A-740003 [N-(1-[(Cyanoimino)(5-quinolinylamino)methyl]amino)-2,2-dimethylpropyl]-2-(3,4-dimethoxyphenyl)acetamide], a Novel and Selective P2X7 Receptor Antagonist, Dose-Dependently Reduces Neuropathic Pain in the Rat


ABSTRACT

ATP-sensitive P2X7 receptors are localized on cells of immunological origin including glial cells in the central nervous system. Activation of P2X7 receptors leads to rapid changes in intracellular calcium concentrations, release of the proinflammatory cytokine interleukin-1β (IL-1β), and, following prolonged agonist exposure, cytolytic plasma membrane pore formation. P2X7 knockout mice show reduced inflammation as well as decreased nociceptive sensitivity following peripheral nerve injury. A-740003 (N-(1-[(cyanoimino)(5-quinolinylamino)methyl]amino)-2,2-dimethylpropyl)-2-(3,4-dimethoxyphenyl)acetamide) is a novel competitive antagonist of P2X7 receptors (IC50 values = 40 nM for human and 18 nM for rat) as measured by agonist-stimulated changes in intracellular calcium concentrations. A-740003 showed weak or no activity (IC50 > 10 μM) at other P2 receptors and an array of other neurotransmitter and peptide receptors, ion channels, reuptake sites, and enzymes.

A-740003 potently blocked agonist-evoked IL-1β release (IC50 = 156 nM) and pore formation (IC50 = 92 nM) in differentiated human THP-1 cells. Systemic administration of A-740003 produced dose-dependent antinociception in a spinal nerve ligation model (ED50 = 19 mg/kg i.p.) in the rat. A-740003 also attenuated tactile allodynia in two other models of neuropathic pain, chronic constriction injury of the sciatic nerve and vincristine-induced neuropathy. In addition, A-740003 effectively reduced thermal hyperalgesia observed following intraplantar administration of carrageenan or complete Freund’s adjuvant (ED50 = 38–54 mg/kg i.p.). A-740003 was ineffective in attenuating acute thermal nociception in normal rats and did not alter motor performance at analgesic doses. These data demonstrate that selective blockade of P2X7 receptors in vivo produces significant antinociception in animal models of neuropathic and inflammatory pain.

P2X7 receptors belong to the family of ATP-sensitive ionotropic P2X receptors, which are composed of seven receptor subtypes (P2X1–P2X7) (North, 2002). Unlike other members of the P2 receptor superfamily, homomeric P2X7 receptors are activated by high concentrations of ATP (>100 μM), and 2’3’-O-(4-benzoylbenzoyl)-ATP (BzATP) has significantly greater potency (EC50 = 20 μM) than ATP (EC50 > 100 μM) (Jacobson et al., 2002). P2X7 receptors are found predominantly on macrophages and other cells of immunological origin, where they can trigger a series of cellular responses such as membrane permeabilization, activation of caspases, cytokine release, cell proliferation, and apoptosis (Panenka et al., 2001; Chakfe et al., 2002, North, 2002; Verhoef et al., 2001; Chakfe et al., 2002).
2003; Kahlenberg and Dubyak, 2004). In addition to agonist-induced changes in intracellular calcium concentrations, activation of P2X receptors stimulates caspase-1 activation, release of interleukin-1β (IL-1β), and activation of p38 mitogen-activated protein kinase (Armstrong et al., 2002; Ferrari et al., 2006). Another feature of P2X7 receptors is that prolonged agonist exposure leads to the formation of large cytolytic pores in cell membranes (Surprenant et al., 1996). The underlying mechanisms and functional significance of P2X7-mediated pore formation remain to be elucidated; however, P2X7-mediated pore formation is dependent on intracellular signaling events involving, at least in part, p38 mitogen-activated protein kinase and caspase-1 (Pannicke et al., 2000; North, 2002; Donnelly-Roberts et al., 2004). Both the localization of P2X7 receptors on proinflammatory cells and the demonstration that activation of P2X7 receptors modulates the release of IL-1 have implicated a role for this P2X7 receptor in inflammatory diseases (North, 2002; Baraldi et al., 2003). A proinflammatory role for P2X7 receptors in inflammation was further supported by the demonstration that P2X7 knockout mice showed reduced inflammation in an experimental passive collagen-induced arthritis model (Labasi et al., 2002). More recently, it was demonstrated that P2X7 knockout mice also show reduced inflammatory thermal hyperalgesia and nerve injury-induced mechanical allodynia as compared with matched wild-type mice (Chessell et al., 2005).

Although the expression of P2X7 receptors has been reported in brain, microglia, and astrocytes based on immunoreactivity studies (Collo et al., 1997; Sim et al., 2004), considerable controversy exists regarding the existence of functional P2X7 receptors on peripheral and central neurons due to the poor specificity of primary antibodies and ligands targeting the rat P2X7 receptor (Anderson and Nedergaard, 2006). However, functional P2X7 receptors have been demonstrated on peripheral glial cells in rat dorsal root ganglion, and this expression may play a role in peripheral sensory transduction of pain perception (Zhang et al., 2005). The present study describes the pharmacological characterization of A-740003 (Fig. 1), a structurally novel and highly specific antagonist for mammalian P2X7 receptors that dose-dependently reduces nociception in rodent models of neuropathic and inflammatory pain.

**Materials and Methods**

**In Vitro Characterization of A-740003**

**Materials.** BzATP, pyridoxal phosphate-6-azophenyl-2-4-disulfonylic acid (PAPDS), and Brilliant Blue G were purchased from Sigma-Aldrich (St. Louis, MO). KN-62 was purchased from Toecris (Ellisville, MO). Fluo-4 dye was purchased from TEF Labs (Austin, TX), and Yo-Pro was purchased from Invitrogen (Eugene, OR). All cell culture medium and Dulbecco’s phosphate-buffered saline (DPBS), pH 7.4, were also obtained from Invitrogen (Grand Island, NY).

**Cell Culture.** 1321N1 human astrocytoma cells stably expressing human and rat P2X7, human P2X6, P2X2, P2X2/3, P2X1, P2Y1, and P2Y2 recombinant receptors were grown according to previously published protocols (Bianchi et al., 1999; Lynch et al., 1999). Briefly, 1321N1 cells expressing transfected P2 receptors were maintained in a humidified 5% CO2 atmosphere at 37°C in Dulbecco’s modified Eagle’s medium containing 1% L-alanyl-L-glutamine, 1% antibiotic/antimycotic, 10% fetal bovine serum, and 300 μg/ml genetin. 1321N1 P2X2/3 cells were grown in Dulbecco’s modified Eagle’s medium containing 1% L-alanyl-L-glutamine, 1% antibiotic/antimycotic, 10% fetal bovine serum, and 100 μg/ml hygromycin B. 1321N1 P2X2Δ/Δ cells were maintained in growth medium containing 150 μg/ml genetin and 75 μg/ml hygromycin B. Cells of the THP-1 monocytic cell line (American Type Culture Collection, Rockville, MD) were maintained in the log phase of growth in RPMI 1640 medium containing high glucose and 10% fetal calf serum (Invitrogen) according to established procedures (Humphrey and Dubyak, 1996). Fresh vials of frozen THP-1 cells were initiated for growth every 8 weeks. To differentiate THP-1 cells into a macrophage phenotype, a final concentration of 25 ng/ml LPS and 10 ng/ml IFN-γ were added to the cells (Humphrey and Dubyak, 1996) either for 3 h for IL-1β release assays or overnight (16 h) for pore formation studies.

**Ca2+ Influx Assay.** Agonist-induced changes in intracellular Ca2+ concentrations were assessed in all of the cell lines using the Ca2+ chelating dye, Fluo-4 in conjunction with a fluorometric imaging plate reader (Molecular Devices, Sunnyvale, CA) as previously described (Bianchi et al., 1999) with minor modifications. The cells were plated out the day before the experiment onto Poly-L-lysine-coated 96-well plates (Becton-Dickinson, Bedford, MA; Sigma-Aldrich). Cell concentration was 5 × 105 cells per plate. Fluo-4 was dissolved in anhydrous dimethyl sulfoxide to a final concentration of 5 μM in DPBS. The dye was loaded onto the adherent cells, and the plates were centrifuged for 5 min at 1000 rpm. Cells were loaded for at least 1 h but not more than 3 h and kept in the dark at room temperature. After loading, the unincorporated Fluo-4 was removed by washing with DPBS using a SkanWasher 400 (Molecular Devices). All compound solutions were prepared in DPBS. After the agonist addition, changes in intracellular Ca2+ concentrations were recorded on a second time scale for 3 min. Ligands were tested at 11 half-log concentrations from 10-10 to 10-4 M. Independent measurements of a positive control (100%) were performed on each plate to normalize values from plate to plate. Agonist concentrations corresponded to their respective EC50 values for each receptor to enable comparison of antagonist potencies across multiple P2 receptor subtypes (Jarvis et al., 2002). Since P2X2/3 receptors have an identical pharmacological profile as homomeric P2X7 receptors and do not rapidly desensitize (Jarvis et al., 2004), the heteromeric receptor was used for selectivity versus P2X7 receptors. Structural analogs of A-740003, as well as other structurally diverse P2X7 antagonists (Nelson et al., 2006), did not activate homomeric P2X7 receptors at concentrations up to 10 μM (D. Donnelly-Roberts, unpublished data).

For measurement of antagonist activity, ligands were added to the cell plate, and fluorescence data were collected for 3 min before the addition of the agonist. Fluorescence data were collected for another 2 min after the agonist addition. Concentration-response data were analyzed using Prism (GraphPad Software, Inc., San Diego, CA); the pEC50 or pIC50 values were derived from a single curve fit to the mean data (n = 4–6) in duplicate.

**Yo-Pro Uptake Assay.** Agonist-induced pore formation was assessed using uptake of Yo-Pro dye (mol. wt. = 629; Invitrogen) in the recombinant rat, human P2X7-1321N1 cell lines, or differentiated THP-1 cells as previously described (Donnelly-Roberts et al., 2004). Yo-Pro iodide dye (1 mM in 100% dimethyl sulfoxide) was diluted to a final concentration of 2 μM in PBS (without Mg2+ or Ca2+) and then placed on the cells immediately prior to agonist addition. Cells were plated the day before at a density of 1 × 104 cells/plate onto poly-L-lysine-coated black-walled 96-well plates to reduce light scat-
For differentiated THP-1 (25 ng/ml LPS and 10 ng/ml IFN-γ overnight), the cells were plated onto poly-L-lysine-coated black-walled 96-well plates to 1 to 2 × 10^6 cells/ml/well. Following the addition of various concentrations of agonists, the Yo-Pro dye uptake was observed in the fluorometric imaging plate reader by capturing the intensity of fluorescence by the CCD camera every 15 s for the first 10 min of agonist exposure followed by every 20 s for an additional 50 min. For antagonist activity measurements, the percent maximal IL-1β-induced release was normalized to that induced by an EC_{50} values for BzATP activation and plotted against the antagonist compound concentration to calculate IC_{50} values and account for plate-to-plate variability. Concentration-response data were analyzed using GraphPad Prism; the pEC50 or pIC50 values were derived from a single curve fit to the mean data of n = 6 in duplicates.

**IL-1β Release in THP-1 Cells.** In the presence of the differentiation media, THP-1 cells were incubated with inhibitors for 30 min at 37°C. Supernatants were collected after a 5-min centrifugation media, THP-1 cells were incubated with inhibitors for 30 min followed by a challenge with 1 mM BzATP for an additional 10 min of agonist exposure followed by every 20 s for an additional 50 min. For antagonist activity measurements, the percent maximal BzATP response, was subtracted from the maximal BzATP-induced release.

**In Vivo Pain Models and Pharmacokinetic Profile**

**Subjects.** Male Sprague-Dawley rats (Charles River, Wilmington, MA) weighing 200 to 300 g were used in all experiments. Animals were group-housed in Association for Assessment and Accreditation of Laboratory Animal Care-approved facilities at Abbott Laboratories (Abbott Park, IL) in a temperature-regulated environment with lights on between 7:00 AM and 8:00 PM. Food and water were available ad libitum except during testing. All animal handling and experimental protocols were approved by an institutional animal care and use committee. All experiments were performed during the light cycle.

**Acute Thermal Nociception.** The response to acute thermal stimulation was determined using a commercially available paw thermal stimulator (UARDG; University of California, San Diego, CA). Rats were placed individually in Plexiglas cubicles mounted on a glass surface maintained at 30°C and allowed a 30-min habituation period. A thermal stimulus, in the form of radiant heat emitted from a focused projection bulb, was then applied to the plantar surface of each hind paw. In each test session, each rat was tested in three sequential trials at approximately 5-min intervals. Paw-withdrawal latencies (PWLs) were calculated as the median of the two shortest latencies.

**Carrageenan and Complete Freund’s Adjuvant-Induced Thermal Hyperalgesia and Edema.** Unilateral inflammation was induced by injecting 100 μl of a 1% solution of λ-carrageenan or 150 μl of a 50% solution of complete Freund’s adjuvant (CFA) (Sigma-Aldrich) in physiological saline into the plantar surface of the right hind paw of the rat. The hyperalgesia to thermal stimulation was determined 2 or 48 h following carrageenan or CFA injection, respectively, using the same apparatus as described above for the noxious acute thermal assay. In addition, in both models, the volume of paw edema was measured using water displacement with a plethysmometer (Buxco, Sharon, CT) by submerging the hind paw up to the ankle hairline (approximately 1.5 cm). The volume of water displacement was measured by a transducer and recorded by a computer.

**Spinal Nerve (L5/L6) Ligation Model of Neuropathic Pain.** As previously described in detail by Kim and Chung (1992), a 1.5-cm incision was made dorsal to the lumbosacral plexus in anesthetized rats. The paraspinal muscles (left side) were separated from the spinous processes, the L5 and L6 spinal nerves were isolated, and tightly ligated with 3-0 silk threads. Following hemostasis, the wound was sutured and coated with antibiotic ointment. The rats were allowed to recover and then placed in a cage with soft bedding for 14 days before behavioral testing for mechanical alldynia. A-740003 was injected i.p. 30 min before testing for mechanical alldynia.

**Mechanical (tactile) alldynia was measured using calibrated von Frey filaments (Stoelting, Wood Dale, IL).** Briefly, rats were placed into individual Plexiglas containers and allowed to acclimate for 15 to 20 min before testing. Paw withdrawal threshold (PWT_{vonfrey}) was determined by increasing and decreasing stimulus intensity and estimated using a Dixon nonparametric test. Only rats with threshold scores ≥ 4.5 g were considered alldynic and utilized in compound testing experiments.

**Sciatic Nerve Ligation Model of Neuropathic Pain.** As previously described in detail by Bennett and Xie (1988), in anesthetized rats, a 1.5-cm incision was made 0.5 cm below the pelvis, and the biceps femoris and the gluteus superficialis (right side) were separated. The sciatic nerve was exposed, isolated, and four loose ligatures (5-0 chronic catgut) with 1-mm spacing were placed around it. The rats were allowed to recover and then placed in a cage with soft bedding for 14 days before behavioral testing for mechanical alldynia as described above. A-740003 was injected i.p. 30 min before testing for mechanical alldynia.

**Chemotherapy-Induced Neuropathic Pain.** As previously described in detail by Lynch et al. (2005), chemotherapy-induced neuropathic pain was induced by a continuous i.v. infusion of vincristine. In anesthetized rats, the right external jugular vein was catheterized (PE60 tubing) with a vincristine filled osmotic pump (0.5 μl/h, 14 days; Alzet model 2002; Durect Corporation, Cupertino, CA) that had been primed overnight to deliver 30 μg/kg/day vincristine sulfate (Sigma-Aldrich). The rats were allowed to recover and then placed in a cage with soft bedding for 14 days before behavioral testing for mechanical alldynia as described above. A-740003 was injected i.p. 30 min before testing for mechanical alldynia. In this model, because of the bilateral alldynia, maximal possible effects were set at 15 g.

**Rotorod Performance.** Rotorod performance was measured using an accelerating Rotorod apparatus (Omnitech Electronics, Inc., Columbus, OH). Rats were allowed a 30-min acclimation period in the testing room and then placed on a 9-cm-diameter rod that increased in speed from 0 to 20 rpm over a 60-s period. The time required for the rat to fall from the rod was recorded, with a maximal score of 60 s. Each rat was given three training sessions. Rotorod performance (latencies to fall from the Rotorod) was determined 30 min following i.p. A-740003 injection.

**Compounds.** A-740003 (Fig. 1) was synthesized at Abbott Laboratories. A-740003 was dissolved in 30% N-methylpyrrolidinone, 30% polyethylene glycol, and 40% hydroxypropyl-β-cyclodextrin for i.p. administration 30 min before testing and in 100% polyethylene glycol for p.o. administration 60 min before testing in a 2-ml/kg dosing volume. As part of an ongoing drug discovery program, doses of A-740003 ranged from 10 to 300 μmol/kg for relative comparisons of compound potency. Doses of A-740003 used in the in vivo studies are expressed as milligrams per kilogram in this report for clarity.

**Statistics.** Analysis of the in vivo data were carried out using analysis of variance. Where appropriate, Fisher’s protected least significant difference was used for post hoc analysis. The level of significance was set at p < 0.05. ED_{50} values were estimated using least squares linear regression. Data are presented as mean ± S.E.M.
Results

A-740003 Potently Blocks P2X7 Receptors. A-740003 potently blocked BzATP-evoked changes in intracellular calcium concentrations in 1321N1 cells stably expressing rat (IC$_{50}$ = 18 nM) or human (IC$_{50}$ = 40 nM) P2X7 receptors (Fig. 2). For comparison, PPADS, Brilliant Blue G (rat selective P2X7 antagonist), and KN-62 (a human selective P2X7 antagonist; Anderson and Nedergaard, 2006) were significantly less potent in blocking BzATP-evoked changes in intracellular calcium concentrations (Fig. 2; Table 1). A-740003 also blocked BzATP-mediated pore formation as measured by the intracellular uptake of Yo-Pro (Fig. 3) at both the rat (IC$_{50}$ = 138 nM) and human (IC$_{50}$ = 93 nM) P2X7 receptors. A-740003 was significantly more potent in blocking P2X7-mediated pore formation as compared with PPADS, KN-62, or Brilliant Blue G (Table 1; Fig. 3).

A-740003 Blocks P2X7 Receptors on Differentiated Human THP-1 Cells. Human THP-1 cells differentiated with LPS and IFN-γ into a macrophage phenotype express P2X7 receptors and receptor activation lead to IL-1β release as well as Yo-Pro uptake (Humphrey and Dubyak, 1996; Donnelly-Roberts et al., 2004). A-740003 potently blocked Yo-Pro uptake in differentiated human THP-1 cells (IC$_{50}$ of 92 nM) (Fig. 4). A-740003 also blocked IL-1β release in the THP-1 cells (IC$_{50}$ of 156 nM) (Fig. 4).

A-740003 Is a Highly Selective P2X7 Antagonist. As shown in Fig. 5, A-740003, at concentrations up to 100 μM, did not significantly reduce agonist-evoked changes in intracellular calcium concentrations mediated by a variety of other P2X and P2Y receptors. We have demonstrated previously that there is a high positive correlation ($r$ = 0.98) between ligand binding at heteromeric P2X2/3 receptors and homomeric P2X3 receptors (Jarvis et al., 2004). Consequently, A-740003 was not tested at homomeric P2X3 receptors since it was inactive at P2X2/3 receptors. Other analogs of A-740003, as well as other structurally novel P2X7 receptor antagonists (Nelson et al., 2006), did not significantly interact at homomeric P2X7 receptors at concentrations up to 10 μM (D. Donnelly-Roberts, unpublished data). A-740003 was also evaluated for its ability to interact with a large array of

![Fig. 2](image-url) A-740003 potently blocks rat and human P2X7 receptors. A-740003 blocked BzATP-mediated changes in intracellular calcium concentrations at rat (10 μM BzATP) (A) and human (5 μM BzATP) (B) P2X7 receptors. PPADS and BBG were significantly less potent, and KN-62 showed no activity (Table 1) to block rat P2X7 receptors. Similarly, PPADS and BBG were significantly less potent, and KN-62 showed no activity (Table 1) to block human P2X7 receptors. Data are represented in percentage of maximal response (control) minus basal fluorescence. See Table 1 for a summary of the pIC$_{50}$ values.

![Fig. 3](image-url) A-740003 potently blocks pore formation mediated by rat and human P2X7 receptors. A-740003 blocked BzATP-mediated Yo-Pro uptake at rat (7.5 μM BzATP) (A) and human (2.5 μM BzATP) (B) P2X7 receptors. PPADS and BBG were significantly less potent, and KN-62 showed negligible activity (Table 1) to block rat P2X7 receptors. Similarly, KN-62 was slightly less potent versus PPADS and BBG, which had weaker activity (Table 1) to block human P2X7 receptors. Data are represented in percentage of maximal response (control) minus basal fluorescence. See Table 1 for a summary of the pIC$_{50}$ values.

**TABLE 1**

<table>
<thead>
<tr>
<th>Antagonist</th>
<th>Rat P2X7</th>
<th>Human P2X7</th>
</tr>
</thead>
<tbody>
<tr>
<td>A-740003</td>
<td>7.75 ± 0.03</td>
<td>7.36 ± 0.04</td>
</tr>
<tr>
<td>PPADS</td>
<td>5.10 ± 0.02</td>
<td>5.45 ± 0.03</td>
</tr>
<tr>
<td>KN-62</td>
<td>&lt;4</td>
<td>4.88 ± 0.18</td>
</tr>
<tr>
<td>BBG</td>
<td>5.08 ± 0.07</td>
<td>&lt;4</td>
</tr>
<tr>
<td>Yo-Pro Activity (pIC$_{50}$ ± S.E.M.)</td>
<td>7.00 ± 0.02</td>
<td>7.03 ± 0.04</td>
</tr>
<tr>
<td>A-740003</td>
<td></td>
<td></td>
</tr>
<tr>
<td>PPADS</td>
<td>5.92 ± 0.09</td>
<td>5.88 ± 0.03</td>
</tr>
<tr>
<td>KN-62</td>
<td>&lt;4</td>
<td>6.7 ± 0.02</td>
</tr>
<tr>
<td>BBG</td>
<td>6.22 ± 0.03</td>
<td>5.7 ± 0.09</td>
</tr>
</tbody>
</table>
The maximal plasma concentration (fluorescence, represented as percentage of maximal response (control) minus basal
used to activate P2X1, P2X4, and P2Y1 receptors, and ATP (10 mM) was
used for P2X2a and P2X2a/3 receptors. UTP (1 mM) was used to activate
P2Y2 receptors. BzATP (5 mM) was used to activate the BzATP binding site. Analysis of these data generated a pA2 value of 7.30 and a slope factor of 0.98

The acute anti-inflammatory effects of A-740003 were assessed in the carrageenan-induced paw edema model. A-740003 reduced carrageenan-induced thermal stimulation with von Frey monofilaments 2 weeks following surgery (PWTvonfrey control, 13.2 ± 0.4 g; PWTvonfrey injured, 2.9 ± 0.1 g; p < 0.01), demonstrating the development of mechanical allodynia. A-740003 attenuated CCI-induced mechanical allodynia in a dose-related manner with an ED50 of 114 mg/kg i.p. and efficacy of 54.7 ± 14.3% at the highest dose tested (Fig. 9A; Table 3).

Chemotherapy-Induced Mechanical Allodynia. Continuous vincristine infusion produced a decrease in PWTvonfrey to mechanical stimulation with von Frey monofilaments 2 weeks following the beginning of the infusion (PWTvonfrey control, 15.0 ± 0.0 g; PWTvonfrey injured, 3.7 ± 0.1 g; p < 0.01), demonstrating the development of mechanical allodynia. A-740003 attenuated vincristine-induced mechanical allodynia in a dose-related manner with 50.8 ± 6.4% effect at the highest dose tested (Fig. 9B; Table 3).

Carrageenan-Induced Acute Inflammatory Thermal Hyperalgesia and Edema. Carrageenan injection into the hind paw induced a significant decrease in PWL to thermal stimulation (PWL control, 9.9 ± 0.2 s versus PWL inflamed, 2.3 ± 0.1 s; p < 0.01), demonstrating the development of thermal hyperalgesia 2 h following carrageenan injection (Fig. 10A). A-740003 reduced carrageenan-induced thermal hyperalgesia in a dose-related manner (ED50 of 54 mg/kg i.p.), with 75 ± 5% antinociceptive effect at the highest dose tested (Fig. 8A). Under the same conditions, A-740003 had no effect on PWL of the contralateral noninflamed paw, indicative of a specific antihyperalgesic effect in this model.

The acute anti-inflammatory effects of A-740003 were assessed in the carrageenan-induced paw edema model. A-740003 (14–142 mg/kg i.p.) attenuated carrageenan-induced paw edema [F(3,47) = 4.75, p < 0.01]. At 142 mg/kg i.p., A-740003 produced a 36 ± 7% decrease in paw volume (p < 0.01, Table 3).

CFA-Induced Chronic Inflammatory Thermal Hyperalgesia and Edema. CFA injection into the hind paw
induced a significant decrease in PWL to thermal stimulation 48 h following CFA injection (PWL control, 10.2 ± 0.3 s versus PWL inflamed, 5.6 ± 0.1 s, p < 0.01), demonstrating the development of thermal hyperalgesia (Fig. 10B). A-740003 attenuated CFA-induced thermal hyperalgesia in a dose-related manner (ED50 of 38 mg/kg i.p.) with 58.7 ± 16.5% effect at the highest dose tested (Fig. 10B). Under the same conditions, A-74003 had no effect on PWL of the contralateral noninflamed paw, indicative of a specific antihyperalgesic effect in this model.

The acute anti-inflammatory effects of A-74003 were also assessed in the CFA-induced paw edema model. A-740003 (14–142 mg/kg i.p.) did not significantly decrease CFA-induced paw edema [F(3, 35) = 2.28, p > 0.05; Table 3].

**Acute Thermal Pain.** A-74003 did not produce significant antinociception on acute thermal pain in naive rats. A-74003 (142 mg/kg i.p.)-treated animals had a PWL to acute thermal stimulation of 10.9 ± 0.1 compared with 10.9 ± 0.5 s in vehicle-treated animals (p > 0.05; Table 3).

**Effects on Motor Activity and General CNS Function.** A-74003 had no significant effect on motor coordination at doses up to 142 mg/kg i.p. [F(3, 31) = 0.35, p > 0.05],
as measured by the ability of rats to run on an accelerating rotating rod (latency to fall for the control group, 58.1 ± 1.0 s; for the highest dose group, 56.3 ± 3.1 s). Rats were fully awake, responsive to stimuli, and retained the righting reflex, consistent with their ability to perform the Rotorod test at all of the doses tested.

Discussion

The present data demonstrate that A-740003 is a potent antagonist of rat and human P2X7 receptors. A-740003 blocked P2X7 receptor-mediated changes in intracellular calcium concentrations in a competitive fashion and was highly selective as compared with its activity at other P2 receptors as well as other cell surface receptors and ion channels. A-740003 also potently blocked other consequences of P2X7 receptor activation including BzATP-evoked IL-1β release in differentiated human THP-1 cells and prolonged agonist-stimulated pore formation.

The discovery of A-740003 represents a significant advance in P2X receptor pharmacology since this compound is both more potent and more selective than other previously described P2X7 receptor antagonists (Romagnoli et al., 2004). A-740003 also shows much less species differences in its ability to block both rat and human P2X7 receptors as compared with the rat selective antagonist BBG and the human selective antagonist KN-62 (Anderson and Nedergaard, 2006). It is interesting to note that both of the latter antagonists are more than 10-fold more potent to block P2X7 receptor-mediated pore formation (Yo-Pro uptake) as compared with their ability to block receptor-mediated calcium influx. The differential potency of these antagonists to block these P2X7 receptor-mediated effects has been attributed to their relatively slow association kinetics in the two different assays (Namovic et al., 2005). In contrast, A-740003 showed equivalent potency to block both human P2X7 receptor-mediated pore formation (Yo-Pro uptake) as compared with their ability to block receptor-mediated calcium influx.

The activation of P2X7 receptors results in the rapid maturation and extracellular release of the proinflammatory cytokine, IL-1β (Perreuxs and Gabel, 1994; Solle et al., 2001; Ferrari et al., 2006). Other biochemical consequences of increased IL-1β concentrations include induction of nitric oxide synthase as well as increased production of cyclooxygenase-2 and tumor necrosis factor (TNF)-α (Woolf et al., 1997; Samad et al., 2001; Parvathenani et al., 2003). All of these proinflammatory events can also contribute to enhanced pain sen-
P2X7 knockout mice show reduced pain sensitivity following et al., 2002). A more recent study has demonstrated that experimental passive collagen-induced arthritis model (Labasi et al., 2002) mice show significantly diminished inflammation in an experimental passive collagen-induced arthritis model (Labasi et al., 2002). A recent study has demonstrated that P2X7 knockout mice show reduced pain sensitivity following both complete Freund’s adjuvant-induced inflammation and partial injury of the sciatic nerve (Chessell et al., 2005). Similar to the nociceptive phenotype of mice lacking P2X7 receptors (Chessell et al., 2005) or lacking both isoforms of IL-1 (Honore et al., 2006), systemic administration of A-740003 produced dose-dependent antinociceptive effects in models of neuropathic pain and inflammatory pain. A-740003 was particularly potent at reducing mechanical allodynia observed 2 weeks following L5/L6 nerve ligation. The antinociceptive activity of A-740003 in this model of neuropathic pain is consistent with recent reports of the analgesic efficacy of two other structurally distinct P2X7 antagonists to neuropathic pain. The antinociceptive activity of A-740003 in this model of neuropathic pain is consistent with recent reports of the analgesic efficacy of two other structurally distinct P2X7 antagonists to neuropathic pain. The exact reasons for the differential potency of A-740003 to reduce tactile allodynia in these neuropathic pain models are unknown. However, these data may illustrate the preferential role of P2X7 receptor modulation of IL-1β in reducing nociception in the SNL model as compared with the robust analgesic efficacy of a P2X7 antagonist to reduce neuropathic allodynia in the chronic constriction injury of the sciatic nerve model (Jarvis et al., 2002).

A-740003 also significantly reduced thermal hyperalgesia in two models of inflammatory pain: intraplantar administration of complete Freund’s adjuvant or carrageenan. The robust antinociceptive effects of A-740003 in these inflammatory pain models does not appear to be secondary to an anti-inflammatory effect since A-740003 was more efficacious at reducing nociception compared with paw edema in both of these models. It should be noted, however, that the anti-inflammatory activity of P2X7 antagonists may be more pronounced in arthritis models as compared with acute (carrageenan) and subacute (CFA) inflammatory models where the contribution of IL-1β to ongoing inflammatory processes may be more prominent in chronic arthritis (Labasi et al., 2002).

Collectively, the present data and the phenotype of the P2X7 knockout mice indicate a specific role for P2X7 receptor activation in chronic pain states. It should be noted that Dell’Antonio et al. (2002a,b) have reported that local administration of oxidized-ATP, a weak and irreversible antagonist of P2X7 receptors (Suprenant et al., 1996), reduced inflammation-induced mechanical hyperalgesia in rats. Although these investigators attributed these results to blockade of P2X7 receptors, it must be appreciated that oxidized-ATP has weak affinity for P2X7 receptors and also has many other pharmacological actions including blockade of P2X1 and P2X2 receptors (Evans et al., 1995) and inhibition of nuclear factor-κB and cytokine release (Murgia et al., 1993; Beigi et al., 1997; Burnstock, 2006). The physiological significance of P2X7 receptor modulation of IL-1β release is evidenced by the reduced inflammatory phenotype of P2X7 knockout mice (Solle et al., 2001; Labasi et al., 2002). ATP stimulation of macrophages from P2X7 knockout mice does not result in the release of mature IL-1β, and P2X7 knockout mice show significantly diminished inflammation in an experimental passive collagen-induced arthritis model (Labasi et al., 2002). A more recent study has demonstrated that P2X7 knockout mice show reduced pain sensitivity following both complete Freund’s adjuvant-induced inflammation and partial injury of the sciatic nerve (Chessell et al., 2005). Table 3 shows the analgesic profile of A-740003 (i.p.) in different pain models. The ED50 values were calculated using the method of least squares in a four-parameter logistic model.

Table 3: Analgesic profile of A-740003 (i.p.)

<table>
<thead>
<tr>
<th>Pain Model</th>
<th>ED50 (mg/kg i.p.)</th>
<th>% Effect at 142 mg/kg i.p.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acute nociception</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rat acute thermal</td>
<td>&gt;142</td>
<td>0 ± 1</td>
</tr>
<tr>
<td>Inflammatory pain</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Carrageenan Acute thermal</td>
<td>54</td>
<td>75 ± 5*</td>
</tr>
<tr>
<td>hyperalgesia</td>
<td>&gt;142</td>
<td>36 ± 7*</td>
</tr>
<tr>
<td>Edema (Complete Freund’s</td>
<td>38</td>
<td>59 ± 16*</td>
</tr>
<tr>
<td>adjuvant)</td>
<td>&gt;142</td>
<td>0 ± 5</td>
</tr>
<tr>
<td>Chronic thermal</td>
<td></td>
<td></td>
</tr>
<tr>
<td>hyperalgesia</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Edema</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Neuropathic pain</td>
<td></td>
<td></td>
</tr>
<tr>
<td>L5/L6 nerve ligation</td>
<td>19</td>
<td>65 ± 11* (47mg/kg)</td>
</tr>
<tr>
<td>Chronic constriction injury</td>
<td>114</td>
<td>55 ± 14*</td>
</tr>
<tr>
<td>Chemotherapy</td>
<td>142</td>
<td>51 ± 6*</td>
</tr>
</tbody>
</table>

* Significantly different (p < 0.05) from vehicle-treated animals (n = 6–12 per group).
findings are further supported by the demonstration that exogenously applied IL-1 produces hyperalgesia when applied peripherally (Ferreira et al., 1988). In addition to the analgesic effects of the IL-1ra in experimental pain models, several genetic approaches have been used to further investigate the pronociceptive actions of IL-1 in mice. These include targeted gene disruption of the IL-1 type I receptor or the IL-1acp, as well as transgenic overexpression of the IL-1ra (Wolf et al., 2004) or IL-1aβ double knockout (Honore et al., 2006). All of these approaches have produced mice that show reduced nociceptive responses relative to wild-type animals. Collectively, these data clearly indicate that P2X7 receptor modulation of IL-1 contributes to nociceptive sensitivity in chronic pain states.

The present data further illustrate the multitude of mechanisms by which ATP can alter nociceptive sensitivity following tissue injury (Burnstock and Wood, 1996). Evidence from a variety of experimental strategies including genetic disruption and the development of selective antagonists has indicated that activation of several P2X receptors, including P2X3, P2X2/3, P2X4 and P2X7, and P2Y (e.g., P2Y2) receptors, can modulate pain. For example, A-317491, a selective P2X3 antagonist (Jarvis et al., 2002), effectively blocks both CFA-induced inflammatory thermal hyperalgesia as well as mechanical and thermal hyperalgesia resulting from chronic constriction of the sciatic nerve. Intrathecally delivered antisense oligonucleotides specifically targeting P2X7 receptors have been shown to decrease tactile allodynia following nerve injury (Tsuda et al., 2003). In addition, activation of P2Y2 receptors leads to sensitization of TRPV1 receptors (Tominaga et al., 2001; Lakshmi and Joshi, 2005). Thus, ATP acting at multiple purinergic receptors either directly on neurons (e.g., P2X3, P2X2/3, and P2Y receptors) or through indirect neural-glial cell interactions (P2X4 and P2X7) can alter nociceptive sensitivity (Tsuda et al., 2003; Zhang et al., 2005).

Taken together, the present data demonstrate that the acute in vivo blockade of P2X7 receptors significantly reduces nociception in animal models of persistent neuropathic and inflammatory pain. Although there is growing appreciation for the role of P2X7 modulation of proinflammatory IL-1 processing (Ferrari et al., 2006), the analgesic activity of A-740003 and other recently described selective P2X7 receptors (Nelson et al., 2006) suggests a specific role for P2X7 in neural-glial cells interactions associated with ongoing pain (Zhang et al., 2005).

**References**


