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Selective Activation of Cannabinoid CB_2 Receptors Suppresses Neuropathic Nociception Induced by Treatment with the Chemotherapeutic Agent Paclitaxel in Rats

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ABSTRACT

Activation of cannabinoid CB_2 receptors suppresses neuropathic pain induced by traumatic nerve injury. The present studies were conducted to evaluate the efficacy of cannabinoid CB_2 receptor activation in suppressing painful peripheral neuropathy evoked by chemotherapeutic treatment with the antitumor agent paclitaxel. Rats received paclitaxel (2 mg/kg i.p./day) on 4 alternate days to induce mechanical hypersensitivity (mechanical allodynia). Mechanical allodynia was defined as a lowering of the threshold for paw withdrawal to stimulation of the plantar hind paw surface with an electronic von Frey stimulator. Mechanical allodynia developed in paclitaxel-treated animals relative to groups receiving the Cremophor EL/ethanol/saline vehicle at the same times. Two structurally distinct cannabinoid CB_2 agonists, the aminoalkylindole (*R*,*S*)-AM1241 [(*R*,*S*)-(2-iodo-5-nitrophenyl)-[1-((1-methyl-piperidin-2-yl)methyl)-1*H*-indol-3-yl]-methanone] and the cannabilactone AM1714 (1,9-dihydroxy-3-(1′,1′-dimethylheptyl)-6*H*-benzo[*c*]chromene-6-one), produced a dose-related suppression of established paclitaxel-evoked mechanical allodynia after systemic administration. Pretreatment with the CB_2 antagonist SR144528 [5-(4-chloro-3-methylphenyl)-1-(4-methylbenzyl)-*N*-(1,3,3-trimethylbicyclo[2.2.1]heptan-2-yl)-1*H*-pyrazole-3-carboxamide], but not the CB_1 antagonist SR141716 [5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-*N*-(piperidin-1-yl)-1*H*-pyrazole-3-carboxamide], blocked the antiallodynic effects of both (*R*,*S*)-AM1241 and AM1714. Moreover, (*R*)-AM1241, but not (*S*)-AM1241, suppressed paclitaxel-evoked mechanical allodynia relative to either vehicle treatment or preinjection thresholds, consistent with mediation by CB_2. Administration of either the CB_1 or CB_2 antagonist alone failed to alter paclitaxel-evoked mechanical allodynia. Moreover, (*R*,*S*)-AM1241 did not alter paw withdrawal thresholds in rats that received the Cremophor EL vehicle in lieu of paclitaxel, whereas AM1714 induced a modest antinociceptive effect. Our data suggest that cannabinoid CB_2 receptors may be important therapeutic targets for the treatment of chemotherapy-evoked neuropathy.

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Painful peripheral neuropathy is a well documented side effect of chemotherapeutic treatment (for review, see Polomano and Bennett, 2001; Aley and Levine, 2002). The major classes of antineoplastic agents, the vinca alkaloids (e.g., vincristine), taxane (e.g., paclitaxel), and platinum-derived (e.g., cisplatin) compounds, are associated with the development of dose-limiting neuropathic pain. The chemotherapeutic agent used, dosing schedule, form of cancer, and presence of additional medical complications can affect the occurrence and severity of chemotherapy-induced neuropathy (for review, see Cata et al., 2006).

Paclitaxel is commonly used for the treatment of solid tumors and ovarian and breast cancers. Paclitaxel induces

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ABBREVIATIONS: CNS, central nervous system; (*R*,*S*)-AM1241, (*R*,*S*)-(2-iodo-5-nitrophenyl)-[1-((1-methyl-piperidin-2-yl)methyl)-1*H*-indol-3-yl]-methanone; AM1714, 1,9-dihydroxy-3-(1′,1′-dimethylheptyl)-6*H*-benzo[*c*]chromene-6-one; SR141716, 5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-*N*-(piperidin-1-yl)-1*H*-pyrazole-3-carboxamide; SR144528, 5-(4-chloro-3-methylphenyl)-1-(4-methylbenzyl)-*N*-(1,3,3-trimethylbicyclo-[2.2.1]heptan-2-yl)-1*H*-pyrazole-3-carboxamide; DMSO, dimethyl sulfoxide; ANOVA, analysis of variance; DRG, dorsal root ganglion; WIN55,212-2, *R*-(+)-[2,3-dihydro-5-methyl-3-[(4-morpholinyl)methyl]pyrrolo[1,2,3-*de*]-1,4-benzoxazinyl]-(1-naphthalenyl)methanone; TRPV, transient receptor potential vanilloid; WIN55,212-3, (*S*)-(−)-[2,3-dihydro-5-methyl-3-(4-morpholinyl]methyl)pyrrolo-(1,2,3-*de*)]-1,4-benzoxazinyl-[1-napthalenyl]methanone.

antimitotic actions by impeding the cell cycle in the late phases of mitosis, stabilizing microtubule formation, and ultimately inducing apoptosis (Schiff and Horwitz, 1980). Paclitaxel preferentially impairs myelinated Aβ and Aδ fibers, which carry sensory information about mechanical stimulation to the central nervous system (CNS) (Dougherty et al., 2004). Paclitaxel-evoked neuropathy is manifested as pain in the distal extremities, forming a glove and stocking pattern (Dougherty et al., 2004). Mitochondrial toxicity is also preferentially localized to long axons innervating distal extremities (Flatters and Bennett, 2006). Thus, effects of paclitaxel are evident in those areas where, due to increased distance of axonal transport and mitochondrial energy demand, disruption in sensation would first be present. Dysfunctional mitochondria could lead to low levels of energy, which could potentially impair ion transporters, resulting in spontaneous neuronal firing with no concurrent receptor stimulation (i.e., paraesthesia) (Flatters and Bennett, 2006).

Peripheral neuropathy can limit dosing and duration of chemotherapeutic treatment (Holmes et al., 1991; Rowinsky et al., 1993). Pharmacotherapies for chemotherapy-induced neuropathy are limited because the underlying cellular mechanisms remain incompletely understood. Amitriptyline, gabapentin, and opioids are used to treat chemotherapy-induced neuropathy. However, none of these drugs has been shown to completely attenuate neuropathic pain (for review, see Lee and Swain, 2006). The absence of approved medications available for preventing or treating this debilitating neuropathy makes the identification of alternative effective analgesics a crucial medical need.

Cannabinoids suppress neuropathic pain induced by traumatic nerve injury, toxic insults, and metabolic changes (for review, see Hohmann, 2002; Guindon and Hohmann, 2008). Both CB_1-specific (Herzberg et al., 1997; Fox et al., 2001) and CB_2-specific (Ibrahim et al., 2003; Beltramo et al., 2006) mechanisms suppress neuropathic nociception evoked by traumatic nerve injury. CB_1 receptors are expressed primarily within the CNS (Zimmer et al., 1999). CB_2 receptors are expressed primarily, but not exclusively, outside the CNS in cells of the immune system (Munro et al., 1993). CB_2 receptors are up-regulated in the CNS in neuropathic pain states (Wotherspoon et al., 2005; Beltramo et al., 2006). CB_2-selective agonists are not associated with psychoactive and motor effects typical of CB_1 receptor activation (Hanus et al., 1999; Malan et al., 2001), making the CB_2 receptor an attractive therapeutic target for the treatment of neuropathic pain.

The mixed CB_1/CB_2 agonist WIN55,212-2 suppresses neuropathic nociception induced by paclitaxel through a CB_1-specific mechanism (Pascual et al., 2005). WIN55,212-2 also suppresses vincristine-induced neuropathy through activation of both CB_1 and CB_2 receptors (Rahn et al., 2007). Activation of CB_2 receptors with (*R*,*S*)-AM1241 partially attenuates vincristine-induced neuropathy (Rahn et al., 2007). However, a role for CB_2 receptor activation in suppressing paclitaxel-evoked neuropathy has not been investigated. This investigation is important because distinct mechanisms may underlie development of neuropathic pain induced by different antineoplastic agents (for review, see Cata et al., 2006). Neuropathic pain symptoms associated with each chemotherapeutic agent vary and can respond differently to pharmacological treatments (Flatters and Bennett, 2004). We used two structurally distinct CB_2-selective agonists, AM1714 and (*R*,*S*)-AM1241 (Fig. 1), to evaluate the contribution of CB_2 receptors to cannabinoid modulation of paclitaxel-induced neuropathy. AM1714 is a novel CB_2-selective agonist (K_i, CB_1 versus CB_2, 400 versus 0.8 nM) from the cannabilactone class of cannabinoids (Khanolkar et al., 2007). AM1714 has recently been shown to induce peripheral antinociception but has not been characterized previously in an animal model of pathological pain. (*R*,*S*)-AM1241 is a CB_2-selective agonist from the aminoalkylindole class of cannabinoids. (*R*,*S*)-AM1241 behaves as a protean agonist in vitro (Yao et al., 2006) and a CB_2 agonist in vivo (for review, see Guindon and Hohmann, 2008). We also compared the ability of (*R*)-AM1241 (K_i, CB_1 versus CB_2, 139.7 versus 1.4 nM) and its less active enantiomer (*S*)-AM1241 (K_i, CB_1 versus CB_2, 2029 versus 160.5 nM) (Thakur et al., 2005) to suppress paclitaxel-evoked neuropathy. Pharmacological specificity was evaluated using selective antagonist/inverse agonists for CB_1 (SR141716) and CB_2 (SR144528). Comparisons were made with the prototypical narcotic analgesic morphine.

Materials and Methods

Subjects. One hundred seventy-five adult male Sprague-Dawley rats (301–396 g; Harlan, Indianapolis, IN) were used in these experiments. All procedures were approved by the University of Georgia Animal Care and Use Committee and followed the guidelines for the treatment of animals of the International Association for the Study of Pain. Bedding containing metabolized paclitaxel was treated as biohazardous waste and disposed of according to the appropriate institutional guidelines.

Drugs and Chemicals. Paclitaxel was obtained from Tecoland Corporation (Edison, NJ). (*R*,*S*)-AM1241, (*R*)-AM1241, (*S*)-AM1241, and AM1714 were synthesized in the Makriyannis laboratory by one of the authors (by A.M.Z. and G.A.T., respectively). The (*R*)- and (*S*)-enantiomers were prepared by chiral synthesis (by A.M.Z). SR141716 and SR144528 were provided by the National Institute on Drug Abuse (Bethesda, MD). Cremophor EL and morphine sulfate were obtained from Sigma-Aldrich (St. Louis, MO). Dimethyl sulfoxide (DMSO) was purchased from Thermo Fisher Scientific (Waltham, MA). Paclitaxel was dissolved as previously described (Flatters and Bennett, 2004) and administered in a volume of 1 ml/kg. In brief,

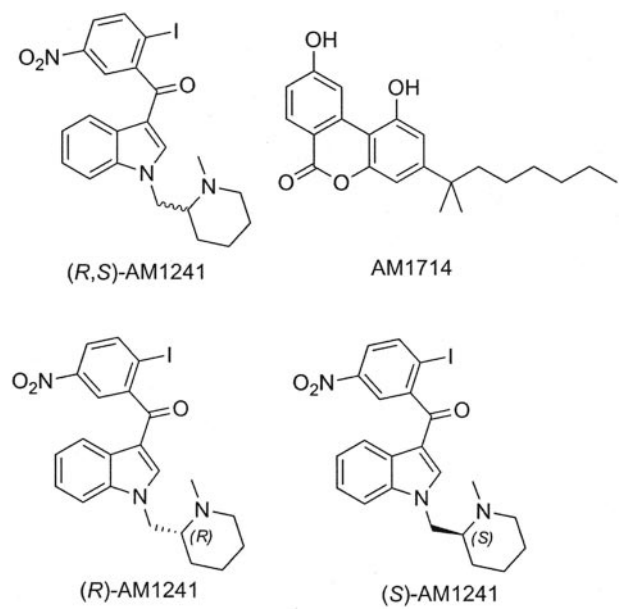

Fig. 1. Chemical structures of (*R*,*S*)-AM1241, (*R*)-AM1241, (*S*)-AM1241, and AM1714.

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paclitaxel was dissolved in a 1:2 ratio of working stock (1:1 ratio of Cremophor EL/95% ethanol) to saline. All other drugs were dissolved in a vehicle of 100% DMSO for systemic administration and administered in a volume of 1 ml/kg body weight.

General Experimental Methods. Baseline withdrawal thresholds to mechanical stimulation of the hind paw were measured on day 0. Rats subsequently received four i.p. injections of either paclitaxel (2 mg/kg/day i.p.) or Cremophor EL/ethanol/saline vehicle (1 ml/kg/day i.p.) on alternate days, immediately after behavioral testing. The injection paradigm consisted of four once-daily injections, administered on days 0, 2, 4, and 6, as described previously (Polomano et al., 2001). Mechanical withdrawal thresholds were measured on days 0, 4, 7, 11, 14, 18, and 21. Behavioral testing was always performed just before paclitaxel administration (except for days 2 and 6, on which paw withdrawal thresholds were not assessed). To evaluate the possible resolution of paclitaxel-induced neuropathy, paclitaxel-treated rats were additionally evaluated weekly for the presence of mechanical allodynia for 86 days after the initial injection of paclitaxel in a pilot study. In all studies, the experimenter was blinded to the drug condition. Moreover, a single experimenter tested all animals in any given study.

Assessment of Mechanical Withdrawal Thresholds. Mechanical withdrawal thresholds were assessed using a digital electronic von Frey anesthesiometer (IITC model Alemo 2290-4; IITC Life Sciences Inc., Woodland Hills, CA) equipped with a rigid tip. Rats were placed underneath inverted plastic cages and positioned on an elevated mesh platform. Rats were allowed to habituate to the chamber for 10 to 15 min before testing. Stimulation was applied to the midplantar region of the hind paw through the floor of a mesh platform. Mechanical stimulation was terminated upon paw withdrawal; consequently, there was no upper threshold limit set for termination of a trial. On the test day (day 21), baseline mechanical withdrawal thresholds were assessed, and effects of pharmacological manipulations were subsequently evaluated. Nocifensive responses were observed in paclitaxel-treated animals at forces (g) that failed to elicit withdrawal responses before chemotherapy treatment. Paclitaxel-induced decreases in mechanical paw withdrawal thresholds (assessed with the electronic von Frey anesthesiometer) were therefore defined as mechanical allodynia.

Preinjection mechanical withdrawal thresholds were measured on day 21 before acute pharmacological manipulations. Paclitaxel-treated animals received systemic injections of (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 7$), AM1714 (10 mg/kg i.p.; $n = 6$), or DMSO ($n = 7$). Mechanical withdrawal thresholds were measured 30, 60, and 90 min postinjection to assess the time course of CB_2 agonist actions. Subsequent studies evaluated dose response and pharmacological specificity by measuring paw withdrawal thresholds at the time point of maximal cannabinoid-induced suppression of paclitaxel-evoked neuropathy (30 min postinjection).

To evaluate dose-response, separate groups of paclitaxel-treated animals received the racemate (*R*,*S*)-AM1241 (1, 5, or 10 mg/kg i.p.; $n = 6$–10 per group), AM1714 (1, 5, or 10 mg/kg i.p.; $n = 6$ per group), or DMSO ($n = 11$). Separate groups of animals received the enantiomers of (*R*,*S*)-AM1241, (*R*)-AM1241 (10 mg/kg i.p.; $n = 6$) or its less active enantiomer (*S*)-AM1241 (10 mg/kg i.p.; $n = 6$), or the opioid agonist morphine (2 or 4 mg/kg i.p.; $n = 6$ per group).

To determine pharmacological specificity, separate groups of paclitaxel-treated rats received (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 6$), AM1714 (10 mg/kg i.p.; $n = 6$), or SR144528 (10 mg/kg i.p.) administered 20 min before (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 6$) or AM1714 (10 mg/kg i.p.; $n = 5$), SR144528 alone (10 mg/kg i.p.; $n = 7$), or DMSO ($n = 6$). In separate groups of animals, SR141716 (10 mg/kg i.p.) was administered 20 min before treatment with either (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 5$) or AM1714 (10 mg/kg i.p.; $n = 8$).

Antagonist pretreatment groups received a double volume of the DMSO vehicle. Therefore, paw withdrawal thresholds were compared in animals receiving dual injections of either DMSO or saline to verify that vehicle effects could not account for the pattern of results obtained. Therefore, additional control groups received (i.p.) either saline 20 min before saline ($n = 6$) or DMSO 20 min before DMSO ($n = 6$). To evaluate possible antinociceptive effects induced by the CB_2 agonists, the maximally effective antiallodynic dose of either AM1714 (10 mg/kg i.p.; $n = 6$) or (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 6$) was additionally administered to Cremophor EL-treated controls. Paw withdrawal thresholds were assessed as described above.

Statistical Analyses. Data were analyzed using analysis of variance (ANOVA) for repeated measures, one-way ANOVA, or planned comparison Student's t tests as appropriate. The Greenhouse-Geisser correction was applied to all repeated factors. Post hoc comparisons between control groups and other experimental groups were performed using the Dunnett test. Post hoc comparisons between different experimental groups were also performed to assess dose-response relationships and pharmacological specificity using the Tukey test. Postdrug thresholds within a given group were compared with either prepaclitaxel (baseline) thresholds or day 21 postpaclitaxel thresholds using paired Student's t tests. $P < 0.05$ was considered statistically significant.

Results

General Results. Body weight did not differ between groups before the treatment with either paclitaxel or the Cremophor EL/ethanol/saline vehicle. Normal weight gain was observed in groups receiving either the Cremophor EL vehicle or paclitaxel ($F_{2,213} = 1.3$, $P > 0.27$; Fig. 2a). However, one fatality was observed in groups receiving paclitaxel.

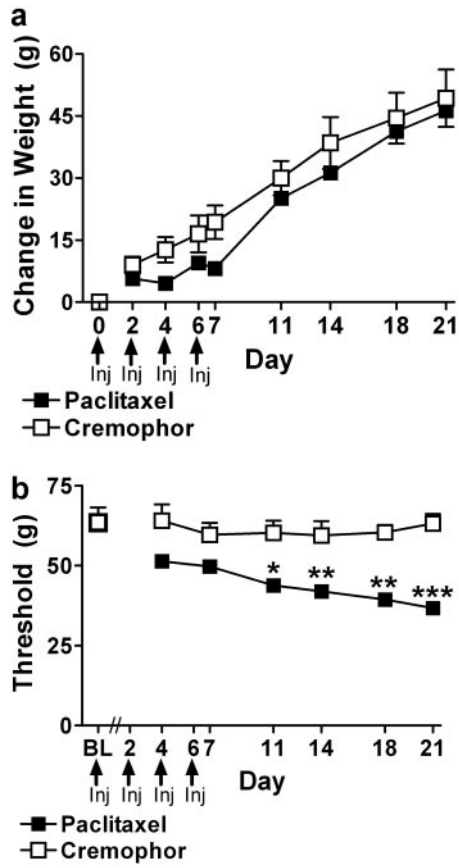

Fig. 2. a, weight gain was observed in groups treated with either paclitaxel or Cremophor EL/ethanol/saline vehicle. b, time course of paclitaxel-induced mechanical allodynia, as demonstrated by a lowering of the threshold for paw withdrawal to punctuate mechanical stimulation. Data are mean ± S.E.M. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$ versus control condition (ANOVA). $n = 6$ to 115 per group.

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TABLE 1
Paw withdrawal thresholds (grams) in paclitaxel-treated control conditions
Data are mean ± S.E.M.

	Prepaclitaxel: Day 0	Postpaclitaxel	
		Day 21	Day 21
Group	Preinjection	Preinjection	Postinjection
Paclitaxel/DMSO-DMSO	66.915 ± 6.58	46.56 ± 4.23†	42.88 ± 3.95$^{\perp\perp}$
Paclitaxel/saline-saline	75.585 ± 6.32	41.20 ± 4.90†††	39.55 ± 4.40$^{\perp\perp\perp}$
Paclitaxel/DMSO	63.88 ± 2.74	32.93 ± 2.43†††	32.08 ± 5.98$^{\perp\perp\perp}$
Paclitaxel/SR141716	63.59 ± 2.20	37.34 ± 1.77†††	37.23 ± 5.05^{X}
Paclitaxel/SR144528	59.92 ± 4.10	32.64 ± 7.42††	39.78 ± 4.86$^{\perp\perp\perp}$

††† $P < 0.001$; ††, $P < 0.01$; and †, $P < 0.05$ vs. baseline prepaclitaxel paw withdrawal thresholds for corresponding group; $^{\perp\perp\perp}$, $P < 0.001$; $^{\perp\perp}$, $P < 0.01$; $^{\perp\perp}$, $P < 0.05$ vs. baseline prepaclitaxel paw withdrawal thresholds for corresponding group (Student's t test).

In a pilot study conducted to evaluate the resolution of paclitaxel-evoked mechanical allodynia, paw withdrawal thresholds were lower than baseline prepaclitaxel thresholds beginning on day 7 ($P < 0.05$, planned comparison). Paclitaxel-induced mechanical allodynia was present, relative to baseline, from days 14 to 72 after the initiation of treatment ($P < 0.05$ for all planned comparisons; data not shown). Paw withdrawal thresholds were also similar from days 14 to 72 postpaclitaxel. Therefore, day 21 postpaclitaxel was used to evaluate CB_2 agonist actions on paclitaxel-evoked mechanical allodynia in all studies reported herein. Paw withdrawal thresholds did not differ between paclitaxel-treated groups before cannabinoid or vehicle treatments on day 21 in any study. In contrast, thermal hyperalgesia was not observed in the present paclitaxel dosing paradigm (data not shown).

Mechanical withdrawal thresholds did not differ between either the right or the left paw for any group on any given day (days 0–21); therefore, withdrawal thresholds are presented as the mean of duplicate measurements, averaged across paws. Paw withdrawal thresholds were similar between groups before administration of paclitaxel in any given study. Paclitaxel lowered mechanical paw withdrawal thresholds (i.e., equivalently in each paw) relative to control conditions receiving the Cremophor EL vehicle ($F_{1,115} = 10.140$, $P < 0.01$; Fig. 2b). Paclitaxel lowered paw withdrawal thresholds in all studies ($P < 0.001$ in each experiment).

Antagonist pretreatment conditions received dual injections of the DMSO vehicle. Therefore, paw withdrawal thresholds were compared in groups receiving DMSO followed by DMSO and saline followed by saline. Postinjection paw withdrawal thresholds did not differ from day 21 preinjection thresholds in either pretreatment group ($P > 0.54$ for both planned comparison Student's t tests; Table 1). Therefore, the volume of DMSO administered did not alter paclitaxel-evoked paw withdrawal thresholds in our study.

The CB_2 Agonists (*R*,*S*)-AM1241 and AM1714 Suppress Paclitaxel-Evoked Mechanical Allodynia. In paclitaxel-treated rats, (*R*,*S*)-AM1241 (10 mg/kg i.p.) and AM1714 (10 mg/kg i.p.) suppressed paclitaxel-induced mechanical allodynia relative to the vehicle condition ($F_{2,16} = 4.05$, $P < 0.05$; $P < 0.05$ for each comparison; Fig. 3). Paclitaxel-induced mechanical allodynia was maximally suppressed by each agonist at 30 min postinjection ($F_{2,16} = 5.34$, $P < 0.05$). At this time point, both (*R*,*S*)-AM1241 and AM1714 normalized thresholds relative to prepaclitaxel levels ($P < 0.05$ for all comparisons). (*R*,*S*)-AM1241 (10 mg/kg i.p.; $n = 6$) failed to induce an antinociceptive effect in animals that received Cremophor EL/ethanol/saline vehicle in lieu of paclitaxel [day 21 paw withdrawal threshold (mean ± S.E.M.) preinjection versus postinjection, 42.14 ± 0.36 versus 40.93 ± 0.78 g; $P > 0.32$; planned comparison t test]. However, AM1714 (10 mg/kg i.p.; $n = 6$) produced a modest antinociceptive effect [day 21 paw withdrawal threshold (mean ± S.E.M.) preinjection versus postinjection, 63.21 ± 2.98 versus 76.92 ± 4.22 g; $P < 0.05$; planned comparison Student's t test]. Moreover, Cremophor EL treatment did not alter day 21 paw withdrawal thresholds relative to day 0 baseline paw withdrawal thresholds in any group. Day 0 baseline paw withdrawal thresholds averaged 46.89 ± 4.23 and 63.60 ± 4.61 g before initiation of Cremophor EL treatment in groups that subsequently received (*R*,*S*)-AM1241 and AM1714, respectively, on day 21. A lower baseline threshold was observed in the former compared with the latter group ($P < 0.05$, Student's t test). Group differences in baseline paw withdrawal thresholds may reflect individual differences combined with the sensitivity of the electronic von Frey device because each animal's threshold was highly reliable and reproducible. No differences between day 0 baseline paw withdrawal thresholds were observed for any groups tested by the same experimenter in any given study.

Effects of (*R*,*S*)-AM1241 and Its Enantiomers on Paclitaxel-Evoked Mechanical Allodynia. (*R*,*S*)-AM1241 increased mechanical withdrawal thresholds in a dose-related fashion relative to the vehicle condition ($F_{3,29} = 3.31$, $P < 0.05$; Fig. 4a). Both the high (10 mg/kg i.p.) and middle (5 mg/kg i.p.) doses of (*R*,*S*)-AM1241 elevated paw withdrawal thresholds relative to vehicle ($P < 0.05$ for both comparisons). Effects of the low dose of (*R*,*S*)-AM1241 (1 mg/kg i.p.) did not differ from vehicle ($P > 0.12$). Both the high (10 mg/kg i.p.) and the middle (5 mg/kg i.p.) doses of (*R*,*S*)-AM1241 also elevated paw withdrawal thresholds relative to preinjection

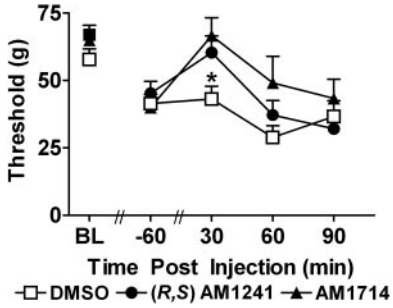

Fig. 3. (*R*,*S*)-AM1241 (10 mg/kg i.p.) and AM1714 (10 mg/kg i.p.) suppressed paclitaxel-induced mechanical allodynia. Both cannabinoid CB_2 agonists normalized thresholds relative to prepaclitaxel levels at 30 min postinjection. BL, baseline (day 0) paw withdrawal thresholds observed before paclitaxel treatment. Data are mean ± S.E.M. *, $P < 0.05$ versus all groups (ANOVA and Dunnett's post hoc test). $n = 6$ to 7 per group.

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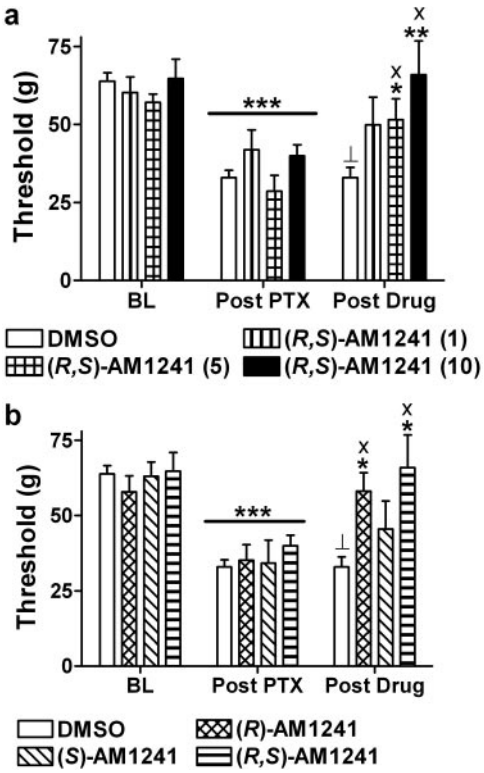

Fig. 4. a, (*R*,*S*)-AM1241 (1, 5, and 10 mg/kg i.p.) produced a dose-related suppression of paclitaxel-evoked mechanical allodynia. In all panels, post-PTX indicates thresholds observed on day 21 postpaclitaxel. b, both (*R*,*S*)-AM1241 (10 mg/kg i.p.) and its enantiomer (*R*)-AM1241 (10 mg/kg i.p.) attenuated paclitaxel-evoked mechanical allodynia. *, $P < 0.05$; **, $P < 0.01$ versus control; ***, $P < 0.001$ versus baseline (ANOVA and Dunnett post hoc test); X, $P < 0.05$ versus corresponding group day 21 postpaclitaxel paw withdrawal thresholds; ⊥, $P < 0.001$ versus corresponding group baseline prepaclitaxel paw withdrawal thresholds (Student's *t* test). $n = 6$ to 11 per group.

thresholds determined 21 days after paclitaxel treatment ($F_{3,29} = 3.54$, $P < 0.05$; $P < 0.05$). Neither the low dose of (*R*,*S*)-AM1241 (1 mg/kg i.p.) nor DMSO altered paw withdrawal thresholds relative to preinjection thresholds assessed on day 21 postpaclitaxel ($P > 0.10$). The middle and high doses of (*R*,*S*)-AM1241 normalized paw withdrawal thresholds relative to baseline (prepaclitaxel) thresholds ($P > 0.16$), whereas DMSO failed to do so.

(*R*)-AM1241 increased paw withdrawal thresholds relative to the vehicle condition ($F_{3,25} = 4.37$, $P < 0.05$; Fig. 4b) in paclitaxel-treated groups. (*S*)-AM1241 (10 mg/kg i.p.) did not significantly elevate paw withdrawal threshold relative to vehicle ($P > 0.43$). However, post hoc comparisons failed to reveal differential effects between (*S*)-AM1241 (10 mg/kg i.p.) and either (*R*,*S*)-AM1241 (10 mg/kg i.p.) or (*R*)-AM1241 (10 mg/kg i.p.) on paw withdrawal thresholds ($P > 0.24$). Both (*R*)-AM1241 (10 mg/kg i.p.) and (*R*,*S*)-AM1241 (10 mg/kg i.p.) significantly increased paw withdrawal thresholds relative to day 21 preinjection thresholds ($P < 0.05$), whereas (*S*)-AM1241 failed to do so. (*R*,*S*)-AM1241 (10 mg/kg i.p.) and (*R*)-AM1241 (10 mg/kg i.p.) also normalized paw withdrawal thresholds relative to day 0 prepaclitaxel thresholds ($F_{3,25} = 3.87$ $P < 0.05$; Fig. 4b). In contrast, normalization of paw withdrawal thresholds was absent in groups receiving DMSO ($P < 0.001$).

The Novel CB_2 Agonist AM1714 Suppresses Paclitaxel-Evoked Mechanical Allodynia. AM1714 suppressed paclitaxel-induced allodynia in a dose-dependent fashion ($F_{3,25} = 5.14$, $P < 0.01$; Fig. 5). All three doses of AM1714 suppressed paclitaxel-evoked mechanical allodynia relative to their vehicle-treated counterparts ($P < 0.05$ for all comparisons). AM1714 (1, 5, and 10 mg/kg i.p.) also normalized paclitaxel-induced mechanical allodynia relative to prepaclitaxel baseline thresholds ($F_{3,25} = 5.63$, $P < 0.01$; $P > 0.14$ for all comparisons; Fig. 5). The high dose (10 mg/kg i.p.; $P < 0.001$) but not the middle (5 mg/kg i.p.) or low (1 mg/kg i.p.) doses of AM1714 elevated paw withdrawal thresholds relative to day 21 preinjection thresholds ($P > 0.23$ for both comparisons).

Pharmacological Specificity. Neither the CB_1-selective antagonist SR141716 (10 mg/kg i.p.) nor the CB_2-selective antagonist SR144528 (10 mg/kg i.p.) altered paclitaxel-evoked mechanical allodynia relative to preinjection thresholds ($P > 0.13$; see Table 1). The CB_2 antagonist SR144528 blocked the antiallodynic effects of both (*R*,*S*)-AM1241 (10 mg/kg i.p.) and AM1714 (10 mg/kg i.p.; $F_{4,23} = 11.155$, $P < 0.001$; $P < 0.01$ for each comparison; Fig. 6a). Paw withdrawal thresholds in agonist groups pretreated with SR144528 did not differ from the vehicle condition ($P > 0.98$ for each comparison). Post hoc comparisons failed to reveal any differences in the antiallodynic effects induced by either AM1714 (10 mg/kg i.p.) or (*R*,*S*)-AM1241 (10 mg/kg i.p.; $P > 0.98$).

SR141716 (10 mg/kg i.p.) failed to block the antiallodynic effects produced by either (*R*,*S*)-AM1241 (10 mg/kg i.p.) or AM1714 (10 mg/kg i.p.; $F_{4,31} = 10.788$, $P < 0.001$; Fig. 6b). Paw withdrawal thresholds in paclitaxel-treated groups receiving DMSO were lower than those observed in groups receiving the CB_2 agonists in either the presence or absence of the CB_1 antagonist ($P < 0.01$ for each comparison). Paw withdrawal thresholds were similar in groups pretreated with SR141716 to those observed in groups receiving either agonist alone ($P > 0.11$ for each comparison). However, animals receiving SR141716 before AM1714 exhibited elevated paw withdrawal thresholds relative to baseline prepaclitaxel thresholds ($P < 0.01$, planned comparison Student's *t* test; Fig. 6b). Postdrug injection paw withdrawal thresholds were higher in all groups relative to day 21 preinjection thresholds

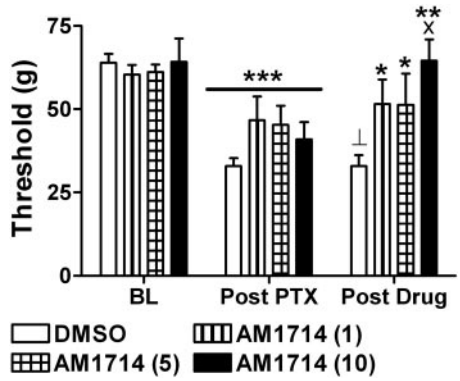

Fig. 5. AM1714 (10 mg/kg) suppressed paclitaxel-induced mechanical allodynia. *, $P < 0.05$; **, $P < 0.01$ different from control; ***, $P < 0.001$ versus baseline (ANOVA and Dunnett post hoc test); X, $P < 0.05$ versus corresponding group day 21 postpaclitaxel paw withdrawal thresholds; ⊥, $P < 0.001$ versus corresponding group baseline prepaclitaxel paw withdrawal thresholds (Student's *t* test). $n = 6$ to 11 per group.

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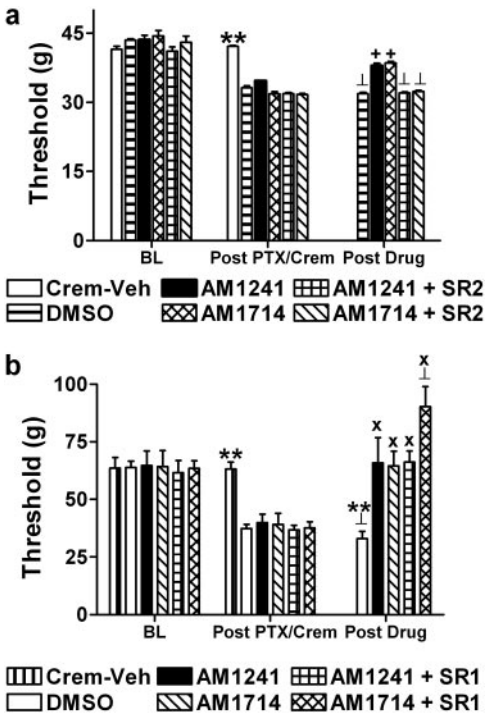

Fig. 6. a, CB_2-selective antagonist SR144528 (SR2) blocked the suppression of paclitaxel-evoked mechanical allodynia induced by the CB_2 agonists, (*R*,*S*)-AM1241 (10 mg/kg i.p.) and AM1714 (10 mg/kg i.p.). b, CB_1-selective antagonist SR141716 (SR1) failed to block the antiallodynic effects of either (*R*,*S*)-AM1241 (10 mg/kg i.p.) or AM1714 (10 mg/kg i.p.) in the same model. **, $P < 0.01$ versus all groups; +, $P < 0.01$ versus DMSO, AM1714 + SR2, and AM1241 + SR2 (ANOVA and Dunnett post hoc test). ⊥, $P < 0.001$ versus corresponding group baseline prepaclitaxel paw withdrawal thresholds (Student's *t* test); X, $P < 0.001$ versus corresponding group day 21 postpaclitaxel thresholds (Student's *t* test). $n = 5$ to 11 per group.

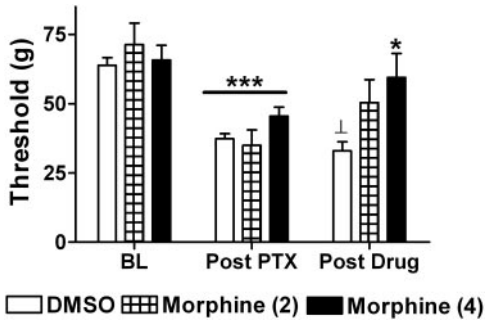

Fig. 7. Morphine (4.0 mg/kg i.p.) blocked mechanical allodynia induced by treatment with paclitaxel. ***, $P < 0.001$ versus baseline; *, $P < 0.05$ versus control (ANOVA and Dunnett post hoc test). ⊥, $P < 0.001$ versus corresponding group baseline prepaclitaxel paw withdrawal thresholds (Student's *t* test). $n = 6$ to 11 per group.

with the exception of vehicle ($P < 0.05$, planned comparison Student's *t* tests).

Effects of Morphine on Paclitaxel-Evoked Mechanical Allodynia. The high dose of morphine (4 mg/kg i.p.) suppressed paclitaxel-induced mechanical allodynia relative to the vehicle condition ($F_{2,20} = 6.023$, $P < 0.01$; $P < 0.01$ for relevant comparison; Fig. 7) and normalized paw withdrawal thresholds relative to prepaclitaxel baseline thresholds ($P > 0.15$). The low dose of morphine (2 mg/kg i.p.) failed to alter postpaclitaxel paw withdrawal thresholds.

Discussion

Two structurally distinct CB_2 agonists attenuated mechanical allodynia induced by treatment with the chemotherapeutic agent paclitaxel. Animals receiving paclitaxel remained in relatively good health as evidenced by the observation of normal weight gain during the course of chemotherapy treatment. However, one fatality was observed after two injections of paclitaxel. Paclitaxel-evoked mechanical hypersensitivity cannot be attributed to sensitization to repeated testing; paw withdrawal thresholds were stable in animals receiving the Cremophor EL/ethanol/saline vehicle in lieu of paclitaxel over the same time course. Mechanical allodynia was observed in paclitaxel-treated animals tested weekly up to 3 months after the initiation of chemotherapy treatment in a pilot study. Paw withdrawal thresholds were similarly reduced relative to baseline from days 14 to 72 postpaclitaxel in this study; therefore, day 21 was selected for the evaluation of drug effects on paclitaxel-evoked mechanical allodynia. Other studies have similarly reported peaks in neuropathic nociception with the present paclitaxel dosing paradigm from days 16 to 27 postinitiation of paclitaxel treatment (Polomano et al., 2001; Flatters and Bennett, 2004). In all subsequent studies, mechanical allodynia developed by day 11 and continued to decrease until the final test day, day 21.

Thermal hyperalgesia was not observed in our study, consistent with previous reports using the present paclitaxel dosing schedule (Polomano et al., 2001). A CB_1-mediated suppression of paclitaxel-induced thermal hyperalgesia has been reported using a cumulative paclitaxel dose of 4 mg/kg (Pascual et al., 2005) compared with our dose of 8 mg/kg. Differences in dosing and timing of paclitaxel injections may account for differences between these studies.

In our study, two structurally distinct cannabinoid CB_2 agonists, the aminoalklyindole (*R*,*S*)-AM1241 and the cannabilactone AM1714, suppressed paclitaxel-evoked mechanical allodynia through a CB_2-specific mechanism. All doses of AM1714 normalized paw withdrawal thresholds relative to prepaclitaxel levels; however, comparisons with day 21 preinjection thresholds suggest that the high dose (10 mg/kg i.p.) was the most reliably effective dose. The high dose of AM1714 (10 mg/kg i.p.) produced a modest antinociceptive effect in animals treated with the Cremophor EL vehicle in lieu of paclitaxel. In contrast, the high (10 mg/kg i.p.) and middle (5 mg/kg i.p.) but not the low (1 mg/kg i.p.) doses of (*R*,*S*)-AM1241 normalized paw withdrawal thresholds to prepaclitaxel levels without inducing antinociception. Thus, AM1714 but not (*R*,*S*)-AM1241 produced antinociception in addition to suppression of allodynia. The mechanisms underlying these differences remain to be explored.

The suppression of paclitaxel-evoked neuropathic nociception induced by AM1241 and AM1714 is likely to be mediated by CB_2 receptors. First, multiple CB_2 agonists from different chemical classes suppressed paclitaxel-evoked neuropathic nociception. Second, (*R*)-AM1241, but not (*S*)-AM1241, suppressed paclitaxel-evoked mechanical allodynia relative to vehicle treatment and preinjection thresholds, consistent with mediation by CB_2. Third, antiallodynic effects of each agonist were blocked by the CB_2 antagonist SR144528. Fourth, the CB_1 antagonist SR141716 failed to block the antiallodynic effects of either (*R*,*S*)-AM1241 or AM1714.

In our study, a trend toward enhanced antihyperalgesic

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efficacy was observed in groups pretreated with SR141716 before AM1714. This observation may suggest that blockade of CB_1 receptors increases endocannabinoid tone and enhances effects of the CB_2 agonist (Zhang et al., 2008). Enhancement of CB_2 agonist efficacy by CB_1 receptor blockade was apparent with AM1714, but not (*R*,*S*)-AM1241, suggesting possible mechanistic differences between the two agonists. More work is necessary to determine whether (*R*,*S*)-AM1241 and AM1714 preferentially activate different signaling pathways or whether off-target effects could contribute to these differences. (*R*,*S*)-AM1241, a racemic compound, may exhibit partial agonist properties that counteract this tendency. Putative changes in endocannabinoid tone may be induced by blockade of CB_1 to enhance the antiallodynic activity of certain CB_2 agonists under conditions in which the balance between CB_1 and CB_2 receptor activation is altered. Blockade of CB_1 may also facilitate interaction of endogenous ananandamide with non-CB_1 receptors [e.g., transient receptor potential vanilloid (TRPV)1] to contribute to the behavioral phenotype. Nonetheless, neither the CB_1 nor the CB_2 antagonist, administered alone, increased paclitaxel-evoked mechanical allodynia. Our data extend previous work documenting that activation of CB_2 suppresses nociception and central sensitization in a variety of tissue and nerve injury models of persistent pain (Ibrahim et al., 2003; Nackley et al., 2003; Beltramo et al., 2006; Jhaveri et al., 2008).

In the present study, we compared the effects of two enantiomers of (*R*,*S*)-AM1241, (*R*)-AM1241 and (*S*)-AM1241, on paclitaxel-evoked mechanical allodynia. (*R*)-AM1241 binds with 40-fold (Bingham et al., 2007) to 114-fold (Thakur et al., 2005) higher affinity to CB_2 receptors than (*S*)-AM1241. This observation is consistent with the ability of (*R*)-AM1241 to preferentially suppress paclitaxel-evoked mechanical hypersensitivity relative to either vehicle or day 21 preinjection thresholds. Similar effects were not observed with administration of (*S*)-AM1241. However, both enantiomers show notable selectivity for CB_2 over CB_1. Thus, it is important to emphasize that (*S*)-AM1241 cannot be considered an inactive enantiomer of (*R*)-AM1241. This property contrasts with that of other aminoalkylinole agonists in which the enantiomer (e.g., WIN55,212-3) of the active compound (WIN55,212-2) fails to bind to cannabinoid receptors. The fact that (*S*)-AM1241 retains activity at CB_2 may account for the efficacy of (*S*)-AM1241 in models of visceral and inflammatory pain (Bingham et al., 2007) and our failure to differentiate between effects of (*R*)-AM1241 and (*S*)-AM1241 in post hoc analyses. Our studies do not preclude the possibility that CB_2-mediated antiallodynic effects of (*S*)-AM1241 could be detected using a higher dose of (*S*)-AM1241 or a larger sample size. It is also possible that differences in enantiomer efficacy reflect differences in agonist-directed trafficking through different G proteins and signal transduction mechanisms (Shoemaker et al., 2005).

In our study, morphine (4 mg/k i.p.) suppressed paclitaxel-induced mechanical allodynia and normalized paclitaxel-evoked paw withdrawal thresholds to prepaclitaxel levels. This same dose was previously reported to be ineffective in suppressing paclitaxel-evoked mechanical hyperalgesia (Flatters and Bennett, 2004). In this latter study, a 2-fold higher dose (8 mg/kg i.p.) than that used here (4 mg/kg i.p.) produced only a 50% reversal of paclitaxel-evoked mechanical allodynia/hyperalgesia, whereas the lower dose (4 mg/kg i.p.) was ineffective. A dose of 8 mg/kg also attenuated vincristine-induced mechanical allodynia in our previous work (Rahn et al., 2007). Differences in the dependent measure (i.e., paw withdrawal frequency versus paw withdrawal threshold in our study), method for assessing mechanical hypersensitivity (i.e., manual von Frey filaments versus electronic von Frey device in our study), and time of testing (i.e., 1 h versus 30 min postmorphine in our study) may account for these differences. Nonetheless, unwanted side effects (i.e., sedation, nausea, altered mental status, constipation) remain associated with activation of the opioid system in humans, warranting development and validation of drug targets that lack these unwanted side effects (Lee et al., 1995).

The mechanism by which paclitaxel induces neuropathic pain symptoms remains unknown. Paclitaxel has been reported to induce neuropathy in the absence of morphological changes in sensory or motor axons in the spinal cord (Polomano et al., 2001). This observation prompted investigations of morphological changes in the periphery. Morphological and immunological changes in sensory nerve fibers have been reported after paclitaxel treatment (Jin et al., 2008). Abnormal calcium homeostasis may also contribute to the development of neuropathic pain symptoms associated with paclitaxel treatment (Siau and Bennett, 2006). Thus, it is noteworthy that blockade of calcium channels is effective in attenuating symptoms of peripheral neuropathy in this model, whereas an *N*-methyl-D-aspartate receptor antagonist was without effect (Flatters and Bennett, 2004). A reduction of mechanical hyperalgesia associated with both paclitaxel and vincristine treatment is also observed in TRPV4 knockout mice, suggesting that TRPV4 may also represent a therapeutic target for treatment of chemotherapy-evoked toxic neuropathy (Alessandri-Haber et al., 2008).

More work is necessary to identify the site of action for CB_2 agonists in suppressing paclitaxel-evoked neuropathy. Up-regulation of the CB_2 receptor in the dorsal horn of the spinal cord has been reported after spinal nerve ligation injury or sciatic nerve sectioning in rats (Walczak et al., 2005; Wotherspoon et al., 2005). Moreover, CB_2 expression is up-regulated in cultured DRG after prior axotomy (Wotherspoon et al., 2005). CB_2 receptors have recently been localized within the CNS, specifically on microglia, which are related to macrophages (Cabral et al., 2008). Thus, it is noteworthy that paclitaxel increased the number of macrophages present in both the spinal cord and the DRG (Peters et al., 2007). More work is necessary to determine whether CB_2 receptors in the CNS or DRG are up-regulated by paclitaxel treatment and contribute to the observed CB_2-mediated suppression of paclitaxel-evoked neuropathy.

The recent observation of increased activation of microglia and astrocytes in paclitaxel-treated rats has led to speculation that these glial cells contribute to chemotherapy-induced neuropathic pain (Ledeboer et al., 2007). Paclitaxel increases levels of activated microglia in laminae III to VI of the spinal cord and astrocytes in laminae I to VI of the spinal cord (Peters et al., 2007). Hypertrophy in both glial cell populations is observed after paclitaxel treatment (Peters et al., 2007). Moreover, pharmacologically induced suppression of glial cell activation abolished and delayed the incidence of mechanical allodynia in paclitaxel-treated rats (Ledeboer et al., 2007). More work is necessary to determine whether CB_2

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agonists suppress paclitaxel-evoked neuropathy by inhibiting microglial activation.

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References

Alessandri-Haber N, Dina OA, Joseph EK, Reichling DB, and Levine JD (2008) Interaction of transient receptor potential vanilloid 4, integrin, and SRC tyrosine kinase in mechanical hyperalgesia. *J Neurosci* **28:**1046–1057.

Aley KO and Levine JD (2002) Different peripheral mechanisms mediate enhanced nociception in metabolic/toxic and traumatic painful peripheral neuropathies in the rat. *Neuroscience* **111:**389–397.

Beltramo M, Bernardini N, Bertorelli R, Campanella M, Nicolussi E, Fredduzzi S, and Reggiani A (2006) CB_2 receptor-mediated antihyperalgesia: possible direct involvement of neural mechanisms. *Eur J Neurosci* **23:**1530–1538.

Bingham B, Jones PG, Uveges AJ, Kotnis S, Lu P, Smith VA, Sun SC, Resnick L, Chlenov M, He Y, et al. (2007) Species-specific in vitro pharmacological effects of the cannabinoid receptor 2 (CB_2) selective ligand AM1241 and its resolved enantiomers. *Br J Pharmacol* **151:**1061–1070.

Cabral GA, Raborn ES, Griffin L, Dennis J, and Marciano-Cabral F (2008) CB_2 receptors in the brain: role in central immune function. *Br J Pharmacol* **153:**240–251.

Cata JP, Weng HR, Lee BN, Reuben JM, and Dougherty PM (2006) Clinical and experimental findings in humans and animals with chemotherapy-induced peripheral neuropathy. *Minerva Anestesiol* **72:**151–169.

Dougherty PM, Cata JP, Cordella JV, Burton A, and Weng HR (2004) Taxol-induced sensory disturbance is characterized by preferential impairment of myelinated fiber function in cancer patients. *Pain* **109:**132–142.

Flatters SJ and Bennett GJ (2004) Ethosuximide reverses paclitaxel- and vincristine-induced painful peripheral neuropathy. *Pain* **109:**150–161.

Flatters SJ and Bennett GJ (2006) Studies of peripheral sensory nerves in paclitaxel-induced painful peripheral neuropathy: evidence for mitochondrial dysfunction. *Pain* **122:**245–257.

Fox A, Kesingland A, Gentry C, McNair K, Patel S, Urban L, and James I (2001) The role of central and peripheral cannabinoid1 receptors in the antihyperalgesic activity of cannabinoids in a model of neuropathic pain. *Pain* **92:**91–100.

Guindon J and Hohmann AG (2008) Cannabinoid CB_2 receptors: a therapeutic target for the treatment of inflammatory and neuropathic pain. *Br J Pharmacol* **153:**319–334.

Hanus L, Breuer A, Tchilibon S, Shiloah S, Goldenberg D, Horowitz M, Pertwee RG, Ross RA, Mechoulam R, and Fride E (1999) HU-308: a specific agonist for CB(2), a peripheral cannabinoid receptor. *Proc Natl Acad Sci U S A* **96:**14228–14233.

Herzberg U, Eliav E, Bennett GJ, and Kopin IJ (1997) The analgesic effects of *R*(+)-WIN 55,212-2 mesylate, a high affinity cannabinoid agonist, in a rat model of neuropathic pain. *Neurosci Lett* **221:**157–160.

Hohmann AG (2002) Spinal and peripheral mechanisms of cannabinoid antinociception: behavioral, neurophysiological and neuroanatomical perspectives. *Chem Phys Lipids* **121:**173–190.

Holmes FA, Walters RS, Theriault RL, Forman AD, Newton LK, Raber MN, Buzdar AU, Frye DK, and Hortobagyi GN (1991) Phase II trial of Taxol, an active drug in the treatment of metastatic breast cancer. *J Natl Cancer Inst* **83:**1797–1805.

Ibrahim MM, Deng H, Zvonok A, Cockayne DA, Kwan J, Mata HP, Vanderah TW, Lai J, Porreca F, Makriyannis A, et al. (2003) Activation of CB_2 cannabinoid receptors by AM1241 inhibits experimental neuropathic pain: pain inhibition by receptors not present in the CNS. *Proc Natl Acad Sci U S A* **100:**10529–10533.

Jhaveri MD, Elmes SJ, Richardson D, Barrett DA, Kendall DA, Mason R, and Chapman V (2008) Evidence for a novel functional role of cannabinoid CB_2 receptors in the thalamus of neuropathic rats. *Eur J Neurosci* **27:**1722–1730.

Jin HW, Flatters SJ, Xiao WH, Mulhern HL, and Bennett GJ (2008) Prevention of paclitaxel-evoked painful peripheral neuropathy by acetyl-L-carnitine: effects on axonal mitochondria, sensory nerve fiber terminal arbors, and cutaneous Langerhans cells. *Exp Neurol* **210:**229–237.

Khanolkar AD, Lu D, Ibrahim M, Duclos RI Jr, Thakur GA, Malan TP Jr, Porreca F, Veerappan V, Tian X, George C, et al. (2007) Cannabilactones: a novel class of CB_2 selective agonists with peripheral analgesic activity. *J Med Chem* **50:**6493–6500.

Ledeboer A, Liu T, Shumilla JA, Mahoney JH, Vijay S, Gross MI, Vargas JA, Sultzbaugh L, Claypool MD, Sanftner LM, et al. (2006) The glial modulatory drug AV411 attenuates mechanical allodynia in rat models of neuropathic pain. *Neuron Glia Biol* **2:**279–291.

Lee JJ and Swain SM (2006) Peripheral neuropathy induced by microtubule-stabilizing agents. *J Clin Oncol* **24:**1633–1642.

Lee YW, Chaplan SR, and Yaksh TL (1995) Systemic and supraspinal, but not spinal, opiates suppress allodynia in a rat neuropathic pain model. *Neurosci Lett* **199:**111–114.

Malan TP Jr, Ibrahim MM, Deng H, Liu Q, Mata HP, Vanderah T, Porreca F, and Makriyannis A (2001) CB_2 cannabinoid receptor-mediated peripheral antinociception. *Pain* **93:**239–245.

Munro S, Thomas KL, and Abu-Shaar M (1993) Molecular characterization of a peripheral receptor for cannabinoids. *Nature* **365:**61–65.

Nackley AG, Makriyannis A, and Hohmann AG (2003) Selective activation of cannabinoid CB_2 receptors suppresses spinal Fos protein expression and pain behavior in a rat model of inflammation. *Neuroscience* **119:**747–757.

Pascual D, Goicoechea C, Suardíaz M, and Martín MI (2005) A cannabinoid agonist, WIN 55,212-2, reduces neuropathic nociception induced by paclitaxel in rats. *Pain* **118:**23–34.

Peters CM, Jimenez-Andrade JM, Kuskowski MA, Ghilardi JR, and Mantyh PW (2007) An evolving cellular pathology occurs in dorsal root ganglia, peripheral nerve and spinal cord following intravenous administration of paclitaxel in the rat. *Brain Res* **1168:**46–59.

Polomano RC and Bennett GJ (2001) Chemotherapy-evoked painful peripheral neuropathy. *Pain Med* **2:**8–14.

Polomano RC, Mannes AJ, Clark US, and Bennett GJ (2001) A painful peripheral neuropathy in the rat produced by the chemotherapeutic drug, paclitaxel. *Pain* **94:**293–304.

Rahn EJ, Makriyannis A, and Hohmann AG (2007) Activation of cannabinoid CB_1 and CB_2 receptors suppresses neuropathic nociception evoked by the chemotherapeutic agent vincristine in rats. *Br J Pharmacol* **152:**765–777.

Rowinsky EK, Chaudhry V, Forastiere AA, Sartorius SE, Ettinger DS, Grochow LB, Lubejko BG, Cornblath DR, and Donehower RC (1993) Phase I and pharmacologic study of paclitaxel and cisplatin with granulocyte colony-stimulating factor: neuromuscular toxicity is dose-limiting. *J Clin Oncol* **11:**2010–2020.

Schiff PB and Horwitz SB (1980) Taxol stabilizes microtubules in mouse fibroblast cells. *Proc Natl Acad Sci U S A* **77:**1561–1565.

Shoemaker JL, Ruckle MB, Mayeux PR, and Prather PL (2005) Agonist-directed trafficking of response by endocannabinoids acting at CB_2 receptors. *J Pharmacol Exp Ther* **315:**828–838.

Siau C and Bennett GJ (2006) Dysregulation of cellular calcium homeostasis in chemotherapy-evoked painful peripheral neuropathy. *Anesth Analg* **102:**1485–1490.

Thakur GA, Nikas SP, Li C, and Makriyannis A (2005) Structural requirements for cannabinoid receptor probes. *Handb Exp Pharmacol* 209–246.

Walczak JS, Pichette V, Leblond F, Desbiens K, and Beaulieu P (2005) Behavioral, pharmacological and molecular characterization of the saphenous nerve partial ligation: a new model of neuropathic pain. *Neuroscience* **132:**1093–1102.

Wotherspoon G, Fox A, McIntyre P, Colley S, Bevan S, and Winter J (2005) Peripheral nerve injury induces cannabinoid receptor 2 protein expression in rat sensory neurons. *Neuroscience* **135:**235–245.

Yao BB, Mukherjee S, Fan Y, Garrison TR, Daza AV, Grayson GK, Hooker BA, Dart MJ, Sullivan JP, and Meyer MD (2006) In vitro pharmacological characterization of AM1241: a protean agonist at the cannabinoid CB_2 receptor? *Br J Pharmacol* **149:**145–154.

Zhang M, Martin BR, Adler MW, Razdan RK, Ganea D, and Tuma RF (2008) Modulation of the balance between cannabinoid CB(1) and CB(2) receptor activation during cerebral ischemic/reperfusion injury. *Neuroscience* **152:**753–760.

Zimmer A, Zimmer AM, Hohmann AG, Herkenham M, and Bonner TI (1999) Increased mortality, hypoactivity, and hypoalgesia in cannabinoid CB_1 receptor knockout mice. *Proc Natl Acad Sci U S A* **96:**5780–5785.

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