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**Analysis of β -subunit-dependent GABA_A receptor modulation and
behavioral effects of valerenic acid derivatives**

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Non-standard abbreviations: VA (valerenic acid)

VA-A (VA-amide; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-2-methylacrylamide),

VA-MA (VA-methylamide; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N*,2-dimethylacrylamide)

VA-DMA (VA-dimethylamide; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N,N*,2-trimethylacrylamide)

VA-EA (VA-ethylamide; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N*-ethyl-2-methylacrylamide)

VA-DEA (VA-diethylamide; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N,N*-diethyl-2-methylacrylamide)

VA-TET (VA-tetrazole; 5-((*E*)-1-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)prop-1-en-2-yl)-1*H*-tetrazole)

VA-CN (VA-nitrile; (*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-2-methylacrylonitrile)

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Abstract

Valerenic acid (VA) - a β 2/3-selective γ -aminobutyric acid (GABA) type A ($GABA_A$) receptor modulator - displays anxiolytic and anticonvulsive effects in mice devoid of sedation, making VA an interesting drug candidate. Here, we analyzed β -subunit-dependent enhancement of GABA-induced chloride currents (I_{GABA}) by a library of VA derivatives and studied their effects on pentylenetetrazole (PTZ)-induced seizure threshold and locomotion. Compound-induced I_{GABA} enhancement was determined in oocytes expressing α 1 β 1 γ 2S, α 1 β 2 γ 2S, or α 1 β 3 γ 2S receptors. Effects on seizure threshold and locomotion were studied using C57BL/6N mice and compared to saline-treated controls. β 2/3-selective VA derivatives such as VA-amide (VA-A) modulating α 1 β 3 γ 2S (VA-A: $E_{max}=972\pm 69\%$, $n=6$, $p<0.05$) and α 1 β 2 γ 2S receptors ($E_{max}=1119\pm 72\%$, $n=6$, $p<0.05$) more efficaciously than VA (α 1 β 3 γ 2S: VA: $E_{max}=632\pm 88\%$, $n=9$ vs. α 1 β 2 γ 2S: VA: $E_{max}=721\pm 68\%$, $n=6$) displayed significantly more pronounced seizure threshold elevation than VA (saline-control: 40.4 ± 1.4 mg/kg PTZ vs. VA 10mg/kg: 49.0 ± 1.8 mg/kg PTZ vs. VA-A 3mg/kg: 57.9 ± 1.9 mg/kg PTZ, $p<0.05$). Similarly, VA's methylamide (VA-MA) enhancing I_{GABA} through β 3-containing receptors more efficaciously than VA ($E_{max}=1043\pm 57\%$, $p<0.01$, $n=6$) displayed stronger anticonvulsive effects. Increased potency of I_{GABA} enhancement and anticonvulsive effects at lower doses compared to VA were observed for VA-tetrazole (α 1 β 3 γ 2S: VA-TET: $EC_{50}=6.0\pm 1.0\mu M$, $p<0.05$; VA-TET: 0.3mg/kg: 47.3 ± 0.5 mg/kg PTZ vs. VA: 10mg/kg: 49.0 ± 1.8 mg/kg PTZ, $p<0.05$). At higher doses (≥ 10 mg/kg), VA-A, VA-MA and VA-TET reduced locomotion. In contrast, unselective VA-derivatives induced anticonvulsive effects only at high doses (30mg/kg) or did not display any behavioral effects. Our data indicate that the β 2/3-selective compounds VA-A, VA-MA and VA-TET induce anticonvulsive effects at low doses (≤ 10 mg/kg), while impairment of locomotion was observed at doses ≥ 10 mg/kg.

Introduction

Interaction of γ -aminobutyric acid (GABA) with GABA type A (GABA_A) receptors enables fast inhibitory neurotransmission in the mammalian brain (Barnard *et al.*, 1998; Olsen and Sieghart, 2008; Sigel and Steinmann, 2012). Like other members of the family of pentameric ligand-gated ion channels (pLGIC), GABA_A receptors are constituted by pseudosymmetrical assembly of five identical or homologous subunits forming a chloride-conducting ion (Miller and Aricescu, 2014). The human genome comprises genes encoding for 19 different GABA_A receptor subunits (α 1-6, β 1-3, γ 1-3, δ , ϵ , ρ 1-3, π and θ (Simon *et al.*, 2004)) theoretically allowing the formation of multiple GABA_A receptor subtypes. The receptor's subunit composition determines its pharmacological properties including agonist sensitivity as well as its sensitivity for drugs (Barnard *et al.*, 1998; Sigel and Steinmann, 2012). In addition, the highly specific cellular and subcellular distribution of GABA_A receptor subunits/subtypes (Pirker *et al.*, 2000; Schwarzer *et al.*, 2001; Olsen and Sieghart, 2008) and in particular the assignment of therapeutic effects of commonly applied GABA_A receptor modulators such as benzodiazepines (e.g. Möhler *et al.*, 2001; Rudolph *et al.*, 2001; Rudolph and Knoflach, 2011; Gallos *et al.*, 2012, 2015; Rudolph and Möhler, 2014) or anaesthetics (Jurd *et al.*, 2003) to single GABA_A receptor subunits raises the possibility to develop drugs selectively targeting specific brain circuits. Such subunit-selective GABA_A receptor modulators are predicted to display the desired therapeutic effects with reduced side effects.

Valerenic acid (VA) – a sesquiterpenoid compound found in common *Valerian* – selectively modulates γ -aminobutyric acid (GABA) type A (GABA_A) receptors containing β 2- or β 3-subunits, while displaying only negligible effects on GABA_A receptors incorporating β 1-subunits (Khom *et al.*, 2007; Benke *et al.*, 2009). *In vivo*, VA induces anxiolytic (Benke *et al.*, 2009; Khom *et al.*, 2010; Hintersteiner *et al.*, 2014) and anticonvulsive (Hintersteiner *et al.*, 2014) effects. Most notably, VA does not significantly reduce locomotor activity even at high doses (Khom *et al.*, 2010).

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These findings combined with a promising pharmacokinetic profile (Sampath *et al.*, 2012) make VA an interesting drug candidate. Previously, we have reported more pronounced anxiolytic and/or anticonvulsive effects by selected VA amide (Khom *et al.*, 2010) and ester derivatives (Hintersteiner *et al.*, 2014). In contrast to VA ester derivatives that are significantly less active *in vitro* and thus might act as prodrugs (Hintersteiner *et al.*, 2014), both VA amide derivatives (Khom *et al.*, 2010; Kopp *et al.*, 2010) as well as VA's nitrile and tetrazole analogues (Kopp *et al.*, 2010) efficaciously enhance GABA-induced chloride currents (I_{GABA}) through receptors composed of $\alpha 1\beta 3$ subunits (Khom *et al.*, 2010) or $\alpha 1\beta 2\gamma 2S$ (Kopp *et al.*, 2010) expressed in *Xenopus* oocytes.

Therefore, to evaluate the potential of VA derivatives as scaffold for the development of novel anticonvulsants, we extended these previous studies by determining β -subunit-dependency of I_{GABA} modulation by a small focused library of VA derivatives in oocytes expressing $\alpha 1\beta 1\gamma 2S$, $\alpha 1\beta 2\gamma 2S$, or $\alpha 1\beta 3\gamma 2S$ receptors followed by an *in vivo* investigation of their effects on pentylenetetrazole (PTZ)-induced seizure threshold and locomotor activity in male C57BL/6N mice.

Our study shows that VA's amide (VA-A) and methylamide (VA-MA) derivatives, respectively, modulating $\alpha 1\beta 3\gamma 2S$ GABA_A receptors more efficaciously than VA displayed also significantly more pronounced elevation of PTZ-induced seizure threshold. In addition, VA's tetrazole derivative (VA-TET) – an efficacious VA derivative already reported in a preceding study by Kopp *et al.* (Kopp *et al.*, 2010) - modulating $\alpha 1\beta 3\gamma 2S$ receptors more potently than VA - displayed anticonvulsive effects at lower doses than the VA. In contrast, unselective VA derivatives such VA-ethylamide (VA-EA) or VA-diethylamide (VA-DEA) displayed anticonvulsive effects only at high doses (30mg/kg) or did not show any effects on either PTZ-induced seizure threshold or locomotion (VA-dimethylamide, VA-DEA).

Materials and Methods

Animals and animal welfare

All experiments involving animals were approved by the Austrian Animal Experimentation Ethics Board in compliance with the European convention for the protection of vertebrate animals used for experimental and other scientific purposes ETS no.: 123, which is in line with the EU Directive 2010/63/EU (GZ 66.006/0019-C/GT/2007, GZ 66.006/0008-II/10b/2008 and GZ 66.006/0009-II/10b/2010). Every effort was taken to minimize the number of animals used.

Female *Xenopus laevis* frogs were purchased from NASCO (Fort Atkinson, USA) and kept in groups in temperature-controlled, continuous-flow water tanks ($20\pm 1^\circ\text{C}$). Male mice (C57BL/6N) were obtained from Charles River Laboratories (Sulzfeld, Germany). Mice were group-housed (maximum 5 mice per type IIL cage) with free access to food and water. At least 24h before the commencement of experiments, they were transferred to the testing facility, continuing *ad libitum* access to food and water.

The temperature in the holding (for mice and frogs) and testing facilities was fixed to $22\pm 2^\circ\text{C}$; the humidity ranged between 40-60%; a 12h light-dark cycle was in operation (lights on from 07.00 to 19.00).

Chemicals

All chemicals used in this study were obtained from Sigma Aldrich (Vienna, Austria) except where stated otherwise. Valerenic acid (VA) was purchased from HWI Pharma Solutions (Rülzheim, Germany). VA derivatives (structural formulae are depicted in Fig.1) were synthesized as previously described: Description of the syntheses of VA-A ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-2-methylacrylamide), VA-MA ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N*,2-dimethylacrylamide), VA-DMA ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-

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1*H*-inden-4-yl)-*N,N*,2-trimethylacrylamide), VA-EA ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N*-ethyl-2-methylacrylamide), VA-DEA ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-*N,N*-diethyl-2-methylacrylamide) can be found in (Khom *et al.*, 2010) and for VA-TET (5-((*E*)-1-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)prop-1-en-2-yl)-1*H*-tetrazole), and VA-CN ((*E*)-3-((4*S*,7*R*,7*aR*)-3,7-Dimethyl-2,4,5,6,7,7*a*-hexahydro-1*H*-inden-4-yl)-2-methylacrylonitrile) see Kopp *et al.*, 2010. Stock solutions (100mM for *in vitro* experiments and 1mg/10μl for *in vivo* experiments, respectively) were prepared in 100% dimethylsulfoxide (DMSO). VA and its derivatives were used up to a concentration of 500μM in *in vitro* experiments. Equal amounts of DMSO were present in control and compound-containing solutions. The maximum DMSO concentration in the bath (0.5%) did not affect I_{GABA}. For *in vivo* experiments, working concentrations were adjusted by dilution with 0.9% sodium chloride; the final concentration of DMSO was fixed to 10% including control solutions. To enhance solubility of the compound Tween 80 (3% final concentration) was added to all solutions. pH was adjusted to 7.2-7.4 with 1M sodium hydroxide. All solutions were freshly prepared every day prior to experiments.

Expression and functional characterization of GABA_A receptors

Preparation of stage V-VI oocytes from *Xenopus laevis* (NASCO, Fort Atkinson, USA), synthesis of capped off run-off poly(A⁺) rat cRNA transcripts from linearized cDNA templates (pCMV vector) was performed as described elsewhere (Khom *et al.*, 2006). Briefly, female *Xenopus laevis* were anaesthetized by exposing them for 15min to a 0.2% solution of MS-222 (methane sulfonate salt of 3-aminobenzoic acid ethyl ester) before surgically removing parts of the ovaries. Follicle membranes from isolated oocytes were enzymatically digested with 2mg/ml collagenase (Type 1A). Oocytes were stored at 18°C in ND96 solution (Methfessel *et al.*, 1986). After isolation, oocytes were injected with about 10-50nl of

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nuclease-free water containing the different rat cRNAs (100-2000ng/ μ l/subunit). For expression of $\alpha 1\beta 3\gamma 2S$ and $\alpha 1\beta 2\gamma 2S$ receptors, respectively, cRNAs were mixed in a ratio of 1:1:10 (Boileau *et al.*, 2002); to avoid formation of homooligomeric $\beta 1$ -receptors in the case of $\alpha 1\beta 1\gamma 2S$ a ratio of 3:1:10 was used (Krishek *et al.*, 1996). Electrophysiological experiments were done using the two-microelectrode-voltage-clamp-technique at a holding potential of -70mV making use of a TURBO TEC 01C amplifier (npi electronic, Tamm, Germany) and an Axon Digidata 1322A interface (Molecular Devices, Sunnyvale, CA). Data acquisition was carried out using pCLAMP v.9.2 (Molecular Devices, Sunnyvale, CA). The bath solution contained 90mM NaCl, 1mM KCl, 1mM MgCl₂, 1mM CaCl₂ and 5mM HEPES (pH 7.4). Microelectrodes were filled with 2M KCl and had resistances between 1 and 3M Ω .

Perfusion system

GABA and drugs were applied by means of a fast perfusion system; drug or control solutions were applied by means of a TECAN Miniprep 60 permitting automation of the experiments ((Baburin *et al.*, 2006) ScreeningTool, npi electronic). To elicit I_{GABA}, the chamber was perfused with 120 μ l of GABA-containing solution at a volume rate between 300 and 1000 μ l/s. The I_{GABA} rise time ranged between 100 and 250ms (Khom *et al.*, 2006). To account for possible slow recovery from increasing levels of desensitization in the presence of high compound concentrations, the duration of washout periods was extended stepwise, i.e. 1min (GABA EC₃₋₇) to 1.5min (co-application of GABA EC₃₋₇ in the presence $\leq 1\mu$ M compound;) to 2.5min (co-application of GABA EC₃₋₇ in the presence of $\leq 10\mu$ M compound) to 5min (co-application of GABA EC₃₋₇ and $\leq 100\mu$ M compound) to 15min (GABA EC₃₋₇ in the presence of 300-500 μ M compound). Oocytes with maximal current amplitudes $> 5\mu$ A were discarded to exclude voltage-clamp errors.

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Analyzing concentration-response curves

Enhancement of chloride currents (I_{GABA}) by VA and VA derivatives was measured at a GABA concentration eliciting between 3 and 7% of the maximal current amplitude (EC_{3-7}). The EC_{3-7} was determined at the beginning of the experiment for each oocyte by application of 1mM GABA followed by submaximal GABA concentrations. I_{GABA} enhancement was defined as $(I_{(GABA+Comp)}/I_{GABA}) - 1$, where $I_{(GABA+Comp)}$ is the current response in the presence of compound and I_{GABA} is the control GABA current. Concentration-response curves were generated and the data were fitted by non-linear regression analysis using Origin software (OriginLab Corporation, USA). Data were fitted to the following equation:

$$Y = \text{min} + (\text{max} - \text{min}) * x^{n_H} / (k^{n_H} + x^{n_H}).$$

In this equation, k corresponds to the EC_{50} value, x -values are logs of concentration, and n_H stands for the Hill coefficient. Each data point represents the mean \pm SEM from ≥ 3 oocytes and ≥ 2 oocyte batches.

In vivo characterization of VA derivatives

Only male C57BL/6N mice (age 3-6 months) were used in the tests described below. Intraperitoneal (i.p.) injection of control or compound-containing solutions was done 30min before the test. Indicated doses in the results and discussion section represent mg/kg bodyweight.

Seizure threshold

Seizure threshold was determined by pentylenetetrazole (PTZ) tail-vein infusion on freely moving animals at a rate of 100 μ l/min (10mg/ml PTZ in saline, pH=7.4). Infusion was stopped when animals displayed generalized clonic seizures. Animals were immediately killed by cervical displacement after onset of seizures. The seizure threshold dose was calculated from the infused dose in relation to body weight (mg/kg).

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Open-field-test

Exploration of a novel environment was tested over 10min in a 50x50cm box built from gray PVC equipped with infra-red beams. Illumination intensity was set to 150lux in the center. Animals' motor activity was analyzed using ActiMot-2 equipment and software (TSE-systems, Bad Homburg, Germany).

Statistical analysis

Statistical significance was calculated using one-way ANOVA followed by a post-hoc mean comparison with Bonferroni (OriginLab Corporation, USA or GraphPad, La Jolla, USA). *p*-values of <0.05 were accepted as statistically significant. All data are given as mean±SEM.

Results

Determination of β -subunit-dependency of I_{GABA} modulation by valerenic acid and VA derivatives

GABA_A receptors composed of $\alpha 1\beta 1\gamma 2S$, $\alpha 1\beta 2\gamma 2S$ or $\alpha 1\beta 3\gamma 2S$ subunits were expressed in *Xenopus laevis* oocytes and modulation of GABA-induced chloride currents (I_{GABA} , GABA EC₃₋₇) by valerenic acid (VA) and seven carboxyl-group modified derivatives was analyzed by means of the 2-microelectrode voltage clamp technique (for structural formulae of investigated VA derivatives, see Fig.1). Amidation of VA (VA-amide, VA-A) resulted in significantly stronger I_{GABA} enhancement of both $\alpha 1\beta 3\gamma 2S$ (VA-A: $E_{max}=972\pm 69\%$, $n=6$ vs. VA: $E_{max}=632\pm 88\%$, $n=9$ $p<0.01$) and $\alpha 1\beta 2\gamma 2S$ receptors (VA-A: $E_{max}=1119\pm 72\%$, $n=6$ vs. VA: $E_{max}=721\pm 68\%$, $n=5$, $p<0.05$, compare Figs.2A and C). Subsequent mono-methylation of VA-A (VA-mono-methylamide, VA-MA) also significantly increased I_{GABA} enhancement of $\alpha 1\beta 3\gamma 2S$ receptors (VA-MA: $E_{max}=1043\pm 57\%$, $n=5$, $p<0.01$, Fig.2E) compared to VA; in addition, a strong trend towards more pronounced I_{GABA} enhancement of $\alpha 1\beta 2\gamma 2S$ channels by VA-MA was observed; however, this effect did not reach statistical significance (VA-MA: $E_{max}=917\pm 36\%$, $n=3$, $p>0.05$, Fig. 2E). Modulation of I_{GABA} through $\beta 1$ -containing receptors by VA-A and VA-MA did not significantly differ – even though slightly enhanced – from I_{GABA} enhancement by VA ($\alpha 1\beta 1\gamma 2S$: VA: $E_{max}=111\pm 16\%$, $n=8$ vs. VA-A: $E_{max}=218\pm 78\%$, $n=6$; VA-MA: $E_{max}=387\pm 56\%$, $n=5$; $p>0.05$; compare Figs.2A, C and E; see also Table 1, see also (Khom *et al.*, 2010; Kopp *et al.*, 2010) for VA-A enhancement of $\alpha 1\beta 1-3$, $\alpha 1\beta 2\gamma 2S$, and $\alpha 3\beta 3\gamma 2S$ and VA-MA enhancement of $\alpha 1\beta 3$ and $\alpha 1\beta 2\gamma 2S$ receptors).

Replacement of the carboxyl-moiety by the bioisosteric tetrazole group (VA-TET) significantly increased efficacy on $\alpha 1\beta 2\gamma 2S$ ($E_{max}=1091\pm 87\%$, $n=5$, $p<0.05$, Fig.2G) receptors though with a slightly reduced potency compared to VA, while efficacy of I_{GABA} enhancement on $\alpha 1\beta 1\gamma 2S$ ($E_{max}=176\pm 43\%$, $n=7$) and $\alpha 1\beta 3\gamma 2S$ ($E_{max}=668\pm 57\%$, $n=8$; Fig.2G) receptors did not differ from that of the parent compound. However, most remarkably, VA-TET displayed

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an approximately 3-fold increased potency on β 3-containing receptors compared to VA (VA: $EC_{50}=20.2\pm 5.2\mu M$; $n=9$ vs. VA-TET: $EC_{50}=6.0\pm 1.0\mu M$; $n=8$; see Fig.2 for representative current traces for β -subunit-dependent I_{GABA} enhancement by (B) VA, (D) VA-A, (F) VA-MA and (H) VA-TET; data for I_{GABA} enhancement of $GABA_A$ channels composed of $\alpha 1\beta 3\gamma 2S$ subunits by VA are taken from (Luger *et al.*, 2015); see also the effect of VA-TET applied at low GABA concentrations (EC_{1-4}) in (Kopp *et al.*, 2010).

Similarly, substituting VA's carboxyl group by a nitrile (VA nitrile, VA-CN) did not affect $\beta 2/3$ -subunit-selective I_{GABA} potentiation: I_{GABA} enhancement by VA-CN through $\alpha 1\beta 1\gamma 2S$ ($E_{max}=55\pm 14\%$, $n=7$), $\alpha 1\beta 2\gamma 2S$ ($E_{max}=765\pm 117\%$, $n=4$), and $\alpha 1\beta 3\gamma 2S$ ($E_{max}=522\pm 114\%$, $n=7$) channels, respectively, was similar to that of VA; however, a trend towards slightly decreased potency on $\beta 2$ - and $\beta 3$ -subunit containing receptors was observed (see Fig.3A; see also (Kopp *et al.*, 2010) for I_{GABA} enhancement through $\alpha 1\beta 2\gamma 2S$ channels by VA-CN at GABA EC_{1-4} concentrations).

In contrast, introducing bulkier residues such as mono-ethylamide (VA-EA), di-methylamide (VA-DMA) or di-ethylamide (VA-DEA) significantly elevated efficacy on $\beta 1$ -containing receptors compared to VA (VA-EA: $E_{max}=458\pm 124\%$, $n=5$, Fig.3B; VA-DMA: $E_{max}=305\pm 67\%$, $n=6$, Fig.3C; VA-DEA: $E_{max}=318\pm 84\%$, $n=7$, Fig.3D; $p>0.05$), while the estimated efficacies on $\beta 2$ - and $\beta 3$ -containing receptors were comparable to VA, indicating a loss of subunit-selectivity (Figs.3B-D). Data for maximal I_{GABA} enhancement (E_{max} , %), EC_{50} (μM), Hill-coefficients (n_H) and number of experiments (n) for all receptor subunit compositions tested are summarized in Table 1.

Effects of VA and VA derivatives on pentylenetetrazole (PTZ)-induced seizure threshold

We have recently reported that the elevation of PTZ-induced seizure threshold by VA (3mg/kg bodyweight) is most pronounced 30min after application (Hintersteiner *et al.*, 2014).

In the present study, dose-dependent effects of VA on seizure threshold were determined. As

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depicted in Fig.4A, VA did not alter seizure threshold at doses <3mg/kg bodyweight; in contrast, pronounced seizure threshold elevation was observed after application of VA at a dose of 3 or 10mg/kg bodyweight (control: 40.4±1.4 mg/kg PTZ, n=6 vs. VA 3mg/kg: 47.7±1.4mg/kg PTZ, n=4, p<0.01 and VA 10mg/kg: 49.0±1.8mg/kg PTZ, n=4; p<0.05, respectively; data for seizure threshold elevation by VA at a dose of 3mg/kg bodyweight are taken from (Hintersteiner *et al.*, 2014)). Seizure threshold of animals treated with VA at a dose of 30mg/kg bodyweight did not differ significantly from diluent-treated control animals (30mg/kg VA: 43.4±1.8mg/kg PTZ, n=3, p>0.05; Fig.4A). Compared to VA, VA-A exerted significantly stronger anticonvulsive activity at doses ≥3mg/kg (see Fig.4B, VA-A 3mg/kg: 57.9±1.9mg/kg PTZ, n=4; VA-A 10mg/kg: 55.4±0.7mg/kg PTZ, n=4, p<0.001). Like VA, VA-A also displayed a trend towards reduced activity at higher doses (30mg/kg), however – in contrast to VA – still retained its anticonvulsive potential (Fig.4B, VA-A 30mg/kg: 50.6±2.2mg/kg PTZ, n=3, p<0.01).

A similar (but left-shifted) bell-shaped dose-response curve on PTZ-induced seizure threshold was observed for VA-TET: at a dose of 0.3mg/kg bodyweight (Fig.4C: VA-TET 0.3mg/kg: 47.3±0.5mg/kg PTZ, n=5, p<0.05) VA-TET's anticonvulsive activity was comparable to that of VA at 10-fold higher doses (3mg/kg) indicating a significantly increased potency. However, VA-TET lost its anticonvulsive properties at doses ≥1mg/kg. The methylated VA-A derivative (VA-MA) induced the most pronounced increase in seizure threshold of all tested compounds; first significant effects were observed at a dose of 10mg/kg (Fig.4D: VA-MA 10mg/kg: 50.4±1.4mg/kg PTZ, n=4, p<0.001). In contrast to VA, VA-A and VA-TET, application of higher doses (i.e. 30mg/kg) VA-MA resulted in an even further elevated seizure threshold (VA-MA 30mg/kg: 63.6±2.5mg/kg PTZ, n=3, p<0.001).

As shown in Fig.4F, G and H, higher doses of VA-EA, VA-DEA, and VA-CN were required for seizure threshold elevation similar to that of VA (compare VA 3mg/kg: 47.7±1.4mg/kg PTZ, n=4, Fig.4A vs. VA-EA 30mg/kg: 55.6±0.4 mg/kg PTZ, n=4, Fig.4F vs. VA-DEA:

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30mg/kg: 48.7±1.7mg/kg PTZ, n=3, Fig.4G vs. VA-CN 10mg/kg: 51.1±0.6mg/kg PTZ, n=3, Fig.4H).

Seizure threshold of mice treated with VA-DMA (Fig.4E) did not significantly differ at any tested dose from diluent-treated control littermates.

Effects of VA and derivatives on locomotion in the open field test

As illustrated in Fig.5A, locomotor activity of VA-treated mice did not differ significantly from control animals at any tested dose in the open field test (control: 38.3±1.5m, n=25 vs. VA 1mg/kg: 34.9±1.3m, n=12 vs. VA 3mg/kg: 38.7±2.1m, n=16 vs. VA 10mg/kg: 38.0±1.4 m, n=16 vs. VA 30mg/kg: 37.6±2.5m, n=16).

Application of VA derivative VA-A at doses of 1mg/kg and 3mg/kg, respectively, did also not affect the total distance covered compared to control littermates; however, reduced locomotor activity in the open field test was measured after application of VA-A at doses ≥10mg/kg (control: 38.3±1.5m, n=25 vs. VA-A 10mg/kg: 29.9±2.6m, n=11, p<0.05 vs. VA-A 30mg/kg: 23.5±2.2m, n=18, p<0.001; see Fig.5B).

Like VA-A, derivatives VA-TET (Fig.5C), VA-MA (Fig.5D) and VA-CN (Fig.5E) did not affect locomotor activity at low doses (≤10mg/kg), however at higher doses reduced ambulation was observed for these compounds (control: 38.3±1.5m, n=25 vs. VA-TET 30mg/kg: 19.2±2.1m, n=13, p<0.001 vs. VA-MA 30mg/kg: 29.7±2.0m, n=12, p<0.01 vs. VA-CN 30mg/kg: 31.5±1.5m, n=14, p<0.05).

Finally, analysis of total distance did not reveal any significantly different behavior of animals treated with any dose of the β-subunit-unselective VA derivatives VA-DMA (Fig.5F), VA-EA (Fig.5G) and VA-DEA (Fig.5H) or diluent-treated control animals.

Discussion

Subunit-selective GABA_A receptor modulators represent interesting lead structures for drug development. The natural compound valerenic acid (VA) selectively modulates GABA_A receptors containing either β 2- or β 3-subunits with only residual modulatory activity on GABA_A receptors incorporating β 1-subunits at high concentrations (Khom *et al.*, 2007; Benke *et al.*, 2009). As opposed to benzodiazepines, VA and also its derivatives do not interact with the benzodiazepine binding site (i.e. I_{GABA} enhancement by VA and VA derivatives does not require the presence of a γ 2S-subunit and VA's modulatory action cannot be blocked by the benzodiazepine site antagonist flumazenil (Khom *et al.*, 2007, Khom *et al.*, 2010)). This selectivity profile combined with pronounced anticonvulsive effects (Hintersteiner *et al.*, 2014) accompanied by anxiolytic activity (Khom *et al.*, 2010; Hintersteiner *et al.*, 2014) and devoid of significant impairment of locomotor activity suggesting lack of sedative side effects makes VA and potentially also its derivatives promising drug candidates.

Previously, we and others have reported more efficacious and/or potent modulation of GABA_A receptors expressed in *Xenopus* oocytes by carboxyl-modified VA derivatives (Khom *et al.*, 2010; Kopp *et al.*, 2010). Furthermore, more pronounced anxiolytic effects by the amide derivative of VA (VA-A) (Khom *et al.*, 2010) as well as stronger anxiolytic and anticonvulsive effects of VA ester derivatives compared to VA were observed (Hintersteiner *et al.*, 2014).

To further evaluate the potential of VA and carboxyl-modified derivatives (for structural formulae, see Fig.1) as scaffolds for the development of novel anticonvulsants with limited impairment of locomotor activity, we extended previous *in vitro* studies of these compounds (Khom *et al.*, 2010; Kopp *et al.*, 2010) by determining their β -subunit-dependency of I_{GABA} modulation (α 1 β 1 γ 2S, α 1 β 2 γ 2S, α 1 β 3 γ 2S) followed by a subsequent analysis of their effects on PTZ-induced seizure threshold and locomotion.

VA derivatization has dual effects on β -subunit-selectivity

An interesting finding of the present study was that VA derivatization had - depending on the substituent introduced - dual effects on β -subunit-selectivity: Amidation of VA (VA-A) as well as mono-methylation of the amide (VA-MA), respectively, strongly increased efficacy of I_{GABA} enhancement of β 2- and β 3-containing receptors compared to VA (compare Fig.2A, 2C and 2E), while efficacy of I_{GABA} enhancement through β 1-containing receptors did not differ significantly from that of VA, thus boosting β -subunit-selectivity. Replacing the carboxyl-function by a tetrazole group (VA-TET) significantly increased potency of I_{GABA} enhancement through β 3-containing receptors and efficacy of I_{GABA} of β 2-containing receptors, while modulation of β 1-receptors by VA-TET was comparable to that of VA (compare Figs.2A and 2C). Intriguingly, this structural modification (replacement of the carboxyl-group by a tetrazole moiety) altered current kinetics (see Fig. 2H for typical I_{GABA} in the presence of 10 μ M VA-TET): compared to I_{GABA} modulated by any of the other studied VA derivatives (Figs. 2 and 3, right panels), receptor activation in the presence of VA-TET appeared to be slowed (see longer time required to reach peak currents) and in addition, current desensitization could not be observed even at high drug concentrations. The parent compound VA - at concentrations $\geq 30\mu$ M - has been reported to directly activate $GABA_A$ channels in the absence of GABA (see (Khom *et al.*, 2007). Compared to I_{GABA} , both activation and deactivation rates of these VA-induced currents were remarkably slower. We speculate that VA-TET - representing the more potent bioisoster of VA in terms of modulatory activity (see Table 1 and Fig. 2A and G)- might also directly activate the channel like VA at lower concentrations and that the observed kinetics reflect a combination of direct activation and modulatory activity. However, these differences in current kinetics warrant further investigations.

Substituting the carboxyl-group by a nitrile function (VA-CN) resulted in a similar β 2/ β 3-subunit dependency as observed for VA (Fig.3A). In contrast, introduction of bulkier

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residues such as dimethylamide (VA-DMA), ethylamide (VA-EA) or diethylamide (VA-DEA) did not alter efficacy or potency of I_{GABA} enhancement of $\beta 2$ - or $\beta 3$ -containing receptors compared to VA, but these compounds efficaciously modulated $\beta 1$ -containing receptors indicating a loss of β -subunit-selectivity (see Figs.3B-D).

Evaluation of anticonvulsive effects and potential impairment of locomotor activity by VA derivatives

In line with the stronger I_{GABA} enhancement by VA-A *in vitro*, VA-A also induced a more pronounced seizure threshold elevation compared to VA (compare Fig.4A, 4B and 4D). However, in contrast to VA that did not affect locomotor activity even at doses up to 30mg/kg compared to saline-treated control animals, application of VA-A (≥ 10 mg/kg) significantly reduced locomotor activity. This finding suggests that VA-A, as opposed to VA, also induces sedative effects (compare Figs. 5A and 5B). A similar profile was observed for VA-MA: compared to VA, I_{GABA} enhancement of $\beta 3$ - and $\beta 2$ -containing receptors *in vitro* was more pronounced, and stronger anticonvulsive effects were also accompanied by a reduction of locomotor activity at higher doses (30mg/kg). VA-TET displaying *in vitro* significantly more potent I_{GABA} enhancement of $\beta 3$ -containing receptors (efficacy comparable to VA) and more efficacious I_{GABA} enhancement of $\beta 2$ -receptors, elevated PTZ-induced seizure threshold at 10-fold lower doses than VA (VA-TET: 0.3mg/kg: 47.3 ± 0.5 mg/kg PTZ vs. VA 3mg/kg: 47.7 ± 1.4 mg/kg PTZ (Fig.4C), but also induced the strongest reduction of locomotor activity of all tested VA derivatives at a dose of 30mg/kg (Fig.5C). These presumably sedative effects by VA-TET were, however, observed at doses more than 100-fold higher than those required for anticonvulsive effects (0.3mg/kg; see also Figs. 4C and 5C).

Efficacy and potency of I_{GABA} enhancement ($\alpha 1\beta 1-3\gamma 2S$) by the nitrile derivative (VA-CN) did not significantly differ from that of VA. A trend towards – although not reaching statistical significance – reduced potency for $\beta 1$ - and $\beta 3$ -containing receptors compared to VA was observed (Fig.3A). However, higher doses of VA-CN (10mg/kg) were required for

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seizure threshold elevation (Fig.4H) and as opposed to VA, VA-CN also slightly reduced locomotion at high doses (30 mg/kg; Fig.5E).

Unaffected locomotor activity and either no effect on PTZ-induced seizure threshold or anticonvulsive effects occurring at rather high doses (30mg/kg) were observed for the unselective VA derivatives (i.e. comparable efficacy and potency on $\alpha 1\beta 1-3\gamma 2S$ receptors) VA-EA (Fig.4F and 5G) and VA-DEA (Fig.4G and 5H) and VA-DMA (Figs.4E and 5F).

These findings – $\beta 2/3$ -selective VA derivatives displaying anticonvulsive effects at low doses ($\leq 10\text{mg/kg}$) and sedative effects at higher doses ($\geq 10\text{mg/kg}$) while unselective VA derivatives (VA-EA, VA-DEA) are anticonvulsive at higher doses (30mg/kg) without affecting locomotor activity - prompted us to probe a possible link between β -subunit selectivity and behavioral effects of this set of VA derivatives. Indeed, plotting maximal seizure threshold elevation (maximal effect or the effect reached at a dose of 30 mg/kg) versus the efficacy of I_{GABA} enhancement at different subunit compositions *in vitro* revealed a significant correlation between the compounds' efficacy at $\beta 3$ -containing receptors and anticonvulsive effects (for details, see Supplementary data). Derivatives enhancing I_{GABA} at $\beta 3$ -containing receptors more efficaciously than VA also displayed stronger protection against PTZ-induced seizures ($r=0.8785$; $p<0.01$, see Fig. 1S Supplementary data). Conversely, efficacy at neither $\beta 2$ - nor $\beta 1$ -containing receptors correlated with the extent of seizure threshold elevation. However, VA derivatives modulating $\beta 2$ -containing receptors more efficaciously also induced more pronounced impairment of locomotor activity ($r=-0.9262$; $p<0.001$, Fig. 1S Supplementary data), while neither efficacy nor potency of VA and its derivatives on $\beta 1$ - or $\beta 3$ -containing receptors apparently correlated with occurring sedation. Therefore, it is tempting to speculate that the motor-impairing effects (probably sedation) of VA derivatives are determined by the interaction with $\beta 2$ -containing receptors, while their anticonvulsive effects might be mediated by receptors containing $\beta 3$ -subunits. This hypothesis would also be in line with previous studies highlighting the role of $\beta 2$ -containing receptors in mediating sedative effects of

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GABA_A receptor modulators such as loreclezole (Groves *et al.*, 2006), etomidate (Reynolds *et al.*, 2003; Zeller *et al.*, 2005) or even benzodiazepines (Antkowiak, 2015). In contrast, anticonvulsive effects of GABA_A receptor ligands apparently might result from a broader, less selective GABA_A receptor modulation including more than a single GABA_A receptor subtype: Rudolph *et al.* have shown that anticonvulsive effects of diazepam are only partially blunted in $\alpha 1H101R$ mice indicating that also $\alpha 2$, $\alpha 3$ and/or $\alpha 5$ -containing GABA_A receptors could contribute to anticonvulsive effects *in vivo* (Rudolph *et al.*, 1999; Löw *et al.*, 2000; Crestani *et al.*, 2002; Fradley *et al.*, 2007). In addition, the $\beta 2/3$ -selective GABA_A receptor ligand loreclezole retained – even though reduced- seizure-protecting activity in $\beta 2N265S$ mice suggesting a complementary role for $\beta 3$ -containing receptors in mediating anticonvulsive effects (Groves *et al.*, 2006). Most notably, the expression of $\beta 3$ -subunits is particularly high in dendritic regions of the hippocampus and dentate gyrus (Sperk *et al.*, 1997; Miralles *et al.*, 1999; Hörtnagl *et al.*, 2013). Assuming the fundamental role of the hippocampus in seizure activity and epilepsy (Schwartzkroin, 1994; Coulter *et al.*, 2011), amplifying GABAergic neurotransmission in the hippocampus via $\beta 3$ -containing receptors might represent an appealing approach to develop novel anticonvulsants with reduced side effects including sedation.

However, it cannot be ruled out that the observed differences in effects on PTZ-induced seizure threshold and locomotion might be also due to distinct pharmacokinetic properties (e.g. tissue distribution, penetration of the blood-brain barrier, different onset of *in vivo* effects etc.). To answer this question warrants further studies. Furthermore, interaction of VA and its derivatives with targets for anticonvulsive acting drugs other than GABA_A receptors has not been analyzed in the present study. VA has previously been reported to bind to 5-HT_{5A}-receptors (Dietz *et al.*, 2005) as well as metabotropic group I/II glutamate receptors (Del Valle-Mojica *et al.*, 2011), and to attenuate physical and psychological stress responses by decreasing the turnover of serotonin to 5-hydroxyindoleacetic acid and turnover of

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norepinephrine to 3-methoxy-4-hydroxyphenylethyleneglycol sulfate in rodent hippocampus and amygdala (Jung *et al.*, 2015). Thus, both anticonvulsive and sedative effects of the studied compounds might also result from interactions of VA and its derivatives with multiple drug targets. Benke *et al.* (2009) demonstrated, however, that VA's anxiolytic effect in mice is mediated exclusively by interaction with β 3-subunit-containing GABA_A receptors based on their observation that VA-induced reduction of anxiety-related behavior was completely absent in β 3N265M mice- a point mutation well-known to abolish I_{GABA} enhancement by VA and other β 2/3-selective GABA_A receptor modulators (Groves *et al.*, 2006; Benke *et al.*, 2009). In line with this, the efficacy of the derivatives on GABA_A receptors *in vitro* highly correlates with the strength of anticonvulsive effects *in vivo* (this study), supports the hypothesis that these actions are likely to be mediated via GABA_A receptors.

Taken together, our study demonstrates that modifications of VA's carboxyl function can profoundly alter potency, efficacy and β -subunit-selectivity of VA derivatives: Replacing the carboxyl group by a tetrazole (VA-TET) (see also Kopp *et al.* (Kopp *et al.*, 2010)) as well as amidation (unsubstituted or mono-methylated amide (Khom *et al.*, 2010; Kopp *et al.*, 2010)) significantly increased efficacy and/or potency of I_{GABA} enhancement of β 2/3-containing receptors, and – most importantly – enhanced β 2/3-selectivity. In contrast, introduction of bulkier substituents (as ethyl for VA-EA, di-methyl for VA-DMA, or diethyl for VA-DEA) resulted in significantly increased efficacy on β 1-containing channels and thus abolished β -subunit selectivity. Increased efficacy and/or potency of I_{GABA} enhancement of β 2/3-containing receptors by VA derivatives VA-A, VA-MA and VA-TET is likely to result in more pronounced anticonvulsive effects at low doses (≤ 10 mg/kg), while sedative effects occur at higher doses (≥ 10 mg/kg). In contrast, relatively high doses (30mg/kg) of unselective VA derivatives were required for anticonvulsive effects. Our data thus suggest that VA and β 2/3-selective VA derivatives represent interesting scaffolds for the development of novel anticonvulsants.

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Authorship contributions

Participated in research design:

Khom S., Schwarzer C. and Hering S.

Conducted experiments:

Khom S., Hintersteiner J., Luger D., Schwarzer C.

Contributed new reagents or analytical tools:

Haider M., Pototschnig G., Mihovilovic MD.

Performed data analysis:

Khom S., Luger D., Schwarzer C.

Wrote or contributed to the writing of the manuscript:

Khom S., Luger D., Schwarzer C., Mihovilovic MD., Hering S.

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Footnotes

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Legends for figures

Figure 1

VA and VA derivatives

Structural formulae of studied valerenic acid (VA) and VA derivatives are illustrated.

Figure 2

β -subunit-dependent I_{GABA} enhancement by VA and VA derivatives

Concentration-dependent modulation of GABA_A receptors composed of $\alpha 1\beta 3\gamma 2S$ (dashed line), (■) $\alpha 1\beta 2\gamma 2S$ and (◆) $\alpha 1\beta 1\gamma 2S$ subunits by (A) VA (data for enhancement of $\alpha 1\beta 3\gamma 2S$ receptors taken from (Luger *et al.*, 2015) (C) VA-A, (E) VA-MA, and (G) VA-TET.

Data were fitted by non-linear regression, as described in Materials and Methods (I_{GABA} enhancement by VA-A at 300 μ M (red symbol) was excluded from the fit). Maximal potentiation of I_{GABA} (E_{max}), EC_{50} values, Hill-coefficients (n_H) and number of experiments for each compound on $\alpha 1\beta 1\gamma 2S$, $\alpha 1\beta 2\gamma 2S$ and $\alpha 1\beta 3\gamma 2S$ receptors are summarized in Table 1. Each data point represents a mean \pm S.E.M from at least 3 different oocytes from 2 different frog batches. I_{GABA} potentiation at 300 μ M ($\alpha 1\beta 3\gamma 2S$ receptors, grey symbols) for VA and VA-A, respectively, were excluded from the fit. Typical traces for the enhancement of GABA-induced chloride currents (I_{GABA} , EC_{3-7} , single bar) by 10 μ M of (B) VA, (D) VA-A, (F) VA-MA and (H) VA-TET (double bar; indicating co-application of GABA and compound) at the indicated GABA_A receptor subtype are illustrated.

Figure 3

β -subunit-dependent I_{GABA} modulation by VA derivatives

Concentration-response curves for the I_{GABA} enhancement through (●) $\alpha 1\beta 3\gamma 2S$, (■) $\alpha 1\beta 2\gamma 2S$ and (◆) $\alpha 1\beta 1\gamma 2S$ channels by VA derivatives (A) VA-CN, (B) VA-EA, (C) VA-DMA and

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(D) VA-DEA. Data were fitted by non-linear regression, as described in Materials and Methods. Maximal potentiation of I_{GABA} (E_{max}), EC_{50} values, Hill-coefficients (n_H) and number of experiments for each compound on $\alpha 1\beta 1\gamma 2S$, $\alpha 1\beta 2\gamma 2S$ and $\alpha 1\beta 3\gamma 2S$ receptors are summarized in Table 1. Each data point represents a mean \pm S.E.M from at least 3 different oocytes from 2 different frog batches. Typical current traces for the enhancement of GABA-induced chloride currents (I_{GABA} , EC_{3-7} , single bar) by 10 μ M of **(B)** VA-CN, **(D)** VA-EA, **(F)** VA-DMA and **(H)** VA-DEA (double bar; indicating co-application of GABA and compound) at the indicated $GABA_A$ receptor subtype are illustrated.

Figure 4

Anticonvulsive effects of VA and VA derivatives

Elevation of seizure threshold upon tail-vein infusion of PTZ 30 min after i.p. application of **(A)** VA is illustrated. The dotted line represents the averaged seizure threshold of saline-treated control animals. Effect on PTZ-induced seizure threshold 30 min. after i.p. application by the VA derivatives **(B)** VA-A, **(C)** VA-TET, **(D)** VA-MA, **(E)** VA-DMA, **(F)** VA-EA, **(G)** VA-DEA and **(H)** VA-CN is compared to VA (dashed line, white circles) and control animals (dotted line). Each data point represents a mean \pm S.E.M from at least 3 mice. Statistical significance (p -values <0.05 were accepted as significant; $*=p<0.05$, $**=p<0.01$ and $***=p<0.001$) against VA was calculated by one-way ANOVA followed by a Bonferroni post-hoc mean comparison.

Figure 5

Effects on locomotion by VA and VA derivatives in the open field test (OF) test

Black bars indicate the total distance covered in the OF test 30 min after i.p. application of **(A)** VA **(B)** VA-A, **(C)** VA-TET, **(D)** VA-MA, **(E)** VA-CN, **(F)** VA-DMA, **(G)** VA-EA and **(H)** (VA-DEA) at the indicated doses compared to saline-treated control animals (white bars

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in all panels). Each bar represents a mean \pm S.E.M from at least 10 different mice. Statistical significance (p-values <0.05 were accepted as significant; $*=p<0.05$ and $***=p<0.001$) against saline-treated control animals was calculated by one-way ANOVA followed by a Bonferroni post-hoc mean comparison.

Table 1

Summary of pharmacological parameters of I_{GABA} enhancement through $\alpha_1\beta_1\gamma_2S$, $\alpha_1\beta_2\gamma_2S$ and $\alpha_1\beta_3\gamma_2S$ receptors including maximal I_{GABA} enhancement (E_{max}), half-maximal effective concentrations (EC_{50}), Hill-coefficients (n_H) and number of experiments for each compound at the tested subunit combinations. Data for I_{GABA} enhancement of $\alpha_1\beta_3\gamma_2S$ receptors by VA at GABA EC_{3-7} concentrations taken from (Luger *et al.*, 2015).

$\alpha_1\beta_1\gamma_2S$				
	E_{max} (%)	EC_{50} (μM)	n_H	n
VA	111±16	74.4±19.3	1.6±0.5	8
VA-A	218±78	66.6±34.6	1.8±0.6	6
VA-MA	387±56	58.1±16.5	1.6±0.3	5
VA-DMA	305±67	52.5±20.1	1.7±0.5	6
VA-EA	458±124	51.4±19.8	1.6±0.3	5
VA-DEA	318±84	97.2±33.8	1.8±0.3	7
VA-CN	55±14	73.0±38.7	1.7±0.7	7
VA-TET	176±43	23.5±10.9	1.8±0.6	7
$\alpha_1\beta_2\gamma_2S$				
	E_{max} (%)	EC_{50} (μM)	n_H	n
VA	721±68	23.1±4.2	1.4±0.2	5
VA-A	1119±72	14.0±2.2	1.4±0.2	6
VA-MA	917±36	9.1±1.4	1.5±0.1	3
VA-DMA	594±36	63.9±23.7	1.3±0.2	5
VA-EA	554±49	11.0±3.6	1.6±0.3	3
VA-DEA	573±33	54.4±11.1	1.6±0.3	5

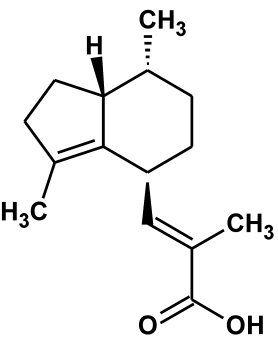
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VA-CN	765±117	56.5±13.3	2.5±0.7	4
VA-TET	1091±87	34.1±6.3	1.0±0.1	5

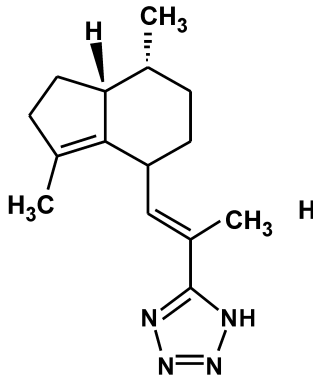
$\alpha 1\beta 3\gamma 2S$

	E_{max} (%)	EC₅₀ (μM)	n_H	n
VA	632±88	20.2±5.2	1.5±0.3	9
VA-A	972±69	7.5±1.8	1.5±0.2	6
VA-MA	1043±57	12.7±0.9	1.5±0.1	6
VA-DMA	415±61	48.0±16.0	1.4±0.2	6
VA-EA	677±64	27.2±6.7	1.3±0.2	12
VA-DEA	374±102	80.8±31.3	1.9±0.5	5
VA-CN	522±114	42.4±15.8	2.1±0.8	7
VA-TET	668±57	6.0±1.0	1.1±0.1	8

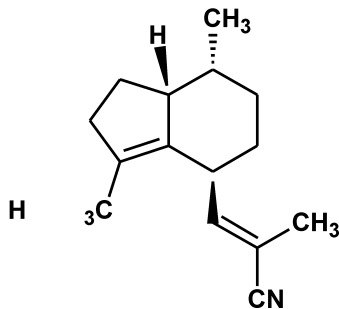
Figure 1



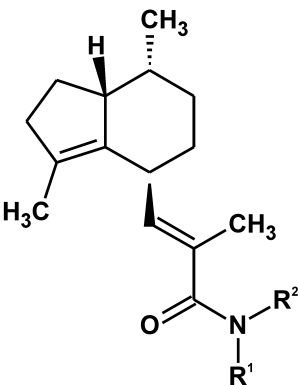
VA



VA-TET



VA-CN



Code	R ¹	R ²
VA-A	H	H
VA-MA	CH ₃	H
VA-DMA	CH ₃	CH ₃
VA-EA	CH ₂ -CH ₃	H
VA-DEA	CH ₂ -CH ₃	CH ₂ -CH ₃

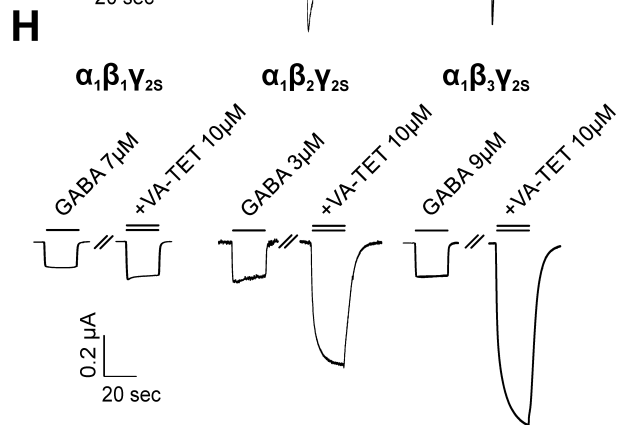
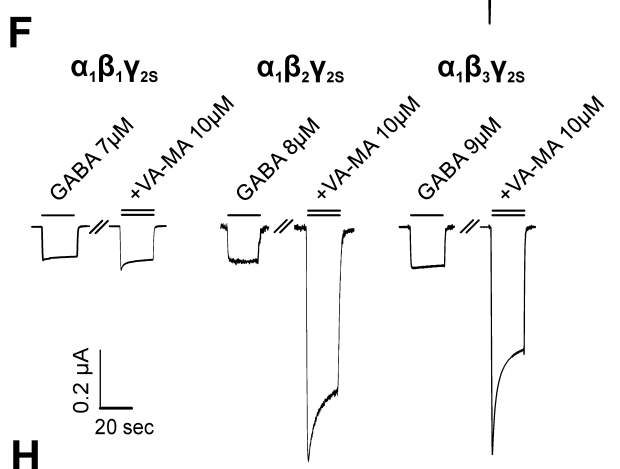
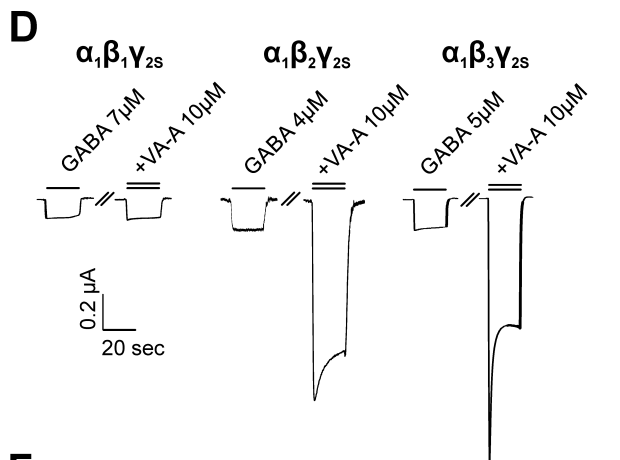
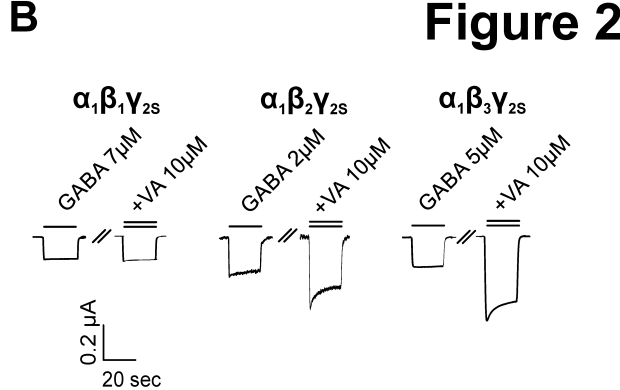
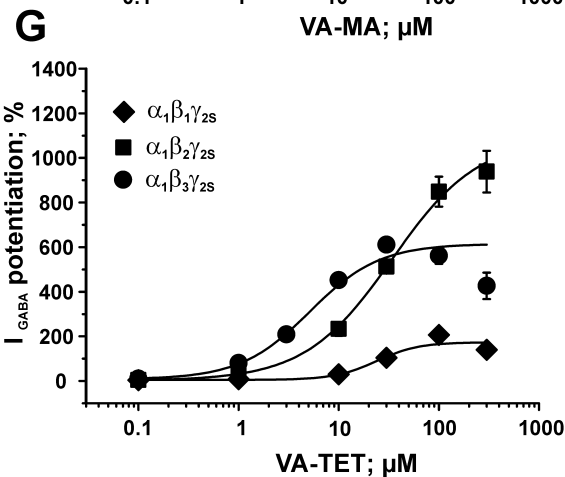
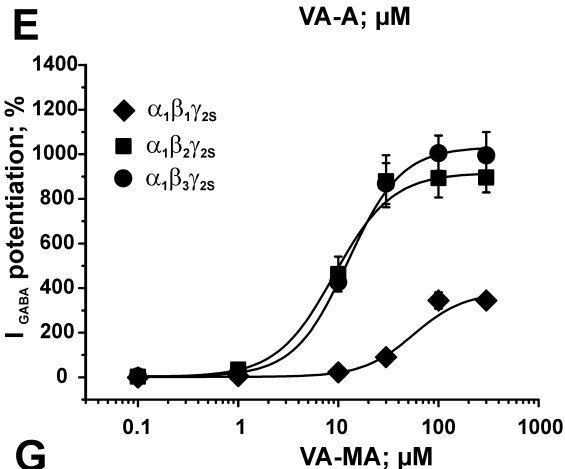
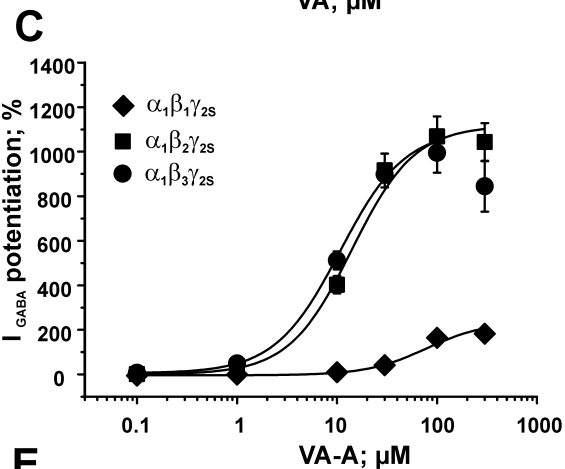
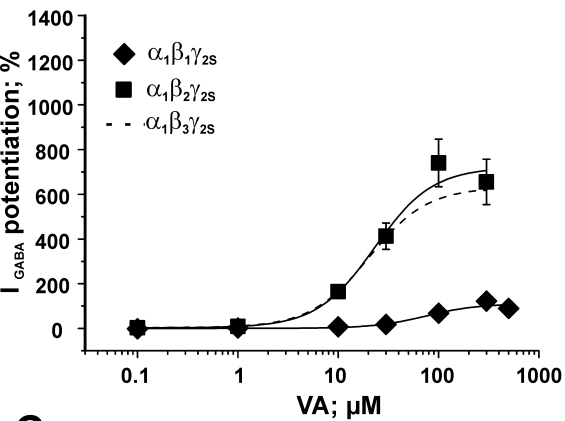
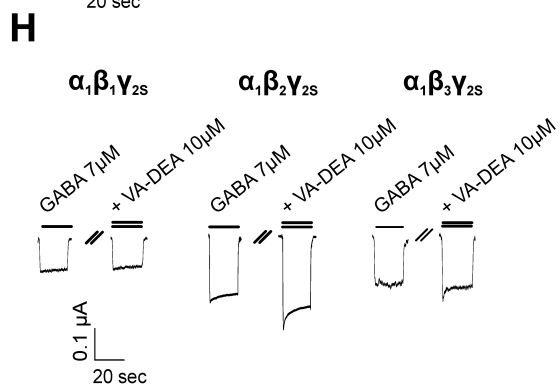
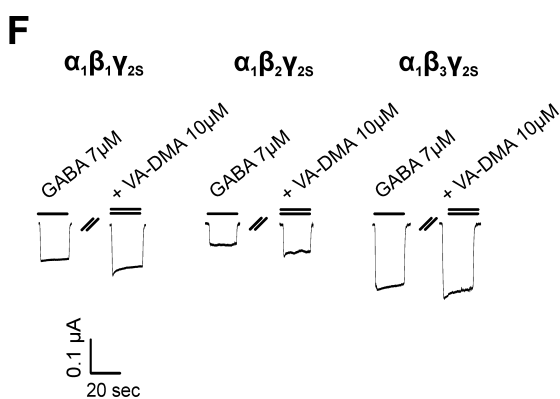
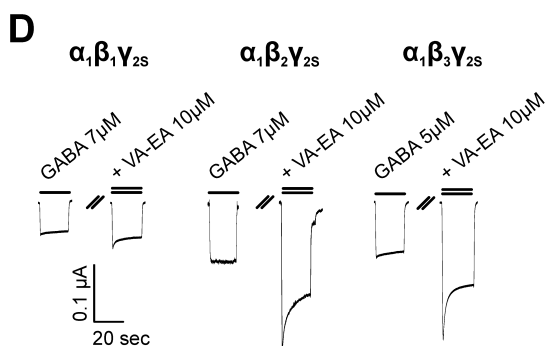
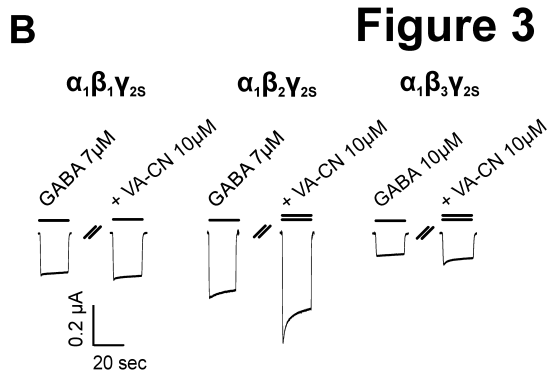
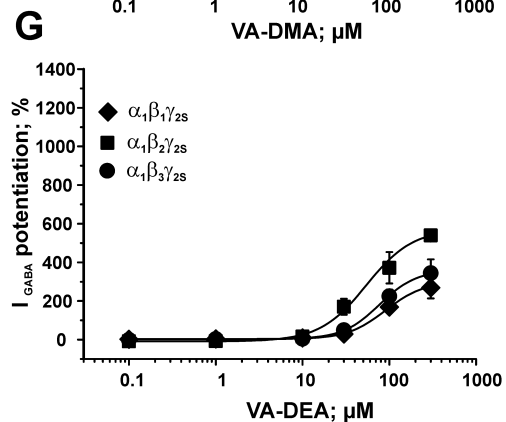
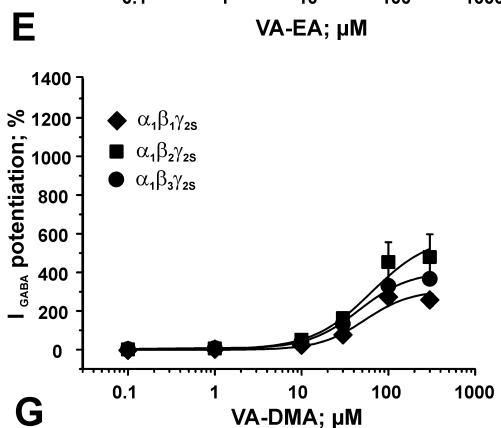
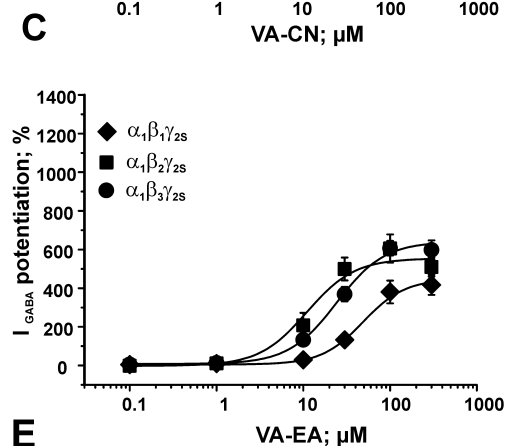
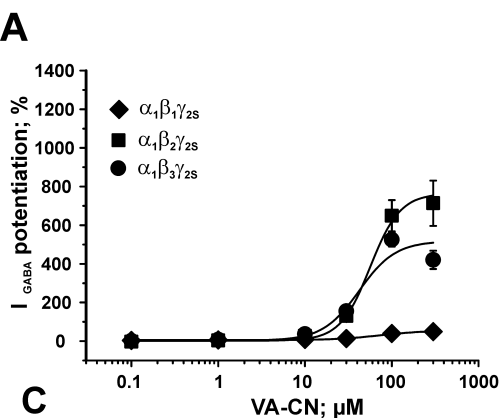
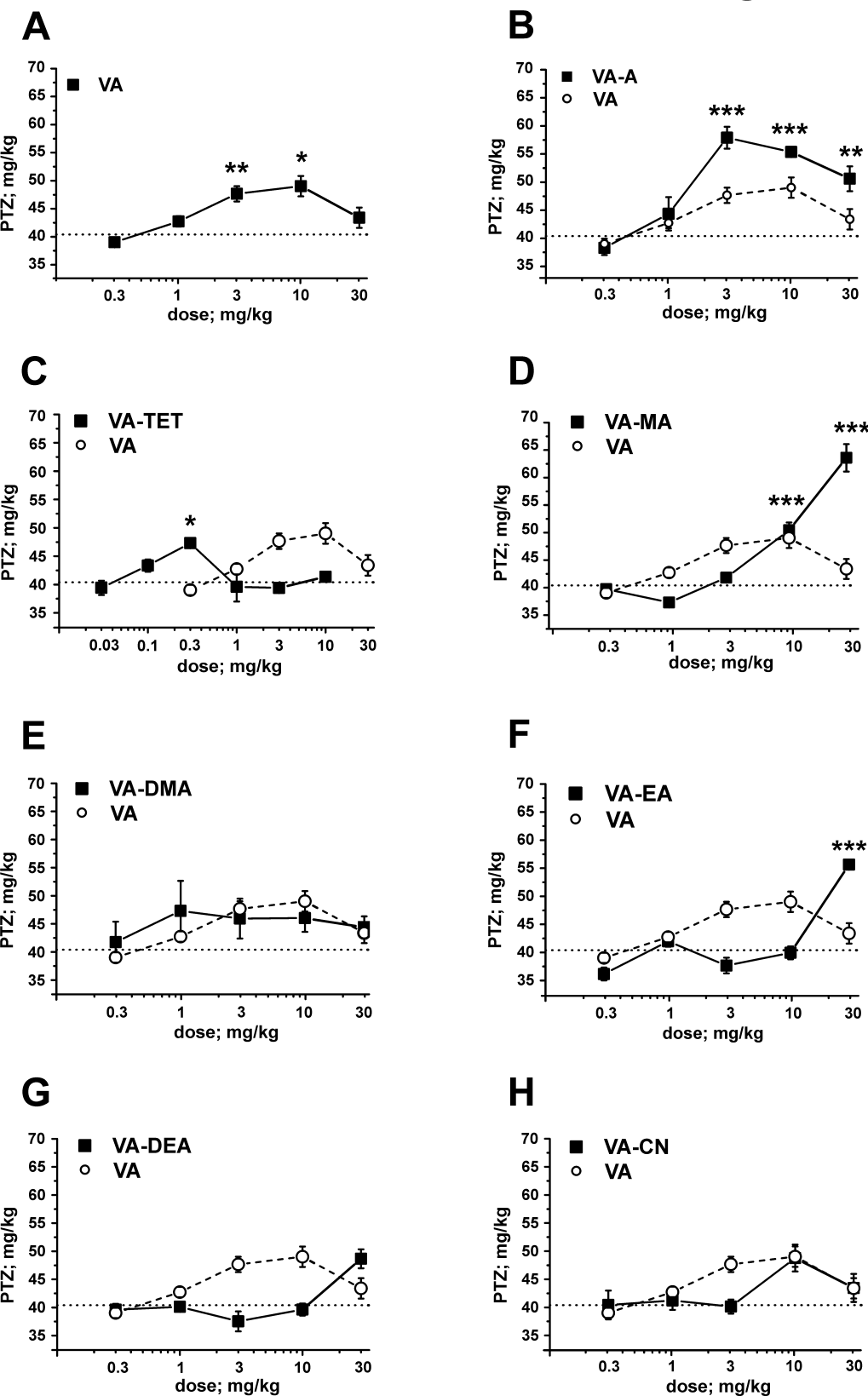
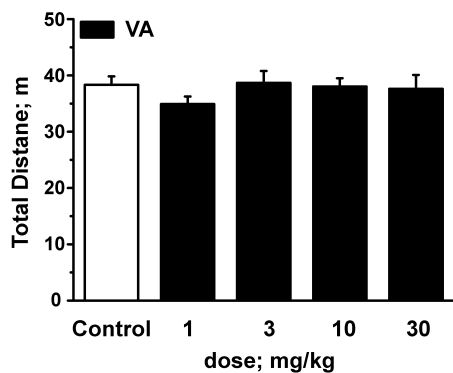
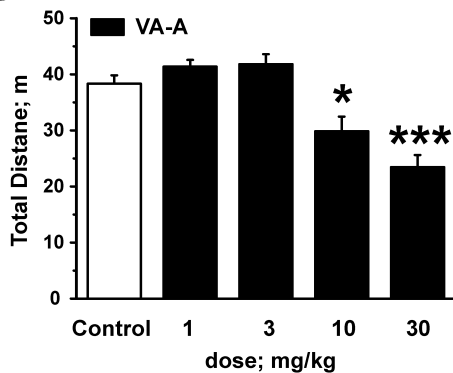
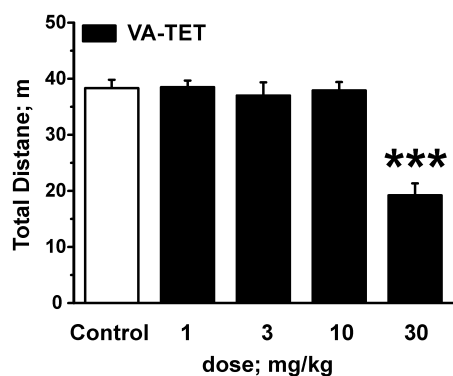
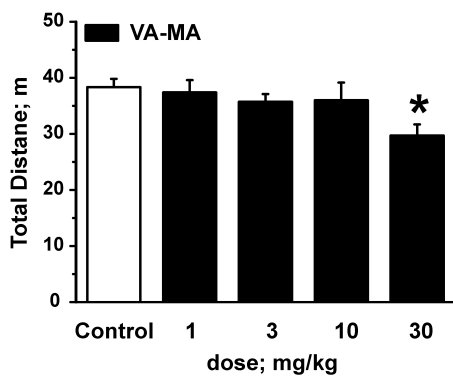
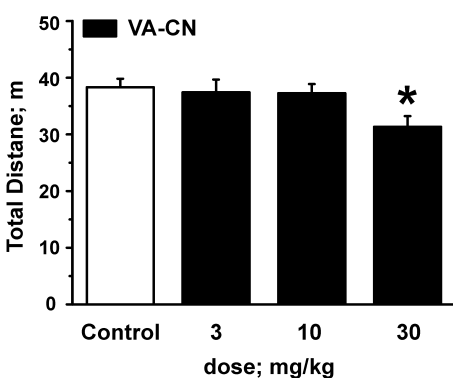
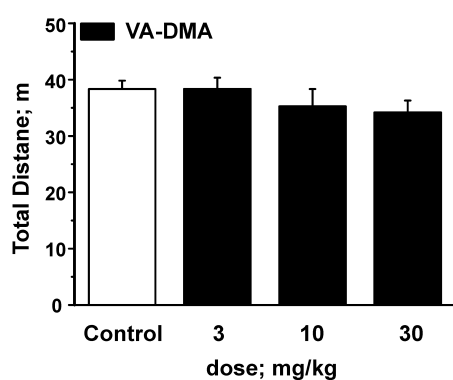
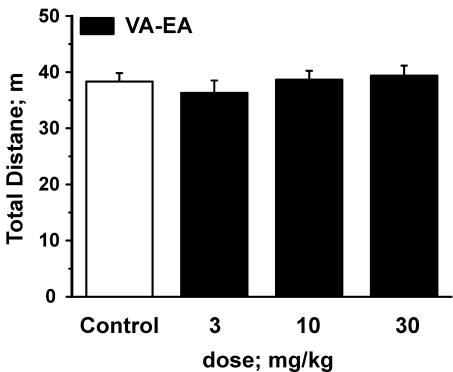


Figure 3





A**B****C****D****E****F****G****H**