Specific Delivery of Captopril to the Kidney with the Prodrug Captopril-Lysozyme

R. J. KOK, F. GRIJPSTRA, R. B. WALTHUIS, F. MOOLENAAR, D. de ZEEUW and D. K. F. MEIJER

Departments of Pharmacokinetics and Drug Delivery (R.J.K., F.G., R.B.W., F.M., D.K.F.M.) and Clinical Pharmacology (D. de Z.), Groningen Utrecht Institute for Drug Exploration, Groningen, the Netherlands

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ABSTRACT

Low-molecular-weight proteins (LMWPs) accumulate in the proximal tubular cells of the kidney, which makes these proteins interesting tools for renal drug targeting. We studied this approach using the LMWP lysozyme as a carrier for the angiotensin-converting enzyme inhibitor captopril. Captopril was conjugated to lysozyme via a disulfide bond. The pharmacokinetics of the captopril-lysozyme conjugate were studied in the rat. Only intact conjugate could be detected in the circulation. The total amount of captopril disulfides in the kidney was six times higher after administration of the conjugate than after the administration of an equivalent amount of free captopril. The conjugate was recovered in the urine partially as intact conjugate and partially as low-molecular-weight disulfides. The excretion of conjugate in the urine was not a consequence of the coupling of captopril to lysozyme because an equivalent bolus dose of native lysozyme was similarly excreted into the urine. By determination of the renal angiotensin-converting enzyme activity, we showed that the conjugate was degraded to the pharmacologically active captopril in vivo. We conclude that the coupling of captopril to the LMWP lysozyme results in increased captopril concentrations in the kidney and reduced captopril concentrations in the circulation.

Materials and Methods

Chemicals. CAP, LZM (3× crystallized, grade I), and Micrococcus lysodeicticus were purchased from Sigma Chemical (St. Louis, MO). Succinimidylloxycarbonyl-α-methyl-α-(2-pyridyldithio)toluene (SMPT) was purchased from Pierce (Rockford, IL). Tributylphosphine (TBP) (purum) was obtained from Fluka (Buchs, Switzerland). All solvents for high-performance liquid chromatography (HPLC) were of HPLC quality (Labscan, Dublin, Ireland). Water was purified with an Elgastat Maxima-HPLC water system (High Wycombe, UK).

Synthesis and Characterization of CAP-LZM. The preparation and characterization of the CAP-LZM conjugate have been described in detail elsewhere (R. J. Kok et al., submitted). The conjugate that was used in this study had a protein content of more than 95% and a degree of drug substitution of 0.4, which indicates that the product consisted a mixture of native LZM and 1:1 CAP-LZM conjugate.
**Pharmacokinetic Experiments.** Male Wistar rats (280–300 g) were obtained from Harlan (Zeist, the Netherlands). The plasma disappearance and urinary excretion of the conjugate were determined in freely moving rats that were equipped with a permanent cannula in the jugular vein 1 week before the experiment (Steffens, 1969). The distribution of CAP to the kidney and the ACE inhibition in the kidney were determined in rats that had received an i.v. bolus dose via the dorsal penile vein under halothane anesthesia.

**Plasma Disappearance and Excretion in the Urine.** A day before the experiment, the rats were placed in metabolic cages, in which they had free access to food and water. From this moment on, the rats received an infusion of 5% glucose at a rate of 2 ml · h⁻¹. At the start of the experiment, the rats received an i.v. bolus dose via the cannula of either the conjugate [(33 mg · kg⁻¹, dissolved in approximately 0.5 ml of 5% glucose (n = 10)], or equivalent doses of uncoupled CAP and LZM (n = 6). Another group of rats received a bolus dose of LZM alone (33 mg · kg⁻¹; n = 6). Blood samples for four of the CAP-LZM rats and all of the CAP plus LZM rats were drawn at several time points via the cannula. Blood samples were heparinized and immediately centrifuged (5 min at 13,000 rpm) to obtain plasma. The rats were killed spontaneously, after which the urine was collected per fraction into preweighed plastic tubes. The collection of the urine was continued up to 10 h after the administration of the drugs. The plasma and urine samples were analyzed as described in the text.

**Distribution of CAP to the Kidney and ACE Inhibition in the Kidney.** Under a light anesthesia of halothane, the rats (280–300 g) received an i.v. bolus dose of the conjugate (33 mg · kg⁻¹, dissolved in approximately 0.5 ml of 5% glucose, n = 4 for each time point) via the dorsal penile vein. A control group of rats received equivalent doses of uncoupled CAP and LZM (n = 4 for each time point). Except for the rats at time point 7.5 min, the rats were allowed to recover from the anesthesia and were placed back into their cages until 5 min before the extirpation of the kidneys. To do so, the rats were anesthetized, a blood sample was taken from the abdominal vena cava, and the rats were perfused blood free via the vena cava with approximately 15 ml of 0.9% NaCl solution, after which the kidneys were removed from the body. The kidney capsule was removed, and the kidneys were briefly stored in liquid nitrogen until homogenization. A 1:5 homogenate in ice-cold potassium phosphate buffer (pH 7.4) was prepared by Turrax homogenization. Aliquots of the homogenates were stored at −80°C before the CAP analysis. Renal ACE activities were determined in the homogenates of rats that had received CAP-LZM (extirpation of kidneys t = 30 min after administration, n = 4) and of rats that had received 500 µl of vehicle (5% glucose, n = 4). The ACE activity analyses were performed within 2 h after homogenization of the kidneys according to a method that has been described in detail elsewhere (Koiter et al., 1998).

**Analysis.** All of the biological samples were analyzed for their total CAP content (the total amount of free CAP and CAP disulfides) by HPLC. For this purpose, the CAP disulfides were converted to the free thiol with the reducing agent TBP. The assay method has been described in detail previously (Kok et al., 1997); this section describes only the details about the sample work-up procedures. The total CAP analysis was performed on sample volumes of 50 µl for plasma samples and 100 µl for urine fractions and kidney homogenates. The samples were mixed with equal volumes of 0.1 N borate buffer, pH 7.4, and TBP solution (1% in methanol); reacted for 30 min, and deproteinized with 3 volumes of methanol (150 or 300 µl, respectively). To discriminate between conjugate-bound CAP and released CAP, an additional analysis was performed on plasma and urine samples that had been deproteinized immediately on collection. The protein-bound CAP was calculated by subtracting the amount of CAP in the deproteinized samples from the total CAP amount.

The urine samples of the rats that had received a bolus dose of 33 mg · kg⁻¹ LZM were also analyzed for the LZM activity in the urine. This analysis was based on the turbidimetric method of Atassi and Habeeb (1969). A 30-µl aliquot of the sample was added to 1 ml of a suspension of M. lysodeicticus in a 0.1 N borate buffer, pH 7.5. The mixture was homogenized, and the decrease in turbidity was measured at 600 nm for 1 min. LZM concentrations were calculated from a calibration curve constructed with native LZM.

**Calculations and Statistics.** Pharmacokinetic analysis of the total CAP plasma disappearance data was performed with the Multifit program (Department of Pharmacokinetics and Drug Delivery, University Center for Pharmacy, Groningen, the Netherlands) using the Simplex algorithm, assuming a constant relative error. The initial plasma clearance, terminal half-life, and volumes of distribution at steady state (Vdss of central and peripheral compartment) were calculated with a two-compartment model, using the combined data of each treatment in one curve fit. The area under the curve of the renal distribution curves was calculated with the Multifit program (trapezoidal rule, up to last data point).
Because freely moving rats were used in this experiment, the animals voided their bladders at different time points; therefore, the averaged excretion curves were calculated by interpolation with the urinary excretion rate at fixed time points.

The statistical significance of differences was tested at a significance level of $p < .05$ (*), $p < .01$ (**), or $p < .001$ (***) using the two-sided Student’s $t$ test.

Results

Figure 3 shows the plasma disappearance curves of the CAP-LZM conjugate and CAP plus LZM. Plasma concentrations were followed during 240 min; after that time point, total CAP concentrations dropped below the detection limit of the applied analytical method (25 ng $\cdot$ ml$^{-1}$). In the group of rats that had received CAP plus LZM, the relative amount of protein-bound CAP ranged from $24 \pm 3\%$ at $t = 5$ min to $53 \pm 4\%$ at $t = 240$ min. No unbound CAP could be detected in the plasma samples of the rats that had received CAP-LZM conjugate.

The calculated kinetic parameters of the plasma disappearance curves are shown in Table 1.

Figure 4 shows the concentration of total CAP that was measured in the kidney. To estimate the amount of CAP that had been accumulated in the kidney, we calculated the area under the curve of the renal distribution curves. These values averaged 97 and 580 (percent dose-minute) for CAP and the CAP-LZM conjugate, respectively.

The excretion of CAP in the urine was followed up to 10 h (Fig. 5). After administration of free CAP plus LZM, only non-protein-bound CAP was excreted in the urine. In contrast, the administration of CAP-LZM resulted in excretion of both protein-bound and non-protein-bound CAP disulfides into the urine. After the administration of free CAP plus LZM, most of the excreted CAP was recovered in the first 2 h after administration. The CAP-LZM conjugate resulted in a sustained excretion of non-protein-bound CAP into the urine with a constant excretion rate of 4.5% $\cdot$ h$^{-1}$. The protein-bound CAP was excreted during the first 5 h after administration (Fig. 6). Figure 6 shows that the urinary excretion of protein-bound CAP was similar to the excretion of intact LZM in the rats that had received an equivalent dose of native LZM. Table 2 gives a summary of the data on the cumulative urinary excretion of the CAP-LZM conjugate and the unconjugated CAP and LZM.

To test whether the accumulation of CAP in the kidney would lead to renal ACE inhibition, we performed a pilot experiment in which we measured the renal tissue ACE activity. At 30 min after administration of the CAP-LZM conjugate, the renal ACE activity was significantly lower than the renal ACE activity of control rats (Fig. 7).

Discussion

The aim of the present study was to increase the renal selectivity of the ACE inhibitor CAP. This can be accomplished either by increasing the amount of drug that accumulates in the kidney or by preventing the extrarenal effects of CAP. Our results show that the CAP-LZM conjugate influences the renal selectivity via both strategies.

The free thiol group of CAP is used for the conjugation of CAP to LZM, but it is also essential for its ACE-inhibiting properties (Ondetti and Cushman, 1981). Therefore, the

![Fig. 3. Plasma disappearance of CAP-LZM and CAP plus LZM. CAP concentrations were determined as total CAP (CAP-SH + CAP-disulfides); all values are presented as the percentage of the injected dose (mean ± S.E.M.). ○, CAP-LZM conjugate ($n = 4$). ●, CAP plus LZM ($n = 6$). The difference between the curves is significant at $p < .001$ at 2, 5, 15, and 30 min and at $p < .01$ at 120 min.]

![Fig. 4. Renal concentrations of CAP. CAP concentrations were determined as total CAP (CAP-SH + CAP-disulfides) and are presented as the percentage of the injected dose per total weight of the two kidneys (mean ± S.E.M.). ○, CAP-LZM conjugate ($n = 4$ per point of time). ●, CAP plus LZM ($n = 4$ per point of time).]

![Table 1. Pharmacokinetic parameters derived from the total CAP plasma data](image)

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Unit</th>
<th>CAP</th>
<th>CAP-LZM</th>
</tr>
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<tbody>
<tr>
<td>AUC</td>
<td>min $\cdot$ % $\cdot$ ml$^{-1}$</td>
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<td>$t_{1/2}$</td>
<td>min</td>
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<td>52.8</td>
</tr>
<tr>
<td>$Cl_{ur}$</td>
<td>ml $\cdot$ min$^{-1}$</td>
<td>3.58</td>
<td>0.74</td>
</tr>
<tr>
<td>$V_{dss}$ (central)</td>
<td>ml</td>
<td>308.1</td>
<td>43.4</td>
</tr>
<tr>
<td>$V_{dss}$ (peripheral)</td>
<td>ml</td>
<td>503.4</td>
<td>62.5</td>
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</table>

The area under the curve (AUC) was calculated up to the last data point with the trapezoidal rule.

$Cl_{ur}$: terminal half-life; $Cl_{ur}$, initial plasma clearance; $V_{dss}$ (central) and $V_{dss}$ (peripheral), volumes of distribution at steady state of the central and peripheral compartment, respectively.
The synthesized CAP-LZM conjugate is an inactive prodrug. Previous experiments have shown that the parent drug can be released by incubation of the conjugate with reduced glutathione (R. J. Kok et al., submitted). Because LZM accumulates rapidly in the kidney and high levels of glutathione are present in the proximal tubular cells but not in the plasma, we anticipated that the release of CAP should predominantly take place in the kidney. This hypothesis is confirmed by the observation that only protein-bound CAP was found in the plasma after administration of the conjugate. Consequently, the total CAP plasma concentration reflects the sum of unchanged CAP and the CAP-containing disulfides and that the calculated parameters are the result of the combined plasma disappearance of these compounds. Because the disulfide metabolite CAP-albumin is cleared more slowly than low-molecular-weight CAP-disulfides, the relative amount of this metabolite in plasma increased in time after a single i.v. injection of free CAP. The excretion of non-protein-bound CAP was measured for 10 h. The excretion of protein-bound CAP and LZM was measured for 5 h. All values are expressed as mean ± S.E.M.

<table>
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<tr>
<th>Cumulative Excretion</th>
<th>CAP</th>
<th>LZM</th>
<th>CAP-LZM</th>
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<tr>
<td>Non-protein-bound CAP</td>
<td>84 ± 1.3</td>
<td>33 ± 3</td>
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</tr>
<tr>
<td>Protein-bound CAP</td>
<td>0</td>
<td>30 ± 4</td>
<td></td>
</tr>
<tr>
<td>LZM</td>
<td>33 ± 2</td>
<td>39 ± 3</td>
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**Fig. 5.** Cumulative excretion of CAP disulfides in the urine. CAP concentrations were determined as total CAP (CAP-SH + CAP-disulfides); all values are expressed as mean ± S.E.M. (n = 6). ○, Total CAP excretion after administration of CAP plus LZM (n = 6). □, Dotted line, total CAP excretion after administration of CAP-LZM conjugate (n = 10); □, solid line, excretion of non-protein-bound CAP after administration of CAP-LZM conjugate (n = 10).

**Fig. 6.** Cumulative excretion of protein-bound CAP and of LZM in the urine. The protein-bound CAP excretion was calculated from the total and non-protein-bound CAP excretion; the LZM excretion was determined with an LZM activity assay. □, Excretion of protein-bound CAP after CAP-LZM administration (n = 10). ▲, Excretion of LZM after LZM administration (n = 6).

**Fig. 7.** Renal tissue ACE activity. ACE activities were measured in kidney homogenates of rats that had received i.v. bolus injections of vehicle (5% glucose, control rats) or CAP-LZM 30 min before the extirpation of the kidneys. Open column, control rats. Closed column, CAP-LZM. Values are expressed as mean ± S.E.M. (n = 4).
As mentioned before, an increase in the renal selectivity of CAP could be obtained by increasing the amount of drug that accumulates in the kidney. Because the amount of total CAP that has distributed to the kidney was increased 6-fold by administering CAP as the CAP-LZM conjugate, we succeeded in delivering more CAP to the kidney. The accumulated conjugate can be degraded and activated in vivo to free CAP, as can be concluded from the observed ACE inhibition in the kidney homogenates. Experiments in which the renal and extrarenal ACE inhibitions of the CAP-LZM conjugate are compared with those of free CAP are ongoing and must reveal whether the administration of the conjugate results in a more renal-selective ACE inhibition. Furthermore, the comparison of the effects of the conjugate and free CAP on blood pressure and renal physiological parameters such as sodium excretion and renal hemodynamics must corroborate the present pharmacokinetic data.

After glomerular filtration, the CAP-LZM conjugate can be either reabsorbed by the proximal tubular cells or excreted into the urine. The tubular reabsorption process of LMWPs is a receptor-mediated endocytotic process with a low affinity and a high capacity (Maack et al., 1979). Our data show that one third of the injected doses of both CAP-LZM and native LZM is not reabsorbed in the kidney. We conclude that the protein-bound CAP in the urine represents conjugate that has been filtered but not reabsorbed in the kidney and that this partial loss of targeted CAP is a consequence of the protein that is used as drug carrier in this study.

The administration of CAP-LZM results in the excretion of non-protein-bound CAP in the urine with a slow and relatively constant excretion rate. This excretion rate should reflect the degradation of the conjugate and subsequent clearance of CAP from the kidney. In contrast, the excretion of non-protein-bound CAP in the urine after administration of free CAP reflects the plasma clearance of this drug. The recovery of total CAP in the urine is in accordance with the recovery of CAP in other studies in the rat (Park et al., 1982; Matsumoto et al., 1986).

In conclusion, this pharmacokinetic study demonstrates that the renal delivery of CAP with the prodrug CAP-LZM resulted in an increased renal selectivity by both a reduced extrarenal distribution and an increased renal accumulation of the drug. We conclude that the delivery of CAP to the kidney has been achieved successfully.

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References


Send reprint requests to: Dr. R. J. Kok, Groningen Utrecht Institute for Drug Exploration, Department of Pharmacokinetics and Drug Delivery, University Center for Pharmacy, A. Deusinglaan 1, 9713 AV Groningen, the Netherlands. E-mail: R.J.Kok@farm.rug.nl