The Free Radical Scavenger Edaravone Rescues Rats from Cerebral Infarction by Attenuating the Release of High-Mobility Group Box-1 in Neuronal Cells[

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Received December 6, 2008; accepted March 16, 2009

ABSTRACT

Edaravone, a potent free radical scavenger, is clinically used for the treatment of cerebral infarction in Japan. Here, we examined the effects of edaravone on the dynamics of high-mobility group box-1 (HMGB1), which is a key mediator of ischemic-induced brain damage, during a 48-h postischemia/reperfusion period in rats and in oxygen-glucose-deprived (OGD) PC12 cells. HMGB1 immunoreactivity was observed in both the cytoplasm and the periphery of cells in the cerebral infarction area 2 h after reperfusion. Intravenous administration of 3 and 6 mg/kg edaravone significantly inhibited nuclear translocation and HMGB1 release in the penumbra area and caused a 26.5 ± 10.4 and 43.8 ± 0.5% reduction, respectively, of the total infarct area at 24 h after reperfusion. Moreover, edaravone also decreased plasma HMGB1 levels. In vitro, edaravone dose-dependently (1–10 μM) suppressed OGD- and H$_2$O$_2$-induced HMGB1 release in PC12 cells. Furthermore, edaravone (3–30 μM) blocked HMGB1-triggered apoptosis in PC12 cells. Our findings suggest a novel neuroprotective mechanism for edaravone that abrogates the release of HMGB1.

Oxygen free radicals are produced under various pathological conditions, including ischemia/reperfusion or stroke. They play a crucial role in brain ischemic injury by exacerbating membrane damage, leading to neuronal death. Therapeutic strategies have been developed to reduce free radical-induced damage processes in ischemic brain injury. Several compounds have been developed, and some have also proceeded into clinical trials, including ebelyn, tirilazad, NXY-059, and edaravone (Wang and Shuaib, 2007). However, the trial with ebelyn and tirilazad in patients with stroke was recently terminated because there was no significant effect (van der Worp et al., 2002; Green and Shuaib, 2006). NXY-059 has been found to be ineffective for the treatment of acute ischemic stroke (van der Worp et al., 2002; Green and Shuaib, 2006).
Edaravone, which is a novel, potent, free radical scavenger, has been widely used for the treatment of acute cerebral infarction in Japan since 2001. Edaravone can quench hydroxyl radicals (·OH) and inhibit both ·OH-dependent and ·OH-independent lipid peroxidation (Watanabe et al., 2004). A clinical trial showed that the use of edaravone alone within 72 h after the onset of a stroke significantly reduced the infarct volume, producing sustained benefits during a 3-month follow-up period (Edaravone Acute Brain Infarction Study Group, 2003; Zhang et al., 2005), whereas clinical treatment within 24 h of stroke onset has been used for patients with lacunae, large-artery atherosclerosis, and cardioembolism cerebral infarctions. Although its neuroprotective role has been widely investigated, no direct mechanism has been proposed for the effect of edaravone on hypoxia-related lethal systemic inflammatory mediators in the brain.

High-mobility group box-1 (HMGB1) is a nonhistone nuclear protein of 30 kDa that is expressed in all eukaryotic cells. HMGB1 seems to play two distinct roles. Its first role is as a regulator of transcription in the maintenance of DNA function. Its second role is as a potent proinflammatory and cytokotoxic cytokine, contributing to the development of disseminated intravascular coagulation and organ failure (Ito et al., 2007). Neutralization of HMGB1 function, even well after the onset of infection, was shown to rescue mice from lethal sepsis (Wang et al., 1999). HMGB1 translocates to the outside of the nucleus in all eukaryotic cells upon necrosis and is released from macrophages through activation by lipopolysaccharide (LPS) and tumor necrosis factor-α. It is also released from cultured macrophages, monocytes, and neutrophils after H₂O₂ exposure, from neuronal cells after ischemia/reperfusion, and it is known to exacerbate brain tissue damage in rats (Kim et al., 2006; Tang et al., 2007). Elevated levels of HMGB1 in the serum of human patients with cerebral infarction have also been reported previously (Goldstein et al., 2006).

To the best of our knowledge, no reports have so far demonstrated a role for HMGB1 in the neuroprotective effect of edaravone on reactive oxygen species (ROS)-mediated ischemic injury. In the present study, we therefore investigated the hypothesis that edaravone plays a neuroprotective role through the abrogation of HMGB1 release triggered by H₂O₂ or OGD and ameliorates HMGB1-induced apoptosis in neuronal cells.

Materials and Methods

Bovine HMGB1 and anti-HMGB1 antibodies were obtained from Shino-Test (Kanagawa, Japan). As determined by Limulus endotoxin assays (Wako Pure Chemicals, Kyoto, Japan), the content of LPS in HMGB1 solutions (5 μg/mL) was found to be 3.11 ng/mL. Edaravone was a kind gift from Mitsubishi Wellpharma (Tokyo, Japan). The S100 mouse monoclonal antibody, mouse anti-oligodendrocyte monoclonal antibody, and mouse neuronal marker anti-MAP-2 monoclonal antibody were purchased from Millipore Bioscience Research Reagents (Temecula, CA). Anti-phospho (p)-extracellular signal-regulated kinase (ERK) 1/2, anti-ERK1/2, and anti-α-actin antibodies were purchased from Cell Signaling Technology Inc. (Danvers, MA). The MAPK or ERK kinase 1/2 inhibitor U0126 was obtained from Calbiochem (San Diego, CA). The anticytchrome c monoclonal antibody was purchased from BioSource International (Camarillo, CA). The H₂O₂ was a product of Santoku Chemical Industries Co., Ltd. (Tokyo, Japan). The AnaeroPuck and its anaerobic jar were purchased from Mitsubishi Gas Chemical Co. (Tokyo, Japan).

Methods

Unilateral Cerebral Hypoxic Ischemia Model. All animal procedures were conducted with the approval of the Animal Care Committee of the Ethics Board of the Institute of Laboratory Animal Sciences of Kagoshima University, Kagoshima, Japan. Adult male Wistar rats (230–260 g; 8 weeks old) were purchased from Japan SLc Inc. (Tokyo, Japan) and maintained under a 12-h light/dark cycle with free access to food and water. Rats were anesthetized intraperitoneally with chloral hydrate (400 mg/kg body weight). Mean tail arterial blood pressure (MABP) and cerebral blood flow (CBF) were monitored by a nonpreheating, noninvasive blood pressure monitor (MK-2000, Muromachi, Tokyo, Japan). Regional CBF (rCBF) was measured over the middle cerebral artery territory by laser Doppler flowmetry (model ALF21 laser flowmeter; Advance Co., Inc., Tokyo, Japan). A rectangular thin probe (7.5 × 3.5 × 1.0 mm, Type-CS; Unique Medical Co., Tokyo, Japan) for the laser Doppler flowmetry was slid through a small scalp incision into the natural pocket between the temporal muscle and the lateral side on the skull over the temporal cortex as reported previously (Harada et al., 2005). Blood gas and glucose levels were determined before and after occlusion of the middle cerebral artery (MCAO) via insertion of a polyethylene-50 tube via the left common carotid artery into the aorta (AVL OPTI critical care analyzer; AVL Scientific Corp., Roswell, GA; and Gluocard GT-1640; Aventis Pharma Ltd., Tokyo, Japan). Rectal and head temperatures were servo-controlled at 37.0°C using an electric blanket (KN-474; Natsume, Tokyo, Japan). The head temperature was determined before and after reperfusion by monitoring the right temporal muscle (model BAT-12 thermocouple thermometer; Physiostem Instruments, Inc., Clifton, NJ). Blood gas analysis and glucose level at 10 min, body temperature and blood pressure at 45 min, and head temperature at 70 min after occlusion were determined.

Experimental left MCAO was induced for 90 min by an intraluminal filament, as reported previously (Longa et al., 1989). Mice that were administered edaravone immediately after reperfusion showed a stronger reduction of infarct volume and a more rapid functional recovery compared with mice that were administered edaravone 3, 6, and 12 h after reperfusion (Zhang et al., 2005). Therefore, in the present study edaravone was administered immediately after reperfusion. Furthermore, we used two different doses of edaravone: 3 and 6 mg/kg. The dose of edaravone was administered as described previously (Nishi et al., 1989; Zhang et al., 2005).

Rats either received no treatment (n = 3) or cervical vein infusion (n = 6) of 3 or 6 mg/kg body weight edaravone immediately after reperfusion. The control sham-operated group (n = 6) underwent the same protocol except for the MCAO procedure. Reperfusion was established after 90 min by withdrawal of the filament. rCBF was measured by a laser Doppler flowmeter (model ALF21) before and during MCAO. Animals without significantly reduced blood flow (more than 20% of the preischemic baseline) during MCAO were excluded from further studies. At 2, 12, 24, and 48 h after reperfusion, the rats were anesthetized, sacrificed by perfusion of 100 ml of saline containing 10 U/ml heparin (Zhang et al., 2005), and their brains were excised. Coronal brain sections (2 mm in thickness) from bregma to 2 mm posterior were stained with 2,3,5-triphenyl tetrazolium chloride (TTC) to evaluate the infarct area; NIH Image 1.63 software (http://rsb.info.nih.gov/nih-image/) was used to calculate the area of the infarct. Sections were fixed with 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.4, at 4°C for histochemical analysis. The left lateral caudoputamen, lower parietal cortex, and...
upper frontoparietal cortex was examined as described by Häberg et al. (2001).

A neurological grading scale (0–4) was used according to Menzies et al. (1992): 0, no apparent deficits; 1, right forelimb flexion; 2, decreased grip of the right forelimb while tail pulled; 3, spontaneous movement in all directions (right circling only if pulled by tail); and 4, spontaneous right circling.

**HMGB1 ELISA.** Brain tissue (120 μg/g) was gently homogenized in 10 mM HEPES, 10 mM KCl buffer with 0.08% Nonidet P40, 0.1 mM EDTA, 0.5 mM diethiothreitol (DTT), and 0.5 mM phenylmethylsulfonyl fluoride, and the soluble fraction derived from the cytoplasm was kept at −80°C until further testing. Serum was obtained by centrifuging (5000 rpm for 5 min) blood samples immediately after collection and storing the supernatant at −80°C until further analysis. All samples were examined for HMGB1 content using an enzyme-linked immunosorbent assay (ELISA) kit (Shino-Test, Tokyo, Japan).

**Immunohistochemistry.** Immunohistochemistry was performed by the indirect immunoperoxidase method. After deparaffinization and hydration, endogenous peroxidase was blocked with methanol containing 0.3% H₂O₂ for 10 min. After three 10-min rinses with 50 mM phosphate-buffered saline (PBS, pH 7.6), the sections were blocked with 10% nonfat milk for 20 min at room temperature and then individually incubated in anti-HMGB1 rabbit antibody (10 μg/ml) overnight at 4°C. After three 10-min PBS rinses, sections were incubated in goat anti-rabbit IgG conjugated to peroxidase-labeled dextran polymer (EnVision; Dako North America, Inc., Carpinteria, CA) for 60 min. After rinsing with PBS, immunoreactivity was visualized with diaminobenzidine/peroxide. As negative controls, isotypic nonimmune serum (1:200 in PBS) was added (final concentration, 0.5 mg/ml) overnight at 4°C. After three 10-min PBS rinses, sections were incubated with an Axioskop microscope (Carl Zeiss, Oberkochen, Germany).

**Cell Culture.** Rat PC12 cells were obtained from the American Type Culture Collection (Manassas, VA) and maintained in Dulbecco’s modified Eagle’s medium supplemented with 10% horse serum. Type Cell Culture Collection (Manassas, VA) and maintained in Dulbecco’s modified Eagle’s medium supplemented with 10% horse serum and 1% FBS (OGD medium was prepared as described previously (Hillion et al., 2005). In brief, the original medium was removed, cells were rinsed twice with glucose-free RPMI 1640 medium (Invitrogen, Carlsbad, CA). The membrane was washed with 5% nonfat milk in Tris-buffered saline containing 0.02% Tween 20 (TBST) for 1 h, followed by incubation with the respective primary antibodies (10 μg/ml anti-HMGB1 antibody, anti-ERK1/2 antibody diluted 1:1000, or 2 μg/ml anti-cytochrome c antibody) in TBST containing 1% nonfat milk for 3 h at room temperature. After three washes with TBST, membranes were incubated with horseradish peroxidase-conjugated anti-rabbit IgG polyclonal antibody (Santa Cruz Biotechnology, Inc.) or horseradish peroxidase-conjugated anti-mouse IgG polyclonal antibody diluted 1:3000 in TBST containing 2.5% nonfat dry milk for 1 h. As negative controls, isotypic nonimmune serum (1:200 in PBS) was added (final concentration, 0.5 mg/ml) overnight at 4°C. After three 10-min PBS rinses, sections were incubated with an Axioskop microscope (Carl Zeiss).

**Immunofluorescence Analysis.** PC12 cells (1 × 10⁶/ml) were cultured on Lab-Tek chamber slides (Nalge Nunc International, Rochester, NY) with or without 40 μM H₂O₂ for 8 h and fixed with 4% paraformaldehyde for 5 min (Kawahara K et al., 1999). Slides were incubated with blocking buffer containing 1% bovine serum albumin and PBS/0.02% Tween 20 (PBST) for 1 h, followed by an incubation of 30 min with anti-HMGB1 rabbit polyclonal antibody (1 μg/ml) for 1 h. Subsequently, cells were washed with PBST and incubated with fluorescein isothiocyanate-labeled anti-rabbit IgG (diluted 1:50) for 20 min. Slides were examined under an Axioskop microscope (Carl Zeiss).

Sections from rat brain tissue were immunostained as described above. Slides were incubated with anti-HMGB1 rabbit polyclonal antibody (1 μg/ml), mouse anti-S100 monoclonal antibody (1:300), mouse anti-oligodendrocyte monoclonal antibody (1:500), and mouse anti-MAP-2 monoclonal antibody (1:200) overnight at 4°C. After washing with PBST, slides were incubated with fluorescein isothiocyanate-labeled anti-rabbit IgG (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) and Alexa Fluor 546-labeled anti-mouse IgG diluted 1:200 in PBS for 1 h. After washing with PBST, the sections were counterstained with 4',6-diamidino-2-phenylindole (1:500; Wako Pure Chemicals) for 5 min. After washing, the slides were examined under an Axioskop microscope (Carl Zeiss).

**Sample Preparation for Western Blot Analysis.** HMGB1-released samples were prepared according to our previous studies (Taniguchi et al., 2003). In brief, PC12 cells (8 × 10⁵/well) were exposed to OGD (2–4 h) or stimulated with 20 to 80 μM H₂O₂ for 12 h at room temperature. Edaravone was either added to the cultures 30 min before OGD treatment, or cells were incubated simultaneously with edaravone, H₂O₂, and HMGB1. U0126 was preincubated for 1 h before H₂O₂ exposure. The supernatant (1 ml) was incubated with 50 μl of heparin-Sepharose 6B beads overnight at room temperature to bind HMGB1. Heparin beads bound to HMGB1 were washed three times with PBS, and SDS sample buffer (62.5 mM Tris-HCl, 2% SDS, and 10% glycerol) was added to the beads. For MAPK activation, cell lysates of stimulated PC12 cells (5 × 10⁵/dish) were obtained by adding 120 μl of SDS sample buffer and lysed as described previously (Tancharoen et al., 2005). To evaluate mitochondrial cytochrome c activation, 50 μg of protein from the cytoplasmic fraction of HMGB1-treated samples was subjected to MAPK activation as described above.

**Western Blot Analysis.** All samples were subjected to 12% SDS-polyacrylamide gel electrophoresis (with the exception of cytochrome c, which was subjected to 15% SDS-polyacrylamide gel electrophoresis) and then transferred to a nitrocellulose membrane (GE Healthcare, Chalfont St. Giles, Buckinghamshire, UK). The membrane was blocked with 5% nonfat milk in Tris-buffered saline containing 0.02% Tween 20 (TBST) for 1 h, followed by incubation with the respective primary antibodies (10 μg/ml anti-HMGB1 antibody, anti-ERK1/2 antibody diluted 1:1000, or 2 μg/ml anti-cytochrome c antibody) in TBST containing 1% nonfat milk for 3 h at room temperature. After three washes with TBST, membranes were incubated with horseradish peroxidase-conjugated anti-rabbit IgG polyclonal antibody (Santa Cruz Biotechnology, Inc.) or horseradish peroxidase-conjugated anti-mouse IgG polyclonal antibody diluted 1:3000 in TBST containing 2.5% nonfat dry milk for 1 h. The membrane was washed twice, and immunoreactive bands were visualized using the enhanced chemiluminescence detection system (GE Healthcare) and measured using Image 1.63 software (National Institutes of Health).

**Nuclear Staining for the Assessment of Apoptosis.** Morphological changes in the nuclear chromatin of cells undergoing apoptosis were detected by staining with Hoechst 33258 (Sigma-Aldrich, Gillingham, UK). In brief, cells were treated for 24 h at room temperature with 3 μg/ml HMGB1 in the presence or absence of edaravone (10 μM), and then cells were washed with PBS and fixed for 30 min with 4% paraformaldehyde. After additional washes with PBS, cells were incubated with Hoechst 33258 (final concentration, 10 μg/ml) for 10 min. Nuclear morphology was examined using an Axioskop microscope. Intact blue nuclei were considered to represent viable cells, and condensed/fragmented nuclei were considered to represent apoptotic cells.

**MTT Assay for Determination of Cell Viability.** PC12 cells were seeded at a density of 2 to 3 × 10⁵ cells per well in 96-well dishes. Cells were treated with HMGB1 (0.5–4 μg/ml) in the presence or absence of edaravone (10 or 30 μM) for 24 h. Subsequently, 3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide (MTT; Sigma-Aldrich) was added (final concentration, 0.5 mg/ml) to each test dish (20 μl per well) and incubated for 2 h at 37°C. After incubation, 20 μl of a 1% SDS, pH 4.8, solution was added to each well, and the crystals were dissolved for 16 h. Absorbance values at a test wavelength of 570 nm and a reference wavelength of 630 nm were determined with an automatic ELISA reader (Immuno Mini NJ-2300, Inter Med, Tokyo, Japan).

**Statistical Analysis.** All experiments were performed at least three times, and representative examples are shown. Experimental values are given as the mean ± S.D. In HMGB1-stained sections, stained cells in three predefined areas (each 0.25 mm²) were counted. Densitometric analysis of protein signals was assessed with the Student’s t test. One-way analysis of variance followed by the post hoc Fisher’s protected least-significant difference test was used to
Results

Physiological Variables. The physiological variables observed in the different groups of rats are shown in Table 1. No significant difference was observed between sham-operated and MCAO animals. Furthermore, administration of edaravone had no significant effect on these variables, as described previously (Zhang et al., 2005). In previous reports, edaravone had no significant effect on body temperature and blood pressure at 45 min, and head temperature at 70 min after occlusion were determined. For comparison of physiological variables, Table 1 describes previously (Zhang et al., 2005). In previous reports, edaravone had no significant effect on these variables, as described and MCAO animals. Furthermore, administration of edaravone i.v. had no significant effect on body temperature and blood pressure at 45 min, and head temperature at 70 min after occlusion were determined. For comparison of physiological variables.

Protein Expression Profile of HMGB1 in Nontreated or Edaravone-Treated Rats. We investigated the expression levels and cellular distribution of HMGB1 2 to 48 h after initiation of ischemic reperfusion in brain tissue of both edaravone-treated and nontreated rats using immunohistochemical analysis. Strong HMGB1 immunoreactivity was observed in the cytoplasm of neurons and glial cells in the infarct area 2 h after reperfusion initiation in the nontreated group. After 12 h of reperfusion, HMGB1 immunostaining was weaker, and small immunopositive cell bodies were found (Supplemental Fig. 1c). Moreover, 24 and 48 h (data not shown) after reperfusion, the HMGB1 signal was further decreased, indicating that nuclear HMGB1 had translocated to the cytoplasm and subsequently to the extracellular space within the infarct area. The infarct area in the brains of edaravone-treated rats showed a marked decrease in HMGB1 immunoreactivity in the cytoplasm and an increased immunoreactivity in cell nuclei 2 h after reperfusion. However, edaravone treatment failed to block the HMGB1 expression profile in nontreated rats. As shown in Fig. 2B, edaravone caused a significant reduction (34 ± 2.2%; \( P < 0.01 \)) of the infarct area at 6 mg/kg but not at 3 mg/kg in the lateral caudoputamen and lower parietal cortex. However, in the upper frontoparietal cortex, a dose of edaravone as low as 3 mg/kg caused a significant reduction (60.6 ± 6%; \( P < 0.01 \)) and a sharp reduction (87.9 ± 4.5%; \( P < 0.005 \)) in infarct area was observed in rats that were administered 6 mg/kg edaravone compared with the nontreated group (vehicle, 0.9% NaCl). Our results are consistent with a previous report (Zhang et al., 2005). Moreover, a significant lower neurological score was observed in the edaravone (3 mg/kg)-treated group compared with the nontreated group [2.3 (2–3)] (\( P < 0.05 \)).

We next examined HMGB1-positive cells in the lateral caudoputamen, lower parietal cortex, and the upper frontoparietal cortex of rats that were administered edaravone (3 or 6 mg/kg i.v.) by double-immunofluorescent labeling using HMGB1 and S100 antibodies (see photographs in Fig. 2B). No immunofluorescence data are shown for edaravone at 3 mg/kg. It has been reported that the S100 protein is localized in the cytoplasm and/or nucleus of neuronal and non-neuronal cells (Donato, 1999; Gonzalez-Martinez et al., 2003). S100 staining was used to identify the subcellular localization of HMGB1 and to determine the cell type of

| TABLE 1 |
| Comparison of physiological variables |
| Blood gas analysis and glucose level at 10 min, body temperature and blood pressure at 45 min, and head temperature at 70 min after occlusion were determined. For experimental group MCAO edaravone (–), \( n = 6 \) (24 h); for MCAO edaravone (3 mg), \( n = 6 \) (24 h); and for MCAO edaravone (6 mg), \( n = 15 \) (6, 12, 48 h; \( n = 3 \) and 24 h; \( n = 6 \)). Values are mean ± S.D. |

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<th></th>
<th>–</th>
<th>3 mg</th>
<th>6 mg</th>
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<tr>
<td><strong>Before occlusion</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MABP (mm Hg)</td>
<td>61 ± 12</td>
<td>62 ± 13</td>
<td>69 ± 10</td>
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<tr>
<td>PaO2 (mm Hg)</td>
<td>62 ± 8</td>
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<td>PaCO2 (mm Hg)</td>
<td>53 ± 8</td>
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<td>pH</td>
<td>7.34 ± 0.06</td>
<td>7.35 ± 0.05</td>
<td>7.37 ± 0.05</td>
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<td>Ht (%)</td>
<td>39 ± 1</td>
<td>38 ± 8</td>
<td>39 ± 4</td>
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<tr>
<td>Glucose (mg/dl)</td>
<td>212 ± 9</td>
<td>211 ± 52</td>
<td>246 ± 60</td>
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<td>Head temp. (°C)</td>
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<td>36.9 ± 0.1</td>
<td>36.9 ± 0.1</td>
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<tr>
<td>Body temp. (°C)</td>
<td>37.0 ± 0.1</td>
<td>37.0 ± 0.1</td>
<td>37.0 ± 0.1</td>
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<tr>
<td><strong>After occlusion</strong></td>
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<tr>
<td>MABP (mm Hg)</td>
<td>75 ± 17</td>
<td>69 ± 20</td>
<td>77 ± 5</td>
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<td>PaO2 (mm Hg)</td>
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<td>76 ± 5</td>
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<td>7.30 ± 0.06</td>
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<td>Ht (%)</td>
<td>41 ± 0</td>
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<tr>
<td>Glucose (mg/dl)</td>
<td>154 ± 46</td>
<td>154 ± 99</td>
<td>179 ± 26</td>
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<td>Head temp. (°C)</td>
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<td>36.9 ± 0.1</td>
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<tr>
<td>Body temp. (°C)</td>
<td>37.0 ± 0.0</td>
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Ht, mean hematocrit; PaO2, mean partial pressure of oxygen; PaCO2, mean partial pressure of carbon dioxide.
HMG1-positive cells. In sham-operated rats, we observed nuclear localization of HMG1 (Fig. 2A, b and d) and cytoplasmic localization of S100 (Fig. 2A, c and d). Anti-MAP-2 and anti-oligodendrocyte antibody staining revealed similar levels of ischemic cerebral damage in the contralateral areas of the edaravone-treated and nontreated groups (data not shown). At 24 h after reperfusion, neither HMG1 nor S100 staining was evident in the lateral caudoputamen and lower parietal cortex, regardless of edaravone treatment (Fig. 2A, f–h and j–l). However, in the nontreated group, HMGB1 (Fig. 2A, n and p) and S100 (Fig. 2A, o and p) colocalized in the cytoplasm and periphery of the nucleus throughout the upper frontoparietal cortex. It is noteworthy that, in this region, no cytoplasmic translocation of HMGB1 was observed in the edaravone-treated group (Fig. 2A, r–t). Furthermore, as shown in Fig. 2B, a significantly increased number of cytoplasmic-HMGB1-positive cells in nontreated groups (45.2 ± 5.4%) compared with edaravone-treated groups (15.3 ± 2.5 versus 45.2 ± 5.4%, P < 0.0001) for edaravone at 3 mg/kg and 2.1 ± 1.9 versus 15.3 ± 2.5%, P < 0.005) for edaravone at 6 mg/kg) indicated that more HMGB1 was shuttled into the cytoplasm and cell periphery of edaravone-nontreated brain tissue 24 h after reperfusion.

We also confirmed the release of HMGB1 in the tissue and serum 24 h after reperfusion by ELISA. A significant increase in HMGB1 (25 ± 4 versus 10.4 ± 1.7 μg/g tissue; P < 0.05) was detected in the upper frontoparietal cortex area of nontreated rats compared with the sham-operated group (10.4 ± 1.7 μg/g tissue), whereas the HMGB1 release was significantly reduced (58.4 ± 6%) in the upper frontoparietal cortex area (10.4 ± 2.5 versus 25 ± 4 μg/g tissue; P < 0.05) of edaravone-treated (6 mg/kg) rats. No significant release of HMGB1 was observed in brain tissue collected from the lateral caudoputamen and lower parietal cortex area compared with sham-operated brain tissue. Moreover, we found a remarkable up-regulation of serum HMGB1 level in nontreated rats 24 h after reperfusion, an effect that was significantly abrogated by administration of either 3 or 6 mg/kg edaravone (Fig. 2C).

Edaravone Inhibits OGD and Reactive Oxygen (H2O2)-Induced HMGB1 Release in PC12 Cells. PC12 cells have been used previously to study neuronal injury in vitro (Hillion et al., 2005), and we therefore used this model system to confirm the in vivo results presented above. We examined whether HMGB1 was released in PC12 cells after different stresses, such as OGD and reactive oxidants (H2O2). Subsequently, we examined whether edaravone could block HMGB1 release. PC12 cells were treated with or without edaravone and then exposed to OGD for 2 to 4 h. As shown in Fig. 3A, cells exposed to OGD for 3 h triggered a marked release of HMGB1 into the culture medium, and this release was further up-regulated by 4 h. Edaravone (3–10 μM) blocked this effect for 3 h strikingly.

ROS that are generated after hypoxia include superoxide, H2O2, •OH, and singlet oxygen (Gechev et al., 2006). In the present study, cells that were exposed to H2O2 released significant levels of HMGB1 in a dose-dependent manner, and, as expected, this release was dose-dependently blocked by edaravone (3 and 10 μM) (P < 0.05) (Fig. 3B). However, no decrease in cell viability was observed after 24 h of exposure to up to 80 μM H2O2 as revealed by MTT assay (data not shown).

We next used fluorescence microscopy to examine the translocation of HMGB1 from the nucleus to the cytoplasm in PC12 cells in response to H2O2. Active expression of HMGB1 was observed in the periphery of the nucleus as well as in cytoplasmic regions 8 h after stimulation of cells with 40 μM H2O2 (Fig. 3C, d–f), whereas most HMGB1 was localized in the nucleus of control cells (Fig. 3C, a–c). Consistent with the in vivo results, 10 μM edaravone treatment (Fig. 3C, g–i) was found to significantly inhibit nuclear translocation of HMGB1 induced by H2O2 in vitro.

Several lines of evidence indicate that MAPKs play an important role in the active release of HMGB1 and are essential in the hypoxic signaling cascade (Haddad, 2004). Therefore, we next examined whether nontoxic doses of H2O2 triggered the activation of MAPKs in PC12 cells. As shown in Fig. 3D, H2O2 dose-dependently triggered the activation of p-ERK1/2 but not p-c-Jun NH2-terminal kinase 1/2 and p-p38 MAPK after doses of up to 100 μM H2O2 (data not shown). We then evaluated the functional role of ERK1/2 activation on H2O2-induced HMGB1 release by pretreating cells with U0126, which is a potent and specific pharmacological inhibitor of ERK kinase 1/2 (an upstream signaling molecule of ERK1/2). We observed that U0126 almost completely blocked the activation of p-ERK1/2 (data not shown) and the release of HMGB1 in response to H2O2 (Fig. 3E).

Edaravone Abrogates HMGB1-Induced Apoptosis in PC12 Cells. PC12 cells are a useful neuron-like model for studying programmed cell death (Mills et al., 1996). To evaluate the role of endogenously produced HMGB1 during cerebral ischemic reperfusion or under experimental hypoxic conditions, we investigated the effect of exogenous administration of HMGB1 on the viability of PC12 cells using recombinant HMGB1. For this purpose, PC12 cells
viability was observed at 4 g/ml. Significantly decreased at 2 g/ml. H9262 viability was initiated at 1 g/ml, which was further significantly decreased at 2 g/ml. A dramatic reduction of cell viability was observed at 4 g/ml (Fig. 4A). Furthermore, HMGB1 but not heat-treated HMGB1 significantly decreased cell viability (Supplemental Fig. 2). The LPS concentration in HMGB1 solutions was 0.581 ng/ml. The viability of PC12 cells did not significantly decrease after treatment with LPS at 10,000 ng/ml (Fig. 4A). Furthermore, HMGB1 but not heat-treated HMGB1 significantly decreased cell viability (Supplemental Fig. 2). The LPS concentration in HMGB1 solutions was 0.581 ± 0.058 ng/ml. The viability of PC12 cells did not significantly decrease after treatment with LPS at 10,000 ng/ml for 24 h, as reported previously (Huang et al., 2002).

Next, we examined the role of edaravone (10–30 μM) on HMGB1-induced cell death. As shown in Fig. 4A (right), edaravone significantly rescued HMGB1-induced cell death in a dose-dependent manner. Moreover, HMGB1-induced cell death was followed by notable features of apoptosis, such as cytoplasmic and nuclear shrinkage, chromatin condensation, and nuclear fragmentation (Fig. 4B). Because mitochondrial-mediated apoptotic cell death involves the release of cytochrome c into the cytosol, we investigated whether this accompanied HMGB1-induced cell death and whether it could be abrogated by edaravone. A significant release of cytochrome c was observed by Western blotting of the cytosolic protein extract 20 h after the exposure of PC12 cells to HMGB1, which was markedly suppressed (by approximately 50%) by 10 μM edaravone (Fig. 4C). These findings indicate that edaravone is a powerful regulator of the HMGB1 cascade in apoptosis.

Discussion

The present study demonstrates that HMGB1 is released from neuronal cells after ischemia and that this release can lead to brain injury via the ROS-ERK1/2-HMGB1 signal transduction pathway. In in vitro and in vivo experiments, the synthetic free radical scavenger edaravone significantly suppressed HMGB1 release, infarct size, neurological deficit, and HMGB1-induced cell death.

In MCAO, the ipsilateral lateral caudoputamen and lower parietal cortex are rendered severely ischemic, whereas the upper frontoparietal cortex represents moderately ischemic tissue. The former is often referred to as the ischemic core, and the latter as the penumbra (Håberg et al., 2001). Necrosis occurs in the core, whereas apoptosis occurs in the penumbra (Benchoua et al., 2001).

We observed that HMGB1 translocated from the nucleus to the cytoplasm in the upper frontoparietal cortex (penumbra), and this translocation did not occur in neurons from healthy control rats or rats treated with edaravone. This implies that HMGB1 is a highly motile protein that shuttles to the cytosol via nuclear pores, that it can be released from the brain...
under pathological conditions (Kim et al., 2005), and that such a release is efficiently inhibited by edaravone.

HMGB1 was increased in the serum of rats 16 to 32 h after LPS stimulation (Wang et al., 1999), implying that HMGB1 is a “magic bullet” targeting inflammatory sites, especially in late-phase inflammation and sepsis. HMGB1 has been identified as an early mediator of hemorrhage after acute lung injury (Kim et al., 2005) and hepatic injury after liver ischemia/reperfusion (Tsung et al., 2005). Furthermore, OGD induced HMGB1 release in cultured neurons (Qiu et al., 2008). It is noteworthy that we found that edaravone significantly suppressed HMGB1 release in OGD-induced PC12 cells. Our results are in agreement with a previous report describing the therapeutic efficacy of edaravone on oxidative damage early after ischemic insult (Zhang et al., 2005).

It is well established that free radicals are produced in the brain during ischemia (Fagan et al., 2005). We found that PC12 cells stimulated with nontoxic doses of H$_2$O$_2$ showed a significant increase in HMGB1 release (Fig. 3B) as was shown previously for monocytes and macrophages (Tang et al., 2007). Extracellular HMGB1 induces receptor for advanced glycation end products (RAGE) signaling, which induces elevated free radicals (Ding et al., 2007), followed by HMGB1 activation of NAD(P)H oxidase and increased ROS production in a toll-like receptor 4-dependent manner (Fan et al., 2007). Furthermore, HMGB1 is released from necrotic...
Edaravone abrogates HMGB1-induced apoptosis in PC12 cells. 

**A** Cell viability was measured by MTT assay 24 h after exposure to the indicated concentrations of HMGB1 and edaravone. Data shown are percentages of control values ± S.D. (+, P < 0.0001). B, Hoechst 33258 staining of PC12 cells treated with HMGB1 in the presence or absence of 10 μM edaravone. The arrowhead indicates fragmented nuclei that are a characteristic feature of apoptotic cell death. Original magnification, ×100. C, edaravone suppresses HMGB1-induced cytochrome c release from mitochondria as shown by Western blot analysis. β-Actin (bottom blot) was used as loading control. The bottom panel shows a graphical representation of the top blot after densitometric analysis. Data shown are relative to the highest cytochrome c signal, which was set to 100. +, P < 0.05.

**Fig. 4.** Edaravone abrogates HMGB1-induced apoptosis in PC12 cells.

Our findings also support the rescue of ischemic/reperfusion-induced apoptosis by edaravone in human choriocarcinoma cells (Yung et al., 2007) and nitric oxide-induced apoptosis in cultured rat astrocytes (Kawasaki et al., 2007), indicating that the interaction between ROS and HMGB1 is important in the pathological mechanism. We propose that ROS triggers the first “bullet” in the ischemic core and subsequently induces the formation of edaravone-suppressible ROS-HMGB1 signaling that causes neuronal damage, increases infarct area, and increases apoptosis in the penumbra, and a continuation of these processes causes severe damage to the brain.

Our study demonstrated a strong association between HMGB1 release and antioxidant activity of edaravone. First, HMGB1 has oxidation-sensitive unpaired cysteines (Cys) at positions 23, 45, and 106 (Sahu et al., 2008). The function of...
the Cys23-Cys45 pair in HMGB1 A-domain is essential for HMGB1 function in the nucleus, and the oxidation of Cys106 in B-domain is caused by HMGB1 release by ROS such as H$_2$O$_2$ (Sahu et al., 2008). We also found that HMGB1 release is caused by oxidative stress in ischemic condition and H$_2$O$_2$-stimulated PC12 cells mediated by ERK1/2 pathway in our condition, suggesting that its release may depend on oxidized conditions. Thus, these evidences indicated that oxidized HMGB1 may be released from apoptosis and enhanced the inflammatory events, demonstrating that HMGB1 release by oxidative stress might be prevented by antioxidant activity of edaravone. In contrast, RAGE, which is the specific receptor of HMGB1, is a multifunctional immunoglobulin superfamily, and its signaling cascade induces a strong ROS (Yamagishi and Takeuchi, 2004), suggesting that inhibition of HMGB1-stimulated cell death might be caused by antioxidant effects of edaravone. However, we are currently unable to elucidate the underlying mechanisms such as the direct effect of edaravone as HMGB1 and/or RAGE antagonist. Therefore, we would prefer to address this matter in our future studies.

HMGB1 short hairpin RNA, transfected 24 h before MCAO, significantly decreased infarct volumes and suppressed microglial activation (Kim et al., 2006). Furthermore, an anti-HMGB1 monoclonal antibody ameliorated brain infarction induced by transient ischemia in rats (Liu et al., 2007). Together with our findings, these results indicate a key role for HMGB1 in both damaged and delayed inflammatory processes in the posts ischemic brain (Kim et al., 2006). Our in vitro and in vivo findings showed that edaravone significantly reduces ischemic-induced cerebral infarction by abrogating the HMGB1 release, and ameliorating HMGB1-induced apoptosis in neuronal cells, suggesting that edaravone could be used in the treatment of ischemia-induced cerebral infarction. However, further studies are needed in ischemic neuronal cell models to explore pharmacological strategies against expanding cerebral infarction, traumatic brain injury, and intracerebral hemorrhage in which ROS act as key mediators (Dohi et al., 2007; Nakamura et al., 2008).

Acknowledgments

We thank N. Uto, T. Nagasato, and H. Sameshima for excellent technical assistance.

References


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