Mechanism of Relaxant Effect of Clonidine in Isolated Bovine Tracheal Smooth Muscle

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ABSTRACT

The relaxant effect of clonidine and the possible involvement of imidazoline I₁ receptors in bovine tracheal smooth muscle (BTSM) were examined. Clonidine caused concentration-dependent significant relaxation in BTSM precontracted with 0.1 or 1 μM carbachol (CCh) but not in 72.7 mM KCl-induced contraction. The relaxation in CCh-contracted BTSM was inhibited by yohimbine (10 μM) and idazoxan (10 and 30 μM) but not by tetrodotoxin, indomethacin and other adrenoceptor antagonists. Oxymetazoline (0.1–100 μM) and phenotolamine (0.1–100 μM) caused concentration-dependent relaxation, which was attenuated by idazoxan (10 μM). Norepinephrine (0.1–100 μM) produced concentration-dependent relaxation, which was completely abolished by propranolol (10 μM) but not by yohimbine (1 μM). In fura-PE3/AM-loaded BTSM, CCh and 72.7 mM KCl increased intracellular calcium concentration ([Ca²⁺]ᵢ) followed by contraction. The high K⁺-induced increase in [Ca²⁺]ᵢ, was not affected by clonidine. In CCh-stimulated BTSM, clonidine decreased [Ca²⁺]ᵢ, and muscle force in parallel, whereas verapamil decreased [Ca²⁺]ᵢ, more strongly than muscle force. Clonidine (100 μM) inhibited the transient increase in [Ca²⁺]ᵢ, induced by CCh but not by caffeine (20 mM) in Ca²⁺-free solution. Clonidine did not change the cAMP content in the presence of either 72.7 mM KCl or CCh. These results indicate that clonidine relaxes CCh-stimulated BTSM through the inhibition of CCh-induced increases in Ca²⁺-influx, Ca²⁺-release and intracellular signal transduction probably via imidazoline I₁ receptors.

Clonidine is a centrally acting A₂R agonist that is used in clinical practice for its antihypertensive, sedative and angesic effects. In the lower brain stem region, clonidine suppresses sympathetic outflow, resulting in a decrease in blood pressure when administered i.p., whereas clonidine administered i.v. causes acute hypertension because of systemic vascular constriction mediated by postsynaptic A₂R activation (Kobinger, 1978).

In the airways, clonidine induces contraction in a canine in situ model (Leff and Munoz, 1981). Furthermore, clonidine enhances agonist-induced bronchoconstriction in conscious (Advenier et al., 1983) and anesthetized (Macquin-Mavier et al., 1988) guinea pigs as well as in conscious humans (Dinh Xuan et al., 1988). Furthermore, histamine-induced contraction was augmented by clonidine in isolated guinea pig trachea (Floh and Advenier, 1985). In contrast to these findings, clonidine has been demonstrated to attenuate the contraction evoked by electrical stimulation in isolated smooth muscles of guinea pig (Wikberg et al., 1982), canine (Tsuchiya et al., 1990) and equine (LeBlanc et al., 1993; Yu et al., 1993) airways. Moreover, in vivo studies have revealed antibronchospastic effects of clonidine in vagal-stimulated (Olsson and Ekdahl, 1985; Anderson et al., 1986) and citric acid-challenged guinea pigs (O’Connell et al., 1994). These complicated and, in part, contradictory results have caused difficulty in evaluating the therapeutic efficacy and safety of clonidine in asthmatic patients.

Although there have been a number of clinical trials investigating clonidine’s effects in patients with asthma (reviewed by Dinh Xuan and Lockhart, 1989), several factors, including dosing methods, made the results conflicting. In particular, it is noteworthy that, unlike oral or transdermal administration, only inhaled clonidine was reported to be beneficial to asthmatic patients (Lindgren et al., 1986). Therefore, clonidine is expected to have several direct peripheral anti-spastic or spasmolytic effects in the airways.

The present study was designed to examine the mechanisms of direct action of clonidine in bovine trachealis smooth

ABBREVIATIONS: A₂R, alpha-2 adrenoceptor; BTSM, bovine tracheal smooth muscle; PSS, physiological salt solution; fura-PE3/AM, acetoxymethyl ester of fura-PE3; [Ca²⁺]ᵢ, intracellular calcium concentration; Kᵓ or Kᵓ, dissociation constant; EGTA, ethylene glycol bis-N,N,N',N'-tetraacetic acid; CCh, carbamylcholine chloride (carbachol); WB 4101, 2-[2,6-dimethoxyphenoxo-ethyl]aminomethyl]-1,4-benzodioxane hydrochloride; TTX, tetrodotoxin; I₁R, imidazoline I₁ receptor.
muscle (BTSM) contracted with muscarinic stimulation and to support its possible therapeutic use in patients with asthma.

Materials and Methods

Tissue preparation. Freshly excised bovine tracheae were obtained from a local abattoir. After cutting free of the cartilage rings, epithelium, fat and connective tissues were carefully removed from the smooth muscle tissues in PSS under a microscope. The smooth muscles were cut into small strips and suspended in PSS maintained at 37°C.

Measurement of mechanical activity. Muscle contraction was recorded isometrically. One end of each muscle was attached by cotton thread to a force displacement transducer (Orientec, Tokyo, Japan), and the other end was tied to a glass holder situated parallel to the tissues with cotton thread under a resting force of 5 mN in a 20-ml tissue bath. After equilibration for 40 to 60 min until passive tension stabilized, high-K⁺ solution (72.7 mM) was repeatedly applied.

Measurement of acetoxymethyl ester of fura-PE3 (fura-PE3/AM) fluorescence. [Ca²⁺]ᵢ was measured simultaneously with muscle contraction as reported previously (Ozaki et al., 1987; Sato et al., 1988). Fura-PE3/AM was added to PSS to make a final concentration of 5 mM, together with noncytotoxic detergent, 0.02% cremophor EL. Muscle strips were loaded with fura-PE3/AM for more than 4 hr at room temperature. After the fura-PE3/AM-loading, muscle strips were washed with PSS in a tissue bath at 37°C for 30 to 40 min to remove the uncleaved fura-PE3/AM. One end of the muscle strip was connected to a force displacement transducer to monitor the mechanical activity. The muscle strips were illuminated alternately (48 Hz) at excitation wavelengths (340 ± 10 nm and 380 ± 10 nm), and the amount of 500 ± 20-nm fluorescence induced by 340-nm excitation (F340) and that induced by 380-nm excitation (F380) were measured using a fluorimeter (CAP-110, Japan Spectroscopic). The absolute Ca²⁺ concentration was not calculated in the present experiment because the dissociation constant ($K_d$) of the fluorescent indicator for Ca²⁺ in cytosol may be different from that obtained in vitro (Karaki, 1989). After equilibration of the force and ratio for 40 to 60 min, high-K⁺ solution (72.7 mM) was repeatedly applied until the response became stable. The ratios obtained in resting state and in high-K⁺ stimulation were taken as 0% and 100%, respectively. In these preparations, approximately 150 min was allowed for measurement because of the leakage of fura-PE3/AM. Therefore, application trials of the antagonists were limited to three times at most in order to obtain reliable data. For the same reason, the effects of drugs in Ca²⁺-free solution were observed immediately (approximately 1 min) after the ratio returned to base line by exchanging the high-K⁺ solution for the Ca²⁺-free solution.

Measurement of adenosine 3’:5’-cyclic monophosphate (cAMP) content. After preincubation in PSS aerated with 95% O₂-5% CO₂ for 3 hr without resting tension, muscle strips were exposed to each drug for 5 min, frozen in liquid nitrogen and homogenized in 6% trichloroacetic acid solution. After centrifugation, trichloroacetic acid in the supernatant was removed by washing with water-saturated ether. cAMP was assayed by a competitive enzyme immunooassay with cAMP peroxidase conjugate.

Solutions and drugs. PSS contained (mM): NaCl 136.9, KCl 5.4, CaCl₂ 1.5, MgCl₂ 1.0, NaHCO₃ 23.8 and glucose 5.5. EDTA (0.01 mM) was added to chelate heavy metal ions contaminating the PSS. High-K⁺ solution was made by substituting equimolar KCl for NaCl. Ca²⁺-free solution was made by removing CaCl₂ from PSS and adding 0.5 mM EGTA. These solutions were aerated with 95% O₂-5% CO₂ mixture at 37°C and pH 7.4.

The following drugs, chemicals and apparatus were used:

Fig. 1. Concentration-response curves of bovine tracheal strips precontracted with 0.1 μM CCh (○), 1 μM CCh (●) and 72.7 mM KCl (□) to clonidine. Each point represents the mean ± S.E.M. Asterisks indicate significant difference (⁎ P < .05, ** P < .01) from control.

Fig. 2. Effects of 1 μM yohimbine (panel A, □), 10 μM idazoxan (B, ▲) and 30 μM idazoxan (panel B, △) on the concentration-response curves of bovine tracheal strips precontracted with 0.1 μM CCh to clonidine compared with control data (● in both). Each point represents the mean ± S.E.M. IC₅₀ values increase from 16.5 ± 2.1 μM to 42.8 ± 1.8 μM, 39.6 ± 1.3 μM and 52.2 ± 2.0 μM, respectively. Asterisks indicate significant difference from control curve (⁎ P < .05, ** P < .01) and between the two idazoxan groups († P < .05, †† P < .01).
Therefore, a combination of high K\(^+\) exposure for 15 min and interval for 15 min was repeated three times before the drug applications.

CCh at concentrations of 0.1 and 1 \(\mu\)M produced sustained contraction amounting to 111 ± 7.9\% and 159 ± 11.4\% of high K\(^+\) (72.7 mM)-induced contraction, respectively. Although no response was observed when it was administered alone, clonidine (0.1–100 \(\mu\)M) exhibited a concentration-dependent suppression of the force induced by 0.1 and 1 \(\mu\)M CCh, with IC\(_{50}\) values of 17.1 ± 2.0 \(\mu\)M and 100 ± 51 \(\mu\)M, respectively (\(n = 4\) each). On the other hand, the relaxant effect of clonidine on high K\(^+\) (72.7 mM)-induced contraction was much weaker (fig. 1).

The A\(_2\)R antagonists yohimbine (1 \(\mu\)M) and idazoxan (10 and 30 \(\mu\)M) shifted the concentration-response curve for clonidine to the right in parallel (fig. 2; \(n = 4\) each). The dissociation constants (\(K_i\)) of these two antagonists were determined as 660 \(\mu\)M and 7,600–15,000 \(\mu\)M, respectively, from the relationship

\[
\log([D]/[D_0] - 1) = \log[I] - \log[K_i]
\]

where \([D]\) is the IC\(_{50}\) of clonidine in the presence of yohimbine or idazoxan, \([D_0]\) is the IC\(_{50}\) of clonidine alone and \([I]\) is the concentration of yohimbine or idazoxan. In contrast, clonidine-induced relaxation was not affected by an alpha\(_1\)\(\lambda\)-adrenoceptor antagonist, WB 4101 (1 \(\mu\)M, \(n = 4\)), by an alpha-1 adrenoceptor antagonist, prazosin (0.01 \(\mu\)M, \(n = 5\)), by a nonselective beta-adrenoceptor antagonist, propranolol (20 \(\mu\)M, \(n = 5\)) or by a beta-2 adrenoceptor antagonist, butoxamine (1 \(\mu\)M, \(n = 4\)). A Na\(^+\) channel blocker, TTX (1 \(\mu\)M), and a cyclooxygenase inhibitor, indomethacin (10 \(\mu\)M), had no effect on clonidine-induced relaxation, either (\(n = 5\) each).

The imidazoline agonists oxymetazoline (0.1–100 \(\mu\)M) and phentolamine (0.1–100 \(\mu\)M), also showed concentration-dependent relaxation in CCh (0.1 \(\mu\)M)-contracted BTSM, with IC\(_{50}\) values of 7.9 ± 2.1 \(\mu\)M and 37.8 ± 7.5 \(\mu\)M, respectively. Additionally, both of these relaxant effects were inhibited by idazoxan (10 \(\mu\)M), with IC\(_{50}\) values of 16.0 ± 3.4 \(\mu\)M and 37.8 ± 7.5 \(\mu\)M, respectively.

**Results**

**Changes in muscle force.** High-K\(^+\) (72.7 mM) solution induced sustained contraction that reached plateau within 10 min in each exposure, and tension obtained in the third exposure trial that showed no significant difference from that in the fourth or later trials was adopted as the control value.
724.4 ± 80.9 μM, respectively, but not by yohimbine (1 μM, data not shown) (fig. 3, n = 4 – 5).

Norepinephrine (0.1–100 μM) exhibited a concentration-dependent relaxation in CCh (0.1 μM)-contracted BTSM, with IC50 of 6.6 ± 1.9 μM, which was completely attenuated by propranolol (10 μM) but not by yohimbine (1 μM) (fig. 4; n = 5 each).

Changes in [Ca2+]i and muscle force. In fura-PE3/AM-loaded trachea, high K+ (72.7 mM) induced sustained increases in [Ca2+]i and muscle force (n = 4 – 5). The control value of the ratio was determined approximately 10 min after high K+ application in the third trial in each strip, when the trace reached plateau. The center of the trace widths was adopted as the point of measurement. In KCl-stimulated tracheae, clonidine induced only slight relaxation without changing [Ca2+]i (fig. 5A). CCh also induced sustained increases in [Ca2+]i and contraction. In CCh-stimulated tracheae, clonidine (10–100 μM) induced concentration-dependent decreases in [Ca2+]i and force (fig. 5B). Atropine (0.1–1.0 μM) also reduced both [Ca2+]i and force (fig. 5C). On the other hand, verapamil (0.01–0.1 μM) more strongly inhibited [Ca2+]i than force in the CCh-stimulated tracheae; although 1.0 μM verapamil almost completely inhibited CCh-stimulated [Ca2+]i, it inhibited CCh-induced contraction only by approximately 50% (fig. 5D).

Figure 6 summarizes the effects of clonidine, atropine and verapamil on the [Ca2+]i-force relationship in the presence of 0.1 μM CCh. CCh induced greater contraction than high K+ (72.7 mM) at a given [Ca2+]i. In the verapamil-induced relaxation, muscle force decreased less steeply than in the clonidine- or atropine-induced relaxation.

In Ca2+-free solution (with 0.5 mM EGTA), CCh (0.1 μM) induced a spike-like transient increase in [Ca2+]i (104.0 ± 11.2% of high K+ induced response, n = 5). Clonidine (100 μM) diminished the transient [Ca2+]i, rise elicited by CCh (0.1 μM) to 45.3 ± 17.5% (fig. 7, A and C). Caffeine (20 mM) also induced the transient increase in [Ca2+]i, in Ca2+-free solution (100.0 ± 9.0%, n = 4). Clonidine (100 μM) had no significant effect on the caffeine-induced Ca2+ transient (92 ± 18.5%, n = 4, fig. 7, B and C).

cAMP assay. Clonidine (100 μM) had no significant effect on cAMP concentration in the presence of CCh (0.1 μM) or KCl (72.7 mM). On the other hand, forskolin (10 μM) greatly increased cAMP concentration in this preparation (fig. 8, n = 4 each).

Discussion

In the bovine trachea, clonidine produced only slight suppression of high K+-induced increases in contraction and did not affect [Ca2+]i. In contrast, clonidine relaxed CCh-induced contraction in association with a decrease in [Ca2+]i. The effects of clonidine were not affected by alpha-1 or beta adrenoceptor antagonists, such as WB 4101, prazosin, propranolol and butoxamine, but were inhibited by yohimbine and idazoxan. TTX and indomethacin also had no effect on the clonidine-induced relaxation. These results suggest that clonidine directly inhibits smooth muscle contraction through A2Rs and/or I1Rs in bovine trachea without any influence of cyclooxygenase-related metabolites. In dogs (Barnes et al., 1983) and in guinea pigs (Floch and Advenier,
Clonidine has been reported to contract isolated airway smooth muscle. By contrast, clonidine did not produce any direct contraction in BTSM, as shown in the present study. On the other hand, that the release of ACh from airways cholinergic nerves is reduced by A2R stimulation has been demonstrated in vitro in horses (Yu et al., 1993) and in guinea pigs (Baker et al., 1994). The variation in these results may be attributable to the difference in species. However, it has been reported that preapplied clonidine (10 and 100 μM) failed to reduce BTSM contraction induced by exogenous ACh (1 mM) (Manning and Brodstone, 1995). Their data seem to conflict with our results. The discrepancy may be caused by differences in the types and concentrations of muscarinic agonists. Potency order in these two agonists, ACh and CCh, varies with species and/or tissue type, which has been suggested to depend on the cholinesterase activity (Mitsui-Saito and Karaki, 1996). ACh and CCh have also been suggested to interact with different muscarinic receptors (Mitchelson and Ziegler, 1984). Furthermore, differences in tissue preparation may contribute to such a discrepancy. Epithelium and mucosa were left intact in their preparation but were removed in ours. Although the effects of clonidine on bovine airway epithelial function are unclear, several reports have indicated that agonist-stimulated epithelial cells modulate airway smooth muscle tone in various species (Farmer and Hay, 1991).

Clonidine stimulates not only A2Rs but also the I1Rs. The imidazoline receptors have been identified in several kinds of smooth muscle cells (Yablonsky and Dausse, 1991; Regunathan et al., 1995) and have been suggested to contribute to regulation of muscle tone. In the present study, the relaxant effects of clonidine were inhibited by either yohimbine (1 μM) or idazoxan (10 and 30 μM). However, the Ki for A2Rs of yohimbine and idazoxan have been reported to be 0.1–12 nM (Blaxall et al., 1994; Chruscinski et al., 1992) and 13–83 nM (Burke et al., 1995), respectively, both of which are approxi-
It has been reported that CCh augmented Ca\(^{2+}\) sensitivity of the contractile element in the canine trachea (Ozaki et al., 1990). In the present study, we also revealed in the bovine trachea that CCh produced greater contraction than high K\(^+\) at a given [Ca\(^{2+}\)], which suggests that CCh increased the Ca\(^{2+}\) sensitivity of the contractile element. In Ca\(^{2+}\)-free solution, 0.1 \(\mu\)M CCh induced a transient increase in [Ca\(^{2+}\)], that may be due to production of inositol-1,4,5-trisphosphate mediated by a muscarinic stimulation (Blüml et al., 1994). An activator of Ca\(^{2+}\)-induced Ca\(^{2+}\) release, 20 mM caffeine (Iino, 1990), also transiently increased [Ca\(^{2+}\)], in Ca\(^{2+}\)-free solution. Clonidine, at the concentration needed to inhibit the 0.1 \(\mu\)M CCh-induced sustained increase in [Ca\(^{2+}\)], and sustained contraction, inhibited significantly the CCh-induced transient increase in [Ca\(^{2+}\)], in Ca\(^{2+}\)-free solution, but not the effect of caffeine. These results suggest that clonidine inhibits the Ca\(^{2+}\) release induced by inositol-1,4,5-trisphosphate but not the Ca\(^{2+}\)-induced release of Ca\(^{2+}\). However, the effect of clonidine against the CCh stimulation was only partial (approximately 50%), which suggests that other mechanism(s) are involved. Because clonidine decreased force more steeply than verapamil in the [Ca\(^{2+}\)]-force relationship, it appears that clonidine inhibits not only Ca\(^{2+}\) increase but also the other downstream mechanisms that enhance Ca\(^{2+}\) sensitivity. Moreover, the relationship was not completely the same as that in the atropine-induced relaxation, which suggests that clonidine may inhibit not only the responses specific to muscarinic stimulation but also the other, nonspecific mechanisms. This hypothesis is supported by the results that clonidine slightly but significantly suppressed the force, but not [Ca\(^{2+}\)], in high K\(^+\)-stimulated BTSM. Thus clonidine may exert its inhibitory effects by blocking the contractile mechanisms related to muscarinic stimulation and the other, nonspecific component(s) in CCh-contracted BTSM.

In summary, clonidine relaxes CCh-contracted BTSM by inhibiting the release of Ca\(^{2+}\), the increase in Ca\(^{2+}\) influx and the increase in Ca\(^{2+}\) sensitivity of contractile elements mainly via \(I_1\)Rs.

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References
Burke J, Kharlamb A, Shan T, Runde E, Padillo E, Manlapaz C and Wheeler L (1995) Adrenergic and imidazoline receptor-mediated responses to UK-14,304-18 (bri- 

expression of the mouse homolog of the human alpha 2C-adrenergic receptor. 

Dinh Xuan A and Lockhart A (1989) Bronchial effects of alpha 2-adrenoceptor 

Effects of clonidine on bronchial responses to histamine in normal and asthmatic 

Farmer S and Hay D (1991) Airway epithelial modulation of smooth muscle function: 
The evidence for epithelium-derived inhibitory factor, in The Airway Epithelium 
(Farmer SG and Hay DWP eds) pp 437–483, Marcel Dekker, New York.

Floch A and Advenier C (1985) Interaction between clonidine and histamine on the 

of stereoselective interactions of catecholamines with the 
a2-adrenoceptor via site directed mutagenesis, in a2 Adrenergic Receptors: Structure, Function and 
Therapeutic Implications (Lanier SM and Limbird LE eds) pp 43–51, Harwood 
Academic Publishers, Amsterdam.

Inoue M (1990) Bihaptic Ca2+ dependence of inositol 1,4,5-trisphosphate-induced Ca 


Kobinger W (1978) Central alpha-adrenergic systems as targets for hypotensive 

stimulation on cholinergic contractions of equine distal arteries. Am J Vet Res 
54:788–792.

Leff A and Munoz N (1981) Evidence for two subtypes of alpha adrenergic receptors 

Lindgren B, Ekstrom T and Anderson R (1986) The effect of inhaled clonidine in 

Macquin-Mavier I, Clerici C and Hart A (1996) Effects of alfenadine on bronchomo-
tor responses in the guinea pig. Am J Cardiol 77:45D–46D.

Manning M and Brodstone R (1995) Effects of a2-adrenoceptor agonist and 
antagonist drugs on cholinergic contraction in bovine tracheal smooth muscle 

on responses to acetylcholine and carbachol in the taenia of the guinea-pig caecum. 
Br J Pharmacol 82:165–175.

Mitsui-Saito M and Karaki H (1996) Carbachol but not acetylcholine inhibits con-
traction by the protein kinase C-dependent and -independent pathways in the 

on induced cough and bronchoconstriction in guinea pigs and healthy humans. 

Olsson O and Ekdahl A (1985) The use of two different pharmacological principles to 
inhibit a cholinergic bronchospasm in guinea pig. Acta Pharmacol Toxicol 
(Copenh) 56:316–320.

signals and mechanical activity using fluorescent dye fura 2 in isolated strips of 

and contraction induced by various stimulants and relaxants in canine tracheal 

human I1-imidazoline binding sites and the high affinity state of alpha-2 adreno-


smooth muscle strip measured simultaneously with contraction using fluorescent 


Takayanagi I, Kawano K and Koike K (1990) a2-Adrenoceptor mechanisms in 

Tsuchiya Y, Hosokawa T and Kauya Y (1990) Involvement of alpha 2-adrenergic 
receptors in the vagal reflex-induced tracheal constriction. J Pharmacobiosyn 
13:30–35.


B-HT 920 in some isolated smooth muscles of the guinea-pig. Acta Pharmacol 
Toxicol (Copenh) 50:266–273.

Yablonsky F and Dausse J (1991) Non-adrenergic binding sites for the alpha2-
agonist [3H]dihazoxan in the rabbit urethral smooth muscle. Biochem Pharma-

acetylcholine release from cholinergic nerves in equine arteries. Am J Physiol 
265:L565–L570.

Zonnenchein R, Diamant S and Atlas D (1990) Imidazoline receptors in rat liver 
cells: A novel receptor or a subtype of alpha 2-adrenoceptors? Eur J Pharmacol 
190:203–215.

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