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The First ‘Universal Opioid Ligand’ (2S)-2-[(5R,6R,7R,14S)-N-cyclopropylmethyl-4,5-epoxy-6,14-ethano-3-hydroxy-6-methoxymorphinan-7-yl]-3,3-dimethylpentan-2-ol (BU08028): Characterization of the In Vitro Profile and In Vivo Behavioral Effects in Mouse Models of Acute Pain and Cocaine-induced Reward.

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Characterization of BU08028

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Abbreviations: CPP, conditioned place preference; PC, place conditioning; N/OFQ, nociceptin/orphanin FQ; DMEM, Dulbecco's Modified Eagle's Medium; MPE, Maximal Percent Effect;

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ABSTRACT

Certain behavioral features of buprenorphine, including a bell-shaped curve for antinociception and attenuation of alcohol consumption, are thought to be mediated by activation of NOP receptors, despite moderate affinity and low efficacy at NOP receptors. We hypothesized that ligands with buprenorphine's physical properties, but possessing increased NOP receptor affinity and efficacy would improve the profile as a drug abuse medication and reduce addiction liability. Using this strategy, we designed several compounds with universally high affinity, i.e., less than 10 nM at mu, delta, kappa, and NOP receptors. Among these, BU08028 has high affinity at all opioid receptors and increased NOP receptor efficacy in vitro in the [³⁵S]GTPγS binding assay, however still being a partial agonist. In vivo, BU08028 was evaluated in an acute thermal antinociception assay, for its ability to induce conditioned place preference (CPP), and for its effect on cocaine-induced CPP. BU08028 is a very potent long lasting analgesic, it produces an increase in locomotor activity, and produces a significant CPP. As a pretreatment to cocaine, BU08028 does not alter cocaine CPP but causes a further increase in cocaine-induced locomotor activity. The analgesic, rewarding, and stimulant effects are likely due to mu receptor stimulation. It is likely that with BU08028, a partial agonist at both NOP and mu receptors, mu-mediated activity overpower NOP-mediated effects. Thus, it is possible that a different buprenorphine analog that is a universal high affinity opioid ligand but with 'full agonist' activity at NOP may counteract traditional opioid-mediated effects such as antinociception and reward.

INTRODUCTION

Buprenorphine has been a successful analgesic and drug abuse medication. It is a potent long-lasting partial agonist with reduced addiction liability, relative to morphine (Jasinski et al., 1978), and is currently used for opiate maintenance as an alternative to methadone. Although it has reduced addiction liability, buprenorphine has been classified a schedule III compound by the Drug Enforcement Agency. Animal models of abuse liability of buprenorphine have been somewhat contradictory; with some experiments suggesting that buprenorphine is rewarding, and other suggesting that it is not (Kosten et al., 1991; Rowlett et al., 1994). Regardless of its own rewarding potential, buprenorphine administration diminishes cocaine self-administration both in animal models and in people (Mello et al., 1989; Montoya et al., 2004), a feature attributed to the mu partial agonist component of its profile (Mello et al., 1993).

Buprenorphine can also display an unusual dose response relationship. In antinociception experiments, it can have an inverted U-shaped dose-response curve, with high doses inducing a decrease in antinociceptive activity (Lutfy et al., 2003). A similar phenomenon has been reported when measuring the ability of buprenorphine to modify ethanol consumption in rats. Buprenorphine increases alcohol consumption at low doses but attenuates consumption at higher doses (Ciccocioppo et al., 2007). In both cases, the high dose reduction in antinociception and alcohol consumption is attributed to activation of NOP receptors (formerly known as ORL1), thereby diminishing the mu-mediated antinociceptive activity and mu-mediated increase in alcohol consumption. This is despite the fact that the affinity of buprenorphine for NOP receptors is modest, and its efficacy at this receptor is extremely low (Spagnolo et al., 2008; Khroyan et al., 2009).

The NOP receptor belongs to the opioid family and binds the endogenous ligand nociception/orphanin FQ (N/OFQ). N/OFQ, through NOP-receptor activation, has been shown to have anti-opiate properties in the brain (Meunier et al., 1995; Reinscheid et al., 1995) and is able to block the reward induced by morphine (Murphy et al., 1999), cocaine (Kotlinska et al., 2002; Sakoori

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and Murphy, 2004), ethanol (Ciccocioppo et al., 1999; Kuzmin et al., 2003), and methamphetamine (Zhao et al., 2003). N/OFQ, when administered icv, also has been shown to block development of tolerance to morphine (Lutfy et al., 2001). These studies, as well as our previous work (Toll et al., 2009), suggest that a compound that activates both the mu-opioid and NOP receptors might be an ideal candidate for a nonaddicting analgesic with reduced tolerance development, and may also have potential efficacy for the treatment of drug addiction. However, the optimum profile of mu and NOP receptor affinity and/or efficacy for an optimum balance of analgesia to antagonism, reward to reward-attenuation, locomotor activation to sedation, for a useful therapeutic profile cannot yet be predicted. We have demonstrated that such mixed NOP/mu compounds can be developed with significant antinociceptive activity and greatly reduced reward, but to attenuate reward in a traditional NOP receptor non-opioid small molecule pharmacophore (see Zaveri et al., 2005), the NOP component must overshadow the mu component with respect to affinity and/or efficacy (Khroyan et al., 2007; Toll et al., 2009). This is not the case with buprenorphine, in which the mu affinity is far greater than the NOP receptor affinity. If beneficial actions of buprenorphine are due to NOP receptor activation, it seems reasonable that a buprenorphine analog with higher affinity and efficacy at NOP would have an in vivo profile preferable to that of buprenorphine itself, with reduced reward and reduced tolerance development. We applied this hypothesis to the buprenorphine class of compounds, and have designed buprenorphine analogs that have higher NOP affinity and efficacy. Here we describe in vitro profile and the behavioral effects of a buprenorphine analog, BU08028, the first “universal opioid ligand” with high affinity at all four opioid family receptors.

METHODS

Drugs. BU08028 (Figure 1) was prepared using the standard techniques for orvinol synthesis (Bentley and Hardy, 1967; Grundt et al., 2003). Treatment of N-cyclopropylmethylidihydronorthevinone with freshly prepared t-pentyl magnesium bromide and subsequent 3-O-demethylation using sodium propane thiolate gave BU08028, which was converted to

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its hydrochloride salt. For behavioral experiments, BU08028 was dissolved in 1-2% DMSO and 0.5% aqueous hydroxypropylcellulose. Morphine sulfate (Lilly, Indianapolis, IN), and buprenorphine HCl (NIDA drug supply) were dissolved in water. Drugs were injected in a volume of 0.1 ml/25g (s.c.). Controls received 0.1 ml/25g of the appropriate vehicle.

Animals. Male ICR mice weighing 25-30g at the start of the experiment were used. Animals were group-housed under standard laboratory conditions and were kept on a 12:12-hr day/night cycle (lights on at 7:00am). Animals were handled for 3-4 days before the experiments were conducted. On behavioral test days, animals were transported to the testing room and acclimated to the environment for 1 hr. Mice were maintained in accordance with the guidelines of SRI International and of the *Guidelines for the Care and Use of Mammals in Neuroscience and Behavioral Research* (National Research Council, 2003).

Experimental Procedures

In vitro Characterization

Cell Culture. All receptors were individually expressed in CHO cells stably transfected with human receptor cDNA, developed in our laboratory. Receptor expression levels were 1.2, 1.6, 1.8, and 3.7 pmol per mg protein for the NOP, mu, kappa, and delta opioid receptors respectively. The cells were grown in Dulbecco's Modified Eagle Medium (DMEM) with 10% fetal bovine serum, in the presence of 0.4 mg/ml G418 and 0.1% penicillin/streptomycin, in 100-mm plastic culture dishes. For binding assays, the cells were scraped off the plate at confluence.

Receptor Binding. Binding to cell membranes was conducted in a 96-well format, as described previously (Dooley et al., 1997; Toll et al., 1998). Cells were removed from the plates by scraping with a rubber policeman, homogenized in 50 mM Tris pH 7.5, using a Polytron homogenizer, then centrifuged once and washed by an additional centrifugation at 27,000 x g for 15 min. The pellet was resuspended in Tris, and the suspension incubated with [³H]DAMGO (51 Ci/mmol, 1.6 nM), [³H]Cl-

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DPDPE (42 Ci/mmol, 1.4 nM), [³H]U69593 (41.7 Ci/mmol, 1.9 nM), or [³H]N/OFQ (120 Ci/mmol, 0.2 nM) for binding to, mu, delta, kappa and NOPr respectively. Non-specific binding was determined with 1 μM of unlabeled DAMGO, DPDPE, ethylketocyclazocine, and N/OFQ respectively. Total volume of incubation was 1.0 ml and samples were incubated for 60 min at 25°C. The amount of protein in the binding assay was 15 μg. The reaction was terminated by filtration using a Tomtec 96 harvester (Orange, CT) through glass fiber filters. Bound radioactivity was counted on a Pharmacia Biotech beta-plate liquid scintillation counter (Piscataway, NJ) and expressed in counts per minute. IC₅₀ values were determined using at least six concentrations of each peptide analog, and calculated using Graphpad/Prism (ISI, San Diego, CA). K_i values were determined by the method of Cheng and Prusoff (Cheng and Prusoff, 1973).

[³⁵S]GTPγS binding. [³⁵S]GTPγS binding was conducted basically as described by Traynor and Nahorski (Traynor and Nahorski, 1995). Cells were scraped from tissue culture dishes into 20 mM Hepes, 1 mM EDTA, then centrifuged at 500 x g for 10 min. Cells were resuspended in this buffer and homogenized using a Polytron Homogenizer. The homogenate was centrifuged at 27,000 x g for 15 min, and the pellet resuspended in Buffer A, containing: 20 mM Hepes, 10 mM MgCl₂, 100 mM NaCl, pH 7.4. The suspension was recentrifuged at 27,000 x g and suspended once more in Buffer A. For the binding assay, membranes (8-15 μg protein) were incubated with [³⁵S]GTPγS (50 pM), GDP (10 μM), and the appropriate compound, in a total volume of 1.0 ml, for 60 min at 25°C. Samples were filtered over glass fiber filters and counted as described for the binding assays. Statistical analysis was conducted using the program Prism.

In vivo Characterization

Assessment of Thermal Nociception

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Tail-Flick Assay. Acute nociception was assessed using the tail flick assay with an analgesia instrument (Stoelting) that uses radiant heat. This instrument is equipped with an automatic quantification of tail flick latency, and a 15 sec cutoff to prevent damage to the animal's tail. During testing, the focused beam of light was applied to the lower half of the animal's tail, and tail flick latency was recorded. Baseline values for tail flick latency were determined before drug administration in each animal. The mean basal tail flick latency was 4.32 ± 0.12 SEM.

Following baseline measures, animals received a s.c. injection of their assigned dose of drug and were tested for tail flick latencies at 30-, 60-, 120-, and 240-min post-injection. The animals were then taken back to their home room and retested 24hr and 48 hr following the injection. Controls received an injection of vehicle prior to testing.

Drug Regimen and Acute Testing. Separate groups of animals (N=8-14/group) received s.c. injections of BU08028 (0.3-30 mg/kg). Since BU08028 is structurally related to buprenorphine, a separate group of animals received s.c. injections of buprenorphine (0.1-10 mg/kg). As positive controls, separate groups of animals received injections of morphine (10.0 mg/kg). Following assessment of baseline values for tail flick latency, animals received an injection of their assigned dose/drug and were tested following the paradigm described above. A group of animals also served as vehicle controls.

In follow-up experiments, we examined whether BU08028-induced antinociception was altered following co-administration with the opiate antagonist naloxone (1mg/kg) or the NOP antagonist SB 612111 (10mg/kg). Doses of the antagonists were chosen based on previous experiments in our laboratory.

Drug Regimen and Tolerance Testing. To assess tolerance development, separate groups of animals (N=8-9/group) received either s.c. injections of vehicle, morphine (10 mg/kg), BU08028 (10 mg/kg), or buprenorphine (10 mg/kg) once daily for 9 days. The 10 mg/kg dose of BU08028 was

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chosen because it produced near maximal levels of %MPE 1 hour following injection. The 10 mg/kg dose of morphine was chosen because it produced similar levels of antinociception as 10 mg/kg BU08028. Finally, 10 mg/kg buprenorphine was used because it produced the highest level of antinociception. This regimen has been used previously in our laboratory (Khroyan et al., 2007). Animals received daily injections of their respective drug and on days 1, 2, 4, and 9 were tested for tail-flick latency 1 hour following their injection using the equipment and methods described above. The mean \pm S.E.M. basal tail-flick latencies were consistent across test days. The 1hr time point was chosen because it captured the maximal effects of BU08028, morphine, and buprenorphine.

Statistical Analyses. Antinociception (% Maximum Potential Effect; % MPE) was quantified by the following formula: $\% \text{ MPE} = 100 * [(\text{test latency} - \text{baseline latency}) / (15 - \text{baseline latency})]$. If the animal did not respond prior to the 15-s cutoff, the animal was assigned a score of 100%. Behavioral results were analyzed using repeated measures ANOVAs with drug treatment as between group variables and post drug-injection time (30-, 60-, 120-, and 240-min; and 24- and 48-hr for acute injection) and drug injection day (1, 2, 4, 9 for tolerance experiments) as repeated measures followed by Student Newman-Keuls post-hoc tests where appropriate. The level of significance was set at $P < 0.05$.

Assessment of Acute Rewarding Effects using the Place Conditioning (PC) Paradigm

PC Apparatus. The apparatus consisted of rectangular Plexiglas chambers divided into two distinct equal-sized compartments (19cm x 22.8cm x 18cm high, Lafayette Instruments). One compartment had cedar-scented bedding underneath a bar grid floor and all but the front walls were black. The other compartment had pine-scented bedding beneath a mesh floor and all but the front wall were white. The front walls were transparent so that the animal's behavior could be monitored. A removable partition divided the two compartments. During conditioning, the compartments were divided by a solid partition. On the PC test day, the solid partition was replaced with a partition that

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had an opening, allowing the animal free access to both compartments. The apparatus is unbiased, as untreated animals do not show a preference for one compartment over the other (Figures 6 and 7).

PC Training. Because of the long-lasting antinociceptive activity of BU08028, we initially used a single trial PC paradigm with conditioning trials separated by 48 hours to allow the drug to have limited behavioral actions. A conditioning trial was composed of two sessions conducted over three consecutive days. During the drug session, animals received a s.c. injection of their respective drug and were confined to one of the compartments for 60 min. 48 h later, the vehicle session, animals received an injection of vehicle and were confined to the alternate compartment for 60 min. The particular compartment paired with the drug and the order of the drug versus the saline session was counterbalanced across groups. Although this single trial PC paradigm was used to prevent BU08028's rewarding effects from being present during the vehicle training session, it should be noted that the rewarding and antinociceptive effects of a drug may not have the same onset and duration.

Assessment of Global Activity. During conditioning, overall activity of the animals after acute drug injection was recorded. These data were captured by the Spontaneous Motor Recording and Tracking software system (SMART; Panlab, Barcelona, Spain), a color image-capturing system that works in real time and tracks all the movements of the animal, for a given amount of time, via a video camera connected to the computer. This system tracks and records all behavior that results in any movement/positional changes. We term this as "global activity," because it encompasses all movement detected including fine movement, movement due to rearing, grooming, sniffing, and locomotor activity.

PC Test Day. 48 hr following the last conditioning session, animals were given access to both compartments simultaneously for 15 min and the amount of time animals spent in each compartment was recorded.

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Drug Regimen. Animals were assigned to groups (N=8/group) receiving s.c. injections of BU08028 (1-10 mg/kg), buprenorphine (1-10 mg/kg), or morphine (15 mg/kg) alone. During the drug session, animals were immediately placed in the drug-paired compartment for 60 min following their s.c. injection. A group of mice received vehicle in both compartments and served as controls.

Statistical Analyses. Global activity was analyzed using a one way ANOVA with drug as the between-subject measure. For the PC test day data, the percent time animals spent in their drug-paired compartment was analyzed using one way ANOVAs and significant effects were further analyzed with post-hoc tests. A conditioned place preference (CPP) was evident if animals spent significantly more time in their drug-paired compartment relative to control animals. The level of significance was set at $P < 0.05$.

Effect of Repeat Administration of BU08028 on Reward and on Cocaine-Induced CPP

Testing and Drug Regimen. To assess whether repeated injections of BU08028 alone would also induce CPP and whether it would alter cocaine-induced CPP, we used a more traditional PC conditioning regimen. The same experimental apparatus and methods as described above were used except for the following: Conditioning trials were composed of two sessions conducted over *two* consecutive days. The drug session and vehicle session were 24 hours apart and animals were confined to their respective compartments for 30min. These two sessions were repeated over 8 consecutive days such that animals received four drug sessions and four vehicle sessions. Global activity was assessed following the first and fourth drug injection using the methods described above. Because increases in global activity produced by one injection of BU08028 had disappeared within 24 hr and were similar to controls (data not shown) we opted to conduct the drug and vehicle sessions over two consecutive days as opposed to every other day as performed in the acute PC experiment described above.

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Animals were assigned to groups (N=6-9/group) receiving s.c. injections of either vehicle, BU08028 (0.3-1 mg/kg) alone, cocaine (30mg/kg) alone or cocaine co-administered with BU08028 (0.1-1 mg/kg). On drug days, animals received their respective injection of drug (or vehicle) and were immediately placed in the compartment for 30min. PC test day was conducted 24hr following the last conditioning session.

Statistical Analyses. Global activity was analyzed using a repeated ANOVA with drug as the between-subject measure and injection day (first versus fourth) as the repeated measure. PC data was analyzed in the same way as described above.

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RESULTS

Receptor and [³⁵S]GTP γ S Binding. BU08028 (Figure 1) is an orvinol with structural similarity to buprenorphine. As seen in Table 1, BU08028 has a K_i of less than 10 nM at each receptor in the opioid receptor family, making it the first universally high affinity opioid ligand. Affinity is highest at the delta receptor (K_i = 1.6 nM), but it has a K_i within a factor of 5 at all four opioid receptors, NOP, mu, delta, and kappa. More importantly, even though the structure is very similar to that of buprenorphine, its affinity at NOP is almost 10 fold higher than that of buprenorphine itself (Table 1). In vitro functional activity, as determined by [³⁵S]GTP γ S binding (Table 2), indicated that BU08028 has virtually no efficacy at delta and kappa receptors. At mu receptors, the functional activity is very similar to buprenorphine. It is slightly more potent, with an EC_{50} of 6.03 nM, but it is a weak partial agonist with only 21% stimulation relative to DAMGO, very similar to that of buprenorphine. At NOP, the EC_{50} is 43.5 nM, approximately 6 times more potent than buprenorphine, and has significantly higher efficacy showing 44% stimulation, compared to buprenorphine, which stimulates only in the range of 15%. These results suggest that the behavioral actions produced by BU08028 administration should have significantly more NOP character than buprenorphine.

Acute Effects of BU08028 on Tail Flick Latency: Comparison with Morphine and

Buprenorphine. The effect of BU08028 on tail-flick latency is shown in Figure 2A. Initial experiments demonstrated that BU08028 has a long lasting effect on tail flick latency, which was therefore assessed over a 2-day period. The overall ANOVA indicated that there was a significant interaction effect [$F(30,235)=5.4$, $P<0.05$]. The positive control morphine produced the anticipated increase in %MPE at the earlier time points of 30-, 60-, 120-, and 240-min post-injection. Morphine antinociception was not evident at the 24- and 48-hr time points. BU08028, at 3, 10 and 30 mg/kg, produced a dose dependent increase in %MPE that was significantly different from vehicle controls up to 24-hr post-injection (Figure 2A). BU08028 was longer lasting, but also slower acting than

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morphine, but by 240-min post-injection all the doses of BU08028 (1-30mg/kg) that were antinociceptive produced a similar increase in %MPE as morphine.

In order to compare the two drugs, we also examined the acute effects of buprenorphine using the same procedure as with BU08028. The effect of buprenorphine on tail-flick latency is shown in Figure 2B. The overall ANOVA indicated that there was a significant interaction effect [$F(15,160)=6.23$, $P<0.05$]. The doses of 0.1-10 mg/kg produced a significant increase in %MPE at the 30-240-min time points relative to vehicle controls. However, these doses of buprenorphine did not produce an increase in %MPE at 24- and 48-hr post-injection. Thus, although buprenorphine produced antinociceptive effects at lower doses relative to BU08028, its effects were neither as great nor long lasting as BU08028. As described previously, buprenorphine-induced antinociception produced a very shallow dose response curve and never reached maximal tail flick latency.

Enhancement of BU08028-induced antinociception by Pretreatment with SB612111 and

Reversal by Pretreatment with Naloxone. The effect of the NOP antagonist SB612111 on BU08028-induced antinociception is shown in Figure 3A-D. BU08028 was tested at low and moderate doses (0.3 to 3.0 mg/kg) to determine whether its NOP agonist activity was attenuating its MOP-mediated antinociceptive activity. The overall ANOVAs indicated a significant interaction [$F(9,216)=2.22$, $P<0.05$] showing that depending on the post-injection time and dose of BU08028, SB612111 had a differential effect. The NOP antagonist SB612111 potentiated BU08028-induced antinociception, however, this potentiation was observed at the 3 mg/kg dose of BU08028 and was statistically significant only at 120- and 240-min post-injection (Figure 3A-D).

The effect of naloxone on BU08028 (3 and 10 mg/kg)-induced antinociception across time is shown in Figure 4A-D. The overall ANOVAs indicated a significant interaction [$F(6, 150)=3.22$, $P<0.05$]. Naloxone (1 mg/kg) attenuated BU08028-induced antinociception, indicating MOP-mediated antinociceptive activity. The effects of naloxone were short-lived since attenuation of BU08028-induced antinociception was only evident at the 30- and 60-min time points, with significant antinociception returning at 2 and 4 h. Because the antagonist activity of SB612111 and naloxone are

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short-lived, with no differences observed beyond the 4 hr time point, the 24 and 48 hr data are not shown

Tolerance Development to Thermal Antinociception Produced by BU08028. The effect of repeated administration of BU08028, buprenorphine, and morphine across 9 days is shown in Figure 5. The overall ANOVA indicated a significant interaction [$F(9,87)=3.69$, $P<0.05$]. To examine the effects of tolerance development, levels of %MPE on test days 2, 4, and 9 were compared to test day 1. As expected, tolerance developed to the antinociceptive effects of 10 mg/kg morphine [$F(3,21)=7.72$, $P<0.05$]. Compared to Day 1 morphine produced a significant reduction in antinociception by day 9. Following repeated injections of BU08028 tolerance development was also evident [$F(3,24)=15.50$, $P<0.05$]. Tolerance to the antinociceptive effects of BU08028 was evident sooner than that observed with morphine, with a significant reduction in antinociception observed on Days 4 and 9. Similar to BU08028, tolerance to buprenorphine-induced antinociception was evident by Day 4 [$F(3,21)=10.22$, $P<0.05$].

Acute Behavioral and Rewarding effects of BU08028. The effect of BU08028 on PC following one drug injection is shown in Figure 6A. The initial PC experiment was conducted with a 48h waiting period between drug and vehicle sessions (rather than the normal 24h period) to avoid any interference with the CPP test due to the long half-life of BU08028. The overall ANOVA indicated a significant main effect of dose [$F(4,35)=5.34$, $P<0.05$]. As demonstrated previously (Marquez et al., 2008), animals that received one injection of 10 mg/kg morphine during conditioning exhibited a significant CPP relative to controls. Similarly, animals that received 1 and 10 mg/kg BU08028 also exhibited a significant CPP relative to controls, whereas 3 mg/kg BU08028 showed a trend for a preference for the drug paired compartment, however, this was not significant. Buprenorphine was also tested using the same parameters, and only the 3 mg/kg dose of buprenorphine produced a significant CPP relative to controls (Figure 6C).

During the PC training phase, drug effects on the animals' global activity was also examined. Acute administration of morphine and BU08028 produced an increase in global activity relative to

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vehicle controls ($[F(4,35)=11.9, P<0.05]$; Figure 6B). In fact, the 3 mg/kg dose of BU08028 produced a greater increase in global activity than morphine. Buprenorphine also produced a significant increase in global activity relative to vehicle controls (Figure 6D), however levels of global activity were lower than that observed in animals that received BU08028.

Effects of Repeat Administration of BU08028 Alone and on Cocaine CPP. The effect of repeated injections of BU08028 on PC is shown in Figure 7A. The overall ANOVA indicated a significant effect $[F(3,19)=3.18, P 0.05]$. The 0.3 and 1.0 mg/kg doses of BU08028 produced a significant CPP, similar to cocaine (30 mg/kg) alone. During conditioning, global activity was also assessed following the first and fourth drug injection (Figure 7B). The overall ANOVA indicated that there was a significant dose by injection day interaction $[F(3,19)=16.7, P 0.05]$. BU08028 (0.3-1.0 mg/kg) produced an increase in global activity following the first injection. In fact, the increase in global activity produced by 1.0 mg/kg BU08028 was greater than that observed with cocaine. Following the fourth injection, the levels of global activity produced by 0.3-1.0 mg/kg BU08028 were attenuated relative to the first injection, indicative of tolerance. Indeed, 0.3 mg/kg was no different than controls, only the 1.0 mg/kg dose of BU08028 produced a significant increase in global activity relative to vehicle controls.

The effect of BU08028 on cocaine-induced behaviors is shown in Figure 7C-D. BU08028 (0.1-1.0 mg/kg) did not attenuate or potentiate cocaine-induced CPP. However, it dose-dependently potentiated cocaine-induced global activity ($[F(4,31)=10.56, P<0.05]$; Figure 7D). Even though cocaine alone produced an increase in global activity relative to controls, BU08028 further potentiated cocaine-induced activity following the first and fourth drug injections.

DISCUSSION

The oripavine buprenorphine has a remarkable activity profile. It is a weak partial agonist at the mu opioid receptor and an antagonist at kappa receptors. Although it is a potent analgesic, it has reduced addiction liability as well as an increased safety margin due to a ceiling effect on the respiratory depression. In addition, buprenorphine is a long lasting compound. These properties

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make buprenorphine useful as an analgesic and also a potential replacement for methadone, in drug abuse pharmacotherapy (Lewis, 1985; Lutfy and Cowan, 2004).

One interesting phenomenon about buprenorphine is its inverted U-shaped dose response curve for antinociceptive activity when countering an appropriate painful stimulus (Cowan et al., 1977). The inverted U-shape has been attributed to its agonist activity at NOP receptors, which presumably attenuates its mu-mediated antinociceptive activity at higher doses, despite moderate binding affinity to NOP receptors (Table 1), and very weak partial agonist activity at this receptor (Lutfy et al., 2003; Khroyan et al., 2009). Other mixed NOP/mu compounds, such as SR14150 and SR16835, which are not in the oripavine family, but bind with greater affinity to the NOP receptor, also have mu-mediated antinociceptive activity that is attenuated by the NOP component (Khroyan et al., 2009). Similarly, buprenorphine enhances alcohol consumption at low doses, presumably through mu opioid receptors, but reduces alcohol consumption at higher doses. This reduction in alcohol consumption is blocked by the NOP receptor antagonist UFP-101, suggesting that the buprenorphine-induced reduction in alcohol consumption is due to activation of NOP receptors (Ciccocioppo et al., 2007). An NOP-mediated reduction in CPP and self-administration of a variety of abused drugs by N/OFQ is already well documented (Murphy et al., 1999; Kotlinska et al., 2002; Ciccocioppo et al., 2003; Zhao et al., 2003). Therefore, it is possible that increasing the NOP agonist efficacy in buprenorphine, or other opioid ligands, may produce an effective non-addicting analgesic and may be more efficacious than buprenorphine for drug abuse pharmacotherapy. The buprenorphine analog BU08028 was designed to have increased affinity and efficacy at the NOP receptors. BU08028 is the first reported “universal opioid ligand” that binds with high affinity (under 10 nM) to each of the receptors in the opioid receptor family. BU08028 is almost 10 times more potent at NOP receptors than buprenorphine and also has higher functional efficacy at NOP compared to buprenorphine in vitro (Table 2).

Since BU08028 has similar mu receptor activity to buprenorphine but increased NOP activity, we expect diminished mu-receptor mediated effects such as antinociceptive activity, tolerance

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development, and reward. In acute antinociception assays, BU08028 appears less potent than buprenorphine, showing virtually no antinociceptive activity at 0.1 mg/kg and little activity at 0.3 mg/kg (Figure 2), possibly suggesting NOP-mediated suppression of mu-mediated antinociception. However, the antinociceptive activity of BU08028 is only potentiated by the NOP receptor antagonist SB612111 at higher doses of BU08028, suggesting NOP receptor agonist activity has little or no discernable effect on antinociception at low doses of BU08028. Alternatively, the lack of antinociceptive activity at low doses might suggest that the higher logP of BU08028 is hindering its access to the CNS. Even though the partial agonist activity of BU08028 at mu receptors in vitro, is similar to that of buprenorphine, the dose response curve of BU08028 for antinociception is much steeper than that of buprenorphine, where the higher doses of BU08028 produced near maximal effects. As demonstrated in many reports, the dose response curve for buprenorphine is very shallow, never approaching 100% MPE (Cowan et al., 1977; Ide et al., 2004). In addition, animals dosed with BU08028 exhibit a very profound Straub tail (unpublished observation), indicative of mu-like excitatory activity, and an obvious increase in locomotor activity, also a mu-mediated effect in mice (Cowan et al., 1977).

BU08028 produces antinociceptive effects that are longer lasting than buprenorphine, consistent with BU08028's higher logP. BU08028 antinociception is still strongly evident 24 hr post injection with virtually 100% activity remaining at 10 mg/kg, whereas buprenorphine antinociceptive activity is no longer evident at this later time (Figure 2A,B). Furthermore, it was anticipated that a compound with higher NOP efficacy, similar to the mixed NOP/mu agonist SR16435, would likely have a reduced tolerance development (Khroyan et al., 2007). However, as shown in Figure 5, administration of BU08028 over 9 days resulted in tolerance to its antinociceptive effects, similar to that seen with morphine and buprenorphine. From these experiments it is evident that BU08028 produces a very long lasting antinociceptive effect but tolerance to this effect develops at a similar pace compared to the parent compound buprenorphine.

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Initial assessment of the rewarding effects of BU08028 was conducted using the one trial PC paradigm with a 48hr period between drug and vehicle sessions (Figure 6). This behavioral protocol was used to avoid any interference with the long lasting effects of BU08028. Similar to morphine, BU08028 produced CPP. BU08028 was also directly compared to its parent compound buprenorphine using this same protocol. Buprenorphine produced an inverted U-shaped dose response curve where the 3 mg/mg dose produced a significant CPP and 1 and 10 mg/kg did not, a phenomenon that has previously been observed in rats (Brown et al., 1991; Rowlett et al., 1994). In mice, CPP to buprenorphine has been reported following repeated injections at the 3mg/kg dose, whereas one trial conditioning with this dose did not result in CPP (Marquez et al., 2007; Marquez et al., 2008). Using the more common repeated injection PC protocol, BU08028 also produced CPP following four drug pairings, when drug sessions were not separated by 48hr (figure 7). Apparently, the effect of BU08028, with respect to “reward”, is not as long lasting as its antinociceptive effect, lasting less than 24 h, being associated only with the drug paired compartment and not interfering with the production of a CPP. HS 599, a dihydro derivative of buprenorphine that is also long lasting, did not produce CPP using the traditional PC training regimen (Lattanzi et al., 2001). It is possible that unlike BU08028, HS 599 may have longer lasting effects that influence the development of CPP, and that the drug effects could have carried over to the vehicle session. In any case, the present findings show that BU08028 produces CPP alone, and suggest that its NOP agonist activity is not attenuating its mu receptor-mediated behavior with respect to antinociception, tolerance development, or reward.

Although buprenorphine is approved for maintenance for heroin addiction, it has been considered as a potential pharmacotherapy for other abused drugs. Previous research has examined the effects of buprenorphine on cocaine-induced behaviors. Chronic buprenorphine administration can decrease cocaine intake during self-administration, drug-induced reinstatement of extinguished cocaine seeking, and cocaine-induced locomotor activity (Sorge et al., 2005; Sorge and Stewart, 2006; Placenza et al., 2008). Using the PC paradigm, buprenorphine has been reported to both

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attenuate and potentiate cocaine-induced CPP (Brown et al., 1991; Kosten et al., 1991). These discrepant findings are probably due to the experimental parameters including the drug regimen used. We were hoping that BU 08208 would reduce cocaine-induced CPP. Not surprisingly, however, since BU08028 produced CPP, it was unable to attenuate cocaine-induced CPP.

The goal of this project was to synthesize and characterize compounds with significant NOP receptor activity to supplement mu (and potentially delta and kappa) activity found in the buprenorphine scaffold. The behavioral results obtained with BU08028 can also be compared to other bifunctional mu/NOP receptor ligands that we have previously characterized. We have demonstrated previously, that SR14150 and SR16435, both with high affinity and partial agonist activity at NOP and mu receptors, have antinociceptive activity that is inhibited by naloxone and potentiated by the NOP receptor antagonist SB 612111 (Khroyan et al., 2009; Toll et al., 2009). SR16435, which has an in vitro profile strikingly similar to BU08028, produces antinociception and CPP, indicating that like BU08028, the NOP component is not sufficient to block the rewarding effect of the mu component. It is interesting to note that SR16435 has one behavioral feature diametrically opposed to that of BU08028. Although BU08028 causes a large increase in locomotor activity, SR16435 does the opposite, resulting in reduced activity levels, a behavior attributed to NOP receptor activation. This difference in global activity observed with SR16435 and BU08028 is difficult to explain since these compounds have a similar in vitro profile with respect to NOP, mu, and kappa receptors, but suggest once again that the mu-mediated activities are strongly represented in BU08028. Alternately, it is possible that the very weak delta component of BU08028, which is not present at all in SR16435, might contribute to the increased global activity seen in BU08028.

As we have shown previously, it is possible to overcome potential mu, kappa, and delta mediated behavior by increasing the relative NOP activity of a compound. For example, we have shown that SR14150, which has increased NOP affinity and is 20 fold selective for NOP over mu receptors, is not rewarding on its own (Toll et al., 2009). To go one step further, SR16835 is a full agonist at NOP receptors and a weak partial agonist at mu receptors. This compound is not

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analgesic on its own and is able to attenuate morphine CPP (Toll et al., 2009). Together, all of these data suggest that the balance of NOP and mu components of mixed-profile compounds such as these can have significant impact in modulating the antinociceptive and rewarding aspects of the compounds. With BU08028, which is a partial agonist at both NOP and mu receptors, mu-mediated activity masks effects that could be mediated by the NOP receptor. It is possible that a different buprenorphine analog with high affinity at all the opioid receptors but with full agonist activity at NOP may counteract other traditional opioid-mediated effects such as reward and antinociception.

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AUTHORSHIP CONTRIBUTIONS

Participated in research design: Toll, Khroyan, Husbands

Conducted Experiments: Khroyan, Polgar, Cami-Kobeci

Performed data analysis: Khroyan

Wrote or contributed to the writing of the manuscript: Toll, Khroyan, Zaveri, Husbands

Other: Toll, Zaveri acquired funding for the research

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FOOTNOTES

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- a) Preliminary reports of these data were presented at the annual meetings of the Society for Neuroscience and International Narcotics Research Conference.
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LEGENDS FOR FIGURES

Figure 1. Chemical structures of BU08028 and buprenorphine.

Figure 2. Acute thermal antinociceptive effect of BU08028 (A) and buprenorphine (B) alone, in the tail flick assay. Data are mean %MPE (\pm SEM). An asterisk (*) represents a significant difference from vehicle control ($P < 0.05$), whereas a dagger (\dagger) represents a significant difference from morphine alone ($P < 0.05$).

Figure 3. The effect of NOP antagonist SB 612111 on BU08028-induced antinociception 30-min (A), 60-min (B), 120-min (C), and 240-min (D) post-injection. Data are mean %MPE (\pm SEM). An asterisk (*) represents a significant difference from vehicle control ($P < 0.05$), and a dagger (\dagger) represents a significant difference from BU08028 alone ($P < 0.05$).

Figure 4. The effect of naloxone on BU08028-induced antinociception 30-min (A), 60-min (B), 120-min (C), and 240-min (D) post-injection. An asterisk (*) represents a significant difference from vehicle control ($P < 0.05$), and a dagger (\dagger) represents a significant difference from BU08028 alone ($P < 0.05$).

Figure 5. Tolerance development following repeated injections of 10 mg/kg BU08028 (A) and 10 mg/kg buprenorphine (B). Animals received daily injections of their respective drug and on days 1, 2, 4, and 9 were tested for tail-flick latency 1 hour following their injection. Data represent mean \pm S.E.M. An asterisk (*) represents a significant difference from vehicle control ($P < 0.05$), and a dagger (\dagger) represents a significant difference from Day 1 ($P < 0.05$).

Figure 6. Effects of BU08028 on place conditioning (PC) (A), global activity (B), and effects of buprenorphine on PC (C) and global activity (D). Data are mean (\pm SEM) % time spent in drug-paired compartment (A,C) or activity (cm) following first and fourth drug injection (B,D). An asterisk (*) represents a significant difference from vehicle control, and a dagger (\dagger) represents a significant difference from morphine alone ($P < 0.05$).

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Figure 7. Effects of BU08028 on PC (A), global activity (B), cocaine-induced CPP (C) and cocaine-induced activity (D). Data are mean (\pm SEM) % time spent in drug-paired compartment (A,C) or activity (cm) following first and last drug injection (B,D). An asterisk (*) represents a significant difference from vehicle control, a triangle (Δ) represents a significant difference from first injection, whereas a dagger (\dagger) represents a significant difference from cocaine alone ($P < 0.05$)

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TABLE 1. Binding affinities K_i (nM) of BU08028, compared to buprenorphine, morphine and other prototypical agonists at NOP and opioid receptors. Data show mean \pm SD for at least 2 experiments conducted in triplicate. SR16435 data is from (Khroyan et al., 2007).

	Receptor Binding $K_i \pm$ SD (nM)			
	NOP	MU	DELTA	KAPPA
Buprenorphine	77.4 \pm 16.1	1.52 \pm 0.8	6.13 \pm 0.4	2.50 \pm 1.2
N/OFQ	0.08 \pm 0.03	133 \pm 30	>10,000	247 \pm 3.4
DAMGO	>10,000	1.59 \pm 0.17	300 \pm 58.6	305 \pm 46
DPDPE	>10,000	503 \pm 10.0	1.24 \pm 0.09	>10,000
U69,593	>10,000	>10,000	>10,000	1.62 \pm 0.26
Etorphine	56.9 \pm 2.61	0.33 \pm 0.05	1.54 \pm 0.6	0.22 \pm 0.05
SR16435	7.49 \pm 1.31	2.70 \pm 0.1	>1,000	31.7 \pm 4.8
BU08028	8.46 \pm 1.31	2.14 \pm 0.79	1.59 \pm 0.28	5.63 \pm 1.30

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TABLE 2. Stimulation of [³⁵S]GTPγS binding by BU08028, compared to buprenorphine, morphine and other prototypical agonists at NOP and opioid receptors. Data show mean ± SD for at least 2 experiments conducted in triplicate. SR16435 data is from (Khroyan et al., 2007).

	NOP		MU		DELTA		KAPPA	
	EC ₅₀ (nM)	%Stim	EC ₅₀ (nM)	%Stim	EC ₅₀ (nM)	%Stim	EC ₅₀ (nM)	%Stim
Buprenorphine	251 ± 94	15.5 ± 5.8	10.2 ± 2.2	28.7 ± 1.0	>10,000		>10,000	
N/OFQ	8.11 ± 1.4	100	-----	-----				
DAMGO	----	-----	35.3 ± 0.5	100				
DPDPE	----	-----	----	-----	6.90 ± 0.4	100	----	-----
U69,593	----	-----	----	-----	-----	-----	78.5 ± 8.8	100
SR16435	28.7 ± 0.6	45.0 ± 5.1	2.73 ± .01	29.5 ± 10	ND	*	>10,000	=*
BU08028	78.6 ± 49	48 ± 13	6.03 ± 2.1	21.1 ± 8.7	*	10.8 ± 6.8	*	0

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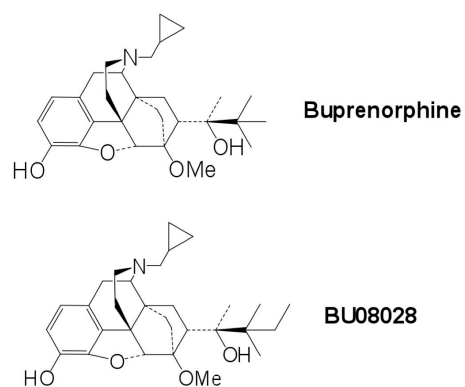


FIGURE 1. STRUCTURE OF BU08028 AND BUPRENORPHINE

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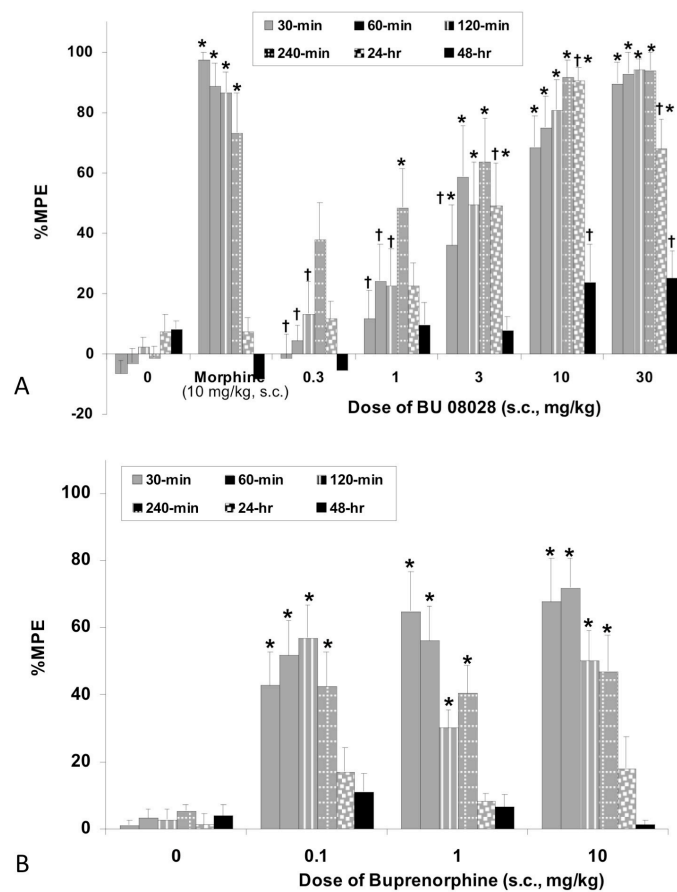


Figure 2

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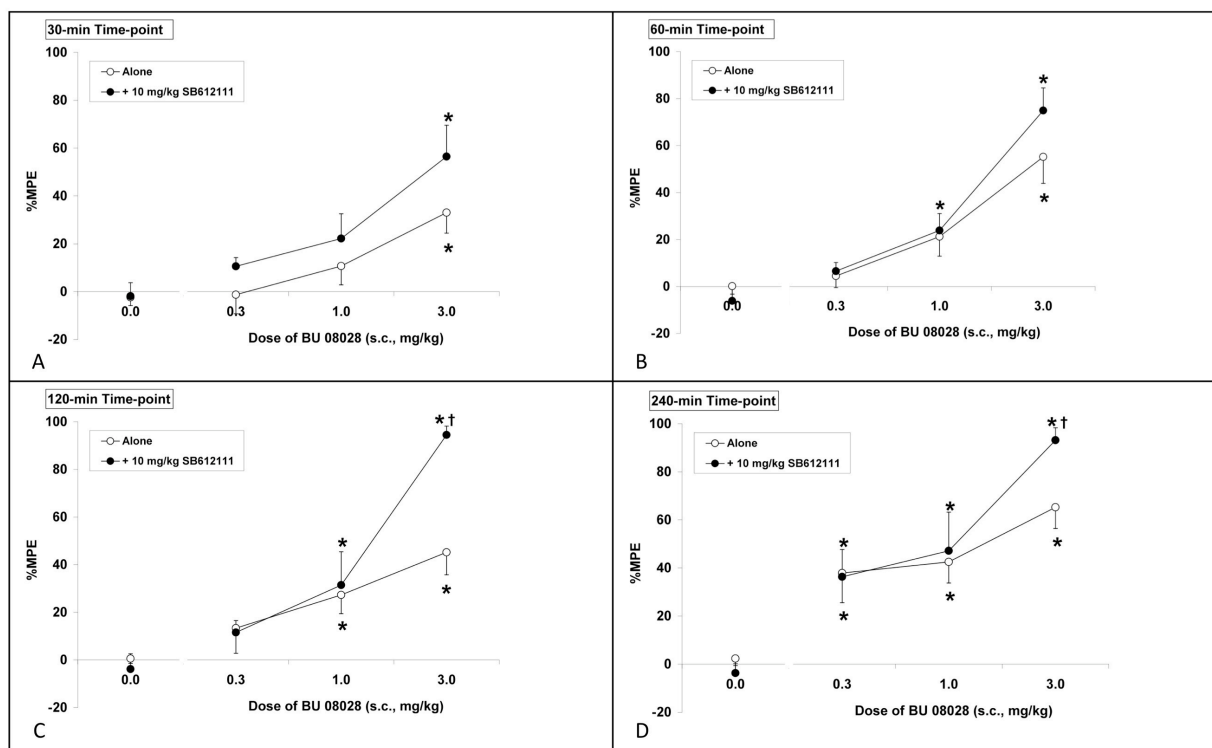


Figure 3

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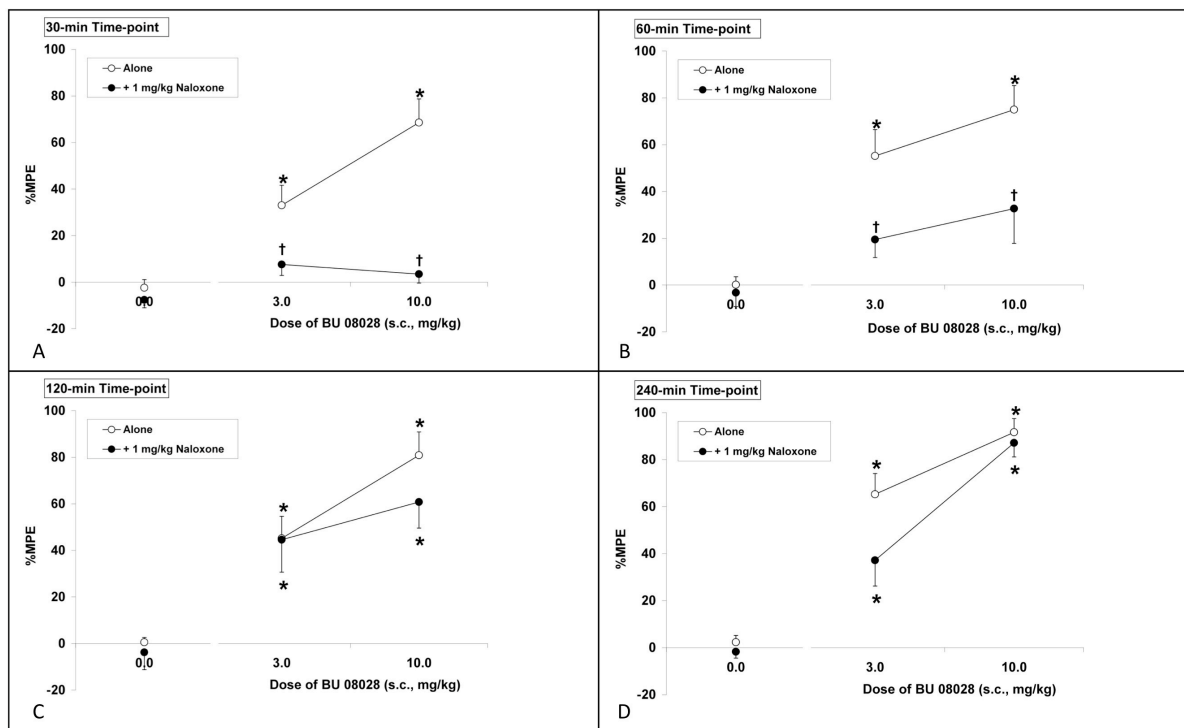


Figure 4

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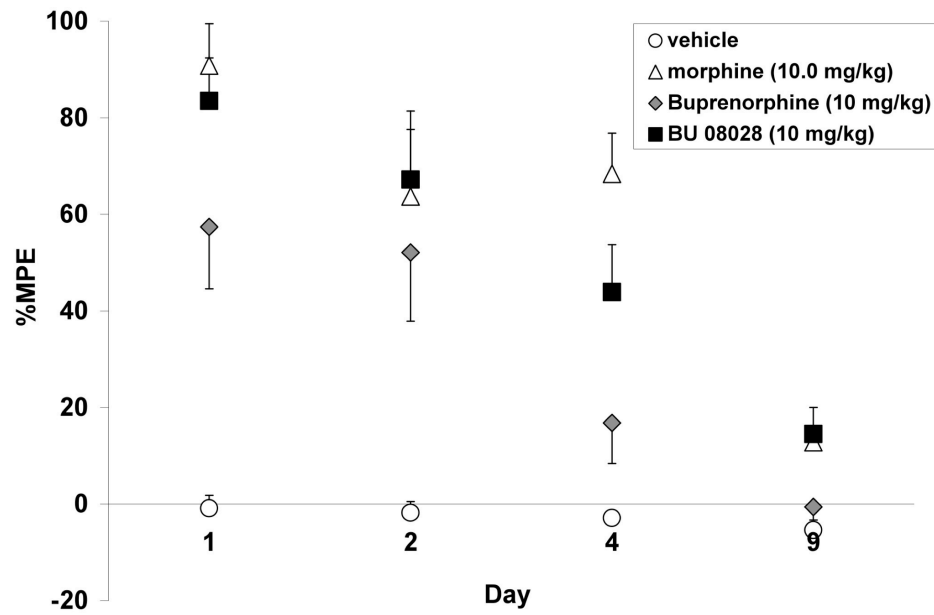


Figure 5

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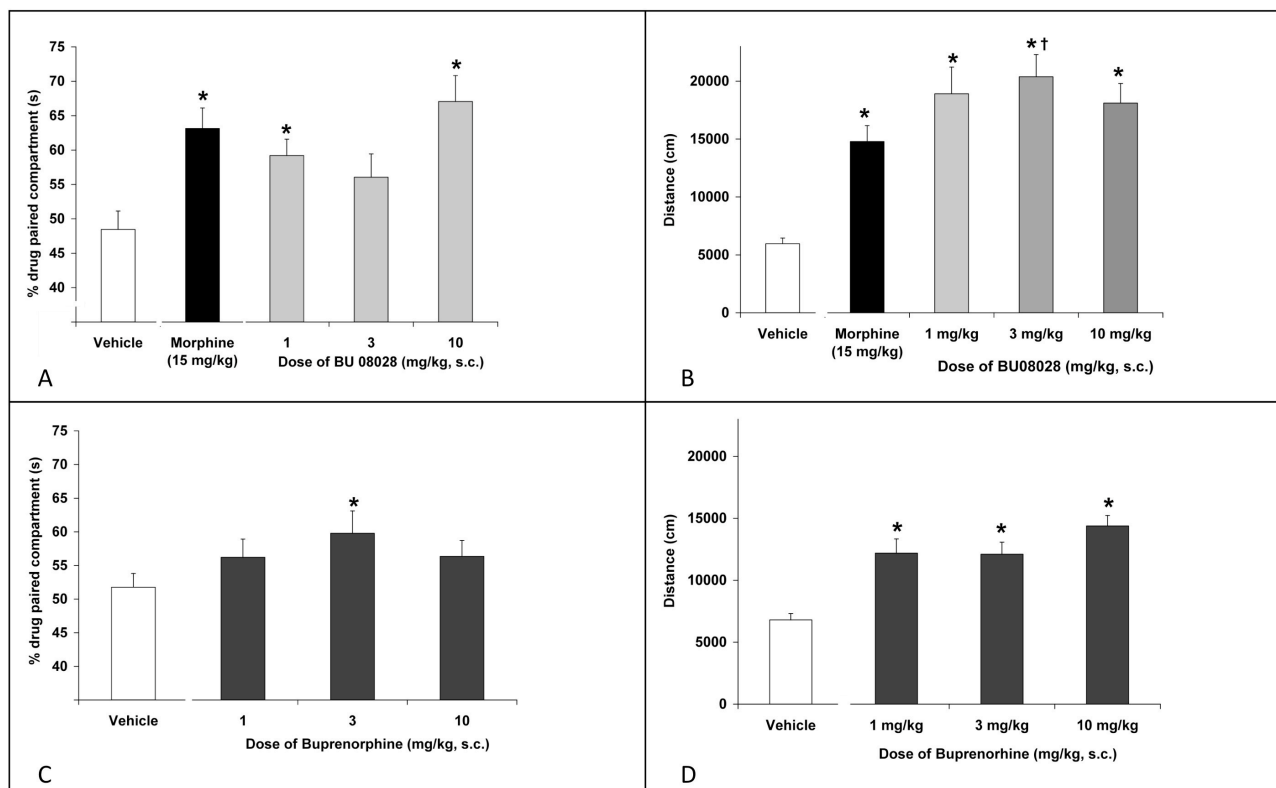


Figure 6

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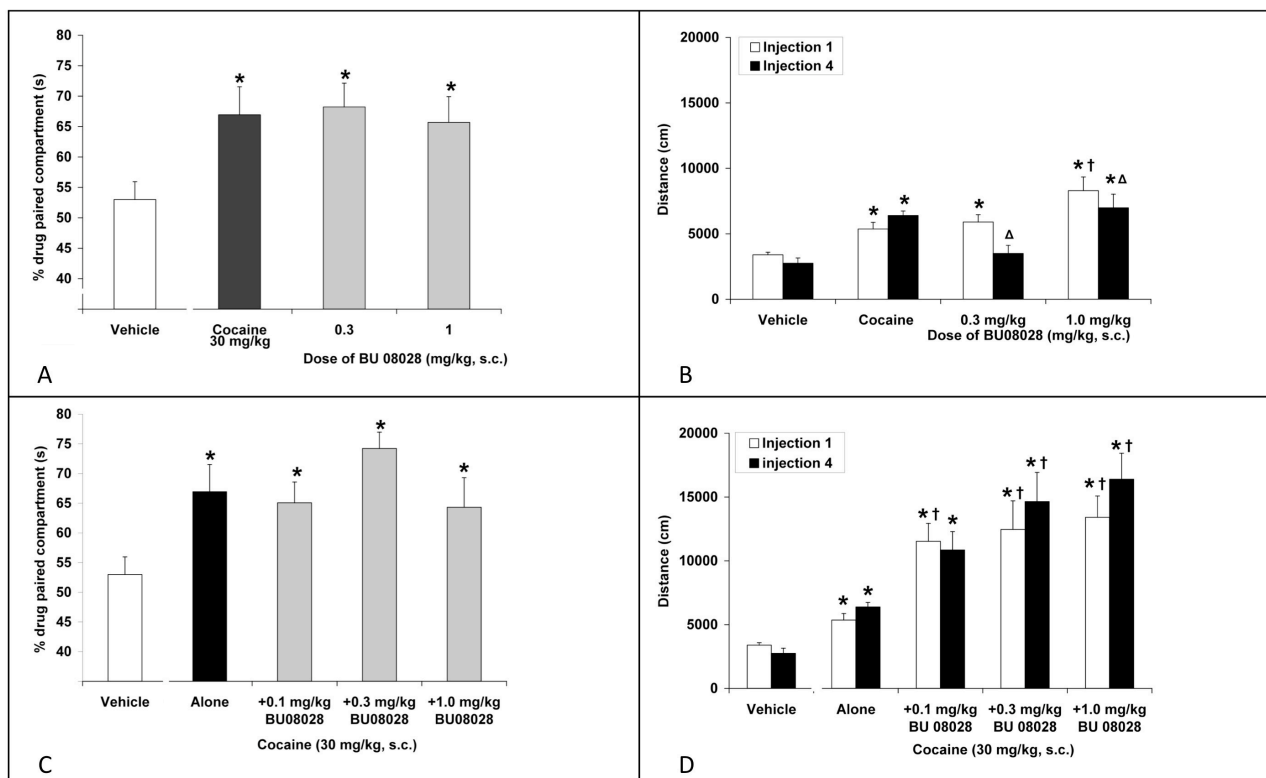


Figure 7