Tretinoin Prevents Age-Related Renal Changes and Stimulates Antioxidant Defenses in Cultured Renal Mesangial Cells

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ABSTRACT

Age-related progressive glomerular sclerosis in the rat is associated with increased expression of tumor necrosis factor-β1, and increased protein content in the renal cortex, enhanced production of H₂O₂ in both renal glomeruli and mesangial cells (MCs) cultured from them, as well as augmented glomerular oxidative damage. We have previously shown that tretinoin-treated old male Fischer 344 rats have 30% lower protein content in the renal cortex than control old rats. Here, we report that this effect may depend on the inhibition of the expression of tumor necrosis factor-β1, a matrigenic cytokine, and osteopontin, a protein with cell adhesive and chemotactic properties. In addition, we show that tretinoin prevents the cytotoxicity of H₂O₂ in cultured human MCs by increasing both the catalase activity and the reduced glutathione content, which are dose- and time-dependent changes. These increases were not dependent on each other: when these effects were previously inhibited with 3-amino-1,2,4-atriazole or L-buthionine-(S,R)-sulfoximine, respectively, tretinoin still induced the increase in the other noninhibited antioxidant defense. An enhanced gene transcription is the most likely mechanism involved in the tretinoin-induced stimulation of MC antioxidant defense systems because 1) preincubation of MCs with actinomycin D or cycloheximide fully abolished it; 2) tretinoin-incubated MCs showed increased levels of catalase mRNA and γ-glutamyl-cysteine synthetase (catalytic subunit) mRNA, the latter being the rate-limiting step in de novo reduced glutathione synthesis; and 3) the stability of both mRNA was unchanged by tretinoin. These results show one strategy of protecting renal cells from H₂O₂-mediated injury based on increasing their antioxidant defenses.

Progressive glomerular sclerosis takes place in aging humans (Lindeman, 1990). Rats also exhibit an age-dependent renal deterioration, and male rats are more susceptible than female rats to age-related glomerulosclerosis (Baylis and Corman, 1998). Reactive oxygen species seem to play a role in the progression of age-related rat renal changes: there is an association between increased glomerular protein content and an augmented oxidative damage (i.e., increased levels of lipid peroxidation) in old rats (Ruiz et al., 1994). In addition, the production of H₂O₂ in glomeruli and cultured glomerular mesangial cells (MCs) is higher in samples from old rats than from those from young animals (Ruiz et al., 1994, 1996). On the other hand, H₂O₂ increases in cultured rat MCs, the production of transforming growth factor-β1 (TGF-β1) (P. Ruiz, personal communication), and this effect could account, at least in part, for the increased expression of this matrigenic cytokine (it contributes to glomerulosclerosis through promoting synthesis of extracellular matrix proteins and inducing inhibitors of metalloproteinases; reviewed by Sharma and Ziyadeh, 1994) in renal cortex of old rats (Ruiz et al., 1998). In addition to resident glomerular cells, leukocytes are other important source of reactive oxygen species and TGF-β1, and under appropriate conditions, these cells may infiltrate the renal glomeruli and contribute to the progression of a glomerular damage. An example of these conditions is the enhanced expression of osteopontin (OP), an arginine-glycine-aspartic acid (RGD)-containing acidic glycoprotein with cell adhesive and chemotactic properties (reviewed by Giachelli et al., 1995), which has been described in some models of renal disease, including progressive glomerulosclerosis (Narita et al., 1997) and age-related glomerulosclerosis (Floege et al., 1997).

Recently, we focused our attention on the treatment of age-related glomerulosclerosis with tretinoin (all-trans-retinoic acid), one of the active metabolites of vitamin A. Because it has antiantiactivator protein-1 activity in glomerular MCs

ABBREVIATIONS: BSO, L-buthionine-(S,R)-sulfoximine; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; SSC, standard saline citrate; γ-GCS, γ-glutamyl-cysteine synthetase; GSH, reduced glutathione; LDH, lactate dehydrogenase; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; RT, reverse transcription; MC, mesangial cell; NAC, N-acetylcysteine; OP, osteopontin; PCR, polymerase chain reaction; TGF-β1, transforming growth factor-β1.
(Simonson, 1994), we hypothesized that it could inhibit the activator protein-dependent effects of H₂O₂, such as cell death (Ishikawa et al., 1997; Xu et al., 1997) In fact, preincubation with tretinoin abolished H₂O₂-induced MC death (Moreno et al., 1997). Although the effects of tretinoin on TGF-β₁ expression are unpredictable a priori (some authors describe that the retinoid increases it in a given cell type, others find the opposite effect in other cell types, and, finally, there are examples in the literature of unaffected TGF-β₁ expression after tretinoin treatment), we found that tretinoin slows the progression of age-related glomerular changes in male Fischer 344 rats (Moreno et al., 1997). Therefore, we expect that its net effect on the expression of the matrigenic cytokine TGF-β₁ at the renal cortex level is inhibitory. In the same way, we also expect that the retinoid will inhibit the expression of other molecules (i.e., OP) involved in the progression of age-related glomerulosclerosis. Taking into account this background, the present work was designed with two objectives. The first objective was to study, in the context of aging, the effect of tretinoin on the renal expression of TGF-β₁ and OP, and the second objective was to examine the effect of the retinoid on antioxidant defenses of cultured glomerular MCs as a possible mechanism of tretinoin in preventing H₂O₂-induced cytotoxicity.

Materials and Methods

Tretinoin (all-trans-retinoic acid) was kindly donated by Productos Roche S.A. (Spain). Unless otherwise stated, all of the biochemical reagents used in this study were purchased from Sigma Chemical Co. (St. Louis, MO). Tissue culture materials, growth media, and serum for cell culture were obtained from GIBCO (Grand Island, NY), and the RNA PCR kit was from Perkin-Elmer (Roche Molecular Systems Inc., Branchburgh, NJ). All other chemicals used were of the purest grade commercially available.

In Vivo Experiments

Animals and Diets. Because we were mainly interested in the early stages of spontaneous glomerulosclerosis, when it is more likely to obtain any benefit from the dietary protocol, the study was not performed in very old rats. Therefore, thirty 18-month-old male Fischer 344 rats were fed 7 days per week with standard chow (control, n = 15) or with standard chow plus tretinoin (tretinoin-treated) for a period of 90 days. Food containing tretinoin (all-trans-retinoic acid) was prepared daily in the following way: a solution of tretinoin in absolute ethanol (1.5 g tretinoin/liter) was mixed in a dark, cold room with standard chow (2 ml tretinoin solution/100 g food). Once ethanol evaporated, 15 g of food (the average daily consumption of food per rat) were given to each rat every day: this renders a daily intake of tretinoin of about 1 mg/kg b.wt. The dose of tretinoin was adjusted each week depending on the body weight consumption of food per rat) were given to each rat every day: this renders a daily intake of tretinoin of about 1 mg/kg b.wt. The dose of tretinoin was adjusted each week depending on the body weight. The dose of tretinoin was adjusted each week depending on the body weight.

After 90 days of treatment, animals received ether anesthesia. Blood taken from the lower aorta was used for hematological studies, and serum isolated from each blood sample was used for biochemical analyses. Pieces of the left kidney cortex were weighed and homogenized in phosphate buffer (pH 7.4) supplemented with 0.1% Triton X-100, 3 mM EDTA, and 2 M NaCl. Aliquots of the lysate were used for the measurement of protein and DNA (see below).

A piece of the renal cortex from the right kidney was collected in a sterile tube containing a denaturing solution for total RNA extraction (Chomczynsky and Sacchi, 1987). Both the quality and the quantity of the RNA were verified by ethidium bromide staining of rRNA bands on an agarose minigel. RNA samples were used to study the expression of TGF-β₁ and OP. A set of RNA renal samples from 21-month-old rats was also used to further study the age-related changes in the expression of OP.

In Vitro Experiments

Cells. Human MCs were obtained from adult nephrectomy specimens as we previously described (Diez et al., 1995). Culture medium was made of RPMI 1640 supplemented with 10% FCS, 200 mM l-glutamine, and antibiotics (100 U/ml penicillin, 100 µg/ml streptomycin, and 0.25 µg/ml amphotericin B). Confluent cells between the 12th and 15th passages were used, and they were made quiescent by 48 h incubation with medium supplemented with 0.5% FCS.

Stimulation of Antioxidant Defenses. Fresh medium with tretinoin or its vehicle (ethanol at a final concentration of 0.09%) was added to quiescent cells. Tretinoin effects on antioxidant defenses (i.e., catalase activity and reduced glutathione (GSH) content) were first tested in dose-response experiments, with the range of retinoid concentration 0.01 to 50 µM and an incubation period of 24 h, and then tested in time-response experiments with 10 µM tretinoin. An approximate value for the highest tretinoin-induced increase in intracellular GSH content was obtained in other experiments, in which the synthesis of glutathione was favored through increasing the availability of cysteine by the addition of 10 mM N-acetylcysteine (NAC) (Deneke and Fanburg, 1989) after incubation with 10 µM tretinoin.

Then, a set of experiments conceived to explore the mechanisms responsible for the stimulation of antioxidant defense systems was designed as described. Effect of dl-α-tocopherol on antioxidant defenses. The contribution of antioxidant properties to the effects of tretinoin on the cellular antioxidant defenses was examined in experiments identical with those described above but cells were incubated with 10 µM dl-α-tocopherol instead of tretinoin.

Effect of blockade of tretinoin-induced stimulation of catalase activity on increase in intracellular GSH. Cells were preincubated for 1 h under control conditions or with 5 mM 3-amino-1,2,4-atriazole, an irreversible inhibitor of catalase activity (Aebi, 1983). Then, they were incubated for 24 h with or without 10 µM tretinoin, and the intracellular GSH content was measured. Cells incubated in parallel in the same experimental conditions were used to assess the cellular activity of catalase.

Effect of blockade of tretinoin-induced increase in intracellular GSH content on stimulation of catalase activity. Cells were preincubated for 1 h in control conditions or with 0.2 mM l-buthionine-(S,R)-sulfoximine (BSO), an irreversible inhibitor of γ-glutamyl-cysteine synthetase (γ-GCS), the rate-limiting step of glutathione synthesis (Griffith, 1982). Then, they were incubated for 24 h with or without 10 µM tretinoin, and the catalase activity was measured. Cells incubated in parallel in the same experimental conditions were used to assess the intracellular GSH content.

Effect of actinomycin D and cycloheximide on tretinoin-induced stimulation of antioxidant defenses. In experiments using these inhibitors of mRNA or protein synthesis, respectively, cells were preincubated for 30 min under three different conditions: control (no inhibitors), 2 µg/ml actinomycin D, and 10 µM cycloheximide. Cells were washed with fresh medium and incubated during 24 h with or without 10 µM tretinoin. Then, catalase activity and GSH content were measured.

Effect of tretinoin on the expression of catalase and γ-GCS. In experiments of dose and time responses to tretinoin, total RNA was extracted to study the expression of catalase and of the catalytic unit of γ-GCS, the rate-limiting step of glutathione synthesis (Griffith, 1982).

mRNA stability. Cells were treated with or without 10 µM tretinoin for 24 h. Actinomycin D (2 µg/ml) was added to the medium, and total RNA was isolated at different time points after treatment.
Northern blot analysis for catalase as well as semiquantitative cDNA amplification γ-GCS (catalytic subunit), normalization, and quantification were carried out as described below.

Prevention by Tretinoin of H₂O₂-Induced Cell Damage. Fresh medium with 1 to 10 μM tretinoin or its vehicle (ethanol at a final concentration of 0.09%) was added to quiescent cells cultured in 96-well microtiter plates (typically 20,000 cells/well). After a 24-h incubation, cells were washed with fresh medium and incubated for 24 h with 0 to 150 μM H₂O₂. Cytotoxicity was quantified as described below by measuring both lactate dehydrogenase (LDH) activity released from the cytosol of damaged cells into the supernatant and the ability of cells to reduce exogenous 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) to formazan. The first test is a measure of plasma membrane integrity, whereas the second test detects living but not dead cells, as the tetrazolium ring is cleaved in active cells (McGahon et al., 1995).

Effect of Tretinoin on Expression of OP and TGF-β₁. Fresh medium with tretinoin or its vehicle (ethanol at a final concentration of 0.09%) was added to quiescent cells. Tretinoin effects on the expression of OP and TGF-β₁ were studied in dose- and time-response experiments similar to those performed to stimulate antioxidant defenses.

Analytical Procedures

Hematological parameters (red blood cell count, hemoglobin concentration, hematocrit, white blood cell count, and platelet count) were measured in a Coulter Counter (model Sar), and serum parameters, including triglycerides, cholesterol, creatinine, glucose, GPT, GPT, bilirubin, sodium, potassium, calcium, and uric acid concentration, were analyzed in an Hitachi 717 (Boehringer Mannheim, Mannheim, Germany).

DNA and Protein Content in Renal Cortex. DNA in renal cortex was measured using the DNA-binding fluorochrome H33258 (purchased from Sigma Chemical) in aliquots of the renal cortex homogenate briefly sonicated (Labarca and Paigen, 1980). In this assay, EDA prevents DNase activity. Aliquots of the homogenate were mixed with the homogenization buffer containing compound H33258 to a final concentration of 1 μg/ml and fluorescent measurements were made in a scanning fluorescence spectrometer (model LS-5B; Perkin-Elmer), with the excitation wavelength set at 365 nm and the emission wavelength set at 460 nm. The DNA content of the samples was calculated from a standard curve made with calf thymus DNA standards at defined concentrations and expressed as mg DNA/g renal cortex. Protein was assayed (Lowry et al., 1951) and results were expressed as mg/g renal cortex.

Catalase Activity and GSH Content. Catalase activity was measured as follows (Aebi, 1983): In a quartz cuvette, 2 ml of sample (previously diluted adequately in phosphate buffer (KH₂PO₄, 50 mM, pH 7.0) containing 0.2% Triton X-100) were added to 1 ml of 30 mM H₂O₂. Changes in absorbance at 240 nm were measured for 30 s. The rate constant of a first order reaction (k) was used as a unit according to the equation k = (1/t₂₀ - t₁₀) × ln(A₁₀/A₂₀), where t₁₀ and t₂₀ is the measured interval in seconds, and A₁₀ and A₂₀ are the absorbances at initial and final measurement points, respectively.

Cells for GSH content measurements were lysed in a cold room. Cellular proteins were precipitated with 0.9 ml of perchloric acid. After neutralization with 0.3 ml 1 M KOH/KHCO₃ and centrifugation, 0.15 ml of supernatant was collected on plastic tubes, o-phthalaldehyde was added (0.15 ml of 7.46 μM solution), and tubes were incubated at room temperature for 15 min. GSH content was measured in a scanning fluorescence spectrophotometer (model LS-5B; Perkin-Elmer) at 420 nm with excitation wavelength of 350 nm, using a standard curve (Hissin and Hilf, 1976).

Results of catalase activity and GSH content were corrected by cellular protein, which was measured according to Lowry et al. (1951).

mRNA Expression. Semiquantitative cDNA amplification of TGF-β₁ and γ-GCS. We used semiquantitative cDNA amplification as a sensitive method to assess the level of TGF-β₁ transcripts in the rat kidney cortex (Ruiz et al., 1998). For the purposes of semiquantification, PCR components were premixed (to generate master mixes) before addition to individual PCR tubes to minimize pipetting errors, and all samples underwent PCR at the same time in the same experiment. One microgram of total RNA was reverse transcribed in a total reaction volume of 20 μl through incubation at 42°C during 30 min. All RT reactions used oligo(dT)₁₅ primer to minimize the variations in RT efficiency seen when using specific RT primers.

The reaction product was amplified by polymerase chain reaction (PCR) using a thermal cycler (MJ Research Inc., Watertown, MA). PCR conditions were determined in the next manner. First, PCR conditions were optimized, and then comparative kinetic analyses (Noonan et al., 1990; Salomon et al., 1992) were performed to determine the phase during which there was exponential generation of PCR product before reaching plateau. It was at this point that the PCR was terminated, allowing for semiquantitative data to be obtained. In summary, semiquantitative cDNA amplification for TGF-β₁ transcripts was performed as follows: it was started with 4 min of denaturation at 94°C followed by 30 PCR cycles. Each cycle consisted of 1 min at 94°C, 1 min at 55°C, and 1 min at 72°C. To quantify PCR products comparatively and confirm the use of equal amounts of the RNAs, we coamplified a house-keeping gene, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (Hughes et al., 1995). Contamination was ruled out by the fact that PCR was negative when the reaction was performed without a prior reverse transcription (RT) reaction.

The upstream and downstream TGF-β₁ primers were 5′-CT-TCACGTCACAGAAGAGACCTC-3′ and 5′-CACAGATCATGTTGGACAACTGCTCC-3′, respectively, which yielded a single band corresponding to a 298-bp cDNA fragment (Qian et al., 1980). Analysis with cycle sequencing revealed that the sequence was identical with position 1266–1564 in rat TGF-β₁ cDNA (Ruiz et al., 1998). The upstream and downstream GAPDH primer sequences were 5′-GTAAGGGCTCGGTGTCACGGATT-3′ and 5′-CACAGATCTCTCAGTGCCAGGTGAT-3′, respectively, which yielded a single band corresponding to a 558-bp cDNA fragment (Hughes et al., 1995). Analysis with cycle sequencing revealed that the sequence was identical with position 3–561 in rat GAPDH cDNA (Ruiz et al., 1998). After amplification, 8 μl of each PCR mix was electrophoresed through a 2% (w/v) agarose gel with ethidium bromide (0.5 μg/ml). Gels were photographed with Polaroid 55 negative film, with the reaction products on the negative film scanned by densitometry using an Image-store Color Onesscanner and analyzed with NIH Image 1.55 Software. The TGF-β₁/GAPDH product ratio (with GAPDH used as an internal standard to correct for sample-to-sample variation in RNA degradation) was calculated and considered an index of TGF-β₁ mRNA expression.

To quantify γ-GCS mRNA expression an RT-PCR, an assay was used in the same conditions as for TGF-β₁, with the only exception that annealing was achieved at 52°C rather than at 55°C and the number of PCR cycles was 24. Oligonucleotide primers were selected from the published sequence of γ-GCS catalytic subunit: the upstream and downstream sequences were 5′-TGCTGTACTCTGTCAC-CAGATGGATCCTC-3′ and 5′-TGATCCAAATGTAACCTTGAGCATT-TCACA-3′, respectively, which yielded a single band corresponding to a 531-bp cDNA fragment (William MacNee, personal communication). To quantify PCR products comparatively and confirm the use of equal amounts of the RNAs, we coamplified a house-keeping gene, β-actin, with primer sequences obtained from Stratagene (catalog no. 302110), yielding a 245-bp fragment.

Northern blot for catalase mRNA and OP mRNA. Catalase and OP mRNA levels were measured in total RNA extracts isolated from cultured human M5Cs. OP mRNA levels were also measured in RNA samples from rat renal cortex. Total RNA was electrophoresed in a denaturing 1% agarose gel, transferred to a nylon membrane, and probed either with ³²P-labeled pCAT10, a purified human liver catalase cDNA clone obtained from American Type Culture Collection
Effect of Tretinoin on MC Antioxidant Defenses. To explore the mechanism through which tretinoin prevents the effects of \( \text{H}_2\text{O}_2 \) in cultured human renal MCs (Moreno et al., 1997), we first studied the catalase activity, and the GSH content; catalase is an obvious candidate to be involved because \( \text{H}_2\text{O}_2 \) is its substrate. GSH is critical for another defense system against hydrogen peroxide and other peroxides: the glutathione peroxidases. Dose-response experiments (tretinoin concentrations tested were 0.1, 1, 5, 10, 25, and 50 \( \mu \text{M} \)) showed that these antioxidant defenses were stimulated within a narrow range of tretinoin concentrations: 1 \( \mu \text{M} \) tretinoin was the lowest effective concentration, whereas toxicity (in terms of LDH release, diminished reduction of MTT to formazan, and diminished cell count) appeared at 25 \( \mu \text{M} \) tretinoin (results are not shown). A concentration of 10 \( \mu \text{M} \) tretinoin was finally chosen for the time-response experiments because 1) it had the maximum stimulatory effect on the antioxidant defenses and 2) we had previously shown that 10 \( \mu \text{M} \) tretinoin prevents the cellular effects of \( \text{H}_2\text{O}_2 \) on cultured human MCs (Moreno et al., 1997).

As shown in Fig. 3, 10 \( \mu \text{M} \) tretinoin had a stimulatory, time-dependent effect on both the activity of catalase and the intracellular GSH content of MCs. Moreover, when the availability of cysteine, one of the amino acids of the tripeptide glutathione (\( \gamma\)-GCS), was increased by the addition of NAC (and consequently the intracellular GSH content was 500% higher than in control cells), preincubation with 10 \( \mu \text{M} \) tretinoin still increased cell GSH by 1200% over control cells (Fig. 4). Overall, these results are consistent with the theory that tretinoin prevents the cellular actions of \( \text{H}_2\text{O}_2 \) through a stimulatory effect on cell antioxidant defenses. In addition, we may reasonably exclude any contribution of retinoid intrinsic antioxidant properties because incubation of MCs with other lipid antioxidants such as \( \alpha\)-tocopherol did not result in any stimulatory effect on catalase activity or on GSH content (results are not shown).

Interdependence between Tretinoin-Induced Increase in Catalase Activity and GSH Content. Given the parallel behavior of catalase activity and GSH content after incubation with tretinoin, we tested the possibility that one of the changes was the consequence of the other. So, we first studied the possible effect of the tretinoin-induced stimulation of catalase activity on the increase of intracellular GSH content found after tretinoin incubation.

Experiments were carried out as described in Table 1: cells were first incubated with aminotriazole, an irreversible inhibitor of catalase activity (Aebi, 1983) that lowers this activity about 99% in MCs, and then with tretinoin. Under these conditions, tretinoin was still able to increase the intracellular GSH content to a similar extent as in the absence of aminotriazole. In a similar way, experiments were designed to study the possible effect of the tretinoin-induced increase of GSH content on the stimulation of catalase activity found after tretinoin incubation. Here, the inhibition of GSH synthesis was achieved by incubation with BSO, which reduces the mesangial GSH content about 60%. Under these conditions, tretinoin still retained its ability to stimulate cellular catalase activity (Table 1). In conclusion, the two effects observed in cells incubated with tretinoin (i.e., increase in both catalase activity and intracellular GSH content) do not appear to be dependent on each other but rather are directly induced by the retinoid.
It is also interesting to mention that the addition of tretinoin to the aminotriazole-treated cells or to the BSO-treated cells increased the catalase activity or the GSH content, respectively, back to levels of the untreated controls (Table 1).

Mechanism of Tretinoin-Induced Stimulation of Mesangial Antioxidant Defenses. To explore the mechanism of tretinoin-induced stimulation of mesangial antioxidant defenses, we first observed the effects of RNA/protein synthesis inhibitors. We added actinomycin D for the inhibition of RNA synthesis and cycloheximide for the inhibition of protein synthesis to the culture media. As shown in Table 2, actinomycin D and cycloheximide fully abolished the stimulatory effect of tretinoin on cell antioxidant defenses. These observations suggest that tretinoin-induced stimulation of mesangial antioxidant defenses requires de novo synthesis of responsible gene products. In fact, further experiments (Fig. 5) showed that tretinoin-treated cells had increased levels of both catalase mRNA and γ-GCS (catalytic subunit) mRNA, with this enzyme being the rate-limiting step in de novo GSH synthesis.

Because a change in mRNA stability could account for the
Changes in steady state levels of both catalase mRNA and γ-GCS mRNA after retinoid treatment, we performed mRNA stability analyses in which cells were first treated with 10 μM tretinoin or its vehicle for 24 h and then treated with actinomycin D to block further mRNA synthesis. We then measured the decay time for catalase mRNA and γ-GCS mRNA after retinoid treatment, we performed mRNA stability analyses in which cells were first treated with 10 μM tretinoin or its vehicle for 24 h and then treated with actinomycin D to block further mRNA synthesis. We then measured the decay time for catalase mRNA and γ-GCS mRNA.

### TABLE 1

<table>
<thead>
<tr>
<th>Catalyst Activity</th>
<th>GSH Content</th>
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<tbody>
<tr>
<td>Catalase Activity</td>
<td>GSH Content</td>
</tr>
<tr>
<td>mK/mg protein</td>
<td>nmol/mg protein</td>
</tr>
<tr>
<td>Control</td>
<td>48.1 ± 4.9</td>
</tr>
<tr>
<td>Aminotriazole</td>
<td>1.2 ± 1.1*</td>
</tr>
<tr>
<td>BSO</td>
<td>47.2 ± 4.5</td>
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<tr>
<td>Tretinoin</td>
<td>102.3 ± 12.2*</td>
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<tr>
<td>Aminotriazole + tretinoin</td>
<td>33.6 ± 6.3</td>
</tr>
<tr>
<td>BSO + tretinoin</td>
<td>99.3 ± 10.6*</td>
</tr>
</tbody>
</table>

*p < .01 versus control.

### TABLE 2

<table>
<thead>
<tr>
<th>Catalyst Activity</th>
<th>GSH Content</th>
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<tbody>
<tr>
<td>Catalase Activity</td>
<td>GSH Content</td>
</tr>
<tr>
<td>mK/mg protein</td>
<td>nmol/mg protein</td>
</tr>
<tr>
<td>Control</td>
<td>47.2 ± 5.4</td>
</tr>
<tr>
<td>Tretinoin</td>
<td>99.1 ± 8.2**</td>
</tr>
<tr>
<td>Actinomycin D</td>
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<tr>
<td>Cycloheximide</td>
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<tr>
<td>Actinomycin D + tretinoin</td>
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</tr>
<tr>
<td>Cycloheximide + tretinoin</td>
<td>37.7 ± 4.6</td>
</tr>
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*p < .05 versus control; **p < .01 versus control.
mRNA and, therefore, in the tretinoin-induced stimulation of mesangial antioxidant defenses.

**Prevention by Tretinoin of H2O2-Induced Cell Damage.** H2O2 induced a dose-dependent damage in MCs. Quiescent human MCs were incubated for 24 h with 1 to 25 μM tretinoin (left) or with 10 μM tretinoin for 0 to 24 h (right), and total RNA was harvested. Top, RNA was reverse-transcribed and used for PCR analysis of γ-GCS mRNA. Bottom, insets (top) show the amplification product obtained after RT-PCR assay of one representative pair of experiments, and the graphs (bottom) represent the mean ± S.D. of the values of the densitometric analysis, obtained from four separate experiments, normalized to β-actin (whose expression was not significantly influenced by tretinoin treatment). The results showed a significant increase in γ-GCS/β-actin from 1 μM tretinoin in the dose-response experiments (left) and from 8-h incubation with 10 μM tretinoin in the time-response experiments (right). B, effect of tretinoin on catalase mRNA expression in human MCs. Quiescent human MCs were incubated for 24 h with 1 to 25 μM tretinoin (left) or with 10 μM tretinoin for 0 to 24 h (right) and total RNA was harvested. Northern blotting (top) and its densitometric analysis (bottom) were carried out as described in Materials and Methods. The autoradiograms (top) are representative of four separate experiments. The graphs represent the mean ± S.D. of the values normalized to GAPDH and expressed as arbitrary units. Densitometric analysis (bottom) showed a significant increase in catalase/β-actin cDNA levels were normalized with a coamplified housekeeping gene, β-actin. 

**Discussion**

Renal function in humans declines with advancing age. The classic structural finding with aging is glomerulosclerosis, leading to complete glomerular obsolescence and glomerular dropout. This bleak outlook, however, has been challenged by the results of the Baltimore Longitudinal Study, which shows that an age-dependent fall in glomerular filtration rate is not inevitable (Lindeman, 1990).

Taken into account that the age-related renal damage may be inevitable, a number of treatments to prevent it have been tested in the rat model. We have previously shown (Moreno et al., 1997) that tretinoin slows the progression of age-related glomerular changes in male Fischer 344 rats (in 18-month-old rats of this strain, we have described that glomerular protein content is about 3-fold higher than that in 3-month-old rats (Ruiz et al., 1994)). Other early age-associated changes in 18-month-old rats include increased glomerular production of H2O2 and increased expression of TGF-β1 in the renal cortex (Ruiz et al., 1994, 1998)). Here, we confirm these previous findings: 18-month-old rats treated with tretinoin have a lower protein content in the renal cortex than do control animals, whereas no differences in the DNA content of this region were observed between the two experimental groups (see Results). Because the DNA values indicate that cell numbers in the renal cortex were similar between the two groups of animals, the higher protein content of this region in the control rats could reflect an expansion of the protein in the extracellular matrix and/or a high cellular dose within a narrow range: 10 μM tretinoin had the maximum protective action against H2O2 and 2.5 μM tretinoin was the lowest dose with a protective effect (Fig. 7B).
protein content (Ruiz et al., 1994). These results suggest that tretinoin may slow the rate of progression of glomerulosclerosis acting on the balance synthesis/degradation of extracellular matrix and/or on the cellular hypertrophic changes found as aging progresses.

We had hypothesized that the action of tretinoin involved an inhibitory effect on the expression of OP and TGF-β1. The expression of OP has been reported to be up-regulated in a diseased kidney before macrophage infiltration and increased fibrosis (Narita et al., 1997), and it is predominantly expressed in chronic and progressive glomerulosclerosis (Floge et al., 1997). Increased expression of TGF-β1 in the renal cortex of Fischer 344 rats during normal aging (Ruiz et al., 1998) may account for glomerulosclerosis because TGF-β1 promotes synthesis of extracellular matrix proteins and induces inhibitors of metalloproteinases (Sharma and Ziyadeh, 1994). Our results indicate that the expression of OP also increases in renal cortex (Fig. 1) during normal aging (therefore, this increase may be related to the progression of age-related glomerulosclerosis) and that tretinoin inhibits this increase (Fig. 1). Moreover, this inhibition has also been specifically found in cultured human MCs from the renal cortex (Fig. 2), a fact that suggests that these cells may be one of the targets of the inhibitory effect of tretinoin in vivo. Renal cortex samples from tretinoin-treated rats also had lower TGF-β1 expression than the corresponding samples from age-matched control rats (Fig. 1). The retinoid had no effect on the TGF-β1 expression of cultured human MCs (see Results), this suggesting that in opposition to OP, these cells may not be the target for the inhibitory in vivo effect of tretinoin on TGF-β1. In summary, the inhibitory effect on OP expression, with OP being a protein with cell adhesive and chemotactic properties (Giachelli et al., 1995), and on TGF-β1 expression, which is a matrigenic cytokine, in old rats may be an important mechanism through which the retinoid slows the progression of age-related renal cortex changes. However, immunostaining for TGF-β1 and OP should be performed 1) to confirm that tretinoin inhibits not only the mRNA expression but also the production of the respective proteins and 2) to show specific localization of TGF-β1 and OP in glomeruli (thus eliminating any concern about material measured in renal cortex being in tubular cells or urine). In addition, our results suggest that although tretinoin toxicity is not negligible, a rational treatment may provide significant benefits in the prevention of age-related renal changes.

Regarding the in vitro studies, we previously described that preincubation with tretinoin abolishes H2O2-dependent cytotoxicity in cultured human MCs (Moreno et al., 1997). This action could be useful in the treatment of several forms of renal damage because 1) both glomeruli and cultured glomerular MCs from 18-month-old rats produce increased amounts of H2O2 and there is an augmented oxidative damage in glomeruli (Ruiz et al., 1994, 1996) during aging and 2) the production of H2O2 might result in the death of glomerular cells found during the late stages of the sclerosis of the glomerular cortex (Schlondorff, 1995).

To elucidate the mechanisms involved in the prevention of H2O2-induced cell responses, we first studied catalase activity and GSH content: with H2O2 being its substrate, catalase is an obvious candidate to be involved in the protective effect of tretinoin (Aebi, 1983). In addition, GSH is critical for another defense system against H2O2 and other peroxides: the glutathione peroxidases (Deneke and Fanburg, 1989). Our results confirmed that tretinoin was able to induce a dose- and time-dependent increase in both the catalase activity and the GSH content in cultured human renal MCs (Fig. 3). The possibility that an antioxidant intrinsic activity of tretinoin could partially contribute to the increase in the antioxidant defenses through a “saving effect” was ruled out because cells incubated with dl-α-tocopherol had no changes in these defenses (results are not shown).
We tested then the possibility that one of the changes in these antioxidant systems was the consequence of the other. Experiments in cells preincubated with aminothiazole, an inhibitor of catalase, showed that tretinoin increased the GSH content to the same extent as that found in the absence of aminothiazole (Table 1). Tretinoin also retained its ability to stimulate catalase activity in cells preincubated with BSO, an inhibitor of GSH synthesis (Table 1). In conclusion, the increases in both catalase activity and intracellular GSH content do not appear to be dependent on each other but rather directly induced by the retinoid.

Interestingly, the addition of tretinoin to aminothiazole- or BSO-treated cells increased the catalase activity or the GSH content, respectively, back to the untreated control levels (Table 1). These results may be reflecting the synthesis de novo of catalase and GSH induced during 24 h by the retinoid after washing out the inhibitors and/or interference of tretinoin with the ability of aminothiazole and BSO to inhibit catalase activity or γ-GCS, respectively.

There is evidence suggesting that tretinoin-induced stimulation of mesangial antioxidant defenses requires de novo synthesis of responsible gene products. First, the relative levels of oxidized and GSH are regulated by a series of coupled reactions involving glutathione peroxidase, glutathione reductase, and glucose-6-phosphate dehydrogenase (Deneke and Fanburg, 1989). Under normal conditions, the balance of the equation is far in the direction of maintaining cellular glutathione in its reduced state: GSH >99% (Deneke and Fanburg, 1989). Taken into account this value, it seems unlikely that the tretinoin-induced increase in cell GSH content, which may reach 250%, could be primarily dependent on an increased activity of the enzymes directly or indirectly involved in the reduction of oxidized glutathione. In fact, when the availability of cysteine, one of the amino acids of the tripeptide glutathione (γ-GCS), is increased by the addition of NAC and, consequently, the intracellular GSH content is 500% higher than that in control cells, preincubation with 10 μM tretinoin still increases cell GSH by 1200% over control cells (Fig. 4). Second, tretinoin-induced stimulation of mesangial antioxidant defenses was fully inhibited by either actinomycin D or cycloheximide (Table 2). Third, the levels of catalase mRNA and γ-GCS (catalytic subunit), the initial and rate-limiting enzyme in the glutathione de novo synthesis pathway (Deneke and Fanburg, 1989), were indeed found to be increased after incubation of MCs with tretinoin (Fig. 5). Fourth, stability analyses of catalase mRNA and γ-GCS mRNA showed that both mRNAs are no more stable in tretinoin-treated cells than they are in control cells (Fig. 6).

These observations suggest that an increased gene transcription of catalase mRNA and γ-GCS mRNA is directly related to the tretinoin-induced stimulation of mesangial antioxidant defenses. This mechanism is not a new one, given that retinoid signaling mechanisms are based on the interaction on the gene expression at the transcriptional level (Giguere, 1994). On the other hand, the transcription, expression, and increasing activity of catalase after ionizing radiation or oxidant stress are well known (Heintz et al., 1991; Hardmeier et al., 1997). The same is true for the catalytic subunit of γ-GCS (Rahman et al., 1996). The interest of our finding is that it provides one strategy by which to protect renal cells from H2O2-mediated injury: to augment cell anti-H2O2 activity levels by increasing their GSH content and their catalase activity through tretinoin treatment. In fact, there is a clear association between the dose-response stimulation of cell antioxidant defenses and the dose-dependent preventive action of tretinoin against the H2O2 cytotoxicity (Fig. 7).

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