GABA<sub>B</sub> Receptor-Positive Modulators: Brain Region-Dependent Effects

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Abstract

This study examined the positive modulatory properties of 2,6-di-tert-butyl-4-(3-hydroxy-2,2-dimethyl-propyl)-phenol (CGP7930) and (R,S)-5,7-di-tert-butyl-3-hydroxy-3-trifluoromethyl-3H-benzo[2,3]furan-2-one (rac-BHFF) at γ-aminobutyric acid B (GABA<sub>B</sub>) receptors in different brain regions. Using quantitative autoradiography, we measured GABA<sub>B</sub> receptor-stimulated binding of guanosine 5′-O-(3-[35S]thiotriphosphate) ([35S]GTP<sub>S</sub>) to G proteins in medial prefrontal cortex (mPFC), hippocampal, and cerebellum. CGP7930 and rac-BHFF enhanced baclofen-stimulated [35S]GTP<sub>S</sub> binding similarly in mPFC and hippocampal, but were more effective in cerebellum. CGP7930 (100 μM) increased [35S]GTP<sub>S</sub> binding stimulated by baclofen (30 μM) from 29 to 241% above basal in mPFC and from 13 to 1530% above basal in cerebellum. Likewise, rac-BHFF (10 μM) increased baclofen-stimulated [35S]GTP<sub>S</sub> binding more in cerebellum (from 13 to 1778% above basal) than in mPFC (from 29 to 514% above basal). rac-BHFF (10 μM) in combination with γ-hydroxybutyrate (20 mM) increased [35S]GTP<sub>S</sub> binding in cerebellum but not in mPFC. rac-BHFF also enhanced the effects of 3-aminopropyl(diethoxymethyl)phosphinic acid (CGP35348). Consistent with its partial agonist properties, CGP35348 stimulated [35S]GTP<sub>S</sub> binding in mPFC when given alone (to 18% above basal), but less extensively than baclofen (140% above basal), and antagonized baclofen when given together. CGP35348 (1 mM) in combination with rac-BHFF (100 μM) produced an increase in [35S]GTP<sub>S</sub> binding that was larger in cerebellum (from 61 to 1260% above basal) than in mPFC (from 18 to 118% above basal). Taken together, the results show that GABA<sub>B</sub> receptor-positive modulators enhance [35S]GTP<sub>S</sub> binding stimulated by GABA<sub>B</sub> receptor agonists in a brain region-dependent manner. This regionally selective enhancement is further evidence of pharmacologically distinct GABA<sub>B</sub> receptor populations, possibly allowing for more selective therapeutic targeting of the GABA<sub>B</sub> system.

Introduction

Metabotropic GABA<sub>B</sub> receptors are present throughout the central nervous system and have been implicated in various psychiatric disorders (Cryan and Kapmann, 2005; Bowery, 2006), including drug dependence (Addolorato et al., 2009; Maccioni et al., 2010; Vlachou and Markou, 2010). These receptors are heterodimers composed of GABA<sub>B1a</sub> or GABA<sub>B1b</sub> subunits in combination with GABA<sub>B2</sub> subunits (Calver et al., 2002; Bettler et al., 2004). GABA<sub>B</sub> receptors are coupled through G<sub>i/o</sub> to the inhibition of adenyly cyclase, the closing of voltage-dependent calcium channels, and the opening of inwardly rectifying K<sup>+</sup> channels (Bowery et al., 2002; Bettler et al., 2004). GABA<sub>B</sub> receptors function as autoreceptors and heteroreceptors, modulating neurotransmitter release and neuronal firing and influencing long-term changes in synaptic strength (Pinard et al., 2010).

Allosteric modulators at GABA<sub>B</sub> receptors are of great interest given the widespread distribution of GABA<sub>B</sub> receptors in the central nervous system and the critical role these receptors play in modulating neuronal excitability and synaptic plasticity. Allosteric modulators bind to regions on the allosteric site of GABA<sub>B</sub> receptors, leading to functional changes in synaptic transmission. The functional significance of these changes is becoming clearer, as GABA<sub>B</sub> receptor modulators have been shown to have therapeutic potential in various neurological and psychiatric disorders. In this context, the development of GABA<sub>B</sub> receptor modulators as potential therapeutic agents is an active area of research.

Abbreviations:
CGP35348, 3-aminopropyl(diethoxymethyl)phosphinic acid; CGP7930, 2,6-di-tert-butyl-4-(3-hydroxy-2,2-dimethyl-propyl)-phenol; rac-BHFF, (R,S)-5,7-di-tert-butyl-3-hydroxy-3-trifluoromethyl-3H-benzo[2,3]furan-2-one; GBH, γ-hydroxybutyrate; DPCPX, 1,3-dipropyl-8-cyclopentylxanthine; [35S]GTP<sub>S</sub>, guanosine 5′-O-(3-[35S]thiotriphosphate); mPFC, medial prefrontal cortex; ACC, anterior cingulate cortex; K<sub>CDT</sub>, potassium channel tetramerization domain-containing; CHO, Chinese hamster ovary; CGP13501, 3,5-bis(1,1-dimethylethyl)-4-hydroxy-α,α-dimethylbenzenepropanol.
receptor that are different from the orthosteric site where the endogenous ligand binds and act by enhancing or attenuating the response elicited by the endogenous transmitter or agonist (Jensen and Spalding, 2004; Conn et al., 2009). By altering the effects of activated receptors without affecting nonactivated receptors, allosteric modulators may have a broader therapeutic window than ligands that indiscriminately alter the activity of all receptors, thereby perhaps offering a potentially attractive alternative to conventional pharmacological agents (Jensen and Spalding, 2004; Pin and Prézeau, 2007; Conn et al., 2009).

Several compounds have been characterized as positive allosteric modulators of GABA<sub>B</sub> receptors. 2,6-Di-tert-butyl-4-(3-hydroxy-2,2-dimethyl-propyl)-phenol (CGP7930) enhances GABA<sub>B</sub> receptor-stimulated [<sup>35</sup>S]GTP<sub>S</sub> binding in rat brain membranes and membranes from GABAB<sub>1b/2</sub>-expressing clonal cells in culture and has modulatory effects in cellular assays of GABA<sub>B</sub> receptor-mediated electrophysiological responses (Urwyler et al., 2001; Adams and Lawrence, 2007). In vitro studies also indicate that (R,S)-5,7-di-tert-butyl-3-hydroxy-3-trifluoromethyl-3H-benzofuran-2-one (rac-BHFF) increases the potency and efficacy of GABA to stimulate [<sup>35</sup>S]GTP<sub>S</sub> binding in membrane preparations from recombinant cells in culture and enhances electrophysiological responses to the GABA<sub>B</sub> receptor agonist baclofen in hippocampal slice preparations (Malherbe et al., 2008). In vivo, rac-BHFF and CGP7930 increase the loss of righting in mice induced by a subthreshold dose of baclofen (Carai et al., 2004; Malherbe et al., 2008). We have recently shown that in rats the GABA<sub>B</sub> receptor-positive modulators CGP7930 and rac-BHFF enhance the loss of righting induced by baclofen or γ-hydroxybutyrate (GHB), which also activates GABA<sub>B</sub> receptors (Koek et al., 2010). The hypothermic response induced by baclofen or GHB, however, is not modulated by CGP7930 and rac-BHFF. The data from this in vivo study suggest that GABA<sub>B</sub> receptor-positive modulators CGP7930 and rac-BHFF act as positive modulators at some, but not all, GABA<sub>B</sub> receptor populations (Koek et al., 2010).

The present study is part of an effort to examine the positive modulatory properties of CGP7930 and rac-BHFF at GABA<sub>B</sub> receptors in different brain regions. Using quantitative autoradiography, we measured GABA<sub>B</sub> receptor-stimulated binding of [<sup>35</sup>S]GTP<sub>S</sub> to G proteins. This approach allowed us to determine the effects of rac-BHFF and CGP7930 on GABA<sub>B</sub> receptor function, specifically the capacity of GABA<sub>B</sub> receptors to activate G proteins, with a high degree of neuroanatomical resolution. We hypothesized that rac-BHFF and CGP7930, although not stimulating [<sup>35</sup>S]GTP<sub>S</sub> binding alone, would increase [<sup>35</sup>S]GTP<sub>S</sub> binding stimulated by the GABA<sub>B</sub> receptor agonist baclofen and by GHB, which has GABA<sub>B</sub> receptor agonist properties (e.g., Mathivet et al., 1997). Given that neurochemical and electrophysiological responses mediated by GABA<sub>B</sub> receptors have been characterized extensively in hippocampus and neocortex (Pinard et al., 2010) and cerebral cortex (Miguel et al., 1995; Vigot and Batini, 1997; Tu et al., 2010), we chose to examine these brain areas in the present study. Taken together, the results show that GABA<sub>B</sub> receptor-positive modulators enhance [<sup>35</sup>S]GTP<sub>S</sub> binding stimulated by GABA<sub>B</sub> receptor agonists in a brain region-dependent manner.

Materials and Methods

**Animals.** C57BL/6J mice (The Jackson Laboratory, Bar Harbor, ME) were bred at the University of Texas Health Science Center at San Antonio. The current study used adult male mice that were group-housed and maintained on a 14:10-h light/dark cycle with continuous access to food and water. These studies were carried out in accordance with the Guide for the Care and Use of Laboratory Animals (Institute of Laboratory Animal Resources, 1996) as adopted and promulgated by the National Institutes of Health.

**Tissue Preparation.** Mice were killed by rapid decapitation. Brains were rapidly removed, frozen on powdered dry ice, and then stored at −80°C until sectioning. Coronal sections of 20-μm thickness were cut at −17°C in a cryostat microtome at the level of the medial prefrontal cortex (mPFC; plates 11–12), anterior cingulate cortex (ACC; plates 21–22), dorsal hippocampus (plates 45–46), and cerebellum (plates 86–87) (Paxinos and Franklin, 1997). Sections were thaw-mounted onto gelatin-coated glass slides, desiccated at 4°C for 18 h under vacuum, and then stored at −80°C until they were used in the autoradiographic experiments.

**[35S]GTP<sub>S</sub> Autoradiography.** Autoradiography of agonist-stimulated [<sup>35</sup>S]GTP<sub>S</sub> binding in brain sections was performed as described previously (Hensler and Durgam, 2001; Advani et al., 2007) with slight modifications. Slide-mounted sections were equilibrated in HEPES buffer (50 mM, pH 7.4), supplemented with 3 mM MgCl<sub>2</sub>, 0.2 mM EDTA, 100 mM NaCl, and 0.2 mM dithiothreitol for 10 min at 25°C. Sections were preincubated in HEPES buffer containing GDP (2 mM) and the adenosine A<sub>1</sub> receptor antagonist 1,3-diisopropyl-8-cyclopentylxanthine (DPCPX; 1 μM) for 15 min at 25°C, and then incubated in HEPES buffer containing GDP (2 mM), DPCPX (1 μM), and 40 pM [<sup>35</sup>S]GTP<sub>S</sub>, either in the absence or the presence of agonist, for 60 min at 25°C. Basal [<sup>35</sup>S]GTP<sub>S</sub> binding was defined in the absence of agonist. Nonspecific [<sup>35</sup>S]GTP<sub>S</sub> binding was defined in the absence of agonist and the presence of 10 μM GTP·S. The incubation was stopped by two washes for 5 min each in ice-cold 50 mM HEPES buffer, pH 7.4, followed by a brief immersion in ice-cold deionized water. Sections were dried on a slide warmer and exposed to Kodak Biomax MR film (Eastman Kodak, Rochester, NY) for 48 h.

**Image Analysis.** Analysis of the digitized autoradiograms was performed with the image analysis program ImageJ, version 1.42q (National Institutes of Health, Bethesda, MD). Tissue sections were stained with thionin, and the brain areas were identified with the atlas of the mouse brain (Paxinos and Franklin, 1997). Autoradiograms of agonist-stimulated [<sup>35</sup>S]GTP<sub>S</sub> binding were quantified by the use of simultaneously exposed [14C] standards (ARC-146; American Radiochemicals, St. Louis, MO). Standard curves were fit to pixel data obtained from [14C] standards, and tissue equivalent values (nanocuries/gram) were provided by American Radiochemicals and used to transform the actual regional densitometric values into relative radioactivity measures. Nonspecific binding of [<sup>35</sup>S]GTP<sub>S</sub> was subtracted from basal binding and binding in the presence of agonist. Specific agonist-stimulated binding was expressed as the percentage above basal. Basal binding of [<sup>35</sup>S]GTP·S varied with brain region, ranging from 266 ± 0.7 nCi/g in cortical areas and 232 ± 6.02 nCi/g in hippocampus to 116 ± 8.51 nCi/g in cerebellum.

**Drugs.** [<sup>35</sup>S]GTP·S (1250 Ci/mmol) was purchased from PerkinElmer Life and Analytical Sciences (Waltham, MA). GDP (disodium salt) was purchased from Sigma (St. Louis, MO). GTP·S (tetraphosphoric acid) was purchased from Roche Diagnostics (Indianapolis, Indiana). Baclofen hydrochloride and DPCPX were purchased from Tocris Bioscience (Ellisville, Missouri). GHB was purchased by the National Institute on Drug Abuse (Bethesda, MD). CGP7930 and rac-BHFF were synthesized by K. Cheng at the National Institute on Drug Abuse, and 3-aminopropyl(diethoxymethyl)phosphonic acid (CGP35348) was synthesized by J. Agyn at the University of Texas Health Science Center (San Antonio, TX).
Data Analysis. \[^{35}\text{S}]\text{GTP}_{\gamma}\text{S} binding was expressed as percentage above basal. Drug effects on this measure were tested by one-factor analysis of variance (followed by comparisons with vehicle control by Dunnett’s test) when several doses were examined or by unpaired \(t\) tests if single doses were used. Differences from 0% basal were analyzed by one-sample \(t\) tests. \[^{35}\text{S}]\text{GTP}_{\gamma}\text{S} binding data obtained in cerebellum were compared with those obtained in mPFC by means of two-factor (brain region, drug treatment) analysis of variance, performed separately on the data obtained with baclofen, GHB, and rac-BHFF, followed by Bonferroni-corrected comparisons between regions.

Results

Before studying the effects of combining the GABA\(_B\) receptor-positive modulators CGP7930 or rac-BHFF with compounds that possess GABA\(_B\) receptor agonist properties (i.e., baclofen or GHB), each was examined alone for its ability to stimulate \[^{35}\text{S}]\text{GTP}_{\gamma}\text{S} binding. Baclofen (Fig. 1A) dose-dependently increased \[^{35}\text{S}]\text{GTP}_{\gamma}\text{S} binding in mPFC and ACC, as well as in subregions of dorsal hippocampus (CA1, CA2/3, dentate gyrus) \((F_{2,17} = 6.71; P < 0.01)\). The binding stimulated by 300 \(\mu\text{M}\) baclofen ranged from 89% above basal in CA1 region of hippocampus to 143% above basal in mPFC. The putative GABA\(_B\) receptor antagonist CGP35348 (1 mM) completely blocked the effect of 300 \(\mu\text{M}\) baclofen. The results obtained with the combination of CGP35348 and baclofen differed significantly from the results obtained with baclofen alone in all regions \((t_6 = 2.84; P = 0.03)\), except dentate \((t_6 = 2.37; p = 0.056)\), and did not differ significantly from basal \((t_6 = 2.02; P = 0.14)\). When given alone at 1 mM,
CGP35348 significantly increased [35S]GTPγS binding in the mPFC (mean ± S.E.M.: 18 ± 3.6% above basal; \( t_9 \) versus 0% above basal = 4.83; \( p = 0.017 \)), but not in the other regions (\( P \leq 0.074 \) data not shown), suggesting that CGP35348 acts as a weak partial agonist in mPFC to stimulate [35S]GTPγS binding. In contrast with baclofen, GHB did not significantly alter [35S]GTPγS binding from basal in any of the regions examined (\( F_{2,11} = 1.39; p \geq 0.29 \)) (Fig. 1B). At the highest concentration tested alone (i.e., 20 mM), GHB significantly decreased the effects of baclofen, but did so only in the mPFC (\( t_6 = 2.50; p = 0.047 \); other regions, \( p \geq 0.09 \) and did so only partially (\( t_3 \) versus 0% above basal = 6.98; \( p = 0.006 \)) (Fig. 1A). Although GHB produces many of its effects by activating GABA\(_B\) receptors (Carter et al., 2009), it was ineffective in stimulating [35S]GTPγS binding.

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The GABA\(_B\) receptor-positive modulators CGP7930 and rac-BHFF alone significantly stimulated [35S]GTPγS binding in the cortical areas and hippocampal subregions examined, with rac-BHFF being more potent and more efficacious than CGP7930. CGP7930 alone seemed to increase [35S]GTPγS binding in several regions (Fig. 1C), but only in ACC did this effect reach statistical significance (\( F_{3,14} = 4.74; p = 0.018 \); other regions: \( p \geq 0.16 \)). In ACC, [35S]GTPγS binding significantly differed from basal at 30 and 100 \( \mu \)M CGP7930 (\( t_3 = 3.34; p = 0.045 \) and \( t_5 = 6.82, p = 0.001 \)), but not at the other concentrations tested (\( p \geq 0.08 \)), and attained a maximum of 33%. None of the results obtained when CGP7930 was combined with the putative GABA\(_B\) receptor antagonist CGP35348 differed significantly from those obtained with CGP7930 alone (\( p \geq 0.12 \)). rac-BHFF dose-dependently increased [35S]GTPγS binding in all regions examined (Fig. 1D) (ACC: \( F_{2,10} = 5.51, p = 0.024 \); other regions: \( F_{2,11} \geq 9.83, p \leq 0.0034 \)). The binding stimulated by 100 \( \mu \)M rac-BHFF ranged from 66% above basal in CA2/3 to 101% above basal in dentate gyrus. Thus, rac-BHFF stimulated [35S]GTPγS binding more potently and to a larger extent than CGP7930. It is noteworthy that [35S]GTPγS binding stimulated by the combination of CGP35348 (1 \( \mu \)M) and rac-BHFF (100 \( \mu \)M) was significantly greater than that obtained with rac-BHFF alone in all forebrain regions examined (\( p \leq 0.033 \)) except ACC (\( p = 0.22 \)) (Fig. 1D). These data are consistent with the modulation of partial agonist activity of CGP35348 by rac-BHFF in mPFC and dorsal hippocampus.

Both positive modulators markedly enhanced the effect of baclofen (30 \( \mu \)M) to increase [35S]GTPγS binding, at concentrations lower than those that stimulated [35S]GTPγS binding when applied alone (Fig. 2). CGP7930 dose-dependently enhanced the effects of baclofen (\( F_{4,25} = 3.04; p = 0.036 \)) (Fig. 2A), with 10 \( \mu \)M producing significant enhancement in three regions (i.e., mPFC, ACC, and hippocampal CA2/3) and 100 \( \mu \)M producing significant enhancement in these regions as well as in dentate gyrus. At 100 \( \mu \)M, the effects of CGP7930 combined with baclofen were significantly greater than those of CGP7930 alone in all regions examined (mPFC: \( t_9 = 3.43, p = 0.0076 \); other regions: \( t_{10} = 2.91, p = 0.016 \)) (Fig. 2A). CGP7930 enhanced baclofen-stimulated [35S]GTPγS binding at minimum significant concentrations of 10 \( \mu \)M in mPFC, ACC, and CA2/3 and 100 \( \mu \)M in CA1 and dentate. Binding stimulated by baclofen (30 \( \mu \)M) in the presence of CGP7930 (100 \( \mu \)M) ranged from 13% above basal in CA2/3 to 241% above basal in mPFC. rac-BHFF dose-dependently enhanced the effects of baclofen (\( F_{4,17} = 5.83; p = 0.0038 \)) (Fig. 2B), with 1 and 10 \( \mu \)M significantly enhancing baclofen-stimulated [35S]GTPγS binding in all regions. As illustrated by the autoradiograms in Fig. 3, rac-BHFF at a concentration (1 \( \mu \)M) that did not stimulate [35S]GTPγS binding above basal markedly enhanced the effect of 30 \( \mu \)M baclofen. At concentrations of 1 and 10 \( \mu \)M, the effects of rac-BHFF combined with baclofen were significantly greater than those of rac-BHFF alone in all regions examined (mPFC: \( t_9 = 3.43, p = 0.0076 \); other regions: \( t_{10} = 2.91, p = 0.016 \)). Binding stimulated by 30 \( \mu \)M baclofen in the presence of 10 \( \mu \)M rac-BHFF ranged from 464% above basal in CA2/3 to 628% above basal in dentate. CGP7930 and rac-BHFF enhanced baclofen-stimulated [35S]GTPγS binding in all regions examined (mPFC: \( t_9 = 3.43, p = 0.0076 \); other regions: \( t_{10} = 2.91, p = 0.016 \)). Binding stimulated by 30 \( \mu \)M baclofen in the presence of 10 \( \mu \)M rac-BHFF ranged from 464% above basal in CA2/3 to 628% above basal in dentate. CGP7930 and rac-BHFF enhanced baclofen-stimulated [35S]GTPγS binding in all regions examined (mPFC: \( t_9 = 3.43, p = 0.0076 \); other regions: \( t_{10} = 2.91, p = 0.016 \)). Binding stimulated by 30 \( \mu \)M baclofen in the presence of 10 \( \mu \)M rac-BHFF ranged from 464% above basal in CA2/3 to 628% above basal in dentate. CGP7930 and rac-BHFF enhanced baclofen-stimulated [35S]GTPγS binding in all regions examined (mPFC: \( t_9 = 3.43, p = 0.0076 \); other regions: \( t_{10} = 2.91, p = 0.016 \)). Binding stimulated by 30 \( \mu \)M baclofen in the presence of 10 \( \mu \)M rac-BHFF ranged from 464% above basal in CA2/3 to 628% above basal in dentate.
GABA<sub>b</sub> Receptor-Positive Modulators: Regional Differences

In contrast to what was observed with baclofen, the modulators did not seem to act in combination with GHB to increase [35S]GTP<sup>γ</sup>S binding in cortical or hippocampal regions. CGP7930 at 100 μM, a concentration that markedly enhanced baclofen-stimulated [35S]GTP<sup>γ</sup>S binding, did not significantly increase the binding of [35S]GTP<sup>γ</sup>S above basal when combined with GHB (mPFC: \( t_\alpha = 0.89, p = 0.40 \); other regions: \( t_\alpha \leq 1.19, p \geq 0.27 \)) (Fig. 2A). rac-BHFF in combination with GHB dose-dependently increased [35S]GTP<sup>γ</sup>S binding (all regions except CA2/3; \( F_{2,11} \geq 4.63, p \leq 0.035 \); CA2/3: \( p = 0.073 \)), with 10 μM having significant effects versus GHB alone in all regions (Fig. 2B). However, only in the CA2/3 region of hippocampus did [35S]GTP<sup>γ</sup>S binding stimulated by 10 μM rac-BHFF in combination with GHB differ significantly from [35S]GTP<sup>γ</sup>S binding stimulated by rac-BHFF alone (\( t_\alpha = 2.51, p = 0.047 \); other regions: \( p \geq 0.10 \)).

Although baclofen seemed to stimulate [35S]GTP<sup>γ</sup>S binding to a similar extent in cortical and hippocampal regions (Figs. 1 and 2), it had a markedly greater effect in cerebellum (molecular layer) (Fig. 4A). Baclofen (100 μM) increased [35S]GTP<sup>γ</sup>S binding in cerebellum to 390% above basal, which is significantly larger than its effect in, for example, the mPFC (i.e., 84% above basal). At a lower concentration (i.e., 30 μM), baclofen stimulated GTP<sup>γ</sup>S binding only modestly and did so similarly in the mPFC and the cerebellum (29 and 13% above basal, respectively; \( p > 0.05 \)). Thus, baclofen seemed to increase GTP<sup>γ</sup>S binding in both brain regions with different efficacy. Although the effects of the modulators alone did not differ significantly between cerebellum and mPFC (\( p > 0.05 \)), large regional differences became apparent when the modulators were given together with baclofen. CGP7930 (100 μM) increased [35S]GTP<sup>γ</sup>S binding stimulated by baclofen (30 μM) significantly more in cerebellum (from 13 to 1530% above basal) than in mPFC (from 29 to 241% above basal) (\( p < 0.0001 \)). Likewise, rac-BHFF (10 μM) increased baclofen-stimulated [35S]GTP<sup>γ</sup>S binding significantly more in cerebellum (from 13 to 1778% above basal) than in mPFC (from 29 to 514% above basal) (\( p < 0.0001 \)). Together, these results indicate that the modulators were

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**Fig. 3.** rac-BHFF markedly enhances the effect of baclofen to stimulate the binding of [35S]GTP<sup>γ</sup>S. Representative autoradiograms of [35S]GTP<sup>γ</sup>S binding to coronal sections of brain taken through the dorsal hippocampus are shown. A, nonspecific binding was defined in the presence of 10 μM GTPγS. B, basal binding was determined in the absence of the GABA<sub>b</sub> receptor agonist baclofen or modulator rac-BHFF. C to E, the binding of [35S]GTP<sup>γ</sup>S in the presence of rac-BHFF (1 μM) (C), baclofen (30 μM) (D), or rac-BHFF (1 μM) + baclofen (30 μM) (E). Data from these experiments are plotted in Fig. 2.

**Fig. 4.** Comparison of the effects of the GABA<sub>b</sub> receptor-positive modulator CGP7930 and rac-BHFF together with baclofen (A) or GHB (B) on [35S]GTP<sup>γ</sup>S binding in cerebellum (molecular layer) and mPFC. Numbers in parentheses are concentrations in microliters, except the concentration of GHB, which is in milliliters. Main effects of brain region and treatment, and brain region × treatment interactions, were statistically significant for the results shown (\( p < 0.0001 \)). * indicates significantly different from the corresponding results obtained in mPFC. Results obtained in mPFC are replotted from Figs. 1 and 2. Data are means ± S.E.M. (n = 4–6). Note that the scale of the abscissa in A is interrupted and differs from the scales used in B and Figs. 1 and 2.
more effective in cerebellum than in mPFC to enhance the effects of baclofen (see also Fig. 5).

As was observed in cortical and hippocampal regions, GHB (20 nM) alone did not significantly affect \[^{35}\text{S}\]GTP\,\gamma\,S binding in the cerebellum (Fig. 4B). And as was observed in these forebrain regions, CGP7930 (100 \,\mu\,M) in combination with GHB did not increase \[^{35}\text{S}\]GTP\,\gamma\,S binding in the cerebellum (Fig. 4B). However, we did observe marked regional differences in the positive modulatory effect of rac-BHFF in combination with GHB comparing cerebellum and forebrain areas. rac-BHFF (10 \,\mu\,M) combined with GHB produced a marked increase in \[^{35}\text{S}\]GTP\,\gamma\,S binding, which was significantly larger in cerebellum (from 13 to 426\% above basal) than, for example, in mPFC (from 7.1 to 91\% above basal) \((p < 0.0001)\) (Fig. 4B). It is noteworthy that this concentration of rac-BHFF by itself did not have significantly different effects in cerebellum than in mPFC.

It is noteworthy that we also observed marked regional differences in the effects of CGP35348 in combination with rac-BHFF on \[^{35}\text{S}\]GTP\,\gamma\,S binding in cerebellum versus mPFC (data not shown). CGP35348 (1 \,mM) significantly increased \[^{35}\text{S}\]GTP\,\gamma\,S binding to 18 \pm 3.6\% above basal in mPFC \((p = 0.017)\) and nonsignificantly \((p = 0.057)\) to 61 \pm 20\% above basal in cerebellum; the difference between the effects of CGP35348 in the two regions was not significant \((p > 0.20)\). Although rac-BHFF (100 \,\mu\,M) stimulated \[^{35}\text{S}\]GTP\,\gamma\,S binding more in cerebellum \((247 \pm 40\% above basal)\) than in mPFC \((77 \pm 9.7\% above basal)\), this difference was not statistically significant \((p = 0.15)\). CGP35348 (1 \,mM) in combination with rac-BHFF (100 \,\mu\,M) produced a marked increase in \[^{35}\text{S}\]GTP\,\gamma\,S binding, which was significantly larger in cerebellum (from 61 to 1260\% above basal) than in mPFC (from 18 to 118\% above basal) \((p < 0.0001)\). Our data indicate that the putative \text{GABA}_B receptor antagonist CGP35348 exhibits agonist activity to stimulate \[^{35}\text{S}\]GTP\,\gamma\,S binding above basal values, and these agonist effects are markedly enhanced by rac-BHFF, particularly in cerebellum.

**Discussion**

The main finding of the present study is that the responses to \text{GABA}_B receptor-positive modulators, as well as to compounds with agonist activity at \text{GABA}_B receptors, differed markedly among brain regions. Baclofen stimulated \[^{35}\text{S}\]GTP\,\gamma\,S binding to a greater extent in cerebellum than in cortical and hippocampal regions. CGP7930 and rac-BHFF were markedly more effective in enhancing the effects of baclofen in cerebellum than in other regions. Previously, we found CGP7930 and rac-BHFF enhance baclofen-induced loss of righting, but not hypothermia \((\text{Koek et al., 2010})\). Together, these results suggest that different \text{GABA}_B receptor populations may differ in their susceptibility to \text{GABA}_B receptor agonist- and -positive modulatory effects, possibly allowing for more selective therapeutic interventions of the \text{GABA}_B system.

Baclofen increased \[^{35}\text{S}\]GTP\,\gamma\,S binding in cortical areas and subregions of dorsal hippocampus, an effect blocked by CGP35348, a putative antagonist at \text{GABA}_B receptors. CGP35348 has been shown to antagonize the effect of baclofen to induce hypothermia and loss of righting in mice \((\text{Koek et al., 2010})\). In general, baclofen was more efficacious in stimulating \[^{35}\text{S}\]GTP\,\gamma\,S binding in cerebellum than in forebrain regions. Large regional differences were also apparent when the modulators CGP7930 or rac-BHFF were given together with baclofen, i.e., the modulators were markedly more effective in cerebellum than in mPFC in enhancing the effects of baclofen. In vivo, the positive \text{GABA}_B receptor modulators CGP7930 and rac-BHFF enhance baclofen-induced loss of righting, but not hypothermia in mice \((\text{Koek et al., 2010})\). Together, these studies provide further evidence of pharmacologically distinct \text{GABA}_B receptor populations.

In the present study CGP35348 seemed to have partial agonist activity in medial prefrontal cortex and cerebellum, producing a modest, but significant, increase in \[^{35}\text{S}\]GTP\,\gamma\,S binding above basal values. This effect of CGP35348 to stimulate \[^{35}\text{S}\]GTP\,\gamma\,S binding in these brain regions was markedly enhanced by the modulator rac-BHFF and to a greater extent in cerebellum than in mPFC. CGP35348 has partial agonist activity to inhibit adenylyl cyclase activity in intact recombinant CHO cells expressing \text{GABA}_B receptors. The potency and efficacy of CGP35348 in this assay system is increased by the positive modulators CGP7930 and GS39783 \((\text{Urwiler et al., 2005})\). Although CGP35348 does not stimulate \[^{35}\text{S}\]GTP\,\gamma\,S binding in membrane preparations from these cells, CGP35348 is a low-efficacy partial agonist in the
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presence of the modulators (Urwyler et al., 2005). Thus, allosteric modulators seem to be useful in revealing the intrinsic properties of agonist ligands at the orthosteric site of GABA<sub>B</sub> receptors.

Positive allosteric modulators act synergistically with agonists to elicit an enhanced response, but have little or no intrinsic agonistic activity of their own (Jensen and Spalding, 2004; Pin and Prézeau, 2007; Conn et al., 2009). In the present study, both CGP7930 and rac-BHFF markedly enhanced the effect of baclofen to increase [<sup>35</sup>S]GTP<sub>S</sub> binding at concentrations lower than those that stimulated [<sup>35</sup>S]GTP<sub>S</sub> binding when applied alone. Likewise, CGP7930 and its analog 3,5-bis(1,1-dimethylbutyl)-4-hydroxy-α,α-dimethylnaphthalene (CGBP13501) (Urwyler et al., 2001) or rac-BHFF (Malherbe et al., 2008) enhance the biochemical and electrophysiological effects of GABA at concentrations that produce little or no GABA<sub>B</sub> receptor activation when applied alone. At higher concentrations, however, CGP7930 and rac-BHFF alone significantly stimulated [<sup>35</sup>S]GTP<sub>S</sub> binding in forebrain regions, with rac-BHFF being more potent and more efficacious than CGP7930. It is noteworthy that the effects of the modulators by themselves did not differ significantly between cerebellum and mPFC. In recombinant CHO cells expressing GABA<sub>B</sub> receptors, the positive modulators CGP7930 and GS39783 alone produce a small inhibition of adenylyl cyclase (Urwyler et al., 2005). In cerebellar granule neurons in culture, CGP7930 has similar effects as baclofen to stimulate ERK1/2/CREB signaling (Tu et al., 2007) and biochemical pathways mediating neuroprotection (Tu et al., 2010). Rac-BHFF has similar effects as GABA in stimulating [<sup>35</sup>S]GTP<sub>S</sub> binding in CHO cells that stably over-express human G<sub>B1a</sub> and human GABA<sub>B1a</sub> subunits (Malherbe et al., 2008). Thus under particular conditions, these GABA<sub>B</sub> receptor-positive modulators have intrinsic agonist activity in the absence of GABA. Our data, collected in brain sections that probably lack GABA due to equilibration during processing or recombinant systems and therefore may have more general relevance. Conceivably, intrinsic agonist activity of these GABA<sub>B</sub> receptor-positive modulators could be involved in their in vivo effects, such as anxiolytic- and antidepressant-like effects (Cryan and Kaupmann, 2005) and inhibition of the reinforcing effects of drugs of abuse (Vlachou and Markou, 2010).

In the present study, GHB was ineffective in stimulating [<sup>35</sup>S]GTP<sub>S</sub> binding in the cerebellum and forebrain regions. GHB binds with weak affinity to the GABA<sub>B</sub> receptor (Mathivet et al., 1997; Lingenhoehl et al., 1999; Wu et al., 2004). Many of the behavioral and physiological effects of GHB seem to reflect weak agonist activity at GABA<sub>B</sub> receptors, in that they are prevented or attenuated by GABA<sub>B</sub> receptor antagonists (Carai et al., 2001; Kaufmann et al., 2003; Quevauviller et al., 2003; Carter et al., 2005; Koek et al., 2007, 2010). GHB-induced hypothermia and sedation are absent in GABA<sub>B</sub> receptor knockout mice (Quevauviller et al., 2003). Although GHB had no effect on the binding of [<sup>35</sup>S]GTP<sub>S</sub> in any area examined in the current study, the combination of GHB and rac-BHFF resulted in increased [<sup>35</sup>S]GTP<sub>S</sub> binding but only in the cerebellum. It is unlikely that these effects involve GHB binding sites distinct from GABA<sub>B</sub> receptors, because these sites, although prevalent in hippocampus and cortex, seem to be absent in cerebellum (Snead, 1996; Gould et al., 2003; Wu et al., 2004). Consistent with the involvement of GABA<sub>B</sub> receptors in the effects of the combination of GHB and rac-BHFF reported here, rac-BHFF enhances GHB-induced loss of righting (Koek et al., 2010), an effect that probably involves cerebellar GABA<sub>B</sub> receptors.

The marked differences between forebrain and cerebellum in the effect of compounds with GABA<sub>B</sub> receptor agonist properties to stimulate [<sup>35</sup>S]GTP<sub>S</sub> binding, or the effects of positive modulators CGP7930 or rac-BHFF, can not be readily explained by a corresponding difference in GABA<sub>B</sub> receptor density. The density of GABA<sub>B</sub> receptors in the cerebellum (molecular layer) is comparable with that in frontal cortex and anterior cingulate cortex, with somewhat lower densities in the hippocampus (Chu et al., 1990). Instead of differences in receptor density, a family of auxiliary proteins that form high-molecular-mass complexes with GABA<sub>B</sub> receptors could conceivably be involved in the brain region-dependent effects reported here. These KCTD (potassium channel tetramerization domain-containing) proteins increase agonist potency and markedly alter the G protein signaling of GABA<sub>B</sub> receptors (Schwenk et al., 2010). The distinct regional expression profiles of KCTD proteins in the brain (Schwenk et al., 2010; Metz et al., 2011) raise the interesting possibility that these proteins in complex with GABA<sub>B</sub> receptors determine not only agonist potency and G protein signaling, but also the effects of positive allosteric modulators.

Positive allosteric modulators act synergistically with agonist or the endogenous neurotransmitter and hold several advantages to the actions of conventional agonists. For example, the modulator will only amplify the neural signal when the neurotransmitter is released into the synapse. The receptor may be less likely to desensitize upon sustained exposure to a positive allosteric modulator compared with an agonist. Because the orthosteric binding sites for a particular endogenous ligand are often highly conserved, it is difficult to achieve high selectivity for conventional ligands targeting this site among specific G protein-coupled receptor subtypes (Jensen and Spalding, 2004; Pin and Prézeau, 2007; Conn et al., 2009). Thus, allosteric modulators offer a potentially attractive alternative to conventional pharmacological agents. Our data suggest that different GABA<sub>B</sub> receptor populations differ in their sensitivity to the effects of positive GABA<sub>B</sub> receptor modulators, which may be related to regional differences in the expression of auxiliary GABA<sub>B</sub> receptor (e.g., KCTD) proteins. That different GABA<sub>B</sub> receptor populations seem to differ in their susceptibility to positive modulatory effects may allow for more selective therapeutic targeting of the GABA<sub>B</sub> receptor system.

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Participated in research design: Hensler, Advani, and Koek.
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