Placental Handling of Fatty Acid Ethyl Esters: Perfusion and Subcellular Studies

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ABSTRACT
The measurement of fatty acid ethyl esters (FAEE) in neonatal meconium is a novel test to confirm prenatal ethanol exposure. The origin of FAEE in the maternal-placental-fetal unit is not known. The objectives of this study were to investigate whether FAEE are transferred and metabolized by the human placenta. Isolated placental cotyledons were perfused with a mixture of four FAEE (palmitic, stearic, oleic, and linoleic acid ethyl esters) commonly detected in the meconium of neonates exposed to ethanol in utero, and the transfer of FAEE to the fetal unit was investigated in the absence and presence of albumin. The metabolic degradation of FAEE by human placental microsomes was subsequently determined. FAEE disappeared from the maternal circulation but remained undetectable in the fetal unit following perfusions. The addition of albumin had no effect on FAEE transfer. The unrecoverable fraction of individual FAEE in the perfusion system accounted for >50% of the initial amount used, suggesting significant metabolic degradation. Subcellular studies documented the enzymatic degradation of FAEE by placental microsomes (mean $K_m$, 35–95 $\mu$M; $V_{max}$, 0.6–1.8 nmol/min/mg for individual FAEE). FAEE at levels found in alcoholics that are originated from the mother are not transferred to the fetus because they are taken up and degraded extensively by the human placenta. Hence, FAEE detected in maternal matrices are likely produced by the fetus from ethanol that has been transferred to and metabolized by the fetus, rendering FAEE a powerful direct biomarker reflective of true fetal exposure to ethanol in utero.

Consumption of alcohol during pregnancy is the cause of Fetal Alcohol Spectrum Disorder (FASD), which affects approximately 1% of all live births (Sampson et al., 1997). According to the current diagnostic guidelines (U.S. Institute of Medicine, 1996), except when the pathognomonic craniofacial dysmorphology is present, FASD can only be diagnosed with confirmed maternal drinking history, which is often difficult to obtain. As a result, large numbers of affected children cannot be properly diagnosed and receive appropriate interventions in early childhood, although studies have shown that early intervention leads to better prognosis of FASD (Streissguth et al., 1996).

Fatty acid ethyl esters (FAEE) are nonoxidative metabolites of ethanol. The production of FAEE from ethanol and its substrates, fatty acyl CoA or free fatty acids, is facilitated by cytosolic and microsomal fatty acid ethyl ester syntheses (Best and Laposata, 2003). FAEE have been proposed as biological markers of acute and chronic alcohol consumption (Laposata, 1999) because they have been reported to accumulate at high levels in the blood of adult drinkers (Doyle et al., 1996) and in critical organs most commonly affected by chronic alcoholism (Laposata and Lange, 1986). Recently, significantly elevated levels of FAEE have been documented in the meconium of neonates of mothers who had used alcohol heavily in pregnancy (Bearer et al., 1999, 2003; Klein et al., 1999; Moore and Lewis, 2001; Chan et al., 2003; Moore et al., 2003). Hence, during the last 5 years, the measurement of FAEE in meconium has become a promising objective and noninvasive method to confirm prenatal exposure to ethanol.

Presently, the source of FAEE found in meconium is unknown. FAEE can be produced in the mother and cross the placenta to the fetus or be produced by the placenta itself. Alternatively, if meconium FAEE are produced in the fetus only, then they would be a biomarker of the amount of ethanol that has circulated in the fetus and has been metabolized by the unborn baby. In such case, meconium FAEE would be superior to other measures such as maternal self-reporting and maternal biochemical markers.

The objectives of the present study were to elucidate whether maternal FAEE cross the human placenta into the
fetus and whether the placenta is capable of metabolizing FAEE.

Materials and Methods

Reagents. All standards and solvents were obtained from Sigma-Aldrich (St. Louis, MO). Acetone and n-hexane were high-performance liquid chromatography grade or better. Bovine albumin (BSA) was essentially fatty acid free (0.005%). FAEE, including palmitic (E16:0), heptadecanoic (E17:0), stearic (E18:0), oleic (E18:1), and linoleic (E18:2) acid ethyl esters, were diluted in methanol and stored at −20°C. A capillary gas chromatography column (ZB-WAX) was obtained from Phenomenex (Torrance, CA). Enzyme-linked immunosorbent assay kits for the analysis of human chorionic gonadotrophin (hCG) were obtained from Alpha Diagnostic International (San Antonio, Texas).

In Vitro Perfusion of Isolated Placental Cotyledons. Placentas (n = 10) were obtained from caesarean sections or vaginal deliveries of uncomplicated, term pregnancies and transported to the laboratory on ice in heparinized phosphate-buffered saline. Perfusion of a peripheral placental lobule was established within 30 min of delivery following a method previously described by our laboratory (Derewnlany et al., 1991). In brief, a chorionic artery and vein supplying a peripheral placental lobule were selected and cannulated. The fetal circulation was perfused at a rate of approximately 3 ml/min and recirculated in a 150-ml reservoir while the maternal circuit was perfused at a rate of approximately 15 ml/min and recirculated in a 250-ml reservoir.

The perfusion medium was maintained at 37°C and consisted of antipyrine (maternal, 1 mM), culture medium M199 (10.9 g/l), dextran (maternal, 7.5 g/l; fetal, 30.0 g/l), glucose (maternal, 2.77 mM), heparin (2000 units/l), kanamycin (100 mg/l), and sodium bicarbonate (maternal, 30 mM; fetal, 25 mM). Sodium bicarbonate was added to maintain the pH of the perfusates within physiologic range (maternal, pH 7.4; fetal, pH 7.35). The maternal circuit was equilibrated with 95% O2 and 5% CO2 whereas the fetal circuit was equilibrated with 95% N2 and 5% CO2.

Perfusion Studies without Albumin (n = 4). In four of the placentae, after an initial 10-min “washout” period, the preexperimental control period proceeded. Both circuits were closed and recirculated with fresh perfusate containing no albumin for 30 min, after which the circuits were switched to albumin-containing perfusate (0.5%, w/v) for another 30 min. At the end of the 1-h control, the fetal reservoir was replaced with 150 ml of fresh albumin-containing perfusate, and the maternal reservoir was replaced with 250 ml of fresh perfusate containing albumin (0.5%, w/v) and the four FAEE at equimolar concentrations [2 μM (n = 3) and 20 μM (n = 3)]. Samples from the maternal and fetal reservoirs were collected every 10 min during the first 0.5 h and then every 30 min until 180 min for the analysis of FAEE. At this stage, the experimental period was shortened from 4 to 3 h based on the data obtained from the studies conducted in the absence of albumin, showing that changes in FAEE disposition plateau after 2 h. At the end of the experiment, both circuits were switched to fresh albumin-containing perfusate for a 1-h postexperimental control. FAEE were measured from the perfused tissue for the determination of the fraction bound to the placenta as described previously.

Sample Analysis. Arterial and venous samples were taken from the maternal and fetal circuits throughout the control and experimental periods for pH and pO2 measurements. The physical integrity of the placental preparation was assessed by frequent monitoring of the fetal perfusate pressure (expressed in mm Hg) and excessive volume loss (reject if >3 ml/h). Samples were taken from both reservoirs (every 15 min during control and every 30 min during perfusion) for the analysis of oxygen and glucose consumption and the production of lactate and hCG. Tissue hCG content was measured in both fresh and perfused tissue.

The perfusate concentrations of pH and pO2 were measured using a blood gas analyzer (ABL30; Radiometer, Copenhagen, Denmark). The concentrations of glucose and lactate in perfusate samples were measured as described previously (Derewnlany et al., 1991). For the analysis of hCG, samples of fresh and perfused placental tissue were homogenized with 5 ml of ice-cold phosphate-buffered saline (0.1 M, pH 7.4) per gram of tissue using a Polytron (Brinkmann Instruments, Westbury, NY). The homogenate was centrifuged at 8000 rpm at 4°C for 15 min, and the supernatant was aliquoted and stored at −20°C until analysis. The perfusate and tissue concentrations of hCG were measured by enzyme-linked immunosorbent assay (Alpha Diagnostic International).

FAEE were extracted from perfusate samples, fresh and perfused placental tissue with 5 ml of hexane:acetone (5:2, v/v) and analyzed by gas chromatography with flame ionization detection using a previously established method (Chan et al., 2003). The limit of detection was 0.5 μmol. Tissue samples were homogenized with 5 ml of ice-cold phosphate-buffered saline (0.1 M, pH 7.4) per gram of tissue using a Polytron (Brinkmann Instruments), aliquoted, and stored at −20°C.
until analysis. The recovery of FAEE from placental perfusate and tissue were determined by extrapolation from standard curves prepared by spiking known concentrations of FAEE in the respective blank matrices (i.e., blank perfusate and tissues) to account for matrix effects.

**Degradation of FAEE by Placental Microsomes**

**Isolation of Placental Cytosol and Microsomes.** Subcellular fractions of the fresh human placenta \((n = 4)\) were isolated using standard differential centrifugation techniques. Pieces of villous tissue (about 70 g) were dissected and rinsed thoroughly in ice-cold saline to remove excess blood and then blotted dry on gauze. The tissues were then homogenized in a blender (Waring Products, Inc., New Hartford, CT) with 5 ml of ice-cold HEGM-D buffer (25 mM NaHepes, 1 mM Na,EDTA, 10% glycerol, 20 mM Na,Monolylde, 2H2O, and 1 mM dithiothreitol) per gram of tissue. The homogenate was aliquoted and centrifuged at 8000 rpm at 4°C for 15 min in a Sigma SK-30 Centrifuge (Sigma Chemie, Deisenhofen, Germany). The resulting pellet was discarded, and the supernatant was centrifuged at 15,000 rpm at 4°C for 15 min. The supernatant from this second spin was pooled and centrifuged at 35,000 rpm at 4°C for 1 h in a Beckman Optima LE-80K Ultracentrifuge (Beckman Coulter, Fullerton, CA). The supernatant (cytosol) was aliquoted into cryovials, frozen in liquid nitrogen, and stored at −80°C until analysis. The pellet (microsomes) was resuspended in HEGM-D buffer using a Teflon pestle homogenizer (Canadian Laboratory Supplies Ltd., Mississauga, Ontario, Canada) and centrifuged at 35,000 rpm at 4°C for 1 h. The final supernatant was discarded, and the microsomal pellet was resuspended with a Teflon pestle homogenizer in HEPES-sucrose buffer (5 mM HEPES and 0.25 M sucrose, pH 7.4), aliquoted into cryovials, frozen in liquid nitrogen, and stored at −80°C until analysis. The amount of proteins in the subcellular fractions was determined by the Bio-Rad microcrither plate method (Bio-Rad, Hercules, CA). The subcellular preparations were assayed within 6 months.

**Assay Conditions.** Initial experiments showed negligible FAEE degradation in cytosolic preparations; thus, only microsomal preparations were investigated in remaining experiments. Two experiments (each in triplicate) were conducted for each of the four placenta studies. Microsomal protein (100 μg) was preincubated in 0.1 M potassium phosphate buffer (0.01 M EDTA and 2 mg/ml bovine albumin, total volume 200 μl) for 20 min at 37°C. A mixture containing equimolar concentrations of palmitic, stearic, oleic, and linoleic EEs (dissolved in methanol) was added (final concentrations of 0, 5, 25, 50, 100, and 200 μM for each FAEE), and samples were incubated for 120 min at 37°C. For 0 μM, 10 μl of methanol was added, and heat-inactivated microsomes were used in control experiments. The reaction was stopped by chilling on ice followed by the addition of 100 μl of ice-cold heptadecanoic EE (internal standard in hexane) and 5 ml of ice-cold extraction solvent (hexane/aceton, 5:2, v/v). FAEE were extracted and analyzed as described previously.

**Kinetic Analysis.** The velocity \((V)\) versus substrate \((\text{FAEE})\) data were linearized by plotting \([\text{FAEE}] / V\) versus \([\text{FAEE}]\) (i.e., Hanes-Wolff Plot). The kinetic parameters \((K_m\) and \(V_{max}\)) of FAEE degradation were derived from the line of best fit as determined by linear regression analysis. The maximum velocity \((V_{max})\) of FAEE degradation was calculated from the reciprocal of the slope and the apparent affinity \((K_m)\) from the product of the y-intercept and \(V_{max}\).

**Statistical Analysis.** The data were analyzed using the Student’s \(t\) test or the Mann-Whitney Rank Sum Test (for nonparametric data) using the SigmaStat software (version 2.03, SPSS Inc., Chicago, IL).

**Results**

**In Vitro Perfusion of Isolated Placental Cotyledons**

**Perfusion Studies without Albumin \((n = 4)\).** The mean mass of the perfused cotyledons was 15.27 ± 6.59 g (mean ± S.D.). There were no significant differences in the fetal arterial pressures between the control and experimental periods (mean ± S.D.): preexperimental control, 38.9 ± 1.4 mm Hg; experiment, 39.5 ± 0.9 mm Hg; and postexperimental control, 43.1 ± 2.1 mm Hg. The fetal flow rate was 2.48 ± 0.37 ml/min, and maternal flow rate was 13.11 ± 0.74 ml/min. The rates of oxygen and glucose consumption and the production of lactate and hCG did not differ in the control and experimental periods (data not shown).

The disposition of FAEE in the maternal and fetal reservoirs in the absence of albumin is shown in Fig. 1A. The levels of each ester decreased steadily during the 4-h perfusion, but none of the four FAEE was detectable in the fetal matrix throughout the experiment at both low (5 μM) and high (20 μM) maternal concentrations. At the end of the low concentration (5 μM) perfusions, only two of the four FAEE species (stearic and oleic EEs) remained in the maternal reservoir at less than 20% of the initial concentration. Between 18 and 88% of the initial concentration of the four esters remained in the maternal matrix at the end of the high concentration perfusions (20 μM). A linear decay was documented for all four FAEE when the maternal concentration versus time data were log transformed (i.e., log [FAEE] versus time), and the respective rates are shown in Table 1. At high concentration, the rates of FAEE elimination from the maternal reservoir were at least 3-fold slower than they were for low concentration perfusions \((p > 0.05)\). The fate of each FAEE postperfusion is shown in Fig. 2A as a mass balance. The amount of FAEE retained in the placenta was less than 6% at the end of low concentration perfusions and less than 17% after perfusing at high concentration. Linoleic EE was not retained by the perfused placenta. Stearic EE was consistently the FAEE the least eliminated from the maternal reservoir and most retained by the perfused placenta at both low and high concentrations. The net disappearance of individual FAEE decreased from greater than 85% at low concentration experiments to approximately 24 to 89% when high initial concentrations (20 μM) were used.

**Perfusion Studies with Albumin \((n = 6)\).** The mean mass of the perfused cotyledons was 9.85 ± 4.80 g (mean ± S.D.). There were no significant differences in the fetal arterial pressures between the control and experimental periods (mean ± S.D.): pre-experimental control, 46.2 ± 12.7 mm Hg; pre-experimental control (with albumin), 49.3 ± 16.3 mm Hg; experiment, 54.7 ± 10.4 mm Hg; and postexperimental control, 56.2 ± 8.1 mm Hg. The fetal flow rate was 3.45 ± 0.67 ml/min, and maternal flow rate was 14.51 ± 0.40 ml/min. The rates of oxygen and glucose consumption and the production of lactate and hCG did not differ in the control and experimental periods (data not shown).

The disposition of FAEE in the maternal and fetal reservoirs in the presence of albumin are shown in Fig. 1B. The levels of each ester decreased steadily during the 3-h perfusion, but none of the four FAEE was detectable in the fetal matrix throughout the experiment at both low (2 μM) and high (20 μM) maternal perfusate concentrations. Less than 15% of the initial concentration of each FAEE remained in the maternal reservoir at the end of the low concentration (2 μM) perfusions, whereas 14 to 49% of the initial concentration remained in the maternal matrix at the end of the high concentration perfusions (20 μM).
A linear decay was documented for all four FAEE when the maternal concentration versus time data were log transformed (i.e., log [FAEE] versus time), and the respective rates are shown in (Table 1). At high concentration, the rates of FAEE elimination from the maternal reservoir were approximately 50% slower than they were for low concentration perfusions \((p < 0.05)\) for stearic EE. In addition, the rate of elimination of stearic EE was significantly slower than that of oleic and linoleic EE \((p < 0.05)\) at 20 \(\mu\)M. The rates of FAEE elimination in the presence of albumin were not significantly different from the values obtained during experiments without albumin. The fate of each FAEE postperfusion is shown in Fig. 2B as a mass balance. The amount of FAEE retained in the placenta
was less than 4% at the end of low concentration perfusions and less than 12% after perfusing at high concentration. Stearic EE was consistently the FAEE the least eliminated from the maternal reservoir and most retained by the perfused placenta at both low and high concentration perfusions. The net disappearance of individual FAEE decreased from greater than 85% at low concentration experiments to 49 to 86% when high initial concentrations (20 μM) were used.

**Degradation of FAEE by Placental Microsomes.** Negligible metabolic activity was observed with cytosolic preparations during initial investigation (data not shown); therefore, only microsomal proteins were assayed in subsequent experiments.

**TABLE 1**  
The rates of elimination (min⁻¹) (mean ± SD) of individual FAEE from the maternal reservoir  
Log-transformed concentration versus time plots (i.e. log [FAEE] versus time) yielded linear relationships for all four FAEE, which suggests elimination following first order kinetics. Similar rates were found in experiments with and without the addition of albumin. At an initial concentration of 20 μM and in the presence of albumin, the rate of stearic EE elimination from the mother was significantly slower than oleic and linoleic EE (p < 0.05) and significantly slower than its rate at 2 μM (p < 0.05).

<table>
<thead>
<tr>
<th>FAEE</th>
<th>Palmitic EE</th>
<th>Stearic EE</th>
<th>Oleic EE</th>
<th>Linoleic EE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial 5 μM (no albumin, n = 2)</td>
<td>0.013 ± 0.011</td>
<td>0.007 ± 0.006</td>
<td>0.012 ± 0.009</td>
<td>0.026 ± 0.024</td>
</tr>
<tr>
<td>Initial 20 μM (no albumin, n = 2)</td>
<td>0.004 ± 0.003</td>
<td>0.002 ± 0.002</td>
<td>0.003 ± 0.002</td>
<td>0.008 ± 0.007</td>
</tr>
<tr>
<td>Initial 2 μM (n = 3)</td>
<td>0.012 ± 0.005</td>
<td>0.011 ± 0.004</td>
<td>0.016 ± 0.008</td>
<td>0.019 ± 0.010</td>
</tr>
<tr>
<td>Initial 20 μM (n = 3)</td>
<td>0.006 ± 0.002</td>
<td>0.004 ± 0.001</td>
<td>0.008 ± 0.002</td>
<td>0.010 ± 0.001</td>
</tr>
</tbody>
</table>

**Fig. 2.** The fate of FAEE postperfusion: A, in the absence of albumin; and B, in the presence of albumin. The addition of albumin had no effects on the metabolic fate of FAEE. Stearic EE was consistently the FAEE the least eliminated from the maternal reservoir and most retained by the perfused placenta at both low and high concentration perfusions.
experiments. The activity of placental microsomes in FAEE degradation is shown in Fig. 3, and the resulting kinetic values are summarized in Table 2. No degradation was observed when heat-inactivated microsomes were used. Among the four placentas studied, there was up to 4-fold variability in the apparent affinities ($K_m$) and 2-fold variability in maximum velocities ($V_{max}$) for individual FAEE. In the same placenta, the variability in these kinetic values was between 2- and 5-fold for individual FAEE. Linoleic and palmitic EE were associated with the highest apparent affinities, whereas the $K_m$ of stearic EE was consistently the lowest among all placentas studied ($p < 0.05$). The $V_{max}$ of stearic EE was significantly lower than the other three FAEE ($p < 0.05$), and it was the FAEE the least degraded by placental microsomes.

Fig. 3. Metabolism of FAEE by human placental microsomes ($n = 4$).
TABLE 2
Kinetic parameters of FAEE degradation by placental microsomes
Data shown are mean ± S.E.M.; n = 4.

<table>
<thead>
<tr>
<th>FAEE</th>
<th>(K_m) (\mu M)</th>
<th>(V_{max}) (\text{nmol/min/mg})</th>
</tr>
</thead>
<tbody>
<tr>
<td>Palmitic EE</td>
<td>95.42 ± 17.18</td>
<td>1.71 ± 0.22</td>
</tr>
<tr>
<td>Stearic EE</td>
<td>35.51 ± 12.53</td>
<td>0.57 ± 0.13</td>
</tr>
<tr>
<td>Oleic EE</td>
<td>66.53 ± 16.47</td>
<td>1.22 ± 0.20</td>
</tr>
<tr>
<td>Linoleic EE</td>
<td>94.47 ± 19.10</td>
<td>1.78 ± 0.27</td>
</tr>
</tbody>
</table>

Discussion

To the best of our knowledge, this is the first study to investigate the fate of FAEE within the maternal-placental-fetal unit and the role of the human placenta in FAEE metabolism. Understanding the origin of meconium FAEE is of critical importance in the development of FAEE as biological metabolites. The variability associated with the kinetic parameters of individual FAEE corroborated the trends observed during perfusion experiments. Linoleic EE was consistently associated with the highest \(K_m\) and \(V_{max}\) values, and it was the ester that was cleared the fastest and least retained by the placenta. As important, the apparent affinities and maximum velocities of stearic EE were the lowest among all placental preparations, which was consistent with the perfusion data that it was the ester associated with the slowest maternal elimination and the most retained by the placenta. The apparent affinity \(K_m\) of a substrate is inversely proportional to its affinity for the enzyme responsible for its metabolism; hence, stearic EE has the highest affinity for the enzymatic activity observed in the human placenta, and low concentrations are sufficient to saturate the enzymes.

The FAEE-hydrolyzing capacities of various organs and tissues have been reported in humans and several animal species, and significant variability was observed. Rapid hydrolysis of FAEE has been observed in rat plasma (oleic EE) (Hungund et al., 1995), rat adipose tissue (linoleic EE) (D’Pergola et al., 1991), and human blood (oleic EE) (Saghir et al., 1997, 1999) with elimination half-lives of less than 24 h. The rates of FAEE hydrolysis were found to be significantly different in human and rat serum \((0.34 ± 0.29\) versus \(15 ± 31\) nmol/min/ml, respectively) (Diczfalusy et al., 2001). Using tissue homogenates, Diczfalusy et al. investigated the capacities of various human organs and tissues to hydrolyze FAEE (palmitic EE) and concluded that the activity was highest in liver and pancreas \((1100–1600\) nmol/min/g) and lowest in heart and adipose \((10–15\) nmol/min/g) (Diczfalusy et al., 2001). There are very limited data on placental metabolism of FAEE that would be commonly detected in a fetal compartment.
FAEE in the literature. Using tissue homogenates, Bearer and colleagues have determined previously the FAEE synthetic activity of the human and mice placenta to be 0.2 and 1.2 mmol/h/mg, respectively, in the presence of 0.4 mM oleic acid and 200 mM ethanol (Bearer et al., 1992). These rates of synthesis are very slow in comparison with the rates of FAEE degradation found in our study ($V_{\text{max}}$ of various esters ranged between 0.6 and 1.8 nmol/min/mg), which supports our hypothesis that maternal FAEE are metabolized extensively by the human placenta and, thus, the net effect is to prevent their transfer to the fetus. The rate of FAEE degradation in our study is most comparable with that observed in purified human pancreatic trypsinogen lipase (oleic EE, 0.5 nmol/min/mg) (Riley et al., 1990) but significantly slower than in liver microsomes of humans (palmitic EE, 40 nmol/min/mg) (Diczfalusy et al., 2001) and rats (palmitic EE, 100 nmol/min/mg) (Diczfalusy et al., 1999), as well as purified porcine pancreatic carboxylester lipase (oleic EE, 13.8 $\mu$mol/min/mg) (Tsujita and Okuda, 1994).

The specific enzymes responsible for the degradation of FAEE in the human placenta are not known at this point. The linear relationships obtained from the kinetic studies may suggest several options: 1) the human placenta contains a single enzyme that is responsible for the degradation of FAEE, 2) multiple enzymes that are kinetically indistinguishable, or 3) one of these enzymes is dominant. The hydrolytic activities of FAEE in rat and human liver microsomes have been attributed to different species of carboxylesterases by the use of specific serine esterase inhibitors (Diczfalusy et al., 1999, 2001). Significant carboxylesterase activity toward the hydrolysis of 1-naphthyl acetate and carboxylic esters of p-nitrophenol has been documented in the microsomal fraction of term human placenta, and at least three isozymes have been identified (Yan et al., 1999). Whether the metabolic capacity observed in our study was associated with carboxylesterase or other enzymes requires further investigation.

Our studies show that FAEE originated in the maternal circulation are not transferred through the placenta to the fetus and that the maternally circulated FAEE are taken up and degraded extensively by the human placenta. Hence, FAEE detected in neonatal meconium or hair are likely produced by the fetus from the ethanol transferred to, and metabolized by the fetus, rendering FAEE as the first direct biomarkers of true fetal exposure to ethanol in utero.

References


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