Pharmacological Properties of MK-3207, a Potent and Orally Active Calcitonin Gene-Related Peptide Receptor Antagonist

Christopher A. Salvatore, Eric L. Moore, Amy Calamari, Jacquelynn J. Cook, Maria S. Michener, Stacey O’Malley, Patricia J. Miller, Cyrille Sur, David L. Williams, Jr., Zhizhen Zeng, Andrew Danziger, Joseph J. Lynch, Christopher P. Regan, John F. Fay, Yui S. Tang, Chi-Chung Li, Nicole T. Pudvah, Rebecca B. White, Ian M. Bell, Steven N. Gallicchio, Samuel L. Graham, Harold G. Selnick, Joseph P. Vacca, and Stefanie A. Kane


Received November 12, 2009; accepted January 7, 2010

ABSTRACT
Calcitonin gene-related peptide (CGRP) has long been hypothesized to play a key role in migraine pathophysiology, and the advent of small-molecule antagonists has clearly demonstrated a clinical link between blocking the CGRP receptor and migraine efficacy. 2-[(8R)-8-(3,5-Difluorophenyl)-10-oxo-6,9-diazaspido[4,5]dec-9-yl]N-[(2R)-2′-oxo-1,1′,2′,3′-3-tetrahydrospiro[indene-2,3′]-pyrrolo[2,3-b]pyridin-5-yl]acetamide (MK-3207) represents the third CGRP receptor antagonist to display clinical efficacy in migraine trials. Here, we report the pharmacological characterization of MK-3207, a potent and orally bioavailable CGRP receptor antagonist. In vitro, MK-3207 is a potent antagonist of the human and rhesus monkey CGRP receptor (Kᵢ = 0.024 nM). In common with other CGRP receptor antagonists, MK-3207 displays lower affinity for CGRP receptors from other species, including canine and rodent. As a consequence of species selectivity, the in vivo potency was assessed in a rhesus monkey pharmacodynamic assay measuring capsaicin-induced changes in forearm dermal blood flow via laser Doppler imaging. MK-3207 produced a concentration-dependent inhibition of dermal vasodilation, with plasma concentrations of 0.8 and 7 nM required to block 50 and 90% of the blood flow increase, respectively. The tritiated analog [³H]MK-3207 was used to study the binding characteristics on the human CGRP receptor. [³H]MK-3207 displayed reversible and saturable binding (Kᵢ = 0.06 nM), and the off-rate was determined to be 0.012 min⁻¹, with a t½ value of 59 min. In vitro autoradiography studies on rhesus monkey brain slices identified the highest level of binding in the cerebellum, brainstem, and meninges. Finally, as an index of central nervous system penetrability, the in vivo cerebrospinal fluid/plasma ratio was determined to be 2 to 3% in cisterna magna-ported rhesus monkeys.

Migraine is one of the most prevalent and disabling neurological disorders, with characteristic symptoms that can last for several days. Despite its severity and high prevalence, migraine is not generally recognized as a serious medical condition and the societal burden is not fully appreciated. Migraine often affects people during their most productive years, which in turn burdens families and employers and ultimately affects the quality of life of the migraine sufferer. Migraine is generally agreed to be underdiagnosed and many migraineurs do not receive appropriate therapy, indicating there is significant room for improvement in the diagnosis and management of migraine.

An overall improvement in migraine treatment occurred

ABBREVIATIONS: CGRP, calcitonin gene-related peptide; BIBN4096BS, olcegepant; MK-0974, telcagepant; MK-3207, 2-[(8R)-8-(3,5-difluorophenyl)-10-oxo-6,9-diazaspido[4,5]dec-9-yl]-N-[(2R)-2′-oxo-1,1′,2′,3′-3-tetrahydrospiro[indene-2,3′]-pyrrolo[2,3-b]pyridin-5-yl]acetamide; CLR, calcitonin receptor-like receptor; RAMP, receptor activity-modifying protein; AM, adrenomedullin; CT, calcitonin; CTR, calcitonin receptor; HEK, human embryonic kidney; HBSS, Hanks’ balanced salt solution; PBS, phosphate-buffered saline; HTRF, homogeneous time resolved fluorescence; DMSO, dimethyl sulfoxide; PK, pharmacokinetic; CIDV, capsaicin-induced dermal vasodilation; LDF, laser Doppler flow; CSF, cerebrospinal fluid; CNS, central nervous system; P-gp, P-glycoprotein; A, apical; B, basal; AMY, amylin; h, human; r, rat.
with the introduction of the 5-hydroxytryptamine1B/1D receptor agonists called triptans that currently represent the antimigraine therapy of choice. However, some patients do not respond optimally to triptans and some only partially respond. Triptans are considered safe when used appropriately but are contraindicated for patients with cardiovascular disease, because they are direct coronary vasoconstrictors. The next-generation antimigraine drugs need to improve upon the shortcomings of triptan therapy.

After successful introduction of the triptans, evidence began to mount linking CGRP to the pathophysiology of migraine. It was found that plasma levels of CGRP were elevated during the headache phase of migraine (Goadsby et al., 1990; Gallai et al., 1995) and the levels were normalized concomitantly with pain relief (Goadsby and Edvinsson, 1993). In addition, intravenous administration of CGRP to migraineurs induced a delayed migraine-like headache (Lassen et al., 1998). These observations suggested antagonism of the CGRP receptor might represent a novel approach to migraine treatment. More definitive evidence for a role of CGRP in migraine came from the original proof of concept studies with olcegepant (BIBN4096BS; Doods et al., 2000). In this study, intravenous administration of olcegepant was shown to be effective in the acute treatment of migraine (Olesen et al., 2004). Subsequently, we described the identification of a novel, orally bioavailable CGRP receptor antagonist, telcagepant (MK-0974; Paone et al., 2007; Salvatore et al., 2008) that was effective as an acute treatment for migraine, with efficacy comparable to zolmitriptan (Ho et al., 2008).

CGRP is a 37-amino acid neuropeptide produced by tissue-specific alternative mRNA splicing of the calcitonin gene (Amara et al., 1982) and belongs to the calcitonin family of peptides, which includes calcitonin, amylin, and adrenomedullin. The CGRP receptor is heterodimeric, and CGRP activity is mediated by the coexpression of a G protein-coupled receptor; calcitonin receptor-like receptor (CLR); a single transmembrane-spanning protein designated receptor activity-modifying protein (RAMP)1 (McLatchie et al., 1998); and an intracellular protein, receptor component protein (Evans et al., 2000). The RAMPs comprise a group of three proteins designated RAMP1, RAMP2, and RAMP3. A functional CGRP receptor requires coexpression of CLR and RAMP1, whereas when CLR is coexpressed with RAMP2 or RAMP3, a receptor with high affinity for adrenomedullin (AM) is produced (McLatchie et al., 1998). In addition, RAMPs can complex with the calcitonin (CT) receptor to form the amylin family of receptors (Christopoulos et al., 1999; Muff et al., 1999).

The distribution of CGRP receptors in the trigeminovascular system is consistent with a role in migraine pathophysiology. CGRP receptor antagonists could act through both peripheral and central sites of action. Blockade of peripheral receptors on blood vessels and mast cells could block neurogenic inflammation and normalize dilated blood vessels, whereas blockade of central receptors in the brainstem may inhibit pain transmission. In addition, CGRP receptors are widely expressed in numerous brain regions, including periaqueductal gray, parabrachial nucleus, nucleus solitarius, cerebellum, hippocampus, and amygdala (Sexton et al., 1986; Sexton, 1991; Christopoulos et al., 1995). Finally, CLR and RAMP1 are located in the spinal trigeminal nucleus of rat and are colocalized in the presynaptic terminals of the spinal dorsal horn, further indicating a potential role in pain transmission (Marvizon et al., 2007; Lennerz et al., 2008).

The work presented here characterizes the preclinical pharmacology of the novel, potent, and orally bioavailable CGRP receptor antagonist MK-3207 (Bell et al., 2010). MK-3207 displayed good oral bioavailability in rats (74%), dogs (67%), and rhesus monkeys (41%). In rhesus monkeys, clearance and intravenous half-life were moderate, with values of 15 ml/min/kg and 1.5 h, respectively. Recently, the efficacy of MK-3207 was evaluated in a phase II adaptive dose-ranging trial for the acute treatment of migraine. In this study, MK-3207 significantly improved migraine pain relief 2 h after dosing compared with placebo (Hewitt et al., 2009), further pointing to the clinical utility of this mechanism for a future migraine therapy.

Materials and Methods

Expression Vector Constructs and Mutagenesis. Human CLR, RAMP1, RAMP2, and RAMP3 expression vector constructs were described by Salvatore et al. (2008). The expression vector construct for the insert-negative human CT receptor (CTR) was described by Salvatore et al. (2006). Rat CLR and RAMP1 expression vector constructs were described by Mallee et al. (2002). Human RAMP1 site-directed mutagenesis (tryptophan at position 74 was replaced with an alanine) was performed using the QuickChange lightning site-directed mutagenesis kit (Stratagene, La Jolla, CA) according to the manufacturer’s protocol.

Cell Culture and Generation of Recombinant Cell Lines. HEK293 and COS-7 cells were cultured in Dulbecco’s modiﬁed Eagle’s medium with 4.5 g/l glucose, 1 mM sodium pyruvate, and 2 mM glutamine supplemented with 10% fetal bovine serum, 100 units/ml penicillin, and 100 μg/ml streptomycin and maintained at 37°C, 5% CO₂, and 95% humidity. Cells were subcultured by treatment with 0.25% trypsin with 0.1% EDTA in Hanks’ balanced salt solution (HBSS).

HEK293 cell lines stably expressing the human CGRP (CLR/RAMP1), AM₁ (CLR/RAMP2), and AM₂ (CLR/RAMP3) receptors were described previously (Salvatore et al., 2008). For transient transfections, 24 h before transfection COS-7 or HEK293 cells were seeded in 500-cm² dishes. Transfections were performed by combining 60 μg/dish DNA with 180 μg/dish Lipofectamine 2000 (Invitrogen, Carlsbad, CA). Human CTR was transfected singly in HEK293 cells or cotransfected in COS-7 cells with equal amounts of RAMP1 (AMY₁ receptor) or RAMP3 (AMY₂ receptor). Transfection cocktail was added directly to the medium, and the cells were harvested for membranes 48 h after transfection.

Membrane Preparation and Radioligand Binding Studies. Transiently or stably transfected cells were washed with PBS and harvested in ice-cold harvest buffer containing 50 mM HEPES, 1 mM EDTA, and Complete protease inhibitors (Roche Diagnostics, Indianapolis, IN). The cell suspension was disrupted with a laboratory homogenizer and centrifuged at 48,000 g to isolate membranes. Membranes from rat and dog brain were prepared similarly. Rhesus cerebellum was disrupted using a laboratory homogenizer in 10 mM HEPES and 5 mM MgCl₂ and used directly in binding experiments. SK-N-MC membranes were purchased from Receptor Biology, Inc. (Beltsville, MD).

CGRP and adrenomedullin competition binding assays were conducted as described by Salvatore et al. (2008). Amylin binding assays were conducted by combining MK-3207 and 40 pM [¹²⁵I]-rat amylin (PerkinElmer Life and Analytical Sciences, Boston, MA), followed by 25 μg of CTR/RAMP1 or 25 μg of CTR/RAMP3 membranes and incubated for 3 h at room temperature in binding buffer (10 mM HEPES, 5 mM MgCl₂, and 0.2% bovine serum albumin) in total
volume of 1 ml. Calcitonin binding assays were set up as described above but with 25 µg of CTR membranes and 30 pM 125I-human calcitonin (PerkinElmer Life and Analytical Sciences) as the radioligand. Incubations were terminated by filtration through GF/B 96-well filter plates that had been blocked with 0.5% polyethyleneimine. Data were analyzed using Prism (GraphPad Software Inc., San Diego, CA), and the Ki value was determined using the equation Ki = IC50/(1 + [ligand]/Ki). The Ki value for each receptor was determined by saturation binding experiments (data not shown).

**[H]MK-3207 Saturation Binding and Association and Dissociation Kinetics.** Saturation binding studies were performed by combining increasing concentrations of [H]MK-3207 (specific activity, 73.7 Ci/mmol), 1 µM compound 25 (Stump et al., 2009) for nonspecific binding, and 50 µg/well SK-N-MC membrane in a total volume of 1 ml of binding buffer. Reactions were incubated overnight (18 h) at room temperature. Association kinetic assays were performed by combining 60 pM [H]MK-3207 with 50 µg/well SK-N-MC membranes in binding buffer and incubating at room temperature for various times from 1 to 300 min. Dissociation kinetic assays were performed by combining 60 pM [H]MK-3207 with 50 µg/well SK-N-MC membranes in binding buffer and incubating at room temperature for 3 h. At that point, 1 µM compound 25 was added, and dissociation was monitored for various intervals from 1 to 390 min. All assays were terminated by filtration through GF/B 96-well filter plates that had been blocked with 0.5% polyethyleneimine.

**In Vitro Functional Studies.** Functional assays were conducted as described previously (Salvatore et al., 2008). In brief, HEK293 cells stably expressing the human CLR receptor were plated at a density of 85,000 cells/well in 96-well poly-l-lysine-coated plates approximately 19 h before the assay. Cells were washed with PBS and preincubated with various concentrations of MK-3207 in the presence or absence of 50% human serum for 30 min at 37°C in a CO2 incubator. Isobutyl-methylxanthine (300 µM) was added to the cells, and then they were incubated for 30 min at 37°C followed by stimulation with 0.3 nM α-CGRP for 5 min at 37°C. After agonist stimulation, cells were washed with PBS and the intracellular cAMP concentration measured using the cAMP SPA Biotrak direct screening assay (GE Healthcare, Piscataway, NJ). Dose-response curves were plotted, and IC50 values were determined. Schild analysis was used as a measure of competitive antagonism by plotting log (DR-1) versus log [B], where DR is the ratio of the EC50 values in the presence and absence antagonist and [B] is the antagonist concentration. The x-intercept is equal to the pA2 and the Kp values calculated using the formula pA2 = − log Kp.

Functional assays with HEK293 cells transiently expressing the CGRP receptor were conducted using the HTRF cAMP dynamic assay kit (CisBio, Bedford, MA). Twenty-four hours before transfection, the cells were plated in 10-cm dishes. For the transfection, 4 µg of CLR and 4 µg of RAMP1 were combined with 23 µg of Lipofectamine 2000 (Invitrogen) and incubated at room temperature for 30 min. The transfection mix was added to the cell medium and incubated for 48 h at 37°C in a CO2 incubator. Cells transiently expressing CLR and RAMP1 were dissociated, centrifuged, and resuspended in assay buffer consisting of Cellgro Complete media (Mediatech, Herndon, VA) with 300 µM isobutyl-methylxanthine. Cells were added to a 384-well assay plate (ProxiPlate Plus; PerkinElmer Life and Analytical Sciences) containing antagonist at a density of 2000 cells/well and incubated at room temperature for 30 min. An EC50 concentration of the peptide agonist CGRP was then added, and the plate was incubated at room temperature for 20 min. Finally, HTRF reagents were added to the plate, incubated for 1 h at room temperature, and the plate was read using an Envision (PerkinElmer Life and Analytical Sciences) plate reader in the HTRF mode. The raw data were converted to nanomolar cAMP using a standard curve.

**Rhesus Monkey Pharmacodynamic Assay.** All procedures related to the use of animals were approved by the Institutional Animal Care and Use Committee at Merck Research Laboratories (West Point, PA) and conform with the Guide for the Care and Use of Laboratory Animals (National Institutes of Health, National Research Council, revised 1996). Six adult rhesus monkeys (either sex; range, 4.8–12.7 kg) were used for both vehicle and active test agent studies. Anesthesia was induced with ketamine (5–30 mg/kg i.m., to effect). Animals were then intubated, and anesthesia was maintained with isoflurane (1–2% carried in 100% oxygen). Animals were placed on a temperature-controlled heating blanket and instrumented with a temporary peripheral catheter for the administration of vehicle or test agent. Four rubber O-rings (8-mm inner diameter) were placed on the ventral side of the forearm and positioned such that they were not directly overlying a visible vessel. Dermal vaso- dilatation was induced within the O-rings in sequential order by topical administration of capsaicin (2 mg in 30% ethanol, 30% Tween 20, and 40% water) and quantitated using a laser Doppler imager (Moor Instruments, Ltd., Millwex, Axminster, Devon, UK), according to the study protocols summarized below. In all vehicle and test agent studies, the order in which capsaicin was administered to the four O-rings was varied from study to study.

To determine the effect of MK-3207 on capsaicin-dependent dermal vasodilatation, each animal was administered three sequential infusions of vehicle or an MK-3207 treatment regimen after a no-treatment capsaicin baseline challenge. Animals were allowed at least a 7-day “washout” between studies. Vehicle administration was as follows: dose 1, intravenous bolus 0.5 ml of 50% DMSO, 50% water followed by 25-min continuous intravenous infusion of 0.025 ml/min 50% DMSO, 50% water (0.625 ml); dose 2, intravenous bolus 0.5 ml of 50% DMSO, 50% water followed by 25-min continuous intravenous infusion of 0.025 ml/min 50% DMSO, 50% water (0.625 ml); and dose 3, intravenous bolus 0.5 ml of 100% DMSO followed by 25-min continuous intravenous infusion of 0.025 ml/min 100% DMSO (0.625 ml). MK-3207 was administered in three different experiments as three dosing regimes to target a range of plasma exposures from 1 to 300 nM. Each regimen was given once to all rhesus monkeys as follows: 1) 0.3 µg/kg i.v. bolus + 0.008 µg/kg/min, 0.6 µg/kg i.v. bolus + 0.025 µg/kg/min and then 2.1 µg/kg + 0.084 µg/kg/min; 2) 0.6 µg/kg i.v. bolus + 0.025 µg/kg/min, 2.1 µg/kg i.v. bolus + 0.084 µg/kg/min and then 9.1 µg/kg i.v. bolus + 0.25 µg/kg/min; and 3) 9.1 µg/kg + 0.25 µg/kg/min, 21.2 µg/kg i.v. bolus + 0.84 µg/kg/min and then 60.6 µg/kg i.v. bolus + 2.5 µg/kg/min.

Within each test period, the dermal blood flow was measured at 20 min after application of capsaicin (i.e., 25 min after initiation of vehicle or test agent administration) as well as baseline: the basal blood flow value acquired in that ring just before the dose of vehicle or test agent. Blood samples for determination of plasma concentration of MK-3207 also were obtained at 20 min after application of capsaicin (i.e., 25 min after initiation of vehicle or test agent administration) during each test agent test period. An empirical Emax model was then used to describe the PK versus efficacy relationship of MK-3207 for inhibition of CIDV in rhesus monkeys. Details of the model structure are provided below. Blood flow is described as a baseline blood flow plus an incremental blood flow as a result of CIDV and blockade of CIDV by MK-3207 through an Emax relationship. The model was fit to the rhesus monkey CIDV data pooled across all MK-3207 treatment regimes, and model parameters for drug efficacy (Emax and EC50) were estimated using least-squares regression method. Two data points at 0.3-µg/kg dose were excluded from the analysis due to plasma concentrations falling below the limit of quantification.

The PK versus efficacy relationship was described using the equation F = F0 + (Fcaps − F0) × (1 × Emax × C/(EC50 + C)), where F is the measured laser Doppler flow (LDF) at the end of MK-3207 infusion and 20 min after capsaicin challenge, F0 is the baseline LDF (no capsaicin or MK-3207), FCaps is the LDF at 20 min after capsaicin challenge (no MK-3207), FCaps is the baseline LDF measured during the capsaicin response test (where Fcaps was measured, no capsaicin or MK-3207), Emax is the maximal percentage of inhibition by MK-3207, C is the plasma concentration of MK-3207, and EC50 is the...
Results

Binding Studies on CGRP Receptors. Competitive binding experiments were carried out to determine the relative affinity of MK-3207 for human, rhesus monkey, rat, and canine CGRP receptors. MK-3207 (Fig. 1) displayed high affinity for the native human CGRP receptor in SK-N-MC cells and for the recombinant human receptor as measured by the ability to compete with 125I-hCGRP binding, with Kᵢ values of 0.024 ± 0.001 nM (n = 3) and 0.022 ± 0.002 nM (n = 14), respectively. MK-3207 displayed a similar affinity (Kᵢ) for the rhesus monkey receptor (0.024 ± 0.001 nM; n = 14) as for human, but it displayed >400-fold lower affinity for the canine and rat receptors, with values of 10 nM (n = 2) and 10 ± 1.2 nM (n = 4), respectively (Fig. 2).

Binding Studies on Human Adrenomedullin, Calcitonin, and Amylin Receptors. Competitive binding experiments were carried out to determine the selectivity of MK-3207 for the human CGRP receptor versus the related human AM, CT, and amylin (AMY) receptors (Fig. 3). MK-3207 was highly selective versus the human AM₁ (CLR/RAMP2) and AM₂ (CLR/RAMP3) receptors, with Kᵢ values of 16,500 nM (n = 2) and 156 ± 17 nM (n = 7), respectively. MK-3207 maintained a high degree of selectivity versus human CTR, with a Kᵢ value of 1.9 ± 0.58 μM (n = 5). MK-3207 also displayed good selectivity versus the AMY₃ (CTR/RAMP3) receptor, with a Kᵢ value of 128 ± 25 nM (n = 3), but it was less selective versus the AMY₁ (CTR/RAMP1) receptor, with a Kᵢ value of 0.75 ± 0.13 nM (n = 3).

Saturation and Kinetic Binding Studies with [3H]MK-3207. Saturation binding experiments using [3H]MK-3207 were conducted on SK-N-MC membranes to determine the Kᵢ and Bₘₐₓ values. [3H]MK-3207 displayed saturable binding, with a Kᵢ value of 60 pM and a Bₘₐₓ value of 350 fmol/mg protein (Fig. 4A).

Fig. 1. Chemical structures of MK-3207 and telcagepant.

Fig. 2. Concentration-dependent inhibition of 125I-hCGRP binding by MK-3207 from SK-N-MC, human (cloned), rhesus cerebellum, dog brain, and rat brain membranes. Mean values ± S.E.M.
Kinetic binding experiments were performed to determine the on rate and off rate for \(^{3}H\)MK-3207 on SK-N-MC membranes. In the association assay, \(^{3}H\)MK-3207 reached saturation very quickly, with an apparent \(k_{on}\) of \(1.5 \times 10^{9} \text{ M}^{-1} \cdot \text{min}^{-1}\) (Fig. 4B). For the dissociation of \(^{3}H\)MK-3207, the \(k_{off}\) was calculated as 0.012 \(\text{min}^{-1}\), with a \(t_{1/2}\) value of 59 min (Fig. 4C).

**Functional Studies on the Human CGRP Receptor.** The effect of MK-3207 on CGRP-induced cAMP production in CLR/RAMP1 cells was investigated. Consistent with the binding data, MK-3207 potently blocked human \(\alpha\)-CGRP-stimulated cAMP responses in human CGRP receptor-expressing HEK293 cells, with an IC\(_{50}\) value of 0.12 ± 0.02 nM (n = 6). Addition of 50% human serum (IC\(_{50}\) = 0.17 ± 0.02 nM; n = 6) had little effect on the apparent potency of MK-3207.

Increasing concentrations of MK-3207 caused a dose-dependent rightward shift in the CGRP dose-response curve, with no reduction in the maximal agonist response (Fig. 5A). Schild regression (Fig. 5B) yielded a \(pA_{2}\) value of 10.3 (n = 2; \(K_s = 0.05 \text{ nM}\)).

**Functional Studies on Mixed Species CGRP Receptors.** To determine whether the species selectivity exhibited by MK-3207 is derived from RAMP1, hybrid human/rat CGRP receptors were generated by transiently transfecting human CLR with rat RAMP1. Human CLR coexpressed with rat RAMP1, or the human RAMP1 mutant that replaces tryptophan 74 with an alanine, resulted in a similar decrease in potency (pIC\(_{50}\) = 8.12 ± 0.1 and 8.66 ± 0.18, respectively (n = 3)) versus the wild-type human receptor (pIC\(_{50}\) = 9.75 ± 0.03; n = 3). MK-3207 displayed significantly lower potency for the rat CGRP receptor, with a pIC\(_{50}\) = 7.31 ± 0.09 (n = 3; Fig. 6).

**Effect of MK-3207 on Capsaicin-Induced Vasodilation in Rhesus Monkeys.** Administration of vehicle alone resulted in somewhat variable and non-time-dependent changes in CIDV responses that were not considered significant from pre-vehicle infusion CIDV responses. Administration of MK-3207 resulted in an exposure-dependent decrease in CIDV. Plasma exposure obtained in this study ranged from less than 1 nM (obtained in the lowest dosing regimen of 0.3 \(\mu\)g/kg i.v. bolus + 0.008 \(\mu\)g/kg/min for 25 min) to approximately 260 nM (obtained in the highest dosing regimen of 60.6 \(\mu\)g/kg i.v. bolus + 2.5 \(\mu\)g/kg/min for 25 min). An empirical E\(_{max}\) model was used to describe the PK versus efficacy relationship of MK-3207 for inhibition of CIDV in rhesus monkeys. The results suggest that MK-3207 has EC\(_{50}\) and E\(_{max}\) values of approximately 0.8 ± 0.3 nM (mean ± S.E.) and 81 ± 5% (mean ± S.E.), respectively, for inhibition of CIDV in rhesus monkeys. The expected EC\(_{50}\) value in rhesus monkey is therefore approximately 7 nM (9-fold higher than the estimated EC\(_{50}\) value). The model fit is presented in Fig. 7.
As shown in Fig. 8B, [3H]MK-3207 displayed minimal localization in rhesus monkey brain slices. The [3H]MK-3207 was labeled with tritium to high specific activity, approximately 2-fold greater than its affinity value (K_i = 0.024 nM). As indicated by B-to-A/A-to-B transport ratios of 33 and 50 in L-MDR1 and L-mdr1a cell lines, respectively. MK-3207 has high passive permeability of 24 × 10^{-6} cm/s.

**Autoradiographic Studies in Rhesus Monkey Brain.** MK-3207 was labeled with tritium to high specific activity, and used for in vitro autoradiographic studies for binding site localization in rhesus monkey brain slices. The [3H]MK-3207 concentration selected for these studies was 0.045 nM, approximately 2-fold greater than its affinity value (K_i = 0.024 nM). As shown in Fig. 8B, [3H]MK-3207 displayed minimal nondisplaceable binding, defined on an adjacent brain slice by 1 µM self-block. In the absence of competing unlabeled ligand, high binding densities were observed in several brain regions examined, including the cerebellum, brainstem, and meninges (Fig. 8A). In the cerebellum, high binding density is mainly located in the gray matter such as cerebellar cortex, with minimal binding in the white matter, consistent with receptor localization in the cerebellum. In the brainstem, high-density binding sites are nonuniformly distributed to regions essential for processing nociceptive stimuli. It is interesting to note that [3H]MK-3207 also showed high binding density in the meninges but minimal binding to the cerebral cortex, providing evidence that a high density of CGRP receptors are localized in the meninges. [3H]MK-3207 binding sites were also observed in the hypothalamus and hippocampus, with minimal binding in the caudate putamen (data not shown; Sur et al., 2009).

**MK-3207 Cerebrospinal Fluid Levels in Rhesus Monkey.** Pharmacokinetic parameters were determined in CSF and plasma after oral dosing in cisterna magna-ported rhesus monkey. After an oral dose of 10 mg/kg MK-3207, the CSF/plasma ratio is 2% to 3% (Table 1). However, the CSF/plasma ratio is approximately 30% of the unbound fraction (9.4%) in plasma, indicating that the central and peripheral compartments are not freely equilibrating.

**In Vitro P-Glycoprotein Transport Studies.** MK-3207 is a substrate for human and mouse P-gp at 1 and 5 µM, as indicated by B-to-A/A-to-B transport ratios of 33 and 50 in L-MDR1 and L-mdr1a cell lines, respectively. MK-3207 has high passive permeability of 24 × 10^{-6} cm/s.

**Discussion**

Approximately 20 years ago, CGRP was first postulated to play a role in the pathophysiology of migraine. It is now evident that CGRP is not only simply a migraine biomarker but also is an important player in migraine pathogenesis. Three CGRP receptor antagonists (olcegepat, telcegepat, and MK-3207) have displayed efficacy in the treatment of migraine; and in this report, we detail the pharmacological characterization of MK-3207, the second orally bioavailable CGRP receptor antagonist to be evaluated in the clinic for the acute treatment of migraine. One potential benefit of the new
CGRP receptor antagonist class of antimigraine treatments is the absence of vasoconstriction (Petersen et al., 2003; Lynch et al., 2010), a current liability of the triptans. Telcagepant, the first orally bioavailable CGRP receptor antagonist tested in clinical trials, has been shown to be effective as an acute treatment of migraine, with efficacy comparable with that of zolmitriptan (Ho et al., 2008). MK-3207, our second orally bioavailable antagonist, is structurally distinct from telcagepant (Fig. 1) and is 50- to 100-fold more potent both in vitro and in vivo. In an adaptive dose ranging trial, MK-3207 demonstrated a positive response on the primary 2-h pain freedom endpoint and the secondary endpoint of 2-h pain relief.

Numerous antagonists of the CGRP receptor exhibit species-selective pharmacology (Doods et al., 2000; Edvinsson et al., 2001; Hasbak et al., 2001), and MK-3207 is no exception. MK-3207 displays approximately 400-fold higher affinity for the human and rhesus monkey CGRP receptors compared with the rat and canine receptors. We previously identified RAMP1 as the primary driver of the species selectivity of olcegepant and telcagepant (Mallee et al., 2002; Salvatore et al., 2008). In addition, we identified W74 of hRAMP1 as a key residue for olcegepant binding (Mallee et al., 2002), and W74 was shown to have a similar effect on telcagepant (Miller et al., 2009). Coexpression of the W74A hRAMP1 mutant or rRAMP1 with hCLR resulted in a 10- to 40-fold reduction in potency implicating RAMP1, specifically residue 74, as playing a key role in MK-3207 binding. The potency of MK-3207 is further reduced on the rat CGRP receptor, suggesting that both RAMP1 and CLR are involved in binding.

CLR can heterodimerize with RAMP2 and RAMP3 to produce high-affinity adrenomedullin receptors. Adrenomedullin is a widely expressed potent vasodilator that, when knocked out in mice, results in death at midgestation due to extreme hydrops fetalis and cardiovascular abnormalities (Caron and Smithies, 2001). Studies to determine which RAMP is responsible for survival identified RAMP2 as being important for survival (Dackor et al., 2007; Ichikawa-Shindo 2008).

### TABLE 1

<table>
<thead>
<tr>
<th></th>
<th>Area under the Curve0–24 h</th>
<th>Cmax</th>
<th>nM</th>
<th>nM × h</th>
</tr>
</thead>
<tbody>
<tr>
<td>CSF</td>
<td>20.0 ± 13.5</td>
<td>979</td>
<td>3265</td>
<td>13.5</td>
</tr>
<tr>
<td>Plasma</td>
<td>979 ± 570</td>
<td>20.0</td>
<td>13.5</td>
<td>96.4</td>
</tr>
<tr>
<td>CSF/plasma ratio, %</td>
<td>2.0</td>
<td>2.0</td>
<td>2.9</td>
<td>2.9</td>
</tr>
</tbody>
</table>

Fig. 8. Distribution of [3H]MK-3207 binding to rhesus monkey coronal brain slices. A, [3H]MK-3207 (0.045 nM) displayed high binding densities in the cerebellum, brainstem, and meninges. B, [3H]MK-3207 displayed minimal non-displaceable binding in the presence of 1 μM unlabeled MK-3207 on an adjacent brain slice.

CGRP receptor antagonist class of antimigraine treatments is the absence of vasoconstriction (Petersen et al., 2003; Lynch et al., 2010), a current liability of the triptans. Telcagepant, the first orally bioavailable CGRP receptor antagonist tested in clinical trials, has been shown to be effective as an acute treatment of migraine, with efficacy comparable with that of zolmitriptan (Ho et al., 2008). MK-3207, our second orally bioavailable antagonist, is structurally distinct from telcagepant (Fig. 1) and is 50- to 100-fold more potent both in vitro and in vivo. In an adaptive dose ranging trial, MK-3207 demonstrated a positive response on the primary 2-h pain freedom endpoint and the secondary endpoint of 2-h pain relief.
et al., 2008), whereas the absence of RAMP3 had little or no effect until old age (Dackor et al., 2007). Based upon these observations, it was important to maintain selectivity for the CGRP receptor versus the related adrenomedullin receptors, most notably AM1. Because adrenomedullin receptors contain CLR, we initially thought that developing highly selective antagonists would be challenging. However, the RAMP1 dependence displayed by MK-3207 provided selectivity for the CGRP receptor versus both AM1 (>600,000-fold selective) and AM2 (6500-fold selective) receptors.

RAMPs can also complex with the related calcitonin receptor (CTR) to form the AMY family of receptors (Christopoulos et al., 1999; Muff et al., 1999). The most well characterized AMY receptors are heterodimers of CTR and RAMP1 (AMY1) and RAMP3 (AMY3). MK-3207 displayed excellent selectivity versus the AMY3 receptor (>5000-fold selective), but selectivity was greatly reduced against the AMY1 receptor. Perhaps this observation should not be surprising because the AMY1 receptor requires coexpression of RAMP1. The RAMP1 dependence that may have helped confer selectivity versus the AM receptors could also be responsible for the reduced selectivity seen against the AMY1 receptor.

MK-3207 was evaluated in a screen of 169 enzyme and binding assays (MDS Pharma Services, Taipei, Taiwan; partial list of assays is summarized in Supplemental Data). MK-3207 was highly selective for the CGRP receptor, with no activity (IC_{50} < 5 μM) in any assay except for the human calcitonin receptor (IC_{50} = 1.75 μM), which is consistent with our in-house determination.

To better understand the binding characteristics of MK-3207, the tritiated analog [3H]MK-3207 was used to define MK-3207 affinity and binding kinetics. Binding was assessed on membranes from the SK-N-MC cell line that constitutively expresses the human CGRP receptor (Semark et al., 1992). [3H]MK-3207 binding was found to be saturable, with an apparent K_{d} value of 60 μM, which is in agreement with the K_{d} values for unlabeled MK-3207 in competition studies using 125I-CGRP as the radioligand. The association of [3H]MK-3207 to SK-N-MC membranes was fast, with equilibrium reached quickly, but the dissociation was significantly slower (t_{1/2} = 59 min) compared with telcagepant (t_{1/2} = 1.3 min; Moore et al., 2009), a likely consequence of the greatly enhanced affinity for the CGRP receptor. The high affinity (K_{d}) and slower dissociation kinetics of MK-3207 could result in a low dose being required for migraine efficacy, but this can only be answered through large clinical efficacy trials.

The CGRP receptor is a class B G protein-coupled receptor that are characterized by having peptide agonists that occupy extensive binding sites. Due to the putative large agonist binding site, there is some discussion centered on whether an antagonist of class B receptors can be strictly competitive. We have shown previously that telcagepant displays behaviors consistent with competitive antagonism (Salvatore et al., 2008; Moore et al., 2009) and therefore interrogated MK-3207 in a similar manner. Increasing concentrations of MK-3207 caused parallel rightward shifts in the CGRP dose-response curves in a cAMP functional assay and the dose-ratio plot displays a straight line, with a slope of 1.1 and a pA_{2} value of 10.3. These behaviors are consistent with competitive antagonism; however it is difficult to determine conclusively whether MK-3207 is strictly a competitive antagonist.

The pronounced species selectivity exhibited by MK-3207 required the utilization of nonhuman primate to assess in vivo pharmacological activity. Therefore, pharmacological studies were conducted in rhesus monkey based upon capsaicin-induced dermal vasodilation (Hershey et al., 2005). Topical application of capsaicin to the rhesus monkey forearm resulted in an increase in dermal blood flow, a direct result of endogenous CGRP release, which is directly measurable via laser Doppler imaging. MK-3207 produced a concentration-dependent inhibition of capsaicin-induced dermal blood flow in the rhesus monkey forearm, affording EC_{50} and EC_{90} values of 0.8 and 7 nM, respectively. MK-3207 (IC_{50} = 0.17 nM) is approximately 65-fold more potent than telcagepant (IC_{50} = 10.9 nM) in the human serum-shifted in vitro functional assay. This in vitro gain in functional potency is maintained upon translation to in vivo activity, where MK-3207 is approximately 100-fold more potent in the rhesus monkey CIDV assay versus telcagepant (EC_{90} = 994 nM).

The prevailing view is that migraine is a neurological disorder, where the primary site of dysfunction resides in the brain. Numerous lines of preclinical and clinical evidence support this hypothesis, including 1) brainstem activation during a migrainous attack (Weiller et al., 1995; Afridi et al., 2005), 2) peripheral application of CGRP to the meningeal dural mater caused an increase in blood flow in rats but did not sensitize meningeal nociceptors (Levy et al., 2005), and 3) intravenous infusion of vasoactive intestinal peptide to migraineurs caused a marked dilation in cranial arteries but did not induce migraine (Rahmann et al., 2008). Although these observations are intriguing, the interpretation of the clinical migraine efficacy results with CGRP receptor antagonists is not clear-cut. To better define potential central nervous system (CNS) sites of action for MK-3207, in vitro autoradiography mapping studies were conducted in rhesus monkey brain slices. The highest expression of [3H]MK-3207 binding sites was found in the cerebellum, brainstem, and meninges. In the cerebellum, high binding density is mainly located in the gray matter, with minimal binding in the white matter. In the brainstem, high-density binding sites are non-uniformly distributed to regions essential for processing noxious stimuli. The next question is does MK-3207 get into the brain at levels that can block the central CGRP receptor binding sites at clinically relevant plasma concentrations? The in vivo CSF level of MK-3207 was evaluated in cisterna magna-ported rhesus monkeys as a surrogate to the clinical experience. A CSF/plasma ratio after oral dosing was computed as an index of CNS penetrability. The CSF/plasma ratio was approximately 2% based upon C_{max} values of 20 nM CSF; 979 nM plasma, suggesting MK-3207 has brain penetration potential. At face value, one could conclude MK-3207 is CNS-penetrant and therefore significantly engaging central receptors. These data must be interpreted carefully because MK-3207 is a P-gp-substrate and therefore CSF levels cannot be equated to receptor occupancy.

In conclusion, we have identified a structurally novel, potent, selective, and orally bioavailable CGRP receptor antagonist. Although the exact mechanism linking CGRP and migraine is yet to be identified, it is quite clear that CGRP is not simply a migraine biomarker but also is a key player in migraine pathogenesis. Three CGRP receptor antagonists to
Salvatore et al.
date have demonstrated clinical efficacy for the treatment of migraine and considered together offer hope for a highly effective new therapy for migraine sufferers.

References


Address correspondence to: Dr. Christopher A. Salvatore, Merck Research Laboratories, WP26A-2000, West Point, PA 19486. E-mail: christopher_salvatore@merck.com.
Pharmacological Properties of MK-3207 [2-[(8R)-8-(3,5-difluorophenyl)-10-oxo-6,9-
diazaspiro[4.5]dec-9-yl]-N-[(2R)-2'-oxo-1,1',2',3'-tetrahydrospiro[indene-2,3'-pyrrolo[2,3-
b]pyridin]-5-yl]acetamide], a Potent and Orally Active CGRP Receptor Antagonist

The Journal of Pharmacology and Experimental Therapeutics

Partial list of enzyme and binding assays performed at MDS Pharma Services (Taipei, Taiwan). MK-3207 had no significant activity (IC$_{50}$ < 5 μM) against any receptor or enzyme system with the exception of the human CGRP receptor (IC$_{50}$ < 10 nM) and the human calcitonin receptor (IC$_{50}$ = 1.75 μM).

<table>
<thead>
<tr>
<th>Assay Name</th>
<th>Species (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adenosine A$_3$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adenosine A$_{2A}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adenosine A$_1$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic β$_2$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic β$_3$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic β$_1$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic α$_2$, Non-Selective</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Adrenergic α$_{2C}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic α$_{2A}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic α$_{2B}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Adrenergic α$_1$, Non-Selective</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Adrenergic α$_{1D}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Androgen (Testosterone) AR</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Angiotensin AT$_2$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Angiotensin AT$_1$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Atrial Natriuretic Factor (ANF)</td>
<td>Guinea pig (2)</td>
</tr>
<tr>
<td>Bombesin, Non-Selective</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Bradykinin B$_2$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Calcitonin</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Calcitonin Gene-Related Peptide CGRP$_{1}$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Calcium Channel L-Type, Benzothiazepine</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Calcium Channel L-Type, Dihydropyridine</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Calcium Channel L-Type, Phenylalkylamine</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Calcium Channel N-Type</td>
<td>Rat (2)</td>
</tr>
<tr>
<td>Cannabinoid CB$_2$</td>
<td>Human (2)</td>
</tr>
<tr>
<td>Cannabinoid CB$_1$</td>
<td>Human (2)</td>
</tr>
</tbody>
</table>
Chemokine CCR1 Human (2)
Chemokine CCR5 Human (2)
Chemokine CXCR2 (IL-8Rβ) Human (2)
Cholecystokinin CCK2 (CCKB) Human (2)
Cholecystokinin CCK1 (CCKA) Human (2)
Cyclooxygenase COX-1 Human (2)
Cyclooxygenase COX-2 Human (2)
Dopamine D3 Human (2)
Dopamine D2S Human (2)
Dopamine D4,4 Human (2)
Dopamine D1 Human (2)
Dopamine D5 Human (2)
Endothelin ETA Human (2)
Endothelin ETB Human (2)
Epidermal Growth Factor (EGF) Human (2)
Estrogen ERβ Human (2)
Estrogen ERα Human (2)
GABA_A, Chloride Channel, TBOB Rat (2)
GABA_A, Muscimol, Central Rat (2)
GABA_B, Non-Selective Rat (2)
Galanin GAL1 Human (2)
Galanin GAL2 Human (2)
Glucocorticoid Human (2)
Glutamate, AMPA Rat (2)
Glutamate, Kainate Rat (2)
Glutamate, NMDA, Agonism Rat (2)
Glutamate, NMDA, Glycine Rat (2)
Glutamate, NMDA, Phencyclidine Rat (2)
Glutamate, Non-Selective Rat (2)
Glycine, Strychnine-Sensitive Rat (2)
Histamine H2 Human (2)
Histamine H3 Human (2)
Histamine H1 Human (2)
Inositol Trisphosphate IP3 Rat (2)
Insulin Rat (2)
Leukotriene, Cysteinyl CysLT1 Human (2)
Lipoxygenase 5-LO Human (2)
Melanocortin MC$_3$  Human (2)
Melanocortin MC$_4$  Human (2)
Melanocortin MC$_5$  Human (2)
Melatonin MT, Non-Selective  Chicken (2)
Monoamine Oxidase MAO-A  Human (2)
Monoamine Oxidase MAO-B  Human (2)
Muscarinic M$_2$  Human (2)
Muscarinic M$_3$  Human (2)
Muscarinic M$_4$  Human (2)
Muscarinic M$_1$  Human (2)
Muscarinic M$_5$  Human (2)
Neuropeptide Y Y$_2$  Human (2)
Neuropeptide Y Y$_1$  Human (2)
Nicotinic Acetylcholine  Human (2)
Nitric Oxide Synthase, Inducible (iNOS)  Mouse (2)
Nitric Oxide Synthase, Neuronal (nNOS)  Rat (2)
Opiate $\delta$ (OP1, DOP)  Human (2)
Opiate $\mu$ (OP3, MOP)  Human (2)
Opiate $\kappa$ (OP2, KOP)  Human (2)
Orphanin ORL$_1$  Human (2)
Peptidase, CASP1 (Caspase 1)  Human (2)
Peptidase, CASP3 (Caspase 3)  Human (2)
Peptidase, CASP6 (Caspase 6)  Human (2)
Peptidase, CASP7 (Caspase 7)  Human (2)
Peptidase, CASP8 (Caspase 8)  Human (2)
Peptidase, CTSG (Cathepsin G)  Human (2)
Peptidase, ELA2 (Neutrophil Elastase 2)  Human (2)
Peptidase, Matrix Metalloproteinase-1 (MMP-1)  Human (2)
Peptidase, Metalloproteinase, Neutral Endopeptidase  Human (2)
Phosphodiesterase PDE1  Bovine (2)
Phosphodiesterase PDE2  Human (2)
Phosphodiesterase PDE3  Human (2)
Phosphodiesterase PDE4  Human (2)
Phosphodiesterase PDE5  Human (2)
Phosphodiesterase PDE6  Bovine (2)
Platelet Activating Factor (PAF)  Human (2)
Potassium Channel [$K_A$]  Rat (2)
Potassium Channel $[K_{ATP}]$ Hamster (2)
Potassium Channel $[SK_{CA}]$ Rat (2)
Prostanoid, Thromboxane A$_2$ (TP) Human (2)
Protein Serine/Threonine Kinase, MAPK14 (p38$\alpha$) Human (2)
Protein Serine/Threonine Kinase, MAPK3 (ERK1) Human (2)
Protein Serine/Threonine Kinase, PKA, Non-Selective Bovine (2)
Protein Serine/Threonine Kinase, PKC, Non-Selective Rat (2)
Protein Serine/Threonine Kinase, PRKCA (PKC$\alpha$) Human (2)
Protein Tyrosine Kinase, EGF Receptor Human (2)
Protein Tyrosine Kinase, Insulin Receptor Human (2)
Protein Tyrosine Kinase, SRC Human (2)
Protein Tyrosine Phosphatase, PTPRC (CD45) Human (2)
Purinergic P$_{2x}$ Rabbit (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_3$ Human (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_2$, Non-Selective Rat (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_{2c}$ Human (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_{2b}$ Human (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_4$ Guinea pig (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_1$, Non-Selective Rat (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_{1a}$ Human (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_{1b}$ Rat (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_6$ Human (2)
Serotonin (5-Hydroxytryptamine) 5-HT$_7$ Human (2)
Sodium Channel, Site 2 Rat (2)
Somatostatin sst5 Human (2)
Tachykinin NK$_2$ Human (2)
Tachykinin NK$_3$ Human (2)
Tachykinin NK$_1$ Human (2)
Thyrotropin Releasing Hormone (TRH) Rat (2)
Transporter, Adenosine Guinea pig (2)
Transporter, Choline Rat (2)
Transporter, Dopamine (DAT) Human (2)
Transporter, GABA Rat (2)
Transporter, Monoamine Rabbit (2)
Transporter, Norepinephrine (NET) Human (2)
Vasoactive Intestinal Peptide VIP$_1$ Human (2)
Vasopressin V$_{1A}$ Human (2)