Silencing Mitogen-Activated Protein Kinase-Activated Protein Kinase-2 Arrests Inflammatory Bone Loss

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Received July 1, 2010; accepted November 15, 2010

ABSTRACT

p38 mitogen-activated protein kinases (MAPKs) are critical for innate immune signaling and subsequent cytokine expression in periodontal inflammation and bone destruction. In fact, previous studies show that systemic p38 MAPK inhibitors block periodontal disease progression. However, development of p38 MAPK inhibitors with favorable toxicological profiles is difficult. Here, we report our findings regarding the contribution of the downstream p38 MAPK substrate, mitogen-activated protein kinase-activated protein kinase 2 (MK2 or MAPKAPK-2), in immune response modulation in an experimental model of pathogen-derived lipopolysaccharide (LPS)-induced periodontal bone loss. To determine whether small interfering RNA (siRNA) technology has intraoral applications, we initially validated MK2 siRNA specificity. Then, gingival tissue surrounding maxillary molars of rats was injected with MK2 siRNA or scrambled siRNA at the palatal regions of bone loss. Intraoral tissues treated with MK2 siRNA had significantly less MK2 mRNA expression compared with scrambled siRNA-treated tissues. MK2 siRNA delivery arrested LPS-induced inflammatory bone loss, decreased inflammatory infiltrate, and decreased osteoclastogenesis. This proof-of-concept study suggests a novel target using an intraoral RNA interference strategy to control periodontal inflammation.

Introduction

Periodontal diseases are chronic bacterial infections manifesting as soft tissue inflammation and alveolar bone loss, which eventually lead to tooth loss. Innate and acquired immune responses both are necessary to clear bacterial pathogens and generate the inflammatory cascade that contributes to osteoclastogenic bone loss, a hallmark of periodontal disease. Lipopolysaccharide (LPS) from Gram-negative periodontal pathogens are recognized by CD14 and Toll-like receptors, triggering intracellular signaling cascades, including the nuclear factor-κB (NF-κB) and mitogen-activated protein kinase (MAPK) pathways (Lee and Young, 1996; Rao, 2001). p38 MAPK, one of three distinct classes of MAPKs, is a nexus for signal transduction, playing a vital role in numerous inflammatory-driven pathological processes including periodontitis. p38 MAPK signaling activation directly or indirectly mediates inflammatory cytokine expression such as interleukin (IL)-1β, IL-6, and tumor necrosis factor-α (TNF-α). These cytokines synergistically stimulate the production of other inflammatory cytokines, matrix metalloproteinases, and prostanooids (Ridley et al., 1997; Ajizian et al., 1999; Dean et al., 1999; Underwood et al., 2000; Mhalavie et al., 2006). Within the periodontal microenvironment, various cell types require p38 MAPK signaling as an integral component in the regulation of expression of proinflammatory cytokines and enzymes induced by inflammatory and...
infectious signals in vitro, including IL-6, matrix metalloproteinase-13, and receptor activator of NF-κB ligand (Patil et al., 2004, 2006; Rossa et al., 2005, 2007). In vivo data suggest that p38 signaling is required for LPS-induced alveolar bone loss because small-molecule p38 inhibitors were effective in reducing periodontitis in rodent models (Kirkwood et al., 2007; Rogers et al., 2007a). p38 MAPK inhibitors have been shown to be efficacious in other small animal inflammatory disease models, but the development of small-molecule inhibitor therapeutics has been hampered by various unwanted side effects, such as dermatoxicity and neurotoxicity, in clinical trials.

MK2 is a direct substrate of p38 MAPK (Stokoe et al., 1992), and studies have suggested a central role of MK2 in the production of proinflammatory mediators (Kotlyarov et al., 1999). One important mechanism by which MK2 increases expression of proinflammatory mediators is via targeting AU-rich elements (ARE) located in the 3′ untranslated region of the mRNA via phosphorylation of RNA stability-regulating proteins such as tristetraprolin (TTP) (Carballo et al., 1998; Chrestensen et al., 2004; Hitti et al., 2006). In vivo data suggest that overexpression of TTP decreased endogenous ARE cytokine levels and was protective against inflammation-induced bone loss via modulation of RNA stability (Patil et al., 2008). Although targeting MK2 with small-molecular inhibitors is complex because of the relatively planar ATP binding site of this critical MAPK, targeting downstream signaling molecules such as MK2 represents a focused approach for regulating post-transcriptional expression of inflammatory mediators. Such an approach could reduce some of the deleterious effects of targeting key signaling intermediates such as p38 MAPK, thereby potentially decreasing side effects and increasing clinical efficacy.

RNAi can control gene expression, and microRNA and siRNA are central components of this technique, which has evolved from a target validation tool to a testing strategy for novel RNAi-based therapeutics. Still, at this time, no studies offer direct evidence that RNAi can be used in the oral cavity, and no studies address the contribution of MK2 signaling in periodontal disease progression. RNAi silencing could provide an innovative anti-inflammatory drug platform to selectively block signaling mechanisms needed for enhanced cytokine mRNA stability/translation in periodontal disease progression. Our in vitro and in vivo data suggest that siRNA targeting of MK2 is a novel therapeutic platform for control of periodontal inflammation and bone loss.

**Materials and Methods**

**Cell Culture and Reagents.** For in vitro studies, the rat macrophage cell line NR8383 was obtained from the American Type Cell Collection (Manassas, VA). Cells were grown in Ham’s F12K medium (American Type Cell Collection) supplemented with 15% heat-inactivated fetal bovine serum (SAFC Biosciences, Lenexa, KS), penicillin (50 IU/ml), and streptomycin (50 μg/ml; MP Biomedicals, Solon, OH) in a 37°C incubator with 5% CO2. For ex vivo studies, rat bone marrow stromal cells were prepared by culturing bone marrow from the femurs/tibiae of donor rats (175–199 g) in α-modified Eagle’s medium (Invitrogen, Carlsbad, CA) containing 10% heat-inactivated fetal bovine serum, 10 ng/ml macrophage– colony-stimulating factor (R&D Systems, Minnepolis, MN), 30% L929 conditioned medium, penicillin (50 IU/ml), and streptomycin (50 μg/ml) in a 37°C incubator with 5% CO2. The cells were cultured for 7 days before use. LPS was extracted from *Aggregatibacter actinomycetemcomitans* strain Y4 (serotype B) by the hot phenol-water method and purified as described previously (Rogers et al., 2007b).

**Small Interfering RNA Delivery.** Accell MK2 siRNA (EQ-092297-00) and scrambled siRNA (D-001910-10) were purchased from Thermo Fisher Scientific/Tharmarca (LaFayette, CO). For in vitro and ex vivo studies, cells were transfected siRNA using the Accell siRNA delivery protocol and incubated for 72 h. For the in vivo study, covalently modified Accell siRNAs were locally injected.

**Western Blot.** Cell lysates were prepared in ice-cold RIPA buffer (Cell Signaling Technology, Danvers, MA) supplemented with protease inhibitor cocktails (Roche Diagnostics, Indianapolis, IN). After adjustment with NuPAGE LDS Sample Buffer (Invitrogen), cell lysates were heated to 95°C for 10 min. Equivalent amounts of proteins were separated by 10% Tris-HCl acrylamide gels and transferred to 0.2 μM nitrocellulose membranes (Bio-Rad Laboratories, Hercules, CA). Membranes were blocked with 5% (w/v) nonfat dry milk and 0.1% (v/v) Tween 20 in PBS, for 1 to 2 h at room temperature. The primary antibodies for total MK2, phospho-MK2 (Thr334), phospho-p38 MAPK (Thr180/Tyr182), phospho-stress-activated protein kinase/JNK (Thr183/Tyr185), phospho-p44/42 MAPK (Erk1/2, Thr202/Tyr204), and β-actin were purchased from Cell Signaling Technology. Primary antibodies were detected with horseradish peroxidase-conjugated secondary antibodies (Cell Signaling Technology) followed by chemiluminescence reaction with SuperSignal West Pico Chemiluminescent Substrate (Thermo Fisher Scientific). Radiographic images were obtained on a GelDoc XR system, and densitometric analysis was performed using Quantity One Software (Bio-Rad Laboratories).

**ELISA.** Cytokine expression in cell culture supernatant was measured using ELISA kits (R&D Systems) according to the manufacturer’s instructions. The protein concentration in cell lysates was measured with a DC Protein Assay Kit (Bio-Rad Laboratories). The concentration of cytokines was normalized by protein concentrations in cell lysates.

**RNA Isolation and Quantitative Reverse Transcription-Polymerase Chain Reaction.** Total RNA was isolated from treated and control cells at designated time points using TRIzol (Invitrogen) according to the manufacturer’s protocol and quantified on a NanoDrop 1000 spectrophotometer (Thermo Fisher Scientific). Using 300 to 500 ng of total RNA in a 20-μl reaction, cDNA was prepared using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems) in a C1000 Thermo Cycler (Bio-Rad Laboratories) according to the manufacturer’s suggested procedures. Quantitative real-time PCR was carried out using TaqMan Universal PCR Master Mix and TaqMan predesigned primers (Applied Biosystems) on a StepOnePlus Real-Time PCR System (Applied Biosystems) under the following conditions: 50°C for 2 min, 95°C for 10 min, and 40 cycles of 95°C for 15 s, 60°C for 1 min. The amplicon primers were obtained from Applied Biosystems for rat Mapkak2 (Rn01648144_g1), IL-6 (Rn01403530_m1), NF-κB (Rn09999017_m1), Ptg2 (Rn01483828_m1), IL-1β (Rn00580432_m1), CXCL1 (Rn00578225_m1), and β-actin (4352931E). Relative quantification was calculated by using the ΔΔCT method, and β-actin as the endogenous control was used to normalize target gene expression between wells. Expression levels were normalized to nontargeting control (β-actin) and reported as a percentage of the nontargeting control expression level.

**Animals.** The Institutional Animal Care and Use Committee at the Medical University of South Carolina approved all animal protocols. Adult male Sprague-Dawley rats (Harlan, Indianapolis, IN) weighing 250 to 274 g were used for in vivo study. Rats were maintained under specific pathogen-free conditions in pairs with food and tap water ad libitum. Once weekly, animals were routinely weighed to ensure proper growth and nutrition.

**Experimental Periodontitis Model and Administration of Accell siRNA In Vivo.** Inflammatory bone loss was initiated according to the aggressive periodontitis model described previously (Rogers et al., 2007b). The rats received one of the following treatments: 2 μl of a 10 mg/ml solution of *A. actinomycetemcomitans* LPS.
three times per week plus mock injection PBS twice per week for 4 weeks (n = 12), LPS three times per week plus Accell MK2 siRNA or Accell Scrambled siRNA (4 nmol/injection) twice per week for 4 weeks (n = 12), and control groups given PBS vehicle five times per week for 4 weeks (n = 12). All direct injections were delivered to the palatal gingiva between the maxillary first and second molars via a 33-gauge Hamilton syringe. At the end of the experimental period, all animals were sacrificed by carbon dioxide asphyxiation. Maxillae were hemisectioned (n = 12/group), and posterior block sections were immersed directly in 10% buffered formalin fixative solution for 24 h and then transferred to 70% ethanol and kept at 4°C.

**Microcomputed Tomography.** As described previously, nonmineralized rat maxillae (n = 12/group) were scanned in 70% ethanol by a cone beam µCT system (GE Healthcare, Chalfont St. Giles, Buckinghamshire, UK), and the images were analyzed using GE Healthcare MicroView software (version viz + 2.0 build 0029). In brief, each scan was reconstructed at a mesh size of 18 × 18 × 18 μm to create a three-dimensional digitized image for each specimen. Then the images were rotated into a standard orientation and threshold at 1621 was used to distinguish between mineralized and nonmineralized tissue. The ROI for the bone volume fraction (BVF) and bone mineral density (BMD) analysis was based on several easily identifiable anatomical landmarks. The BVF/BMDs were then calculated using the GE MicroView software’s innate bone analysis module (Kirkwood et al., 2007; Patil et al., 2008; Sartori et al., 2009). All µCT scans were measured and assessed by an independent masked examiner in a blinded manner.

**Laser Capture Microdissection.** To measure expression of the siRNA target gene and inflammatory cytokine genes at the end of the experimental period in vivo samples, quantitative real-time PCR was performed combined with LCM. Gingival tissue from sites of palatal injection were removed and immediately embedded in Tissue-Tek OCT compound (Sakura Finetek USA, Torrance, CA), snap-frozen in liquid nitrogen, and kept at −80°C. Serial 8-μm-thickness frozen sections were cut on a Digital Microtome Cryostat (Microm,vana, Tampere, Finland) and mounted on superfrost-plus microscope slides. After the sections were stained with the HistoGene LCM Frozen Section Staining Kit (Molecular Devices, Sunnyvale, CA), LCM using the ArcturusXT Microdissection System (Arcturus; Molecular Devices) was used to separate and collect connective tissue in which siRNA and LPS injections were performed. The tissue was transferred into a 0.5-mL tube and stored at −80°C until it was used for RNA extraction. Total RNA was extracted and purified from the microdissected tissue with a RNAqueous-Micro Kit (Applied Biosystems) according to the manufacturer’s instructions. The quantity and quality of RNA was assessed using an RNA 6000 Pico kit on an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA). qRT-PCR was performed to measure target mRNA as described above.

**Conventional Histology and Tartrate-Resistant Acid Phosphatase Staining.** Formalin-fixed, decalciﬁed maxillae were parafﬁn-embedded, and serial sagittal sections (7 μm) were prepared. Some slides were stained routinely with hematoxylin and eosin for descriptive histology. Histological images were acquired using an Olympus BX61 microscope (Olympus Research, Center Valley, PA) outfitted with a DP71 digital camera. The degree of inﬂammation was quantiﬁed by counting the number of nuclei as a measure of cellular inﬁltrate using the VIS image analysis module (Visopharm Inc., Hoersholm, Denmark). Pixel information that corresponded to “nuclei” was used to create a universal category that was then applied to all images, with the resulting objects that ﬁt this classiﬁer under a Bayesian analysis for counted nuclei. Approximately the same maxillary sections were used for histological analysis. Serial sections slides were submitted for the TRAP staining method as described in the protocol from BD Biosciences (Technical Bulletin 445) to determine the number of osteoclasts. The multinucleated TRAP-positive cells that contained three or more nuclei and made contact with the bone surface were deﬁned as osteoclasts. The number of osteoclasts within the eroded area was measured, and data were expressed as the total number of osteoclasts per section.

**Statistical Analysis.** Between-group analyses were conducted using Student’s t test for two-group comparison and analysis of variance for multigroup comparison, applying Prism (version 4; GraphPad Software Inc., San Diego, CA). P values of 0.05 or less were considered significant.

**Results**

**MK2 siRNA Target Validation Is Selective for MK2 after LPS Challenge.** To validate knockdown of MK2 by siRNA before in vivo use, target MK2 protein and mRNA were measured in rat macrophages. As seen in Fig. 1a, MK2 expression was signiﬁcantly reduced after MK2 siRNA delivery. Densitometric analysis of data from four independent experiments revealed ~70% decrease in MK2 compared with scrambled siRNA control samples. Likewise, MK2 mRNA was reduced >90% in macrophages transfected with MK2 siRNA compared with scrambled siRNA samples (Fig. 1b). A similar reduction was also observed in primary rat bone marrow stromal cells transfected with MK2 siRNA (data not shown).

To investigate the effects of MK2 knockdown on activated MAPKs, phosphorylated MAPKs were analyzed by immunoblot. As shown in Fig. 1c, total MK2 were reduced at all times in all experimental conditions in MK2 siRNA-treated cells compared with controls. Phospho-MK2 peaked at 60 min post-LPS treatment and then decreased at 120 min in either scrambled siRNA transfected or untreated cells. It is noteworthy that after MK2 siRNA delivery phospho-MK2 expression was lower than in scrambled siRNA transfected or untreated cells. Phospho-jNK (p54, p46) protein was signiﬁcantly induced 30 to 120 min after LPS stimulation in MK2 siRNA-treated cells. Phospho-ERK (p44, p42) expression increased modestly at 30 min after LPS stimulation in all treated cells and peaked from 60 to 120 min.

**MK2 siRNA Arrested Inflammatory Mediator Production and Stimulated Anti-Inflammatory Cytokine Expression.** MK2 plays a central role in cytokine production, and IL-6 and TNF-α are key cytokines in the pathogenesis of periodontitis. As depicted in Fig. 2, a and b, compared with scrambled siRNA group MK2 siRNA-treated cells had a 60.2% reduction in IL-6 protein and a 62.1% reduction for TNF-α protein 24 to 48 h after LPS stimulation. However, prostaglandin E2 was undetectable (data not shown). Corresponding mRNA of IL-6, TNF-α, COX2, IL-1β, CXCL1, and IL-10 were quantified post-LPS stimulation for the indicated time periods in MK2 siRNA-treated cells. As shown in Fig. 2c, LPS-induced IL-6 expression in the MK2 siRNA-treated group decreased by 55.3% at 4 h and 22.0% at 8 h, compared with controls. Likewise, TNF-α mRNA expression (Fig. 2d) was reduced by 61.6 and 53.1% at 4 and 8 h, respectively, compared with scrambled siRNA controls. Comparable data were obtained with other cytokines COX2 (Fig. 2e) and IL-1β (Fig. 2f) and the chemokine CXCL1 (Fig. 2g).

**MK2 siRNA Intraradial Delivery Silenced Target MK2 Gene Expression and Impeded Alveolar Bone Loss in an A. actinomycetemcomitans LPs-Induced Periodontitis Model.** The significance of silencing MK2 was determined in an experimental model of periodontal disease. Initially, local MK2 mRNA knockdown in gingival tissues after...
MK2 siRNA intraoral delivery was measured. MK2 or scrambled siRNA was bilaterally microinjected in the palatal regions between the maxillary first molar and the second molar (n/H110056 rats/group; 4 nmol/injection; two injections/week). On days 9 and 16 after siRNA delivery, rats were sacrificed and mRNA from gingival tissues in the immediate area was used to perform qRT-PCR for MK2 mRNA expression. As shown in Fig. 3b, MK2 was decreased 30.5/H11014.97 and 50.76/H1109.6% after the third and fifth injections, respectively, compared with scrambled siRNA samples. These differences were significant (P/H1100.05) for both treatment groups compared with the scrambled siRNA injection group.

Next, we investigated the ability of injected MK2 siRNA to prevent A. actinomycetemcomitans LPS-induced alveolar bone loss in an established 4-week model of rat periodontal bone loss (Rogers et al., 2007b) and siRNA delivery as described above. A schema of the experimental protocol is presented in Fig. 3a. A. actinomycetemcomitans LPS-induced experimental periodontitis led to robust and significant bone loss (P/H110< 0.001) between the vehicle group and the LPS-injected group (Fig. 3, d and e). The CT-determined BVF (Fig. 3d) of a standardized ROI (as highlighted) was reduced from 0.77 ± 0.03 in the vehicle group to 0.64 ± 0.07 in the LPS group. Injections of MK2 siRNA effectively attenuated LPS-induced alveolar bone loss as evidenced by a BVF of 0.69 ± 0.05, which is significantly larger than the BVF associated with scrambled siRNA injections (P/H110< 0.05). BMD data (Fig. 3e) further corroborated BVF result in this in vivo study.

**Fig. 1.** a, MK2 protein expression was inhibited by specific MK2 siRNA in rat macrophage NR8383. b, in rat macrophage NR8383, densitometer scans of MK2 and β-actin were performed and recorded as the ratio of MK2 to β-actin, and density analysis showed significant knockdown of MK2 protein by MK2 siRNA delivery (left). qRT-PCR data validated the knockdown effect on MK2 mRNA expression after specific MK2 siRNA delivery (**, P < 0.01; ***, P < 0.001) (right). c, a total of 3 × 10⁶ rat macrophages NR8383 were grown in each well of a six-well plate and either transfected with MK2 siRNA or scrambled siRNA for 72 h. Then the cells were either unstimulated (0 min) or stimulated with 1 μg/ml of LPS for various time periods (10, 30, 60, and 120 min). The MK2, phospho-MK2, phospho-p38, phospho-JNK (p54, p46), and phospho-ERK (p44, p42) were examined. β-Actin was used for loading control. Data are representative of three separate experiments.

Inflammatory Gene Expression Is Attenuated by MK2 Gene Silencing In Vivo. In vivo suppression of cytokine mRNA by MK2 siRNA in periodontal connective tissues was measured by qRT-PCR from tissues harvested by LCM
Connective tissues were used for these experiments because LPS and siRNA injections occurred in this region (Fig. 4, b–f). We observed a trend toward reduced expression of MK2, TNF-α, IL-1β, COX2, and CXCL1 mRNAs in gingival connective tissue samples from MK2 siRNA-injected sites in comparison with scrambled siRNA-injected sites. We did not detect IL-6 and interferon-γ mRNA expression in these samples.

**MK2 siRNA In Vivo Delivery Attenuated A. actinomycetemcomitans LPS-Induced Inflammatory Infiltrate and Osteoclastogenesis.** Histological examination of periodontal tissues was used to evaluate the extent of inflammatory infiltrate (Fig. 5a). Hematoxylin and eosin staining indicated low levels of inflammatory cells in control samples. In contrast, a large amount of inflammatory cells, primarily neutrophils with scattered lymphocytes and macrophages, were associated with LPS injections with/without scrambled siRNA injections. Local delivery of MK2 siRNA clearly reduced the number of inflammatory cells, particularly neutrophils. A significant increase ($p < 0.01$) in the number of cells was observed in LPS groups with/without scrambled siRNA treatment compared with the vehicle group. A significant reduction ($p < 0.05$) in cell number occurred in the MK2 siRNA-injected group.
Fig. 3. Specific MK2 siRNA in vivo delivery silenced target gene expression and reduced LPS-induced bone loss. a, a schematic showing overall
experimental protocol. b, MK2 mRNA expression in palate gingiva after three and five times siRNA in vivo delivery. Results are expressed as mean ±
S.E. (n = 5 or 6 rats/group; *, P < 0.05). c, representative µCT images of rat maxillae from the indicated treatment groups. ROI for quantitative
analysis is highlighted. d, volumetric analysis of bone loss levels. e, BMD analysis of bone loss levels. *, P < 0.05; ***, P < 0.001.
compared with LPS groups with/without scrambled siRNA treatment, indicating that MK2 siRNA delivery repressed the inflammatory response (Fig. 5b). TRAP staining for osteoclast formation (Fig. 6a) indicated that MK2 siRNA treatment significantly reduced osteoclasts \((p < 0.001)\), which were identified by morphological features (size and multinucleation) and compared with scrambled siRNA injection samples (Fig. 6b).

**Discussion**

Our main findings are as follows: 1) silencing MK2 modulates the innate immune response triggered by bacterial LPS; 2) intraoral delivery of covalently modified siRNA is effective for inhibiting target gene expression in vivo; and 3) siRNA-mediated inhibition of MK2 decreases periodontal disease
severity in vivo. To our knowledge, ours is the first study to show that MK2 is critical in the pathogenesis of periodontitis and to evaluate the therapeutic potential of modulating MK2 activity in a periodontal disease model using a covalently modified siRNA.

Local delivery of siRNA therapeutics is a promising platform for the treatment of orally related diseases, and until now, direct evaluation of intraoral siRNA delivery has not been reported in the literature. However, because oral mucosa is similar to vaginal mucosa, other studies may provide insight into oral disease mechanisms. For example, in a study with mouse vaginal mucosa, siRNA vaginal instillation silenced gene expression in the mouse vagina and ectocervix for at least 9 days (Palliser et al., 2006). In our study, Accell siRNA was used to perform local delivery. This covalently modified siRNA has been systemically delivered by intraperitoneal injection in an ovarian cancer model (Difeo et al., 2009). We observed that 9 or 16 days post-MK2 siRNA delivery target gene expression was significantly decreased. However, further work should be done to elucidate the knockdown effects on distant tissues from the injection site and more peripheral tissues. In related studies of locally delivered siRNA, target gene levels by siRNA intrathecal injection lasted 5 days (Christoph et al., 2006). siRNA against apolipoprotein B was active in mice for only a few days, and after 9 days apolipoprotein B approached 70% of its initial level, whereas apolipoprotein B knockdown in nonhuman primates was still effective after 11 days (Zimmermann et al., 2006).

In the periodontal microenvironment, microbial-associated molecular patterns derived from bacteria in the dental biofilm activate leukocytes, macrophages, osteoblasts, and fibroblasts to produce cytokines such as TNF-α, IL-6, and prostaglandin E2. Thus, in vivo, various cell types participate in the pathogenesis of periodontal diseases. To study the potential

Fig. 5. a, MK2 silencing reduced inflammatory infiltrate induced by A. actinomycetemcomitans LPS in the area adjacent to bone loss. The representative sections on the left showed histological features from indicated treatment group (bars = 1500 μm). The sections on the right show an enlarged box area from the left at high magnification (bars = 60 μm). b, enumerated nuclei in each treatment group are presented. Results are expressed as mean ± S.E. (n = 5 or 6 rats/group; *, P < 0.05; **, P < 0.01).

Fig. 6. MK2 siRNA delivery attenuated osteoclastogenesis induced by A. actinomycetemcomitans LPS. a, TRAP staining of representative tissue sections of rat maxillae from the indicated treatment groups (bars = 100 μm). TRAP+ osteoclasts are stained red. B, bone; T, tooth. Dotted line shows area of bone, and the arrow indicates osteoclast. b, graphic representation of enumerated TRAP+ multinucleated formation in each tissue section is presented. Horizontal bar indicates mean TRAP+ cell counts. **, P < 0.01; ***, P < 0.001.
role of siRNA therapeutics, total bone marrow cultures comprised of monocytes/macrophages, lymphocytes, and osteoblasts/stromal cells, all part of the periodontal microenvironment, were used to evaluate the effect of MK2 knockdown by siRNA on LPS-induced cytokine expression. Our data show that LPS-induced IL-6 expression was significantly decreased, both at the mRNA and protein levels, a result consistent with previous observations in MK2−/− mice (Kotlyarov et al., 1999; Hegen et al., 2006). We also observed that MK2 siRNA delivery significantly reduced TNF-α mRNA and protein expression. Regulation of IL-6 and TNF-α expression via MK2 occurs at two different levels: IL-6 synthesis is regulated at the level of mRNA stability, and TNF-α is regulated mainly through an ARE-dependent translation efficiency mechanism (Neininger et al., 2002). However, other data also show that TTP is a direct substrate of MK2, controlling both the stability and translation of TNF-α mRNA (Carballo et al., 1998). The role of MK2 in the regulation of LPS-induced inflammatory cytokine gene expression is confirmed by significant reductions of mRNA expression for COX2, IL-1β, and the chemokine CXCL1 in cells transfected with MK2 siRNA. In the present study, MK2 siRNA gene knockdown changed the activation of JNK and ERK MAPKs, suggesting the existence of cross-talk and compensatory mechanisms and underscoring the complexities of Toll-like receptor signaling pathways.

In vivo evidence presented here suggests that silencing MK2 is a valid therapeutic target for the management of periodontal diseases. Local delivery of MK2 siRNA reduced alveolar bone loss in an LPS-induced periodontitis model. Multiple inflammatory signals can modulate the expression of receptor activator of NF-κB (RANK; receptor activator of NF-κB ligand), and osteoprotegerin, which comprise the cytokine network controlling osteoclastogenesis. It is well established that the production of proinflammatory cytokines correlate with inflammatory periodontal bone loss resulting from osteoclast differentiation and activation. MK2 siRNA attenuated the inflammatory infiltrate associated with A. actinomycetemcomitans LPS-induced bone loss in the present study. Concomitantly, MK2 siRNA resulted in a decrease in osteoclast formation and a subsequent reduction in bone resorption. At the end of the experimental period, mRNA expression of MK2 and other inflammatory genes was examined in the gingival connective tissue by qRT-PCR. We observed a nonsignificant, but distinct, reduction on the expression of MK2 and the inflammatory genes, findings that were consistent with in vitro results. Although MK2 silencing in this study resulted in a reduction of osteoclast numbers and activity, we cannot rule out another plausible mechanism whereby MK2 silencing reduced the absolute numbers of osteoclast precursors recruited into the periodontal microenvironment. Ongoing experiments are addressing this possible mechanism.

In summary, our study using locally delivered siRNA for treatment of an oral-related disease provides strong evidence for proof-of-principle. Our data show that MK2 has a fundamental role in the pathogenesis of periodontitis and MK2 is a potential therapeutic target. Future studies with larger animal models or nonhuman primates with naturally occurring periodontitis can be exploited to address the therapeutic effect of MK2 silencing and develop a clinical approach for intraoral delivery of siRNA.

References


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