Impact of High and Low Folate Diets on Tissue Folate Receptor Levels and Antitumor Responses Toward Folate-Drug Conjugates

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ABSTRACT
Herein, we present a detailed analysis on the effects of feeding laboratory mice both high and low folic acid (folate)-containing diets as related to associated changes in serum and red blood cell (RBC) folate levels, tissue-derived folate receptor levels, and the ability of folate-drug conjugates to bind and effectuate activity against folate receptor (FR)-positive tumor xenografts. Our data show that serum and RBC folate concentrations sharply drop immediately after mice are switched to low folate diets; however, both parameters reach steady-state, “human-like” levels after 6 weeks. Interestingly, tissue-related folate binding capacities were also lowered during the dietary modulation period, whereas the net uptake of a radiolabeled folate conjugate was simultaneously increased 2.6- and 5-fold in FR-positive kidney and tumor tissue, respectively. Finally, the performances of several clinically and preclinically relevant folate-drug conjugates were evaluated against tumors in mice that were fed high or low folate diets. Except when administered at a dose level 6-fold less than that required to saturate endogenous FRs, no significant loss of antitumor activity was observed. From these findings, we conclude that lowering the dietary intake of folates in mice has little impact on the biological activity of repetitively dosed folate-targeted agents but that low folate diet regimens will reduce serum and RBC folate levels down to levels that more closely approximate the normal human ranges.

Folic acid (FA, folate, vitamin B9) is an essential nutrient required by all living cells for proper metabolic maintenance of 1-carbon pathways as well as for nucleotide biosynthesis (Clifford et al., 1998). Hence, acquisition of folates is extremely important for maintaining the viability of both normal and proliferating (e.g., cancer) cells. Many have studied the impact of modulating dietary FA intake in rodents on the activities of various chemotherapeutic agents, including 5-fluorouracil, mitomycin C, cytostine arabinoside, doxorubicin, cyclophosphamide, and the antifolates aminopterin, methotrexate, dideazatetrahydrofolic acid, and pemetrexed (Mendelsohn et al., 1996; Raghunathan et al., 1997; Worzalla et al., 1998; Branda et al., 2002). In some cases, lowering the intake of folate was found to potentiate antitumor activity, whereas in others (e.g., antifolates), greater toxicities were observed compared with when animals were fed high folate diets. Such preclinical modeling had proved to be, in part, useful for guiding the clinical development of some agents (Azzoli et al., 2007); however, increasing dietary folate levels has also been met with problems, since rapid hematologic and clinical relapses were noted in some leukemia patients (Lennard et al., 1986).

At the cellular level, uptake of folates mainly occurs by two independent transport mechanisms, namely, 1) transport of reduced folates by a carrier-mediated, low-affinity but high-capacity anion transport protein (Antony, 1992); or 2) by an active endocytic process mediated by the high-affinity membrane-bound glycoprotein receptor, commonly referred to as the folate-binding protein (FBP) or the folate receptor (FR) (Kamen et al., 1988; Antony, 1996). The FR is a known biomarker protein that is expressed in high quantities by many primary and metastatic cancers (Parker et al., 2005) but not on most normal cells (Parker et al., 2005). Recogniz-
ing this attribute, we have focused our efforts on the development of small-molecule targeting systems, based on FA, to deliver covalently linked therapeutic and diagnostic imaging agents to cells that express the FR (Leamon and Low, 1991; Leamon et al., 2002; Leamon and Reddy, 2004; Reddy et al., 2004). In recent years, our laboratory and research collaborators have reported on the preclinical pharmacology results for a variety of FA-drug conjugates, including those constructed with mitomycin C, desacetylvinblastine monohydrize, epothilone B, and the maytansinoid DM1 (Leamon et al., 2005, 2006, 2007a,b; Reddy et al., 2006, 2007a,b; Covello et al., 2008). In each case, strong evidence was presented for FR-mediated cytotoxicity and in vivo antitumor effect. Importantly, three related agents (EC145, EC0225, and BMS-753493 a folate-epothilone conjugate) are currently being evaluated for safety and activity in multiple phase 1 and 2 clinical trials.

Supportive preclinical data for each of the aforementioned compounds were collected using animals that had been acclimated on a low folate diet for two or more weeks before test article administration. This technique is commonly done because normal rodent laboratory chow contains high levels of FA (6 mg/kg chow) that allow for excessively high serum folate concentrations to persist (Mathias et al., 1996). Importantly, such supraphysiological levels of FA in the sera of mice were predicted to decrease the uptake of FA-based radiodiagnostic agents in FR-expressing tumor tissue; therefore, protocols using short dietary modulation with non-FA-supplemented chows were developed for the purpose of lowering mouse serum folate levels down to the normal human range (Mathias et al., 1996). As a consequence of that finding, nearly all in vivo studies reported for FA-drug conjugates have involved the use of these low FA diets. What is not known, however, is whether the antitumor activities of frequently administered FA-drug conjugates would truly be compromised in mice that were maintained on the more common high folate-containing diets.

In our current study, we present a detailed analysis of modulating dietary folate levels in laboratory mice on the effects of serum and red blood cell (RBC) folate levels, tissue-derived folate receptor levels, and the ability of both clinically and preclinically relevant FA-drug conjugates to bind and effectuate activity against FR-positive tumor xenografts.

**Materials and Methods**

**Materials.** Folate-free RPMI 1640 medium and PBS were obtained from Invitrogen (Carlsbad, CA). Bovine soluble milk FBP was purchased from Scirpps Laboratories (San Diego, CA). [3H]Thymidine (71.9 Ci/mmoll and [3H]folic acid (30 Ci/mmoll) were purchased from Moravek Biochemicals (Brea, CA). Sera samples were obtained from Harlan (Indianapolis, IN; BALB/c mice, Lewis rats, and New Zealand White rabbits), BAS Bioanalytical Systems (Evansville, IN; beagle dog and cynomolgus monkey), and human donors. All test diets were produced by Harlan Teklad (Madison, WI) and included 1) Teklad Diet 2188S Global 16% Protein Rodent Diet (Sterilizable); 2) Test Diet 00434, which is the standard chow devoid of added folic acid; and 3) TD 00633, the folate-free chow supplemented with 0.5 mg/kg leucovorin. All other common reagents were purchased from Sigma-Aldrich (St. Louis, MO) or other major suppliers.

**Test Articles.** EC20, EC145, and EC0305 were produced by Endocyte, Inc. (West Lafayette, IN). Their syntheses, purifications, and analytical characterizations have been described in detail previously (Leamon et al., 2002, 2005, 2006; Vlahov et al., 2006, 2007, 2008).

**Mouse Animal Housing/Conditions.** Female BALB/c or nu/nu mice (4–5-week-old; BALB/c background) were purchased from Harlan and housed for study in polycarbonate shoebox cages with shredded aspen beddings at the Endocyte, Inc. Animal Facility for duration of study. Bedding was replaced twice a week in addition to fresh cages biweekly by a qualified technician. The diet and drinking water (municipal) were provided ad libitum throughout the study periods. Mice were housed in an environmentally controlled room with temperature settings that ranged from 70 to 74°F and relative humidity that ranged from 30 to 70%. Light cycles were set to provide a 12-h light/12-h dark photoperiod. Test diets were provided to mice for the duration of study. Mice were euthanized at end of diet duration using carbon dioxide. All housing, care, and procedures were followed according to Endocyte, Inc.-approved animal care and use protocols.

**Serum and RBC Total Folate Quantitation.** Mouse blood was obtained by cardiac puncture as part of a terminal procedure. For whole blood, samples were collected in pediatric-sized K$_2$EDTA-coated tubes (BD Microtainer; BD Biosciences, San Jose, CA). For sera, blood was collected in uncoated tubes and allowed to sit at room temperature for half an hour. Samples were spun two consecutive times at 6600g at 4°C for 10 min. All samples collected on a particular day were then shipped for analysis (overnight on cold packs) to Ani-lytics, Inc. (Gaithersburg, MD).

Methods used for measuring serum and RBC folates were proprietary to Ani-lytics, Inc. Briefly, total folate levels in sera or RBC samples were measured using Dualcount reagent kit (KDCDSF) supplied by Diagnostic Products (Los Angeles, CA). Whole blood samples were lysed in 1% ascorbic acid solution. Whole blood hemolysis samples, serum samples (diluted in ascorbate solution), and standard folate samples were then treated with dithiothreitol plus an 125I-folic acid tracer followed by an extraction step performed at alkaline pH. Folates were extracted by mixing milk folate-binding protein immobilized to a solid support. After a brief centrifugation and removal of the supernatant, bound radioactivity was measured with a gamma counter. The total folates in the sample were quantitated by comparison with a standard curve.

**Folate Receptor Assay.** Tissue FR levels were quantitated according to a previously reported procedure (Parker et al., 2005). Briefly, membrane pellets were isolated from homogenized tissue samples and then solubilized in an octyl-β-D-glucopyranoside-containing buffer. Aliquots of solubilized membranes were placed inside each of two Microcon-30 microcentrifuges (30,000 molecular weight cut-off; Millipore Corporation, Billerica, MA) and then acid-stripped to remove endogenously bound folates. [3H]Folic acid binding reagent (in the presence or absence of a 1000-fold excess of unlabeled folic acid) was added to the appropriate concentrators. Following a 10-min incubation at room temperature, the concentrators were washed, and the retentates containing the solubilized FRs were recovered from the membrane surface of the microcentrifuges. The samples were then counted in a liquid scintillation counter (PerkinElmer Life and Analytical Sciences, Boston, MA). The cpm values were converted to picomoles of FR based upon the CMP of a known standard, and the final results were normalized with respect to the sample protein content.

**Relative Binding Assay Using Immobilized Folate-Binding Protein.** One hundred microliters of a soluble milk FBP solution (10 µg/ml in PBS) was added to each well of a Reacti-Bind microtiter plate. Plates were incubated at 4°C overnight and then washed three times with ice-cold PBS containing 0.05% Tween 20 (PBS-T). Plates were equilibrated to room temperature, blocked for 1 h at room temperature with 100 µl/well of freshly prepared PBS-T containing 0.2% gelatin, and then washed three additional times with PBS-T. Undiluted serum samples or bovine serum albumin (43 mg/ml in PBS) was added to FBP-coated ELISA plates for 2 h at 37°C. Wells were washed three times with PBS, and then 100 µl of 100 nM [3H]FA in PBS (per well) was added in triplicate. Plates were incubated at 37°C for 30 min with gentle shaking and then rinsed three
times with PBS-T. Bound [3H]FA was then stripped from the plates with 100 μl of acid-saline solution (20 mM sodium acetate, pH 3.0) for up to 20 min at room temperature, and the acidic samples were transferred into individual scintillation vials containing 3 ml of scintillation cocktail. Values are expressed as the percentage of maximum binding relative to PBS-treated (control) wells, which bound 107,671 ± 3514 dpm.

**FolateScan (99mTc-EC20) Uptake in KB Tumors and Kidneys of nu/nu Mice.** An EC20 kit was used for the preparation of radioactive drug substance (Leamon et al., 2002). Each kit contains a sterile, nonpyrogenic lyophilized mixture of 0.1 mg of EC20, 80 mg of sodium α-d-glucopentanoate dihydrate, 80 μg of tin(II)chloride dihydrate, and sufficient sodium hydroxide or hydrochloric acid to adjust the pH to 6.8 ± 0.2. Chelation of 99mTc to EC20 was done by injecting 1 ml of sodium pertechnetate 99mTc injection (<50 mCi) into this vial and heating it for ~18 min in a boiling water bath. Samples of the 99mTc-EC20 solution were analyzed for radiochemical purity using a high-performance liquid chromatography system consisting of a 600E multisolute delivery system and 490 UV detector (Waters, Milford, MA), an FC-3200 radiodetector, with Laura version 1.5 radiochromatogram software (BioScan, Washington, DC), and a Nova-Pak C18 (3.9 × 150 mm) column (Waters). Injected samples were eluted isocratically using an aqueous mobile phase containing 20% methanol and 0.1% trifluoroacetic acid at a flow rate of 1 ml/min. The radiochemical purity of 99mTc-EC20 was greater than 90% (Leamon et al., 2002).

Female nu/nu mice (BALB/c background; 6–7 weeks of age; Harlan) were maintained on the folate-rich or folate-free diets for a total of 3 weeks before the biodistribution experiment. FR-positive KB tumor cells (1 × 104 cells per animal) were inoculated in the subcutis of the dorsal medial area of the right axilla. Stock 99mTc-EC20 solutions containing 100 μg of agent per milliliter were prepared on the day of use. Mouse with 300- to 500-mm3 KB tumors were divided into the following groups: 1) the former parameter is a measure for long-term folate status, whereas the latter parameter is a measure for short-term folate status, 2) the high serum folate levels in mouse sera could negatively affect the performance of folate-drug conjugates used for cancer therapy, we chose to study the impact of placing mice on low folate chows on the levels of folates in the sera as well as RBCs. Importantly, the former parameter is a measure for short-term folate status, whereas the latter parameter is a measure for long-term folate status.

**Tumor Model and Therapy.** Six- to 8-week-old female nu/nu mice (Charles River Laboratories, Inc. (Wilmington, MA) were maintained on a standard 12-h light/dark cycle for the duration of the experiment. Mice used in these studies were fed defined chows beginning 2 weeks before tumor implantation and maintained throughout the experiment. FR-positive KB cells (1 × 103 nu/nu mouse) in 100 μl of folate-free RPMI 1640 medium were injected in the subcutis of the dorsal medial area. Tumors were measured in two perpendicular directions every 2 to 3 days using a caliper, and their volumes were calculated as 0.5 × L × W2, where L is measurement of longest axis in millimeters, and W is measurement of axis perpendicular to L in millimeters. Dosing solutions were prepared fresh each day in PBS and administered through the lateral tail vein of the mice. Importantly, dosing was initiated when the subcutaneous tumors were approximately 133 ± 26 mm3 in volume.

**Results**

**Common Laboratory Animals Have High Serum Folate Levels.** Humans are known to have low serum folate levels, typically in the 20 nM range (Venn et al., 2002). For comparison, sera obtained from five commonly used laboratory species, along with freshly isolated human donor sera, were tested for total folate content using a commercial radioimmunoassay. As shown in Fig. 1, sera taken from BALB/c mice were found to contain the highest level of folates at nearly 250 nM. Importantly, this level is approximately 10-fold higher than that measured in human sera (see below). Serum folates from Lewis rats and New Zealand White rabbits were also very high (range, 130–180 nM), whereas the folate levels in beagle dog and cynomolgus monkey sera were in the same range as that measured in normal human sera (~25 nM). These data collectively show that the choice of animal model could be critical when studying the biological effects of folate-based compounds.

**Low Folate Diets Decrease Serum and Red Blood Cell Total Folate Levels.** Because 1) mice are most commonly used for evaluating the performance of most anticancer agents and 2) the high serum folate levels in mouse sera (Fig. 1) could negatively affect the performance of folate-drug conjugates used for cancer therapy, we chose to study the impact of placing mice on low folate chows on the levels of folates in the sera as well as RBCs. Importantly, the former parameter is a measure for short-term folate status, whereas the latter parameter is a measure for long-term folate status. As shown in Fig. 2A, pre-study serum folate levels for mice ~8 weeks of age were again very high (241 ± 7 nM). Serum folate levels did actually drop somewhat (to 180 ± 63 nM) while maintaining these mice on high folate chow for 18 weeks, probably due to natural age-related changes. In contrast, serum folate levels abruptly fell in mice that were fed a low folate chow, and the time-related decrease was independent of whether the chow was supplemented with 0.5 mg/kg leucovorin, which is a reduced form of folic acid that does not significantly interfere with FA-drug targeting (Reddy et al., 2004). Furthermore, 6 weeks after the start of these “low” folate diets, serum folate levels seemed to stabilize, because the concentration remained steady at ~20 nM during weeks 6 through 18. The cause for this stabilization is merely speculative, but it could be due to the low amount of folates supplied naturally by the intestinal flora of the mice (Klipstein and Lipton, 1970). Regardless, the data in Fig. 2A
indicate that dietary modulation can lower the serum folate levels of laboratory mice down to nearly the same levels as humans within a short time.

In addition to serum folate, RBC folate levels were also found to abruptly drop when mice were fed low folate chows. As shown in Fig. 2B, prestudy RBC folate levels for mice ~8 weeks of age were initially very high (4329 ± 44 nM), but they fell to ~1000 nM after 6 weeks on the low folate chows. Interestingly, these RBC folate levels are within the 347 to 1167 nM range measured in humans (Pfeiffer et al., 2005). Thus, the data in Fig. 2 show that both serum and RBC folate levels decrease in mice to concentrations that closely resemble that found in humans when the animals are fed low folate diets for at least 6 weeks.

**Tissue Folate Receptor Levels Decrease When Animals Are Fed Low Folate Chows.** Besides serum and RBC changes, we also examined the effects of dietary folate modulation on major organ tissue-derived FRs. As shown in Fig. 3A, negligible FR levels (i.e., >2.5 pmol/mg membrane protein; Parker et al., 2005) were found in all tissues prestudy except for kidney. Renal cortex is known to be rich with functional FR expression (Holm et al., 1992); therefore, this finding was expected. However, following 18 weeks of eating low folate chow, FR levels surprisingly did not rise in any murine tissue examined; instead, FR levels actually decreased to undetectable levels in all tissues except, again, for the kidney.

Further examination of the dietary effects on kidney FR expression is shown in Fig. 3B. Kidney FR levels were measured at 12.31 ± 1.45 pmol/mg when the mice were ~8 weeks of age, but these levels naturally dropped to 3.28 ± 0.19 pmol/mg while maintaining the mice on high folate chow for 24 consecutive weeks (again, probably due to natural age-related changes). Interestingly, feeding the younger mice low folate chows was found to cause a rapid drop in kidney-associated FR levels by as much as 88% to ~1.5 pmol/mg within 6 weeks. Further decline in FR levels was not observed up to the 24-week period. This relationship between low folate intake and kidney FR expression seems paradoxical, especially since kidney FRs are believed to function as “salvage” receptors for this easily-filtered vitamin (Birn et
al., 1997). But, this effect was observed by others previously (see Discussion) (Gates et al., 1996; da Costa et al., 2000).

**High Serum Folate Levels in Mice Interfere with [3H]FA Binding to the FR in Vitro.** To ascertain the biological significance of lowering serum folate levels through dietary modulation, we tested the ability of radiolabeled folate to bind to its receptor in the presence of different test sera. Thus, normal BALB/c and nu/nu (BALB/c background) mice were fed either high folate or low folate chows for 3 weeks. Sera obtained from these animals were then tested along with pooled human sera and an albumin/PBS control solution for the ability to block [3H]FA from binding to immobilized folate-binding protein. As shown in Fig. 4, maximal [3H]FA binding was measured in wells treated with either the albumin control or 100% pooled human sera. Serum samples obtained from the “high folate” animals were found to block [3H]FA binding by >80%. In contrast, serum samples obtained from the “low folate” animals had blocked [3H]FA binding by no greater than 23%. Taken together with the data in Figs. 1 and 2, these results confirm that 1) mice fed standard laboratory chows have high serum folate levels that can competitively block the binding of folates to the FR, and 2) short dietary modulation with low folate chows will reduce serum folates to “human-like” (Fig. 2), which minimally interfere with ligand binding to the FR.

**Folate-Targeted Radiopharmaceutical Tissue Retention Increases When Animals Are Fed Low Folate Chows.** We next explored the effect of modulating dietary folate levels on the ability of 99mTc-EC20, a folate-based radiopharmaceutical (Leamon et al., 2002; Reddy et al., 2004; Fisher et al., 2008), to target FR+ tissues in vivo. In addition to kidney, which is the predominant FR+ tissue in mice (Fig. 3), we also implanted and assayed FR+ KB tumors in the test animals for 99mTc-EC20 retention to better assess the dietary effects on FR targeting. As shown in Table 1, 99mTc-EC20 retention was prominent in the kidneys but low in the tumors of mice fed the high folate diet. However, we measured a 2.6- and 4.9-fold increase in kidney and tumor retention, respectively, when the mice were fed the low folate diet. Notably, the former result was nearly identical to that observed for folic acid’s kidney retention in a similar study (Gabizon et al., 2003). No significant changes were noted in 99mTc-EC20 retention by all other major (FR-negative) organs. Collectively, these results indicate that although access to FR+ tissues is not completely blocked for animals on high folate diets, upward of 5 times more folate-labeled compounds can be delivered to these FR+ tissues if the laboratory chow is not heavily supplemented with folic acid.

**A High Folate Diet Has Minimal Impact on the Antitumor Response toward Potent Folate-Drug Conjugates.** There are currently three distinct folate-chemotherapeutic drug conjugates being clinically evaluated in multiple phase 1 and 2 oncology trials across the United States, and clinical development is being planned for at least two additional agents in the near future (Leamon et al., 2007a; Reddy et al., 2007a; Sausville et al., 2007; Covello et al., 2008). Related to this study, EC145 and EC0225 are examples of clinically evaluated conjugates, whereas EC0305 represents one that is under consideration for development. The impact of feeding mice high folate diets on the performances of these potent therapeutic agents was next evaluated using the KB nu/nu model. In our first experiment, tumor-bearing mice feeding on the low folate chow had endured complete re-
spontaneous responses and remained tumor-free for >90 days after being briefly treated with 2 μmol/kg EC145 or 1 μmol/kg EC0225 (Fig. 5). Interestingly, the performance of EC0225 was not altered if the mice had been fed the high folate diet. However, tumors in one of five mice in the EC145 cohort did recur while on the high folate diet.

In a second experiment, we evaluated the impact of each diet on the performance of increasing dose levels of EC0305. This agent is a potent folate conjugate of the microtubule inhibitor, tubulysin B monohydrazide (Reddy et al., 2008; Vlahov et al., 2008). As shown in Fig. 6A, 100% of the animals fed the low folate chow and treated with 1 or 2 μmol/kg EC0305 had endured complete responses and remained tumor-free for >90 days. Mice in the low 0.5 μmol/kg cohort also responded well, with four of five having complete responses (two of which remained tumor-free at the end of study) and one of five having a partial response. Similar to what was observed for EC145 and EC0225 (Fig. 5), maintaining mice on the high folate diet had no negative consequences for animals treated at ≥1 μmol/kg EC0305, because each of these animals remained tumor-free for >90 days (Fig. 6B). However, mice that were fed the high folate diet and treated with the low 0.5 μmol/kg dose level did not fare as well as their counterparts that had been maintained on the low folate chow. Although a small antitumor signal was present during this brief 2-week treatment (on the high folate diet), we observed only one partial responder in that cohort (Fig. 6A). Collectively, the data presented in Figs. 5 and 6 suggest that a high folate diet has a small but measurable impact on the antitumor response toward potent folate-drug conjugates, and such negative effects on pharmacological responses may be overcome by slightly increasing the dose of the administered folate-drug agent.

**Discussion**

As with any promising therapeutic approach, use of relevant preclinical models becomes paramount to the successful identification of clinical candidates and, ultimately, to the development of effective products for human use. Our laboratory has been actively researching and now developing FA-targeted drug therapies for nearly 20 years. We recognized long ago that high serum folate concentrations, like that found in the sera of laboratory rodents, could potentially block the binding of FA-drug conjugates to FR-positive tissues, e.g., tumors (Mathias et al., 1996). Because of that finding, the majority of “folate-targeted” pharmacological studies published to date had involved the use of animals that were acclimated to some form of low folate chow. Since we have been fortunate to witness the start of clinical testing for a number of folate-based agents, the impact that a patient’s diet might have (i.e., net folate intake) on the performance of these novel agents has been called into question. Therefore, the purpose of our current work was to directly examine such dietary effects using well known animal models and to determine whether changes in dietary folate could alter the antitumor effect of some clinically and preclinically relevant compounds.

Dietary modulation, using a standard rodent laboratory chow devoid of supplemental FA, was found to quickly lower both serum and RBC folates in mice to concentrations that better approximate those levels found in human sera. In fact, human-like folate levels were reached in the mice within 6 weeks (Fig. 2). At that point, and at least up to 18 weeks thereafter, serum and RBC folate levels reached steady-state levels. This observation was important, because our tumor therapy protocols allow for responding animals to remain on study for up to 120 days, suggesting that animal nutrition and welfare should never be compromised. Notably, therapy with folate-drug conjugates typically begins 2 or 3 weeks after the start of the low folate diet (Figs. 5 and 6) (Leamon et al., 2005, 2006, 2007a,b; Reddy et al., 2006, 2007a,b). Because serum and RBC folate concentrations do not reach their neohomeostatic levels until week 6, one may argue that better therapeutic outcomes could result if therapy were delayed until that point. However, because 1) housing costs are expensive, 2) serum folate levels are 75% lower in mice after only 2 weeks on the low folate diet, and 3) tumor-free responses are often observed with folate-drug therapies when initiated at that earlier time, we believe it is not necessary to delay the start of in vivo studies until that 6-week point.

Throughout our investigation, we noted that while mice were maintained on the low folate diet, 99mTc-EC20 uptake in the kidney was observed to increase 2.6-fold; yet, their kidney FR levels concomitantly dropped by 88% (compare Table 1 and Fig. 3, respectively). The increase in 99mTc-EC20 kidney uptake can be explained by the presence of empty FRs.
in that tissue relative to a minuscule amount under high dietary folate conditions. The massive loss of kidney FRs observed in response to low folate diets, although intriguing, has actually been observed by others previously (Gates et al., 1996; da Costa et al., 2000), but the cause for this dramatic change was not defined. Although this apparent paradox is not the subject of this report, we propose that the diet-induced drop in kidney FR levels may be the result of FR naturally “shedding” into the blood, in ligand-bound form, to promote uptake and retention of the ligand/FR complex by the liver. In other words, such a process could serve as a means for better salvaging folates under more limiting conditions. Certainly, it is known that liver uptake of FA is warranted to better determine the full translational impact.

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References


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