OPC-41061, a Highly Potent Human Vasopressin V₂-Receptor Antagonist: Pharmacological Profile and Aquaretic Effect by Single and Multiple Oral Dosing in Rats¹

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ABSTRACT

The pharmacological profile and the acute and chronic aquaretic effects of OPC-41061, a novel nonpeptide human arginine vasopressin (AVP) V₂-receptor antagonist, were respectively characterized in HeLa cells expressing cloned human AVP receptors and in conscious male rats. OPC-41061 antagonized [³H]-AVP binding to human V₂-receptors (Ki = 0.43 ± 0.06 nM) more potently than AVP (Ki = 0.78 ± 0.08 nM) or OPC-31260 (Ki = 9.42 ± 0.90 nM). OPC-41061 also inhibited [³H]-AVP binding to human V₁a-receptors (Ki = 12.3 ± 0.8 nM) but not to human V₁b-receptors, indicating that OPC-41061 was 29 times more selective for V₂-receptors than for V₁a-receptors. OPC-41061 inhibited cAMP production induced by AVP with no intrinsic agonist activity. In rats, OPC-41061 inhibited [³H]-AVP binding to V₁a-receptors (Ki = 325 ± 41 nM) and V₂-receptors (Ki = 1.33 ± 0.30 nM), showing higher receptor selectivity (V₁a/V₂ = 244) than with human receptors. A single oral administration of OPC-41061 in rats clearly produced dose-dependent aquareesis. In treatment by multiple OPC-41061 dosing for 28 days at 1 and 10 mg/kg p.o. in rats, significant aquaretic effects were seen throughout the study period. As the result of aquareesis, hemoco ncentration was seen at 4 hr postdosing although, no differences were seen in serum osmolality, sodium, creatinine and urea nitrogen concentrations at 24 hr postdosing. Furthermore, there was no difference in serum AVP concentration, pituitary AVP content or the number and affinity of AVP receptors in the kidney and liver at trough throughout the study period. These results demonstrate that OPC-41061 is a highly potent human AVP V₂-receptor antagonist and produces clear aquareesis after single and multiple dosing, suggesting the usefulness in the treatment of various water retaining states.

The regulation of body fluid volume is important in almost every area of medicine. AVP is well known to play a major role in water metabolism by inducing water reabsorption at the renal collecting duct through stimulation via V₂ receptors. In various water retaining conditions, such as advanced cardiac failure (Cas et al., 1995; Goldsmith et al., 1983; Szatalowicz et al., 1981), cirrhosis (Bichet et al., 1982), nephrotic syndrome and syndrome of inappropriate ADH secretion (Barter and Schwartz, 1967; Zerbe et al., 1980), the kidneys fail to excrete the amount of water ingested, causing water retention and leading to an excessive water state. Thus, water diuretics (aquaretics) have exciting therapeutic implications in the management of patients with water excess and consequent dilutional hyponatremia (Schrier and Niederberger, 1993; Verbalis, 1993).

Although several potent peptide vasopressin antagonists are currently available (Laszlo et al., 1991; Manning and Sawyer, 1989), none has emerged as a clinically useful antidiuretic antagonist because of their low oral bioavailability, species differences and especially, antidiuretic (agonistic) effects when tested in humans (Albrightson-Winslow et al., 1989; Allison et al., 1988; Brooks et al., 1988; Mah and Hofbauer, 1988). Because two nonpeptide, orally effective AVP antagonists, OPC-21268 and OPC-31260, were respectively reported in 1991 and 1992 (Yamamura et al., ), there has been great interest in AVP research and clinical use, encouraging

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ABBREVIATIONS: AVP, arginine vasopressin; cAMP, cyclic adenosine monophosphate; BSA, bovine serum albumin; DMEM, Dulbecco’s modified Eagle’s medium; HeLa, human endocervical carcinoma cells; V₂-HeLa cells, HeLa cells expressing human AVP V₂-receptors; V₁a-HeLa cells, HeLa cells expressing human AVP V₁a-receptors; V₁b-HeLa cells, HeLa cells expressing human AVP V₁b-receptors; ANOVA, analysis of variance; CHF, congestive heart failure.
the development of various nonpeptide AVP antagonists (Freidinger and Pettibone, 1997; Serradeil-Le et al., 1993, 1996; Tahara et al., 1997) and some of them have been clinically developed in patient.

The AVP receptor subtypes originally proposed by Michell et al. (1979) were based on primary intracellular signaling mechanisms: cyclic AMP-independent (V1) and cyclic AMP-dependent (V2) pathways. Recently, cDNAs encoding the AVP receptors have been cloned and their primary structure identified. AVP receptors were further classified into at least three subtypes: V1a- (Morel et al., 1992; Thibonnier et al., 1994), V1b- (De Keyzer et al., 1984; Sugimoto et al., 1984) and V2-receptors (Birnbaumer et al., 1992; Lollard et al., 1992).

Species differences between humans and rats have been reported with peptide AVP antagonists to V2-receptors and with nonpeptide antagonists to V1a-receptors (Liu et al., 1994; Hirasawa et al., 1994; Pettibone et al., 1992). In these previous reports, crude membrane preparations or V1a-receptors expressed only transiently in COS-7 cells were used to determine the species differences. We designed the stable expression of three cloned human AVP receptors in HeLa cells and attempted to develop more potent nonpeptide antagonists to human AVP V2-receptors. OPC-41061, 7-chloro-5-hydroxy-1-[2-methyl-4-(2-methylbenzylamino)benzoyl]-2,3,4,5-tetrahydro-1H-1-benzazepine, was selected as the most potent human V2-receptor antagonist through a series of structural conversions of OPC-31260 (fig. 1). In this study, we determined the pharmacological profile of OPC-41061 and its antagonistic action on human AVP receptors using intact HeLa cells stably expressing each subtype of human AVP receptors and on rat AVP receptors using plasma membrane preparations.

Furthermore, aquaretics may be well-suited for chronic therapy devoid of the well-known complications of conventional diuretics (saluretics) (e.g., hyponatremia or hypokalemia), and aquaretics may provide the optimum diuretic ther-

Materials and Methods

Materials. OPC-41061 and OPC-31260 were synthesized by the Tokushima 2nd Factory of Otsuka Pharmaceutical Co., Ltd. (Tokushima, Japan). AVP was purchased from Peptide Institute Inc., Japan, lipofectamine and fetal calf serum from Gibco BRL., penicillin, streptomycin and BSA from Sigma, DMEM from Nissui Pharmaceutical Co., [3H]-AVP from New England Nuclear, AVP RIA kit from Mitsubishi Petrochemical Co., and G418 (Geneticin disulfate) and other chemicals from Wako Pure Chemicals, Japan.

Animals. Male Sprague-Dawley rats (SD rats) were purchased from Charles River Japan, Inc., at 6 or 7 wk of age and housed during the experiment in an air-conditioned (temperature-, humidity- and light-controlled) animal room with free access to food and water. All experiments were performed under the regulations of the Guideline for Animal Experimentation in Otsuka Pharmaceutical Co., Ltd.

Preparation of HeLa cells expressing human AVP receptors. The cDNAs for human vasopressin V1 receptor was cloned in our laboratory (Hirasawa et al., 1994). For V1a- and V2-receptors, cDNA synthesized from human pituitary and kidney RNA was amplified the full length of coding region by using primers for V1a- (Sugimoto et al., 1994) and V2- (Birnbaumer et al., 1992) receptors. Cloned cDNAs and PCR products were subcloned into the Bluescript KS II (+) plasmid vector, La Jolla, CA. Nucleotide sequence analysis was performed with the ABI 373A DNA Sequencer (Applied Biosystems, Inc., Foster City, CA) for both complete strands. We confirmed that the clones obtained were identical to the previously reported human V1a- and V2-receptor cDNA by sequencing, respectively. These cDNA fragments were ligated in SmaI promoter based mammalian expression vector pMe18S and resulting constructs were used for transfection.

HeLa cells, grown in 5% CO2 at 37°C, were plated at a density of 1 × 106 cells/10-cm plate in 10 ml of DMEM supplemented with 10% fetal calf serum and penicillin/streptomycin, and allowed to attach overnight. Each expression vector for vasopressin receptors and pSV2neo vector (Clontech), which contained neomycin resistance gene for selection of stable transformants, were cotransfected into HeLa cells by lipofectamine method (Hawley-Nelson et al., 1989).

After overnight transfection, cells were fed with fresh medium, allowed to grow for 2 days, adjusted to a density of 1 × 106 cells/plate, and incubated for an additional 24 hr. Cells were continually selected for 3 to 4 wk in medium containing 400 to 800 µg/ml of the antibiotic G418. Single colonies were then isolated, expanded and harvested for radioligand binding assays to measure expression of receptors. Cells were maintained in the medium containing 200 µg/ml of G418. The medium was changed every 3 days, and the cells were subcultured after trypsinization.

Preparation of rat liver and kidney plasma membranes. Liver and kidney plasma membranes were prepared from SD rats (300–400 g, Charles River, Yokohama, Japan). Details of the methods used have been published previously (Yamamura et al., 1991, 1992). Protein concentration was measured by the method of Bradford using BSA as a standard.

Radioligand binding assay to HeLa cells. After reaching confluence in 12-well (V1a-, V1b-HeLa) or 24-well (V2-HeLa) dishes, the cells were washed twice with ice-cold PBS and incubated for 2 hr with [3H]-AVP at 4°C in DMEM medium adjusted to pH 7.4 using 10 mM HEPES-NaOH containing 0.3% BSA. For the competition experiment, OPC-41061 and OPC-31260 were dissolved in dimethyl sulfoxide, diluted with DMEM medium and added into the wells at several appropriate concentrations. After 2 hr of incubation, the cells were rinsed twice with ice-cold phosphate-buffered saline, lysed in 500 (12-well) or 250 (24-well) µl of 0.1 N NaOH containing 0.1% sodium dodecyl sulfate, transferred into scintillation vials, combined with 5 ml of Aquazol II (NEN, Boston, MA) and detected using a liquid scintillation counter (LSC-1050, Aloka, Tokyo, Japan). The protein content of the lysed cells was determined by the method of Bradford.

Radioligand binding assay to rat kidney and liver plasma membranes. Binding assay to rat kidney and liver plasma membranes was performed as described previously (Yamamura et al., 1991, 1992).

cAMP production in V2-HeLa by AVP. After reaching confluence in 24-well dishes, the V2-HeLa cells were washed twice with ice-cold PBS and incubated for 10 min with AVP at 1 nM in the presence or absence of OPC-41061 at 37°C in DMEM medium adjusted to pH 7.4 using 10 mM HEPES-NaOH containing 1 mM 3-isobutyl-1-methylxanthine and 0.3% BSA. For the competition-ex
periment, OPC-41061 and OPC-31260 were dissolved in DMSO, diluted with DMEM medium and added into the wells at several appropriate concentrations. After 10 min of incubation, the medium was evacuated and the cells were rinsed once with ice-cold PBS. cAMP was extracted from the cells using 250 μl of 0.1 N HCl solution and stored at −20°C until determination using a radioimmunoassay kit (Yamasa, Tokyo, Japan).

Single-dosing experiments in rats. OPC-41061 suspended in 1% hydroxymethyl cellulose or the solvent was administered orally at doses of 0.3 to 10 mg/kg to rats at 8 wk of age (body weight: 280–340 g) by a gastric tube. Seven animals in each dosing group were then placed individually in metabolic cages and spontaneously voided urine was collected for the periods of 0 to 2, 2 to 4, 4 to 6, 6 to 8 and 8 to 24 hr. Another five animals in each group were decapitated at 4 hr postdosing and trunk blood was collected. After centrifugation at 3000 rpm for 10 min, the serum was obtained and frozen until use. Osmolality was determined by freezing point depression using a Fiske osmometer (Model 3400, Boston, MA). Electrolytes were measured by the ion-electrode method (CX-3, Beckman Instruments, Fullerton, CA), and creatinine and urea nitrogen were measured using a COBAS autoanalyzer (COBAS FARAII, Roche, Basel, Switzerland). AVP was measured by radioimmunoassay using an AVP-RIA kit (Mitsubishi Petrochemical Co. Ltd., Tokyo, Japan).

Multiple-dosing experiments in rats. Male SD rats at 7 wk of age (body weight: about 260 g) were divided into two groups. One group (group A) were periodically examined for aquaretic effect and the other (group B) were decapitated for determination of AVP-receptors and AVP content. Six animals were used in each group. OPC-41061 was suspended in 1% hydroxyethyl cellulose solution and orally administered at doses of 1 and 10 mg/kg once daily at around 9:30 AM. The rats in group A were placed individually in metabolic cages and spontaneously voided urine was periodically (day –1, 1, 3, 7, 10, 14, 21 and 28) collected for the periods of 0 to 4 and 4 to 24 hr. The rats in group B were decapitated at 24 hr postdosing on days 7, 14 and 21 and those in group A were decapitated at 24 hr postdosing on day 28. In the control group, rats were decapitated at predosing (day 1) to obtain basal values. The kidney and liver (for analysis of AVP receptors) and the pituitary (for determination of AVP content) were quickly removed and immediately cooled with ice-cold saline. The removed pituitary was weighed and then stored frozen at −80°C until extraction.

Kidney and liver plasma membranes of each rat were prepared as described above. The Kd and Bmax were determined by Scatchard analysis of [3H]-AVP saturation binding. The pituitary was combined at doses of 1% hydroxyethyl cellulose and 0.1 N HCl solution for 30 sec using a sonicator (Microson, output: 6), and then centrifuged. The supernatant was diluted 500- to 5000-fold with saline containing 0.1% BSA. AVP content was determined by radioimmunoassay using an AVP-RIA kit.

Binding data analysis. The dissociation constant (Kd) and the number of binding sites (Bmax) were determined by Scatchard analysis of [3H]-AVP saturation binding. The pituitary was combined at each concentration for the optimum number of receptors for [3H]-AVP binding assay (Yamasa, Tokyo, Japan).

Table 1. Characterization of human AVP receptors expressed on cultured HeLa cells for [3H]-AVP binding

<table>
<thead>
<tr>
<th>AVP (logM)</th>
<th>V0</th>
<th>V1a</th>
<th>V1b</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.2 M</td>
<td>1.09 ± 0.05</td>
<td>2.29 ± 0.11</td>
<td>1.68 ± 0.06</td>
</tr>
<tr>
<td>Bmax</td>
<td>1021 ± 98</td>
<td>519 ± 45</td>
<td>69 ± 5.7</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± S.E.M. of at least four separate determinations performed in duplicate.

Results

Antagonistic affinities of OPC-41061 for AVP receptors. As shown in table 1, HeLa cells transfected with each subtype of human AVP receptors constantly expressed a sufficient number of receptors for [3H]-AVP binding and these data were from one typical experiment of the three performed.

![Fig. 2. Effects of AVP on cyclic AMP production (A) and inhibitory effect of OPC-41061 on cAMP generation by AVP (B) in V2-HeLa cells](image-url)
ANTAGONISTIC POTENCY OF OPC-41061 AND OPC-31260 FOR HUMAN AND RAT AVP RECEPTORS

These data clearly show that OPC-41061 possesses a more potent affinity for human V2-receptors than native AVP and that OPC-41061 can clearly antagonize V2-receptors with no intrinsic agonistic activity.

Aquaretic effect by single-dosing in conscious rats. A single oral administration of OPC-41061 increased urine volume and decreased urine osmolality in a dose-dependent manner at doses of 0.3 to 10 mg/kg in normally hydrated conscious rats (fig. 4). The maximum urine output for 2 hr postdosing was 18.0 ± 2.6 ml, which was 12 times higher than the control, and urine osmolality reached a minimum of 175 ± 15 mOsm/kg (vs. 714 ± 136 mOsm/kg for the control). No significant increase in urine output was seen after 4 hr postdosing (data not shown). Urinary Na excretion during the 4 hr postdosing increased dose-dependently. However, the magnitude of the increase was considerably smaller than that by natriuretic agents such as furosemide [data shown previously (Yamamura et al., 1992)].

At 4 hr postdosing, serum osmolality and Na and creatinine concentrations were significantly elevated presumably by the decreased body fluid resulting from the aquarexia induced by OPC-41061 (table 3). Serum AVP concentration also increased dose-dependently after the elevation of serum osmolality. In spite of the hemoconcentrated status at 4 hr postdosing, blood urea nitrogen was dose-dependently decreased presumably by OPC-41061 inhibition of urea reabsorption at the terminal of the inner medullary collecting duct, leading to an increase in urinary urea excretion.

Aquaretic effect by multiple-dosing in conscious rats. OPC-41061 was orally administered once daily at 1 and 10 mg/kg starting from day 1 to day 7 in normally hydrated conscious rats. OPC-41061 was administered orally in 1% hydroxymethyl cellulose solution. Values are expressed as the mean ± S.E.M. of at least four separate determinations performed in duplicate.

Changes in urine volume and osmolality were analyzed by linear regression analysis and significance level was confirmed. Differences vs. the control group were then analyzed by t test using a closed testing procedure (Marcus et al., 1976; Hothorn and Lehmacher, 1991). * P < .05, ** P < .01.

Although serum osmolality and Na, blood urea nitrogen and AVP concentrations were significantly changed at 4 hr postdosing (table 3), there were no differences in the net urinary excretion of creatinine between the OPC-41061 groups and the control group (10 mg/kg: 41.8 ± 1.1 mg/kg/24 hr, 1 mg/kg: 39.4 ± 1.7 mg/kg/24 hr, control: 42.2 ± 1.4 mg/kg/24 hr at day 28), indicating that OPC-41061 did not affect glomerular filtration rate. Urinary AVP excretion was significantly increased in the urine collected during 0–4 hr and 0–24 hr postdosing and remained almost constant throughout the study. It is important to note that, in addition to the constant change in urine osmolality, the constant AVP excretion during the study period further support the conclusion that repeated administration did not alter the aquaretic effect of OPC-41061.

Table 2

Antagonistic potency of OPC-41061 and OPC-31260 for human and rat AVP receptors

<table>
<thead>
<tr>
<th></th>
<th>Human</th>
<th>Rat</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$K_i$ (nM)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>$V_{1a}$</td>
<td>$V_{1b}$</td>
</tr>
<tr>
<td>AVP</td>
<td>0.78 ± 0.08</td>
<td>0.84 ± 0.08</td>
</tr>
<tr>
<td>OPC-41061</td>
<td>0.43 ± 0.06</td>
<td>12.3 ± 0.8</td>
</tr>
<tr>
<td>OPC-31260</td>
<td>9.42 ± 0.90</td>
<td>150 ± 15</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± S.E.M. of at least four separate determinations performed in duplicate.
fmol/mg for control. The dissociation constant \( K_d \) of control, 1 and 10 mg/kg at day 7 was 1.94 \( \pm \) 0.19, 1.73 \( \pm \) 0.30 and 1.70 \( \pm \) 0.42 nM and at day 28 was 1.10 \( \pm \) 0.05, 1.22 \( \pm \) 0.13 and 1.23 \( \pm \) 0.10 nM, respectively. The \( B_{\text{max}} \) and \( K_d \) values of \( V_2 \)-receptors (liver) also did not change at trough throughout the experiment. Furthermore, pituitary AVP content was constant throughout the experiment (data not shown).

These results suggested that although oral administration of OPC-41061 increased AVP secretion from the pituitary, the secretion recovered to the basal level by 24 hr postdosing. In contrast, serum renin activity was gradually decreased and serum aldosterone concentration was significantly lowered through repeated administration of OPC-41061 for 28 days (table 5).

In conclusion, orally administered OPC-41061 is a powerful aquaretic, and the aquaretic effect was unchanged during repeated administration for 4 wk.

**Fig. 5.** Effects of multiple dosing of OPC-41061 on body weight in male SD rats. Values are expressed as the mean \( \pm \) S.E.M. \((n = 6)\). Differences between the OPC-41061 groups and the control group were analyzed by one-way ANOVA followed by two-tailed Dunnett’s test \(* P < .05, *^{*} P < .01\ vs. \ the \ control\). Differences in serum AVP levels were not statistically analyzed, because the control value was below the detection limit.

### Discussion

Aquaretics have exciting therapeutic implications in the management of patients with water excess and consequent dilutional hyponatraemia, as in patients with congestive heart failure and cirrhosis, or in patients with euvoletic hyponatraemia, as in patients with syndrome of inappropriate ADH secretion. There is an obvious need for a potent \( V_2 \)-receptor antagonist that can be safely administered orally over the long term in a clinical setting. Previous reports of chronic blockade of vasopressin receptors by a peptide \( V_2 \)-antagonist did not show persistent aquareisia (Hofbauer et al., 1986; Mah et al., 1988). A peptide \( V_2 \)-antagonist, \( d-(CH_2)_3-D-Tyr^{(CH_2)}-r(Et)VAVP \), increased urine volume and decreased urine osmolality after i.v. or s.c. administration in normally hydrated or dehydrated Sprague-Dawley rats in acute experiments. However, by chronic administration by i.v. infusion or repeated s.c. dosing, water excretion and intake increased markedly on the first day but then subsequently reverted to normal, although the marked initial water loss was fully compensated for by an increased water intake. The lack of a chronic effect with peptide antagonists is thought to be due to the activation of endogenous compensatory mechanisms such as hormones, prostaglandins and renal and systemic hemodynamics. Alternatively, the lack of effect may be due to an intrinsic agonism which might have contributed to the normalization of water balance by limiting the maximum antidiuretic effects of renal tubular AVP receptors.

OPC-41061 did not increase cyclic-AMP production in \( V_2 \)-HeLa cells and did not show any antidiuretic effect in water-loaded, alcohol-anesthetized rats (data not shown), indicating that OPC-41061 possesses no agonistic activity for \( V_2 \)-receptors. Therefore, OPC-41061 is expected to produce a chronic aquaretic effect during multiple dosing. In this report, we examined changes not only in the aquaretic effect of

### Table 3

**Effect of single oral administration of OPC-41061 on serum parameters at 4 hr postdosing in normally hydrated male SD rats**

<table>
<thead>
<tr>
<th>Dose (mg/kg)</th>
<th>Osmolality (mOsm/kg)</th>
<th>Na (mEq/l)</th>
<th>BUN (mg/dl)</th>
<th>Creatinine (mg/dl)</th>
<th>AVP (pg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>293 ± 2</td>
<td>145 ± 0</td>
<td>17.1 ± 0.7</td>
<td>0.58 ± 0.01</td>
<td>&lt;0.2</td>
</tr>
<tr>
<td>1 mg/kg</td>
<td>294 ± 1</td>
<td>147 ± 1</td>
<td>14.7 ± 0.6a</td>
<td>0.60 ± 0.02</td>
<td>0.4 ± 0.1</td>
</tr>
<tr>
<td>3 mg/kg</td>
<td>294 ± 2</td>
<td>148 ± 1a</td>
<td>12.0 ± 0.4a</td>
<td>0.58 ± 0.01</td>
<td>1.2 ± 0.2</td>
</tr>
<tr>
<td>10 mg/kg</td>
<td>301 ± 3</td>
<td>152 ± 1</td>
<td>12.6 ± 0.5a</td>
<td>0.71 ± 0.02b</td>
<td>4.4 ± 0.9</td>
</tr>
</tbody>
</table>

Values are expressed as the mean \( \pm \) S.E.M. \((n = 5)\). Differences between the OPC-41061 groups and the control group were analyzed by one-way ANOVA followed by two-tailed Dunnett’s test \(* = P < .05, *^{*} = P < .01\ vs. \ the \ control\). Differences in serum AVP levels were not statistically analyzed, because the control value was below the detection limit.
TABLE 4

<table>
<thead>
<tr>
<th>Day</th>
<th>Control</th>
<th>1 mg</th>
<th>2 mg</th>
<th>4 mg</th>
<th>10 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>395 ± 1</td>
<td>307 ± 2</td>
<td>304 ± 1</td>
<td>300 ± 1</td>
<td>306 ± 3</td>
</tr>
<tr>
<td>2</td>
<td>308 ± 0</td>
<td>307 ± 2</td>
<td>307 ± 2</td>
<td>308 ± 0</td>
<td>306 ± 1</td>
</tr>
<tr>
<td>4</td>
<td>305 ± 0</td>
<td>307 ± 2</td>
<td>307 ± 2</td>
<td>298 ± 0</td>
<td>306 ± 2</td>
</tr>
<tr>
<td>6</td>
<td>311 ± 3</td>
<td>307 ± 2</td>
<td>307 ± 2</td>
<td>308 ± 0</td>
<td>306 ± 2</td>
</tr>
<tr>
<td>8</td>
<td>311 ± 3</td>
<td>307 ± 2</td>
<td>307 ± 2</td>
<td>308 ± 0</td>
<td>306 ± 2</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± S.E.M. (n = 6). Differences between the OPC-41061 groups and the control group were analyzed by ANOVA based on repeated measurements and any significant differences were then analyzed by two-tailed Dunnett’s multiple-comparison test at each measurement day. *P < 0.05 vs. the control.

OPC-41061 but also in the circulating AVP system and targeted organ function by chronic oral administration of OPC-41061 for 28 days in rats. Urine volume did not change throughout the study period in the 1-mg/kg group, but it decreased slightly in the 10-mg/kg group. The slight decrease in urine volume was considered to be the result of activated compensatory mechanisms. There have been similar reports that the natriuresis by chronic hydrochlorothiazide or furosemide administration markedly abated and sodium excretion was returned to the control level in a so-called “braking effect” (Kahn et al., 1983; Walter and Shirley, 1986). However, in our study, urine volume was decreased only slightly in the first few days and did not return to the control level as was seen in the experiments with peptide AVP antagonists and with hydrochlorothiazide and furosemide. Furthermore, urinary osmolality and urinary AVP excretion showed constant change throughout the study period, indicating that the aquaretic effect of OPC-41061 was unchanged during the 28 days of repeated administration. Although OPC-41061 increased the excretion of Na and urea nitrogen in the urine collected during 0 to 4 hr postdosing, serum Na concentration was increased and serum urea nitrogen was decreased significantly at 4 hr postdosing. This discrepancy seems attributable to the fact that hemoconcentration by the increased free water clearance exceeds the Na excretion but not the urea excretion. These changes then returned to the control levels at 24 hr postdosing, because there were no differences in the urine collected during 0 to 24 hr postdosing between the treated and control groups. The excretion of AVP was increased during 0 to 4 hr postdosing and remained almost the same throughout the study period. Serum AVP concentration was increased at 4 hr postdosing and subsequently returned to the control values. There were also no differences in serum AVP concentration, pituitary AVP level, or the number and dissociation constant of AVP receptors in the liver and kidney between the treated and control groups. Although oral administration of OPC-41061 increased AVP secretion from the pituitary and increased circulating AVP for several hours after dosing, both parameters recovered to the basal level by 24 hr postdosing. Because the marked aquarexia by OPC-41061 did not last past 4 hr postdosing, and the initial water loss was fully compensated for by an increasing water intake, there seemed to be no significant changes in the AVP producing and secreting organ and the AVP target receptors. In other circulating hormone systems, serum renin activity tended to be decreased and aldosterone was significantly decreased following by chronic administration. The suppression of renin and aldosterone secretion might be caused by a direct action of the increased AVP on the juxtaglomerular apparatus (Reid et al., 1983) or secondary to an apparent increases in serum sodium after OPC-41061 dosing.

Congestive heart failure is a complex clinical syndrome characterized by a number of neuroendocrine responses. Recent work has established the importance of the renin-angiotensin systems and angiotensin-converting enzyme inhibitors have emerged as distinctly useful drugs in CHF. Therefore, it may be desirable for drugs used in the treatment of CHF to not activate the renin-angiotensin-aldosterone system. Diuretic therapy is a mainstay in the treatment of edematous conditions in CHF. The conventional diuretics (natriuretics) now in use are effective for the treatment of...
emodematous states, but they are associated with activation of the renin-angiotensin-aldosterone systems. Furthermore, natriuretics may cause electrolyte imbalance, such as hypokalemia or hypokalemia. Serum sodium concentration is one of the most powerful predictors of cardiovascular mortality, with hypotension patients showing substantially shorter survival than patients with a normal serum sodium concentration (Lee and Packer, 1986). It is not known whether neurohormonal activation and decreased serum Na level are the results of a more advanced degree of heart failure or whether they contribute directly to the progression of mortality. As a result of the aquarexia demonstrated in this report, natriuretics have a potential medical benefit for the treatment of edematous conditions in CHF by removing excess water from the body without activating the renin-angiotensin-aldosterone system or causing serum electrolytes imbalances.

In conclusion, OPC-41061 was characterized as a highly potent nonpeptide human AVP V$_2$-receptor antagonist with no agonistic properties. The compound’s aquaretic effect by single administration were confirmed by an increase in urine volume and a decrease in urine osmolality followed by an increase in serum sodium concentration. Chronic administration for 28 days also confirmed a sustained aquaretic effect with no change in pituitary AVP content or target AVP receptors. Thus, these results support the contention that OPC-41061 may be therapeutically useful for the treatment of various diseases involving fluid retention. It is also expected that natriuretics will be well-suited for chronic therapy without the well-known side effects of conventional diuretics (saliuretics) on the balance of electrolytes, such as hypokalemia or hypokalemia, and that aquaretics may make possible optimum diuretic therapy in water-retaining states.

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References


TABLE 5

Effects of multiple dosing of OPC-41061 on serum hormone levels and pituitary AVP content at 24 hr post dosing in male SD rats.

<table>
<thead>
<tr>
<th>Day</th>
<th>Control</th>
<th>1 mg</th>
<th>10 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0.3 ± 0.1</td>
<td>0.3 ± 0.2</td>
<td>0.2 ± 0.0</td>
</tr>
<tr>
<td>7</td>
<td>0.3 ± 0.1</td>
<td>0.3 ± 0.0</td>
<td>0.2 ± 0.0</td>
</tr>
<tr>
<td>14</td>
<td>0.4 ± 0.4</td>
<td>0.4 ± 0.4</td>
<td>0.6 ± 0.2</td>
</tr>
<tr>
<td>21</td>
<td>0.6 ± 0.1</td>
<td>0.6 ± 0.1</td>
<td>1.8 ± 1.0</td>
</tr>
<tr>
<td>28</td>
<td>2.1 ± 1.3</td>
<td>0.5 ± 0.1</td>
<td>0.4 ± 0.0</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± S.E.M. (n = 6). Differences between the OPC-41061 groups and the control group were analyzed by two-tailed Dunnett’s multiple-comparison test at each measurement day (*P < .05, **P < .01 vs. the control).


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