Regression of Fibrosis after Chronic Stimulation of Cannabinoid CB2 Receptor in Cirrhotic Rats

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Received September 19, 2007; accepted November 19, 2007

ABSTRACT

Two cannabinoid (CB) receptor subtypes, CB1 and CB2, have been cloned and characterized. Among other activities, receptor activation by cannabinoid ligands may result in pro- or antifibrogenic effects depending on their interaction with CB1 or CB2, respectively. In the current study, we investigated whether selective activation of hepatic CB2 modifies collagen abundance in cirrhotic rats with ascites. mRNA and protein expression of CB receptors in the liver of control and cirrhotic rats was assessed by reverse transcription-polymerase chain reaction, Western blot, and immunohistochemistry. The effect of chronically activating the CB2 receptor was investigated in cirrhotic rats with ascites treated daily (9 days) with the CB2 receptor-selective agonist 3-(1,1-dimethylbutyl)-1-deoxy-D8-tetrahydrocannabinol (JWH-133). At the end of treatment, mean arterial pressure and portal pressure were measured, and liver samples were obtained to evaluate infiltrate of mononuclear cells, hepatic apoptosis, α-smooth muscle actin (SMA) expression, collagen content, and matrix metalloproteinase (MMP)-2 abundance in all animals. JWH-133 improved arterial pressure, decreased the inflammatory infiltrate, reduced the number of activated stellate cells, increased apoptosis in nonparenchymal cells located in the margin of the septa, and decreased fibrosis compared with cirrhotic rats treated with vehicle. This was associated with decreased α-SMA and collagen I and increased MMP-2 in the hepatic tissue of cirrhotic rats treated with the CB2 agonist compared with untreated cirrhotic animals. Therefore, selective activation of hepatic CB2 receptors significantly reduces hepatic collagen content in rats with pre-existing cirrhosis, thus raising the possibility of using selective CB2 agonists for the treatment of hepatic fibrosis in human cirrhosis.

The discovery of specific membrane receptors of the marijuana component Δ8-tetrahydrocannabinol in the early 1990s led to the identification of a novel endogenous signaling pathway, now known as the endocannabinoid system (Felder et al., 1992). This system is made up of the cannabinoid receptors, their endogenous ligands (or endocannabinoids), and the proteins for their synthesis and inactivation. The endogenous cannabinoid family includes anandamide (AEA), 2-arachydonyl glycerol, virodhamine, noladin ether, and Δ9-arachidonoyl-dopamine. These substances promote their action through cannabinoid (CB) receptors. Two CB receptors, CB1 (Matsuda et al., 1990) and CB2 (Munro et al., 1993), have been cloned and characterized. Pharmacological evidence has suggested the presence of another as yet uncloned cannabinoid receptor (Begg et al., 2005). Moreover, AEA also interacts with the transient receptor potential (TRP) vanilloid type 1 protein, which is also known as the VR1 receptor and belongs to the large family of TRP ion channels (Zygmont et al., 1999). CB1 receptor is abundant in the brain, and it is involved in the control of motor activity.

ABBREVIATIONS: AEA, anandamide; CB, cannabinoid; TRP, transient receptor potential; HSC, hepatic stellate cell(s); HPRT, hypoxanthine-guanine phosphoribosyl transferase; bp, base pair(s); RT, reverse transcription; PCR, polymerase chain reaction; JWH-133, 3-(1,1-dimethylbutyl)-1-deoxy-D8-tetrahydrocannabinol; MAP, mean arterial pressure; MMP, matrix metalloproteinase; SMA, smooth muscle actin; TUNEL, terminal deoxynucleotidyl transferase dUTP nick-end labeling.
memory and cognition, emotion, sensory perception, and autonomic and endocrine functions. In addition, the CB1 receptor is expressed in peripheral nerve terminals and vascular endothelium. In contrast, CB2 receptors are expressed mainly in immune and hematopoietic cells, the modulation of cytokine release being one of their roles (Pacher et al., 2006).

Endocannabinoids may behave as pro- or antifibrogenic substances depending on their interaction with CB1 or CB2 receptors, respectively (Julien et al., 2005; Siegmund et al., 2005; Pacher et al., 2006). This raised the possibility that pharmacological modulation of the hepatic endocannabinoid system could be a valuable therapy in cirrhosis. In fact, activation of the CB1 receptor triggers fibrosis progression (Teixeira et al., 2006), whereas CB2 activation promotes antifibrogenic actions in the liver (Julien et al., 2005). In vitro and in vivo studies with CB2 knockout mice demonstrate that CB2 receptors inhibit the proliferation of hepatic stellate cells (HSC) and stimulate apoptosis through two different signaling routes involving induction of cyclooxygenase-2 and intracellular oxidative stress, respectively (Julien et al., 2005). In addition, selective blockade of CB1 receptors has been shown to inhibit fibrogenesis in mice (Teixeira et al., 2006). In the current study, we alternatively investigated whether selective long-term activation of hepatic CB2 receptors in rats with pre-existing cirrhosis and ascites decreases hepatic collagen abundance and therefore reverses fibrosis.

Materials and Methods

Induction of Cirrhosis in Rats. Studies were performed in male adult Wistar rats with cirrhosis, without or with ascites and in male adult Wistar control rats (Charles-River, Saint Aubin les Elseuf, France). Cirrhosis was induced by CCl4 following a method described previously (Claria and Jimenez, 1999). Cirrhotic rats without ascites were studied between 11 and 12 weeks after starting the cirrhosis induction program. The absence of ascites was confirmed by laparotomy. Cirrhotic rats with ascites were studied between 13 and 17 weeks after ascites had fully developed. Control rats were studied after a similar period of phenobarbital administration. The study was performed according to the criteria of the Investigation and Ethics Committee of the Hospital Cliníc Universitari.

CB1 and CB2 mRNA Expression in Hepatic Tissue. Total RNA was extracted from the middle liver lobe of control and cirrhotic rats and brain and spleen from control rats using a commercially available kit (TRizol Reagent; Invitrogen, Carlsbad, CA). One microgram of total RNA was reverse transcribed (RT) by using a complementary DNA synthesis kit (Roche Applied Science, Indianapolis, IN). Primers for the CB1-receptor (sense, 5′-TGT-GGGCAGGCTGTCCCTCA-3′; antisense, 5′-GGGTTTGGCCAGCCTAATGGTC-3′) and for the CB2-receptor (sense, 5′-TCCCCCTT-GATCCTCCCCGACCTA-3′; antisense, 5′-CTTCCACTCCTCGCACGGCATAATG-3′), were prepared according to rat CB1 and CB2 mRNA sequences (GenBank accession numbers: NM_012784 and NM_012785, respectively). Primers were also synthesized to amplify the cDNA encoding HPRT, a constitutively expressed gene, as control. HPRT primers were designed as described previously (Tugues et al., 2005), giving rise to a 264-bp polymerase chain reaction (PCR) product from the cDNA base sequence of rat HPRT. PCR was performed for CB1, CB2, and HPRT using a DNA amplification kit (Invitrogen). The PCR products were sequenced to check correct amplification.

CB1 and CB2 Protein Expression in Hepatic Tissue of Control and Cirrhotic Rats. Samples were individually homogenized (PT 10-35 Polytron; Kinematica, Krienli-Luzern, Switzerland) in a 20 mM Tris-HCl, pH 7.4, containing 1% Triton X-100, 0.1% SDS, 50 mM NaCl, 2.5 mM EDTA, 1 mM Na3PO4·10H2O, 20 mM NaF, 1 mM Na2VO4, 2 mM Pefabloc (Roche Diagnostic, Mannheim, Germany), and a cocktail of protease inhibitors (Complete Mini; Roche Diagnostics, Basel, Switzerland). To detect the CB1 receptor, 80 μg of the denatured proteins was run on a 10% SDS-polyacrylamide gel, and then they were transferred to nitrocellulose membranes (Transblot transfer medium; Bio-Rad, Hercules, CA), which were blocked with 5% powdered nonfat milk in TTBS buffer (50 mM Tris-HCl, pH 8, containing 0.05% Tween 20 and 150 mM NaCl) overnight at 4°C. Next, they were incubated with a primary rabbit polyclonal antibody against the CB1 receptor (1:250; Cayman Chemical, Ann Arbor, MI), followed by incubation with horseradish peroxidase-conjugated anti-rabbit antibody (1:5000; GE Healthcare, Little Chalfont, Buckinghamshire, UK). To detect the CB2 receptor, 80 μg of the denatured proteins was run on a 10% SDS-polyacrylamide gel, and then they were transferred to nitrocellulose membranes, which were blocked with 5% powdered nonfat milk in TTBS buffer (50 mM Tris-HCl, pH 8, containing 0.05% Tween 20 and 150 mM NaCl) overnight at 4°C. Next, they were incubated with a primary rabbit polyclonal antibody against the CB2 receptor (1:750; Cayman Chemical), followed by incubation with horseradish peroxidase-conjugated anti-rabbit antibody (1:2000; GE Healthcare). Bands were visualized by chemiluminescence (ECL Western blotting analysis system; GE Healthcare). The relative expression of CB1 and CB2 receptors was determined by densitometric scanning.

Selective Activation of CB2 Receptors in Cirrhotic Rats with Ascites. Cirrhotic rats with ascites were included in the protocol after developing stable ascites. Thereafter, CCl4 treatment was discontinued, and animals were randomly assigned to one of the following groups: group A, daily s.c. injection of the CB2 receptor-selective agonist JWH-133 (1 mg/kg b.wt.; Toceir Cookson, Inc., Bristol, UK; Huffman et al., 1999) for 9 days beginning the second week after the detection of sustained ascites; and group B, daily s.c. injection of saline solution (1 mg/kg b.wt.) containing 5% ethanol. At the end of the treatment, animals were anesthetized with inactin (50 mg · kg−1) and mean arterial pressure (MAP) and portal pressure were recorded as described previously (Ros et al., 2005). Liver specimens were obtained from each animal, washed in 0.1% diethyl pyrocarbonate-treated phosphate-buffered saline salt solution (140 mM NaCl, 8.5 mM Na2HPO4, and 1.84 mM NaH2PO4·H2O; pH 7.4), immediately frozen in dry ice, and stored in liquid nitrogen to evaluate collagen type I and matrix metalloproteinase (MMP)-2 expression. Liver samples from treated and untreated animals were also fixed in 10% buffered formalin for further hematoxylin and eosin and immunostaining analysis.

Immunodetection of α-SMA, CD68, and α-SMA-Positive Cells. Liver sections from cirrhotic rats with ascites chronically receiving the CB2 agonist or vehicle underwent microwave antigen retrieval to unmask antigens hidden by cross-linkage occurring during tissue fixation. Endogenous peroxidase activity was blocked by hydrogen peroxide pretreatment for 10 min, and it was then further blocked by incubation with 5% goat serum for 45 min. Sections were then stained with rabbit polyclonal anti-rat CB2 antibody (1:300; Cayman Chemical), mouse anti-rat CD68, a lysosomal protein present in activated macrophages (clone ED1, 1:150; Serotec, Kidlington, Oxford, UK), or with mouse anti-rat α-smooth muscle actin (SMA) (1:1200; Dako Denmark A/S, Glostrup, Denmark) and incubated overnight at 4°C or 1.5 and 1 h, respectively, at room temperature. The LSAB 2 System-HRP (Dako Denmark A/S) was used for antigen detection, and antigen visualization was achieved with streptavidin peroxidase and counterstained with hematoxylin. Macrophages (CD68-positive cells) in the middle and margin of the septa were assessed by counting 16 random fields (magnification 1000×) per each section. The mean cell count for each sample was calculated.

The area of α-SMA-positive staining was visualized using a digital microscope (Eclipse E-600; Nikon, Kawasaki, Japan). Images were processed using a morphometric analysis system (AnalySIS, version...
3.2 Soft Imaging System (Olympus, Tokyo, Japan). The percentage of immunostained/fields areas of digital photomicrographs was then quantified. As negative controls, immunostaining was performed without the first antibody.

**Apoposis in Hepatic Tissue.** We used the terminal deoxynucleotidyl transferase dUTP nick-end labeling (TUNEL) assay to detect cell death using the fluorescein-FragEL DNA fragmentation detection kit (Calbiochem, San Diego, CA) according to the manufacturer's protocol. To quantify and compare the rates of cell death between groups, a semiquantitative scoring method was used. For each sample, the number of TUNEL-positive cells was counted per 200× high-power field. At least eight representative fields were evaluated for each treatment group, from which an average value was calculated.

**Fibrosis Quantification.** Liver sections (4 μm) were stained in 0.1% Sirius red F3B (Sigma-Aldrich, St. Louis, MO) in saturated picric acid (Sigma-Aldrich). Relative fibrosis area (expressed as a percentage of total liver area) was assessed by analyzing 36 fields of Sirius red-stained liver sections per animal. Each field was acquired at 10× magnification [E600 microscope (Nikon) and RT-Slider SPOT digital camera (Diagnostic Instruments, Inc., Sterling Heights, MI)], and then it was analyzed using a computerized Bioquant Life Science morphometry system. To evaluate the relative fibrosis area, the measured collagen area was divided by the net field area and then multiplied by 100. Subtraction of vascular luminal area from the total field area yielded the final calculation of the net fibrosis area. From each animal analyzed, the amount of fibrosis as percentage was measured and the average value presented.

**Western Blot Analysis of Collagen Type I, MMP-2, α-SMA, and Activated Caspase-3.** Hepatic tissue from treated and nontreated cirrhotic rats with ascites was individually homogenized as described previously. To detect collagen type I and MMP-2, 120 and 80 μg, respectively, of total denatured proteins were loaded on a 7.5% SDS-polyacrylamide gel. To detect α-SMA, 120 μg of total proteins was separated on a 10% SDS-polyacrylamide gel. To detect activated caspase-3, 80 μg of total proteins was separated on a 12% SDS-polyacrylamide gel (Mini Protein III; Bio-Rad). Gels for collagen type I and activated caspase-3 were transferred to nitrocellulose membranes for 2 h and blocked with 5% powdered nonfat milk in TTBS buffer overnight at 4°C. Gels for MMP-2, and α-SMA, were transferred overnight at 4°C to nitrocellulose membranes. All membranes were stained with Ponceau S Red as a control for protein loading, and then they were incubated at room temperature with mouse monoclonal anti-collagen type I (1:500 dilution; Abcam plc, Cambridge, UK), MMP-2 (1:750 dilution; Neomarkers, Fremont, CA), and α-SMA (1:250 dilution; Dako Denmark A/S) for 2 h or incubated at 4°C with rabbit polyclonal anti-activated caspase-3 (1:300 dilution; Abcam plc) for 48 h. The bands for collagen type I, MMP-2, α-SMA, and activated caspase-3 were visualized by chemiluminescence (ECL Western blotting analysis system; GE Healthcare).

**Immunofluorescence.** For immunofluorescence, tissues were fixed in 10% buffered formaldehyde solution and embedded in paraffin. Sections were underwent microwave antigen retrieval, blocked with 5% normal goat serum, and incubated with rabbit anti-active caspase-3, mouse anti-α-SMA, or mouse anti-CDB68 antibodies. In addition, 4,6-diamidino-2-phenylindole (Vectashield; Vector laboratories, Burlingame, CA) was used to counterstain cell nuclei. Controls without primary antibodies were used as negative controls. Binding sites of the primary antibodies were revealed with cyanine-3-conjugated goat anti-rabbit IgG and with fluorescein isothiocyanate-conjugated goat anti-rabbit IgG (Jackson ImmunoResearch Laboratories Inc., West Grove, PA). Samples were visualized with a fluorescence microscope (Eclipse E600).

**Measurements and Statistical Analysis.** Serum osmolality was determined from osmometric depression of the freezing point (Osmometer 3300; Advanced Instruments, Needham Heights, MA) and sodium concentration by flame photometry (IL 943; Instrumentation Laboratory, Lexington, MA). Serum albumin, alanine aminotransferase and lactate dehydrogenase were measured by the ADVIA 1650 Instrument (Siemens Medical Solutions Diagnostics, Tarrytown, NY).

Statistical analysis of results was performed by unpaired Student's t tests when appropriate. Data are expressed as mean ± S.E.M., and they are considered significant at a p level of 0.05 or less.

**Results**

In total, the liver of the animals treated with CCl4 included in the study had a finely granulated surface, and histological examination showed the characteristic features of cirrhosis. In those animals with ascites, the ascites volume ranged between 5 and 60 ml. Control rats displayed no appreciable alterations in the liver histology.

**CB1 and CB2 mRNA Expression.** DNA amplification products obtained from hepatic tissue of six cirrhotic and three control rats are shown in Fig. 1A. Total RNA was also obtained from brain and spleen of normal rats for positive controls of CB1 and CB2 receptors, respectively. Bands of 425, 369, and 264 bp corresponding to CB1 and CB2 receptors and HPRT mRNAs, respectively, were detected in all the samples analyzed, thus demonstrating expression of these transcripts in the liver tissue. Densitometric analysis of these results is shown in Fig. 1B. Both CB1 and CB2 transcript abundance was significantly higher in samples obtained from cirrhotic rats, regardless of whether they were obtained from animals without or with ascites. Therefore, gene expression of cannabinoid receptors was markedly activated in the cirrhotic liver.

**CB1 and CB2 Protein Expression.** Western blot analysis of CB1 receptor yielded a specific band at the expected molecular mass of ~53 kDa in the positive control. In contrast to the clear band found in the RT-PCR assays, no signal was detected in protein extracts isolated from hepatic tissue of both cirrhotic and control animals (Fig. 2). Because these findings were reproduced after using a different type of monoclonal anti-CB1 antibody (1:250; Chemicon International, Temecula, CA), these results probably reflect the relatively low abundance or dilution of this protein in whole hepatic tissue because the majority of cells (i.e., hepatocytes) did not express the receptor. Samples from liver of cirrhotic and control rats showed a specific band of ~51 kDa that was identified as CB2 protein, based on identical size as the positive control (Fig. 2A). Paralleling the increased CB2 mRNA, enhanced abundance of CB2 protein was detected in cirrhotic livers compared with controls. It is of interest that this increase in CB2 protein expression was mainly localized in portal tracts and fibrous septa (Fig. 2B).

**Liver Function Tests, Mean Arterial Pressure, and Portal Pressure in Treated and Nontreated Cirrhotic Rats with Ascites.** Table 1 shows biochemical tests of liver function and serum electrolytes in both groups of cirrhotic rats. No significant differences were observed in any of these parameters between treated and nontreated cirrhotic rats. Furthermore, cirrhotic rats receiving vehicle had significant hemodynamic dysfunction, as reflected by marked portal hypertension (13.9 ± 1.5 mm Hg) and arterial hypotension (80 ± 4 mm Hg). Administration of the CB2 receptor agonist to cirrhotic rats for 9 days resulted in a significant improvement in MAP (93 ± 2 mm Hg; p < 0.05) in the absence of any change in portal pressure (14.1 ± 1.0 mm Hg).
Effects of CB2 Receptor Activation on Infiltrating and Fibrogenic Cell Density and Apoptosis. Density of infiltrating monocytes/macrophages in the liver tissue of cirrhotic rats with ascites was assessed by determining the amount of CD68-positive cells within and at the margin of the portal tracts and septa. CD68 antigen was expressed in both JWH-133-treated and nontreated cirrhotic rats. The morphology of cells expressing CD68 in these regions of the liver tissue of three control and six cirrhotic rats. A, HPRT was amplified as a housekeeping gene. B, densitometric analysis of RT-PCR is shown at the bottom.
liver sections predominantly occurred as round/oval-shaped cells (Fig. 3A), in accordance with infiltrating monocytes/macrophages. Of note was that the density of CD68+ staining cells within and around the margins of the fibrous bands was significantly lower in the cirrhotic livers chronically treated with the CB2 receptor agonist compared with the cirrhotic livers receiving vehicle (Fig. 3B).

Activated HSC acquire a myofibroblastic phenotype that is characterized by expression of α-SMA, which is not detected in quiescent HSC. Thus, α-SMA is a well-validated marker of activated HSC (Schmitt-Graeff et al., 1991). We detected α-SMA as linear staining in the portal tracts and fibrous septa, which was intense in the severely fibrotic tissue of both groups of cirrhotic rats (Fig. 4A). However, staining was more diffuse in liver tissue of rats receiving JWH-133 than in the liver of vehicle-treated cirrhotic animals. As shown in Fig. 4B, the percentage of the liver tissue that stained for α-SMA was significantly reduced in rats after CB2 receptor agonist stimulation compared with paired untreated cirrhotic animals.

To explore whether any cell populations were driven to apoptosis by the CB2 agonist, we performed in situ detection of nuclear DNA fragmentation by the TUNEL assay in liver sections of treated and nontreated cirrhotic rats. As a positive control of the TUNEL assay, apoptosis was induced by incubation of liver sections with DNase I. No staining was observed in the negative control in which terminal deoxynucleotidyl transferase enzyme was omitted (data not shown). Liver sections from cirrhotic rats showed few positive TUNEL staining cells, with immunoreactivity localized to the margin of the fibrous septa (Fig. 5A). However, the number of positive cells for TUNEL staining significantly increased in hepatic sections of animals treated with JWH-133 compared with the vehicle group (8.4 ± 0.4 versus 6.4 ± 0.3 positive cells/field; respectively; p < 0.05). In addition, we measured the amount of active caspase-3 in livers of vehicle- or JWH-133-treated animals. As shown in Fig. 5B, the amount of activated caspase-3 was significantly higher in cirrhotic rats treated with JWH-133 than in the vehicle group. It is of interest that activated caspase-3 immunostaining colocalized with α-SMA and CD68-positive cells (Fig. 6).

Effect of CB2 Receptor Activation on Liver Fibrogenesis in Cirrhotic Rats with Ascites. Sirius red, a dye that selectively binds collagen proteins, was used to stain the collagen fibrils in the liver of CCl4-treated rats (Jimenez et al., 1985). As shown in Fig. 7A, both groups of rats had abundant fibrosis showing a characteristic pattern of perivenular and periportal deposition of connective tissue with development of portal-to-portal septa, ending blind in the parenchyma, and marked architectural distortion resulting in micronodular cirrhosis. However, biopsies obtained from cirrhotic rats receiving JWH-133 displayed thinner septa and more preserved hepatic parenchyma than nontreated cirrhotic animals. This was confirmed by the morphometric analysis of all Sirius red-stained sections in which samples of rats after chronic CB2 receptor stimulation showed a significant reduction in the percentage of cirrhotic parenchymal area.

### Table 1

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<th>Vehicle (n = 7)</th>
<th>JWH-133 (n = 8)</th>
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<tr>
<td>Body wt (g)</td>
<td>482 ± 3</td>
<td>459 ± 15</td>
</tr>
<tr>
<td>Alanine aminotransferase (U/l)</td>
<td>53.6 ± 8.7</td>
<td>89.0 ± 10.0</td>
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<tr>
<td>Lactate dehydrogenase (U/l)</td>
<td>620 ± 81</td>
<td>578 ± 98</td>
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<tr>
<td>Total bilirubin (mg/dl)</td>
<td>1.10 ± 0.26</td>
<td>1.10 ± 0.60</td>
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<tr>
<td>Albumin (g/l)</td>
<td>23.2 ± 1.3</td>
<td>26.3 ± 0.9</td>
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<tr>
<td>Serum sodium (mEq/l)</td>
<td>144 ± 1</td>
<td>143 ± 2</td>
</tr>
<tr>
<td>Serum potassium (mEq/l)</td>
<td>4.7 ± 0.4</td>
<td>4.0 ± 0.5</td>
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<tr>
<td>Serum osmolality (mOsm/kg)</td>
<td>293 ± 3</td>
<td>299 ± 3</td>
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![Image](https://via.placeholder.com/150)
fibrosis area than sections of vehicle-treated cirrhotic rats (Fig. 7B).

**Effect of CB2 Receptor Activation on Protein Expression of α-SMA, Collagen I, and MMP-2 in the Liver of Cirrhotic Rats.** In brief, we examined the effect of JWH-133 on hepatic expression of α-SMA, as a marker of HSC activation; collagen I, a fibril-forming collagen that predominates in chronic liver disease; and MMP-2, a matrix metalloproteinase that degrades collagen and other matrix proteins. As shown in Fig. 8, α-SMA and collagen I levels were significantly reduced in cirrhotic rats treated with vehicle or JWH-133 (1 mg/kg b.wt. for 9 days). Eighty micrograms of protein extracts was loaded per lane. Numbers below the panels indicate relative levels based on densitometry.

**Discussion**

Achieving regression of fibrosis has been a major challenge in patients with advanced liver disease. Although it was initially thought that hepatic cirrhosis was an irreversible phenomenon, there are currently numerous experimental indications suggesting that regression of hepatic cirrhosis is a realistic endpoint of therapy. Indeed, several resident hepatic cell types can degrade cellular matrix through the enzymatic action of MMPs, particularly those possessing type I collagenase activity (Han et al., 2004; Iredale, 2004; Siller-López et al., 2004). Moreover, different degrees of fibrosis regression have been demonstrated in a wide array of liver diseases, including viral hepatitis (Dufour et al., 1997; Kweon et al., 2001, Dienstag et al., 2003), biliary obstruction (Hammel et al., 2001), and non-alcoholic steatohepatitis (Dixon et al., 2004).

Recent studies in patients with chronic hepatitis C have demonstrated that daily cannabis smoking is an independent predictor of fibrosis progression (Hézode et al., 2005) and
that steatosis severity is closely related to cannabis use (Hézode et al., 2007). These effects seem to be mediated through the interaction with the CB1 receptor, because activation of this receptor induces fatty acid synthesis and contributes to diet-induced obesity in mice (Osei-Hyiaman et al., 2005). Altogether, these results suggest that CB1 receptor activation could mediate fibrogenesis, thereby establishing the rationale for CB1 receptor blockade as an antifibrogenic therapeutic strategy in liver disease. This has been further supported by the results of Teixeira-Clerc et al. (2006), demonstrating that CB1 receptor antagonism decreases wound healing response to acute liver injury and the progression of fibrosis in different experimental models of liver disease. In contrast, increased cardiac and hepatic content of endogenous cannabinoids has been reported in cirrhosis (Bátkai et al., 2007a). This is probably the consequence of the increased

Fig. 7. Effect of CB2 receptor activation on liver fibrosis. A, Sirius Red staining of a representative liver section obtained from a cirrhotic rats with ascites treated with vehicle (n = 7) or receiving JWH-133 (1 mg/kg b.wt. for 9 days; n = 8). Original magnification, 200x. B, quantification of relative fibrosis area was assessed in 36 fields per animal.

Fig. 8. Effect of CB2 receptor activation on protein expression of α-SMA, collagen I, and MMP-2. Left, representative Western blot for α-SMA, collagen I, and MMP-2 in the liver tissue of cirrhotic rats with ascites receiving vehicle or chronically treated with JWH-133 (1 mg/kg b.wt. for 9 days). Protein extract (120, 120, and 80 g) was loaded per lane, respectively. Protein extracts from rat thoracic aorta, skin, and thoracic aorta were used as positive control for α-SMA, collagen I, and MMP-2, respectively. Right, densitometric analysis of all samples (seven nontreated and eight treated cirrhotic rats).
endocannabinoid production by decreasing hepatic inflammation molecules in endothelial cells adjacent to areas of local inflammatory infiltrate, which favors the attraction of circulating cells to the activated vasculature where they adhere to and migrate into the liver parenchyma. This pattern of endothelial activation after liver injury is quite similar to that observed after hepatic ischemia reperfusion. In this latter condition, CB2 receptor agonists protect against the hepatic insult by decreasing endothelial cell activation, inflammatory response, expression of adhesion molecules, inflammatory cytokines and recruitment, and adhesion and activation of inflammatory cells (Bátkai et al., 2007b; Rajesh et al., 2007). Therefore, the antifibrogenic effect of CB2 stimulation in the cirrhotic animals could also be likely related to the anti-inflammatory effect associated with CB2 receptor stimulation.

Although the mechanisms of its action are not fully clarified, JWH-133 could act mainly by direct action on hepatic mesenchymal, fibrogenic cells. This view is supported by Julien et al. (2005), showing that CB2 receptors are strongly expressed in nonparenchymal cells and biliary cells located within and at the edges of fibrotic septa. These authors also showed that the growth arrest and apoptotic effects of cannabinoid receptor ligands are selectively mediated by CB2 receptors in human hepatic myofibroblasts. However, the current investigation could not exclude the possibility that JWH-133 exerts its beneficial effect through mechanisms other than those mediated by CB2 receptors. Siegmund et al. (2005) recently reported that the endocannabinoid AEA is a killer of activated HSC in vitro, a phenomenon not dependent on CB1, CB2, or TRPV vanilloid type 1 protein receptor activation. Rather, the authors suggested that AEA induces cellular necrosis through its interaction with the membrane cholesterol, resulting in reactive oxygen species formation, intracellular 

There are numerous experimental data indicating that by interacting with vascular CB1 receptors, endocannabinoids are involved in the pathogenesis of the circulatory dysfunction occurring in advanced liver disease (Bátkai et al., 2001, 2007a; Ros et al., 2002; Domenicali et al., 2005). It is possible that among the most conclusive evidence is the fact that CB1 receptor blockade increases MAP in cirrhotic rats (Bátkai et al., 2001; Ros et al., 2002). Therefore, it is noteworthy that in the present investigation administration of the CB2 receptor agonist also improved blood pressure in cirrhotic animals. It has been demonstrated that endocannabinoid release directly correlates with hepatic damage and inflammation in ischemia/reperfusion injury (Bátkai et al., 2007b). Therefore, it is possible that in our study the CB2 agonist reduced endocannabinoid production by decreasing hepatic inflam-
mation, thereby reducing in endocannabinoid-mediated hypotension. This would be further supported by the fact that CB2 agonist administration did not produce significant hemodynamic effects in normal rodents (Batkai et al., 2007b). In summary, the results of the present investigation demonstrate that selective activation of hepatic CB2 receptors for 9 days significantly increases MAP, reduces the density of monocyte/macrophages within and around the margins of the portal tracts, decreases the amount of staining-positive α-SMA cells, increases apoptosis in myofibroblastic and in monocyte cell types, diminishes the tissue content of collagen I and α-SMA, and increases the abundance of the pro- teolytic enzyme MMP-2 in the liver of rats with pre-existing cirrhosis and ascites. These effects result in a significant reduction in the hepatic collagen content of cirrhotic animals, thus raising the possibility of using selective CB2 agonists for the treatment of hepatic fibrosis in advanced human liver disease, including cirrhosis.

References