative lack of tolerance to side effects such as respiratory depression (Marks and Goldring, 1973; Ling et al., 1989; Paronis and Woods, 1997; Athanasos et al., 2006). Moreover, morphine is thought to cause lethality via respiratory depression (White and Irvine, 1999). With long-term use, differential tolerance to morphine analgesia and respiratory depression would decrease the therapeutic index, the ratio between LD\(50\) and ED\(50\) (White and Irvine, 1999). Therefore, morphine would be less lethal if tolerance to respiratory depression could be increased relative to analgesic tolerance. More than 10,000 people died of unintentional nonheroin prescription opioid overdose in the United States in 2007, and the majority of deaths occurred in long-term opioid users (Hall et al., 2008; Okie, 2010). Thus, there is the need to develop a safer opioid that has similar levels of tolerance to analgesia, respiratory depression, and lethal overdose.

The preBo\(\ddot{t}\)C initiates respiration rhythm and also mediates opioid-induced respiratory depression (Janczewski and

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ABBREVIATIONS: preBo\(\ddot{t}\)C, pre-Bötzer complex; MOR, \(\mu\)-opioid receptor; CAPKC\(\alpha\), constitutively active PKC\(\alpha\); CAPKC\(\varepsilon\), constitutively active PKC\(\varepsilon\); WTPKC\(\alpha\), wild-type PKC\(\alpha\); WTPKC\(\varepsilon\), wild-type PKC\(\varepsilon\); PKC, protein kinase C; vlPAG, ventrolateral periaqueductal gray; siRNA, small interfering RNA; Hb Sat, hemoglobin oxygen saturation; HEK, human embryonic kidney; VRG, ventral respiratory group; pFRG/RTN, parafacial respiratory group/retrotrapezoid nucleus; Bo\(\ddot{t}\)C, Bötzer complex; MOR, \(\mu\)-opioid receptor; WT, wild type; CA, constitutively active; OCT, optimal cutting temperature; BSA, bovine serum albumin; %MPE, percentage maximal possible effect; AP, anteroposterior; ML, mediolateral; DV, dorsoventral; TBS, Tris-buffered saline; GFP, green fluorescent protein; ANOVA, analysis of variance; qRT-PCR, quantitative reverse transcription-polymerase chain reaction.
Feldman, 2006; Montandon et al., 2011). Somatostatin is also both a functional and anatomical marker for preBoTc since abrupt silencing of somatostatin-expressing neurons at the preBoTc results in apnea and death (Stornetta et al., 2003; Tan et al., 2008). preBoTc also stains positive for the μ-opioid receptor (MOR), and microinjection of [D-Ala²,N-Me-Phe³, Gly⁵-ol] enkephalin, a MOR agonist, slows down the respiration rate (Gray et al., 1999). Furthermore, preBoTc is involved in mediating an enhanced respiratory drive in response to hypoxic or acidic conditions (Solomon et al., 2000; Krause et al., 2009). preBoTc is sandwiched by Bötzinger complex (BoTc) and parafacial respiratory group/retrotrapezoid nucleus (pFRS/RTN) rostrally and the ventral respiratory group (VRG) caudally—all of them involved in respiration. Morphone is a well known respiratory depressant, and it is thought to cause lethality by depressing respiration (Pattinson, 2008). If cellular responses to morphine at preBoTc can be altered, it might be possible to increase overdose and respiratory depression tolerance during long-term morphine treatment. One such cellular response is the μ-opioid receptor-mediated PKC activation.

PKC activity is required for both the initiation and maintenance of morphine antinoceptive tolerance. Intracerebroventricular injection of PKC inhibitors before long-term morphine administration prevented the development of antinoceptive tolerance (Gabra et al., 2008). After tolerance has developed, intracerebroventricular injection of either nonspecific PKC inhibitor or subtype-specific inhibitors of α, γ, and ε each reversed morphine antinoceptive tolerance (Javed et al., 2004). Inhibiting PKCα has been shown to block MOR desensitization in locus coeruleus neurons, whereas inhibiting PKCε blocked MOR desensitization in HEK293 cells expressing MOR (Bailey et al., 2009; Chu et al., 2010). Furthermore, development of morphine tolerance was attenuated in PKCγ or ε knockout mice (Zeitz et al., 2001; Newton et al., 2007). Therefore, because of preBoTc’s role in initiating respiration, mediating opioid respiratory depression, and chemoreception, we reasoned that if PKC activity at the preBoTc is increased by expressing constitutively active PKC, tolerance to respiratory depression and lethality will be increased. Although several if not all of the classic or novel PKC subtypes could be involved, we decided to examine one classic subtype, PKCα, and one novel subtype, PKCε, in controlling respiratory depression and lethality tolerance. The reason for choosing these two PKC subtypes over others was also based on their relative abundance within the preBoTc and ventrolateral periaqueductal gray (vIPAG) as demonstrated in our current study. By increasing morphine tolerance in respiratory depression and lethal overdose through PKCα or ε activation at the preBoTc, the safety of morphine might be improved.

The effect of increasing PKC activity at preBoTc on morphine respiratory depression and lethality tolerance was tested in this study. Lentiviruses carrying wild type, constitutively active, or small interfering RNA (siRNA) against either PKCα or PKCε were stereotaxically microinjected into the preBoTc bilaterally. Activation of PKCα or ε at preBoTc increased tolerance to morphine respiratory depression and lethality. We observed that rats expressing constitutively activated PKCε (CAPKCε) developed the highest level of tolerance to morphine respiratory depression in terms of the time to onset of tolerance, level of hemoglobin saturation, and fold increase in ED50. We also observed that wild-type PKCε (WTPKCε) expressing rats developed overdose tolerance but not rats that received WTPKCα lentivirus injections. Therefore, there might be PKC subtype selectivity in morphine respiratory depression and lethality tolerance development.

Materials and Methods

Animals. Male Sprague-Dawley rats between 250 and 275 g were purchased from Harlan (Indianapolis, IN). All surgeries and behavioral tests were approved by the University of Minnesota Institutional Animal Care and Use Committee (protocol 0902A00823). Rats were housed in a 12-h light/dark cycle with food and water ad libitum. Only well healed rats were used in subsequent experiments because approximately 25% of rodents either died or suffered neurological deficits from stereotoxic brain surgery.

Chemicals. Morphine sulfate was obtained from the National Institute on Drug Abuse (Bethesda, MD). Ketamine was purchased from Phoenix Pharmaceuticals (Belmont, CA), xylazine was from Lloyd Laboratories (Shenandoah, IA), isoflurane was from Phoenix Pharmaceuticals, and cafestol was from Apotex Corporation (Weston, FL). Triton X-100, bovine serum albumin, paraformaldehyde, and iodixanol were purchased from Sigma-Aldrich (St. Louis, MO). Toluidine blue was from Ricca Chemical (Arlington, TX). Advanced Dulbecco’s modified Eagle’s medium-reduced serum medium, penicillin, and streptomycin were purchased from Invitrogen (Carlsbad, CA).

Cell Culture. NS20Y neuroblastoma cells were grown in advanced Dulbecco’s modified Eagle’s medium-reduced serum medium (Invitrogen) with 5% fetal calf serum, 100 units/ml penicillin, and 100 µg/ml streptomycin in a 10% CO₂ incubator. For lentiviral transductions, virus was applied at a multiplicity of infection of 500 when the cells were 25% confluent.

Lentivirus Production. Plasmids containing CAPKCα (A25E point mutation), CAPKCε (pseudosubstrate inhibitory site amino acid 154–163 deletion), WTPKCα, and WTPKCε were generously provided by Drs. Allen Samarel and Jody Martin from Loyola University (Maywood, IL) (Strait et al., 2001). Deleting or mutating the pseudosubstrate inhibitory site causes PKC to become constitutively activated (Pears et al., 1990). WTPKCα and CAPKCα were cloned into the EcoRI sites of the pCDHGF vector to generate pCDHGF-WTPKCα and pCDHGF-CAPKCα (System Biosciences, Mountain View, CA). Correct gene orientation was confirmed by nucleotide sequencing. WTPKCε and CAPKCε plasmids were amplified with primers 5’-TCAAGCTAGCACCATGGTAGTGTTCAATGGCCT-3’ and 5’-TCATGGCGGCCTCAAGGATCAGTTCACC-3’ and cloned into the NheI and NotI sites of pCDHGF vector to generate pCDHGF-WTPKCε and pCDHGF-CAPKCε. Plasmids containing siRNA against PKCα were constructed using Invitrogen’s Block-it Pol II RNAi expression kit (5’-AGGTGAAAGACCACAAATTCA-3’). Plasmids containing siRNA against PKCε were either purchased from Open Biosystems (Huntsville, AL) or constructed using Invitrogen’s Block-it Pol II RNAi expression kit (5’-AAGCATGCGGCTGTGCTTTT-3’; 5’-AGAGC-CAATCTTACACTTG-3’; 5’-GCAAGATCCACCGAGAATT-3’; K4935-00). Vector for scramble siRNA was obtained from Open Biosystems (PGL3 NsiI silencing negative control). HEK293T cells were purchased from Open Biosystems. Fifteen-centimeter plates were coated with polylysine for 15 min at 37°C and washed thoroughly with distilled water. Cells (20 million) were plated on each 15-cm dish 24 h before transfection. All plasmids were harvested using QIAGEN’s Endotoxin-free Plasmid Maxi kits and kept in 0.3 M sodium acetate/ethanol until ready to use (QIAGEN, Valencia, CA). On the day of transfection, plasmids were ethanol precipitated and dissolved in Tris-EDTA, pH 8.0. Invitrogen’s ViraPower Lentivirus Expression System was used to produce lentivirus (Invitrogen).
Three microliters of Lipofectamine 2000 was used to transfect each microgram of the four-plasmid mixture: pLP1, pLP2, pLP/VSVG, and pCDHGFp (ratio 3:1:1:5:5). Supernatants were collected every 24 h for 4 days and stored at 4°C. Cell debris were cleared by centrifuging at 3000 rpm for 5 min and then filtered through 0.45-μm filters (SCHVU01RE, Millipore, Billerica, MA). To concentrate lentivirus, 30 ml of supernatant was put into sterilized ultracentrifuge tubes (Beckman Coulter, Brea, CA). At 20 μl of 60% iodixanol (Sigma-Aldrich) was layered on the bottom using a metal cannula. After ultracentrifugation for 2.5 h at 4°C, 55,000g, the virus appeared as a visible band between the clear iodixanol and red medium. Afterward, medium was removed from the top until the last 3 ml. For larger volumes, ultracentrifugation was repeated until the suspension decreased to less than 30 ml, which underwent a last round of ultracentrifugation for 180 min 4°C at 75,000g using only 30 μl of an iodixanol underlayer. All of the supernatant was removed very carefully until the last 200 μl (Coleman et al., 2003). The concentrated lentivirus solution was measured in aliquots and frozen at –80°C. Lentivirus was titered using flow cytometry with HT-1080 cells following the manufacturer’s protocols (K4975-00; Invitrogen). All of the lentivirus used had a titer between 2 and 3 × 10^9 transducing units/ml.

**Morphine Respiratory Depression and Lethal Overdose.**

Dixxon’s up and down method was used to determine the morphine ED_{50} dose for respiratory depression and lethal overdose (Dixon, 1965). The up and down method was used because it required fewer animals for determining ED_{50} without compromising accuracy and because its use has been supported by other investigators (Lichtman, 1998; Kao et al., 2010). This method requires consecutive testing of rats using doses spaced at constant log intervals where a negative response elevates the dose for the next animal, but a positive result decreases the dose for the next rat. The log interval was 0.15 for respiration and 0.11 for lethality experiments. The initial dose of morphine used on the first rat in the sequence was an estimation of the ED_{50}, and the second dose would either be a log interval higher or lower than the first dose depending on the response of the first rodent. Each sequence required between six and nine animals to assess ED_{50}. All rats were used only once in each up and down sequence. After completing the sequence with six to nine animals, the equation ED_{50} = X_{1} + K × d was used to calculate the ED_{50}, where X_{1} was the value of the last dose injected, K was a tabular number obtained from a table reported by Dixon, and d was the interval in log scale (Dixon, 1965).

To determine the ED_{50} value of morphine to cause respiratory depression, the baseline hemoglobin oxygen saturation (Hb Sat) without any opioid treatment was measured for 5 min for all animals using the MouseOx (Starr Life Sciences, Oakmont, PA). The baseline Hb Sat was consistently between 94% and 98%. Since Dixon’s up and down method required a cutoff threshold to differentiate positive responses to morphine from negatives, a decrease in Hb Sat to <85% was deemed to be a positive response for morphine respiratory depression. The threshold of 85% was chosen because it represented a significant and robust decrease in Hb Sat from baseline (P < 0.001). After measuring the baseline Hb Sat, morphine was injected intraperitoneally, and the rats were returned to home cages for 25 min. Hb Sat then was recorded for 3 min with a data collection rate of 60 Hz. With the exclusion of motion artifacts, the average Hb Sat during the 3-min interval was recorded. If the average Hb Sat was <85%, that rat exhibited a positive response to morphine respiratory depression, and the subsequent rodent would be tested with a smaller dose. The ED_{50} of morphine to cause respiratory depression was determined for both morphine-naive rats and ones that received four daily injections of 20 mg/kg morphine. Morphine tolerance to respiratory depression is defined by one of two methods in this report: a significant increase in ED_{50} or significant attenuation in the severity of respiratory depression in response to 20 mg/kg morphine as explained in the following paragraph.

In addition to measuring morphine respiratory depression ED_{50} on the 1st and 5th day, the Hb Sat of rats after injection with 20 mg/kg morphine was measured every day for 5 days. Attenuation in the severity of Hb Sat decrease compared with day 1 would also be considered as respiratory depression tolerance in addition to ED_{50} changes. Hb Sat was measured at 25 and 50 min after morphine injections, and the lower of the two time points was recorded. Two time points were taken (25 and 50 min) instead of only one (25 min) because rats were measured in parallel instead of sequentially, such as during ED_{50} determination, allowing more time for data acquisition. Hb Sat was used to monitor respiration as it integrates both the breathing frequency and amplitude.

To determine morphine LD_{50}, Dixon’s up and down method was also used. Lethality was defined as death within 24 h of morphine injection. A time of 24 h was chosen as the cutoff for comprehensiveness even though most deaths occurred between 1.5 and 5 h after morphine injection. The LD_{50} of morphine was determined for both morphine-naive rats and ones that had been chronically treated with 4 days of morphine (5 mg/kg b.i.d.) for a total of eight injections.

**Morphine Antinociception.**

Dixon’s up and down method was used again to determine the ED_{50} of morphine to elicit antinociception because this approach required fewer animals. Antinociception was measured with the hot-plate test. The hot plate was set to 55°C, and the cut off time was 30 s. Thirty minutes after morphine injection, rats were tested on the hot plate. Licking of the hind paw was deemed a positive nociceptive response. Percentage maximal possible effect (%MPE) was calculated according to the formula: (test latency-baseline latency)/(cut off-baseline latency) × 100, where >50% MPE was noted as a positive morphine antinociceptive response. This threshold was instituted because Dixon’s up and down method to assess ED_{50} required a binary outcome of positives and negatives. The threshold of 50% MPE was chosen because it represented robust morphine antinociception. The ED_{50} of morphine to cause antinociception was determined for both morphine-naive rodents, and rats that were chronically treated with 4 days of morphine (5 or 1.5 mg/kg b.i.d.). Morphine antinociception and respiratory depression were measured in different groups of animals.

**Sterotoxic Injections.**

For microinjections at the preBötC, rats between 250 and 275 g were anesthetized with 90 mg/kg ketamine and 10 mg/kg xylazine. Depth of anesthesia was monitored every 15 min, and additional doses were given as needed. After mounting on a Stoelting stereotaxic frame (Stoelting, Wood Dale, IL), a midsagittal incision was made to expose the bregma and lambda. Bregma was positioned 5 mm below lambda, and the muscles on the back of the head were removed to uncover the skull. The connective tissue was cut away with a scissor, and a dental drill cleared away the skull to expose the obex. With obex as zero, coordinates for preBötC were anterior-posterior (AP) 0.9 mm, mediolateral (ML) ±2 mm, and dorsoventral (DV) 2.7 mm (Tan et al., 2008). One microliter of the lentiviral solution was injected on each side over 5 min for a total of two injections, and the cannula was kept in place for another 5 min before withdrawal to prevent backflow (Quintessential Stereotaxic Injector; Stoelting). Viral solution levels in the cannula tubing were marked before and after the injection to confirm virus delivery. For lentivirus microinjections at the vIPAG, four injections of 1 μl each (two injections/side) were made to each rat. Coordinates were AP -8.16 mm and AP -7.44 mm, ML ±0.8 mm, DV -6 mm with respect to bregma. Four injections were made to cover the entire rostrocaudal extent of vIPAG as the structure is longer in the anterior-posterior axis. Rats received 5 mg/kg i.p. b.i.d. ketoprofen for antinociception and 20 mg/kg i.p. t.i.d. cefazolin for antimicrobial prophylaxis after surgeries.

**Immunohistochemistry.**

Rats were anesthetized with 90 mg/kg ketamine and 10 mg/kg xylazine, perfused with phosphate-buffered saline, and then with 4% paraformaldehyde (Sigma-Aldrich). Brains were postfixed in 4% paraformaldehyde overnight at 4°C, immersed in 30% sucrose until they sank to the bottom, and frozen in OCT (Sakura Finetek, Tokyo, Japan), and 25-μm thick sections were cut.
on a cryostat (Leica Microsystems, Wetzlar, Germany). Brains of rats that died from morphine overdose were removed from the skull, directly frozen in OCT, cut into 25-µm thick sections with a cryostat, and fixed with 4% paraformaldehyde for 3 min before staining. Sections were blocked with 10% normal goat serum (Jackson ImmunoResearch, West Grove, PA), 0.3% Triton X-100 (Sigma-Aldrich), and 0.5% BSA (Sigma-Aldrich) for 1 h at room temperature. All primary antibodies were dissolved in TBS (25 mM Tris, 3 mM KCl, 140 mM NaCl, 0.05% Tween 20, pH 7.4) containing 0.3% Triton X-100 and 0.5% BSA. GFP was stained with mouse anti-GFP 1:1000 (Invitrogen), somatostatin was stained with rabbit anti-somatostatin 1:600 (ImmunoStar, Hudson, WI), and MOR was stained with rabbit anti-MOR 1:10,000 (custom antibody; GeneTex, Irvine, CA). To characterize the rabbit anti-MOR, rat and wild-type mice with cervical spinal cords served as the positive control, whereas MOR knockout mice served as negative control (Supplemental Fig. 1). Sections were incubated with primary antibodies with gentle agitation overnight at 4°C and washed three times, 15 min each. Secondary antibodies were goat anti-mouse Alexa Fluor 488 1:200 (Invitrogen) and goat anti-rabbit Alexa Fluor 594 1:200 (Invitrogen) and were diluted in TBS with 0.3% Triton X-100 and 0.5% BSA. Tissue was incubated in secondary antibody for 1 h at room temperature and then washed three times for 15 min each before mounting on slides using FluoroMount-G (SouthernBiotech, Birmingham, AL). Fluorescence images were captured with a Leica DMIRE2 microscope connected to a BD CARVII confocal imager and a Hamamatsu EM CCD camera with laser light sources from Spectral Applied Research (green: 473 nm; Richmond Hill, ON, Canada). Successful lentivirus laser light sources from Spectral Applied Research (green: 473 nm; Richmond Hill, ON, Canada). Successful lentivirus microinjection to the preBo¨tC or vlPAG was confirmed.

Laser Microdissection and Quantitative Reverse Transcription-Polymerase Chain Reaction. Rats were anesthetized with 5% isoflurane/95% O2 and decapitated. The brains were immediately removed and frozen in OCT (Sakura Finetek). On the day of microdissection, 8-µm-thick sections were cut with a cryostat, mounted on 2.0-µm-thick polyethylene naphthalate membrane coated slides (Leica Microsystems), stained with 1% toluidine blue (Ricca Chemical, Arlington, TX) for 5 s, fixed in acetone at 4°C for 3 min, and stored on dry ice until microdissection. Laser microdissection was performed with a Leica AS LMD (Leica Microsystems), and mRNA was isolated using TRI Reagent according to manufacturer’s protocol (Molecular Research Center, Cincinnati, OH). Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) was performed using QIAGEN’s Quantitect SYBR Green RT-PCR kit and the Bio-Rad iCycler (Bio-Rad, Hercules, CA). qRT-PCR primers to amplify PKCα were 5'-GGTTCACGGGCAACAGCT-3' and 5'-GGCCGATAGTTGTGCTTCT-3'; qRT-PCR primers to amplify PKCγ were 5'-GACCTCCGCAACAGCAAG-3' and 5'-GATTTCAGTTGCAAGCAGTCC-3'; qRT-PCR primers to amplify PKCε were 5'-AGCCGCGTCTCTGGAAACTCCCT-3' and 5'-AGCTGCTTCTTGCTTACACCTTGAT-3'; and qRT-PCR primers to amplify β-actin were 5'-GACGGATAGGAAGATTGGGCCAC-3' and 5'-GAGGGCATACAGGGCAACACAGC-3'. To translate Ct values into absolute copy numbers, standard curves were obtained for PKCα, γ, and ε values. To construct the standard curve with comparable levels, which are significantly lower than the PKCε abundance (P < 0.001). The preBo¨tC, PKCε was the highest followed by PKCγ and PKCε. Furthermore, preBo¨tC had more PKCε transcripts than vlPAG (P < 0.01), whereas preBo¨tC had less PKCε than vlPAG (P < 0.001). PKCγ transcripts were present at similar levels in the two nuclei (Fig. 1). Although all three PKC subtypes have been implicated in morphine antinociceptive tolerance development (Smith et al., 2007), we focused on the probable roles of PKCα and PKCε in current studies because these two PKC subtypes represent the classic and novel PKC subtypes, respectively. Furthermore, the higher PKCε levels in vlPAG compared with preBo¨tC (P < 0.001) might reflect PKCε’s role on the observed greater morphine tolerance in antinociception than respiratory depression or lethal overdose (Fig. 1).

Lentiviral Expression of PKC at the preBo¨tC. Immunohistochemistry was carried out to confirm transduction of PKC in the preBo¨tC. preBo¨tC is located in the ventrolateral medulla, and somatostatin is both a functional and anatomical marker for preBo¨tC neurons (Stornetta et al., 2003; Tan et al., 2008). GFP delivered by the lentivirus confirms transgene expression in the ventrolateral medulla cells (Fig. 2A). Higher magnification colocalization studies indicated that a subset of GFP-positive cells also expressed the µ-opioid receptor and somatostatin, markers for preBo¨tC neurons (Fig.
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2, B and C). The intracellular staining of MOR is attributed to permeabilization of the plasma membrane with 0.3% Triton X-100, which was required in co-staining with GFP and is in accord with earlier reports (Arvidsson et al., 1995). By the 2nd week after lentivirus injection, robust GFP expression was readily observed. Such GFP expression continued for at least 6 months. All experiments were completed within the time frame between 2 weeks to 6 months after lentivirus injection.

**The Role of PKCα in Tolerance to Morphine-Induced Respiratory Depression.** PKCα is known to be important in maintaining morphine antinociceptive tolerance (Smith et al., 2007). Whether PKCα is involved in respiratory depression tolerance was investigated by expressing the wild-type, constitutively active, or siRNA against PKCα at preBo¨tC through lentivirus injection. siRNA knockdown of PKCα was confirmed by laser-microdissecting rat preBo¨tC, and its mRNA abundance was quantitated via qRT-PCR. Lentiviral infection was confirmed both in mouse NS20Y neuroblastoma cells and rat preBo¨tC. Knockdown in rat preBo¨tC was 88.2% compared with GFP, scramble siRNA, or PKCα siRNA lentivirus at the preBo¨tC, 20 mg/kg morphine consistently depressed Hb saturation below 79% throughout the five daily administrations, indicating a lack of respiratory tolerance (Fig. 4B). In contrast, CAPKCe-injected rats started to develop significant tolerance to morphine respiratory depression on the 3rd day with Hb Sat of 91.7% (P < 0.01) [Fig. 4C; F(3,260) = 216.1; P < 0.0001]. CAPKCe also developed tolerance faster than CAPKCa as CAPKCa rats exhibited tolerance from the 4th day instead of the 3rd, similar to CAPKCs (Figs. 3C and 4C). Alternatively, tolerance can be quantified as increases in ED50, morphine-naive ED50 for respiratory depression was compared with the ED50 after four injections of 20 mg/kg morphine sulfate. Rats that received CAPKCe at preBo¨tC demonstrated a 3-fold increase in respiratory depression ED50 from 16.8 ± 1.81 to 51.4 ± 8.55 mg/kg (P < 0.001) (Fig. 4D). When both were treated with morphine long term, the ED50 of CAPKCe was significantly higher than that of WTPKCs (P < 0.01) [Fig. 4D; F(10,60) = 8.74; P < 0.01]. Injection of GFP, scramble siRNA, PKCε siRNA, and WTPKCe into preBo¨tC did not result in significant increases in the respiratory depression ED50 after long-term morphine treatment (Fig. 4D). Thus, similar to PKCα, activation of PKCε rather than simple overexpression is needed to cause morphine tolerance in respiratory depression.

**PKCα and PKCε Increase Morphine Tolerance to Lethal Overdose.** Lethal overdose is the most feared adverse effect during opioid treatment, and respiratory depression is thought to be the cause of morphine lethality (White and Irvine, 1999). Increasing tolerance to overdose would make morphine a safer drug for long-term use. Given that activating PKCκ and PKCe increased tolerance to respiratory depression, tolerance to lethal overdose might also be increased. In rats that received GFP, scramble siRNA, PKCα siRNA, PKCε siRNA, or WTPKCs lentivirus injections at preBo¨tC bilaterally, LD50 did not increase significantly after 4 days of 5 mg/kg b.i.d. morphine. When rats received siRNA against PKCe at the preBo¨tC, there was a nonstatistically significant decrease in LD50 after long-term use of morphine.

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**Fig. 1.** Relative abundance of PKC subtypes at the vlPAG and preBo¨tC. vlPAG and preBo¨tC were laser microdissected to compare the relative quantities of PKCα, γ, and ε and β-actin transcripts in the two nuclei. Copy numbers were normalized to that of β-actin, which was set as 1. *, P < 0.05, compared with PKCα vlPAG, PKCγ vlPAG, and PKCe preBo¨tC. †††, P < 0.001, compared with PKCy preBo¨tC and PKCe preBo¨tC. **, P < 0.01 compared with PKCe vlPAG. Data were analyzed with two-way ANOVA with Bonferroni’s post-test and presented as mean ± S.E.M (n ≥ 5).
With the same 4 days of 5 mg/kg b.i.d. morphine, rats injected with WTPKCε had their LD₅₀ value increased by 2.02-fold ($P < 0.01$), whereas those injected with CAPKCα and CAPKCε had their LD₅₀ value increased by 2.12- ($P < 0.001$) and 2.76-fold ($P < 0.001$), respectively, compared with the morphine-naive LD₅₀ values within the same gene [Fig. 5A; $F_{(16,107)} = 6.49; P < 0.001$]. In addition, after long-term morphine treatment, the LD₅₀ of CAPKCα was significantly higher than WTPKCα, and the LD₅₀ of CAPKCε was higher than WTPKCε ($P < 0.05$) (Fig. 5A). To complement LD₅₀ quantifications, a Kaplan-Meier curve illustrates how GFP and CAPKCε-expressing rats survived when injected with increasing doses of morphine. GFP and CAPKCε rodents were first treated with 5 mg/kg b.i.d. morphine for 4 days, and then both groups were challenged to increasing doses of morphine each day to measure the percentage survival. CAPKCε-expressing rats survived higher doses of morphine than GFP-expressing rats (Fig. 5B). Seventy-eight percent of the rats injected with GFP virus died at 301 mg/kg, and the remaining 22% died when injected with 390 mg/kg the next...
day. In contrast, only 43% of CAPKε-expressing rats died when injected with 301 mg/kg morphine, and the remaining 57% all survived dosage of 645 mg/kg with one rat able to survive dosage of 1071 mg/kg. When morphine caused lethality, most rats died between 1.5 and 5 h after morphine injection.

Expressing CAPKε at the vIPAG Decreased the Dose Required to Develop Morphone Antinociceptive Tolerance. One possible explanation of why expressing PKC at the preBotC increased tolerance to respiratory depression and lethal overdose is that PKC lowered the threshold to develop tolerance. The threshold required to develop morphine tolerance is different for antinociception versus lethality. In wild-type rats treated with 5 mg/kg b.i.d. morphine for 4 days, the ED50 of morphine to cause antinociception increased 3-fold from 4.04 to 12.87 mg/kg ($t = 3.17; P < 0.01$), but their LD50 did not change significantly (Fig. 6, A and B). Thus, lethality has a higher threshold to develop tolerance than antinociception. Expressing CAPKε at preBotC might increase tolerance to respiratory depression and lethality by decreasing the threshold to develop tolerance. If this is the case, injection of CAPKε could also alter tolerance development to morphine’s other effects. Hence, to determine whether CAPKε is able to decrease the threshold for morphine antinociceptive tolerance development, we injected either saline or CAPKε lentivirus into the vIPAG (Fig. 6C). When both groups were treated with 1.5 mg/kg b.i.d. for 4 days, only the CAPKε-expressing rats developed tolerance—$ED_{50}$ increased 2-fold to 11.9 ± 1.73 mg/kg ($P < 0.01$) [Fig. 6D; $F_{1,26} = 20.04; P < 0.001$]. Since CAPKε decreased the threshold to develop morphine tolerance, we sought to investigate whether CAPKε could further increase tolerance after morphine antinociceptive tolerance developed. Rats given saline injections or CAPKε at the vIPAG were treated with 5 mg/kg b.i.d. morphine for 4 days, a regimen that results in significant antinociceptive tolerance. Both saline- and CAPKε-injected rats developed similar levels of antinociceptive tolerance ($P < 0.05$ for saline and $P < 0.01$ for CAPKε) [Fig. 6E; $F_{1,26} = 22.48; P < 0.0001$]. Thus, CAPKε expression at the vIPAG allowed rats to develop significant antinociceptive tolerance when injected with morphine doses that normally do not cause tolerance, but CAPKε would not increase the degree of tolerance once antinociceptive tolerance occurred.

Discussion

The hypothesis that activating PKCα or ε at the preBotC could lead to increased morphine tolerance in respiratory depression and lethal overdose was tested. Expression of CAPKα and ε at preBotC increased morphine tolerance to respiratory depression and lethal overdose. The level of morphine tolerance in respiratory depression was higher in CAPKα ε rats than CAPKα lentivirus-injected rodents. Past studies on PKC’s role in morphine tolerance have focused on inhibiting PKC to decrease antinociceptive tolerance. Our current study is the first report that has investigated PKC’s ability to increase morphine tolerance in respiratory depression and lethal overdose. Because respiration and overdose tolerance is increased, augmenting PKC activity at the preBotC could be one approach to improve the safety of morphine in long-term use.
The goal of this study was to test whether enhancing PKC activity at an important respiration control center could increase tolerance to opioid lethal overdose and respiratory depression. Although the stereotaxic coordinates targeted the preBoëtC, our lentivirus carried the cytomegalovirus promoter, which meant that exogenous PKC expression was not limited to the preBoëtC. Although immunohistochemistry showed that some viral transduced GFP-positive cells also stained for somatostatin and MOR, colocalization was not 100%. Caudal neighbors to preBoëtC include the rostral and caudal VRG. The rostral VRG is involved in inspiration by sending projections to the phrenic nucleus, whereas caudal VRG is implicated in expiration by projecting to expiratory motor neurons (Kirkwood and Sears, 1973; Dobbins and Feldman, 1994). Rostral to preBoëtC are BoëtC and pFRG.

Fig. 4. PKCε in tolerance to morphine respiratory depression. Rats were microinjected bilaterally at the preBoëtC with either lentivirus carrying GFP, scramble siRNA, PKCε siRNA, WTPKCε, or CAPKCε. A, PKCε lentiviral siRNA knockdown. In vitro, NS20Y cells were transduced with PKCε siRNA lentivirus. In vivo, rat preBoëtC injected with siRNA lentivirus was laser microdissected for qRT-PCR analysis. β-Actin was set as 1. *, P < 0.05 and ***, P < 0.001 compared with NS20Y PKCε siRNA (one-way ANOVA). ††, P < 0.01 and †††, P < 0.001 compared with rat preBoëtC PKCε siRNA (one-way ANOVA). B and C, respiratory depression due to 20 mg/kg morphine. **, P < 0.01 compared with day 1 CAPKCε (two-way ANOVA). D, morphine respiratory depression (Resp Dep) ED50. The ED50 values to cause respiratory depression were determined before and after long-term morphine treatment. ***, P < 0.001 compared with naive CAPKCε and †††, P < 0.001 compared with chronic WTPKCε using two-way ANOVA. Data are presented as mean ± S.E.M. (n ≥ 6).

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RTN. BöC neurons are glycinergic and project to the ventral respiratory group and phrenic nucleus. The inhibitory input from BöC suppresses inspiratory neurons during expiration (Dobbins and Feldman, 1994; Ezure et al., 2003). pFRG/RTN is involved in both expiration and CO2 chemosensation (Janczewski and Feldman, 2006). Dorsal to preBöC is the ambiguous nucleus, which controls muscles of the larynx and pharynx and contains premotor parasympathetic neurons that regulate cardiac function (Dergacheva et al., 2010). Respiratory neurons in the ventrolateral medulla are known to project extensively to each other, and preBöC is known to have reciprocating projections with VRG, BöC, and pFRG/RTN (Ellenberger and Feldman, 1990; Tan et al., 2010). Because many adjacent neurons to preBöC are involved in respiration, unintended viral transduction of preBöC neighbors would still allow us to test the ability of PKC in enhancing morphine tolerance to respiratory depression and lethal overdose.

WTPKCa or ε rodents did not develop significant morphine tolerance in respiratory depression; however, but CAPKCa or ε expressing rats did. These results can be explained by the higher PKC activity of the constitutively active mutants. It is well known that deleting or mutating the PKC pseudosubstrate inhibitory region increased PKC activity both in vitro and in vivo as judged by increased translocation to the plasma membrane (Pears et al., 1990; Takeishi et al., 2000). Although μ-opioid receptor-mediated activation of PKC has been reported in cell models (Chu et al., 2010), our current result implies that lower PKC activation by morphine at preBöC could be one of the reasons for less morphine respiration and overdose tolerance than antinociception. At the preBöC, the lack of effect from PPKCa and ε siRNA lentivirus indicated that PKC activation was minimal, and PKC might not be an intrinsic mechanism to regulate MOR (Figs. 3, B and D, and 4, B and D). Both CAPKCa and CAPKCe were able to increase respiration and overdose tolerance, and this is in agreement with both subtypes’ role in mediating receptor desensitization after morphine treatment (Bailey et al., 2009; Chu et al., 2010). Whether other members of the classic and novel PKC subtypes would elicit similar responses remains to be demonstrated. In addition, CAPKCe was able to increase respiratory depression tolerance to a higher extent than CAPKCa, which can be caused by differences in kinase activity levels or downstream phosphorylation targets. An
alternative explanation for this difference could stem from the nonspecific promoter, cytomegalovirus, used to drive the expression of the transgenes causing the expression of the constitutively active PKC in nonopioid receptor and nonosmo-matostatin expressing neurons (Fig. 2). Whether CAPKCa and CAPKCr were expressed at identical neurons after injection was assumed and not determined.

Several mechanisms have been hypothesized as causing morphine tolerance, and this article investigated PKC because of the kinase’s role in opioid receptor signaling when morphine was the agonist. Morphine is known to cause PKC activation, which in turn participates in opioid receptor desensitization and tolerance (Smith et al., 2007; Bailey et al., 2005; Chu et al., 2010). In addition to PKC, β-arrestin-mediated opioid receptor desensitization and opioid receptor recycling have been proposed as mechanisms to explain opioid antinociceptive tolerance (He and Whistler, 2005). β-Arrestin 2 knockouts are known to experience less respiratory depression than antinociception (He and Whistler, 2005). Activation of the kinase’s role in opioid receptor signaling when morphine tolerance would improve the safety of current opioids.

There is correlation between the development of tolerance to morphine respiratory depression and lethality for CAPKCa and CAPKCr. In contrast, when tolerance did not develop to respiratory depression, such as in WTPKCa, there was a corresponding lack of tolerance to morphine lethality. This correlation between respiratory depression and lethality agrees with the widely believed idea that respiratory depression is the main cause of morphine lethality (White and Irvine, 1999). The majority of morphine lethality occurred between 1.5 and 5 h after injection, which is later than the morphine peak effect at 30 min for intraperitoneal injections. This observation is consistent with the fact that morphine does not cause complete cessation of breathing even at lethal doses but a precipitous reduction instead. Although total respiratory arrest would probably cause death close to the peak effect of morphine at 30 min after injection, a longer time course is required to cause death when rats are breathing at less than adequate minute ventilation. The cumulated effect of depressed respiration such as respiratory acidosis did eventually cause lethality but with slower kinetics than complete respiratory arrest.

Although therapeutic applicability of the findings in this article is limited by current technology, our results indicate that PKCe activation could greatly narrow or equalize tolerance between morphine lethality and antinociception. Morphine lethality normally has a higher threshold to develop tolerance than antinociception (Fig. 6, A and B); however, expressing CAPKCe would lower the threshold and allow morphine lethality tolerance to develop. On the other hand, morphine antinociceptive tolerance would be unchanged as antinociceptive tolerance easily develops, and CAPKCe could not further increase the magnitude of tolerance after it has occurred (Fig. 6E). Equalizing antinociceptive and lethality tolerance would improve the safety of current opioids.

This article tested the hypothesis that selective PKC activation in preBotC could increase tolerance to morphine respiratory depression and lethal overdose. Future opioid therapeutics that take advantage of differences in PKC subtype distribution and morphine-induced PKCe activity between analgesics versus respiratory centers could maintain or improve the therapeutic index of morphine during long-term pain treatment. As annual lethal overdoses from prescription opioid analgesics have increased to more than 10,000 in the United States, developing approaches to improve the safety of opioids for long-term use is an important health care issue (Okie, 2010).

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Authorship Contributions

Participated in research design: Lin, Law, and Loh.
Conducted experiments: Lin.
Performed data analysis: Lin and Law.
Wrote or contributed to the writing of the manuscript: Lin, Law, and Loh.

References


**Address correspondence to:** Hong-You Li, Department of Pharmacology, University of Minnesota, 6-120 Jackson Hall, 321 Church St. S.E., Minneapolis, MN 55455-0217. E-mail: linxx102@umn.edu