Special Section on Therapeutic Approaches to Treat Disorders of the Urinary and Gastrointestinal Tracts—Minireview

Nitric Oxide Resistance in Priapism Associated with Sickle Cell Disease: Mechanisms, Therapeutic Challenges, and Future Directions

Dalila Andrade Pereira, ⁽¹⁾ Fabiano Beraldi Calmasini, Fernando Ferreira Costa, Arthur L. Burnett, and ⁽¹⁾ Fábio Henrique Silva

Laboratory of Pharmacology, São Francisco University Medical School, Bragança Paulista, SP, Brazil (D.A.P., F.H.S.); Universidade Federal de São Paulo, Escola Paulista de Medicina, Department of Pharmacology, São Paulo, SP, Brazil (F.B.C.); Hematology and Hemotherapy Center, University of Campinas, Campinas, SP, Brazil (F.F.C.); and The James Buchanan Brady Urological Institute and Department of Urology, The Johns Hopkins School of Medicine, Baltimore, Maryland (A.L.B.)

Received October 20, 2023; accepted January 5, 2024

ABSTRACT

Patients with sickle cell disease (SCD) display priapism, a prolonged penile erection in the absence of sexual arousal. The current pharmacological treatments for SCD-associated priapism are limited and focused on acute interventions rather than prevention. Thus, there is an urgent need for new drug targets and preventive pharmacological therapies for this condition. This review focuses on the molecular mechanisms linked to the dysfunction of the NO-cyclic guanosine monophosphate (cGMP)-phosphodiesterase type 5 (PDE5) pathway implicated in SCD-associated priapism. In murine models of SCD, reduced nitric oxide (NO)-cGMP bioavailability in the corpus cavernosum is associated with elevated plasma hemoglobin levels, increased reactive oxygen species levels that inactive NO, and testosterone deficiency that leads to endothelial nitric oxide synthase downregulation. We discuss the consequences of the reduced cGMP-dependent PDE5 activity in response to these molecular changes, highlighting it as the primary pathophysiological mechanism leading to excessive corpus cavernosum relaxation, culminating in priapism. We also further discuss the impact of intravascular hemolysis on therapeutic approaches, present current pharmacological strategies targeting the NO-cGMP-PDE5 pathway in the penis, and identify potential pharmacological targets for future priapism therapies. In men with SCD and priapism, PDE5 inhibitor therapy and testosterone replacement have shown promising results. Recent preclinical research reported the beneficial effect of treatment with haptoglobin and NO donors.

SIGNIFICANCE STATEMENT

This review discusses the molecular changes that reduce NOcGMP bioavailability in the penis in SCD and highlights pharmacological targets and therapeutic strategies for the treatment of priapism, including PDE5 inhibitors, hormonal modulators, NO donors, hydroxyurea, soluble guanylate cyclase stimulators, haptoglobin, hemopexin, and antioxidants.

Introduction

Sickle cell disease (SCD) is the most common hereditary hematologic disorder, affecting millions worldwide (Kavanagh et al., 2022). It arises from a mutation in the β -globin gene, producing an abnormal hemoglobin variant called hemoglobin S (HbS) (Telen et al., 2019). Within red blood cells, HbS tends to polymerize, increasing intracellular viscosity (Kim-Shapiro and Gladwin, 2018). This process not only compromises the integrity of the red blood cell membrane, making it less flexible and more fragile, but also triggers a morphologic shift from a biconcave to a sickle shape, a phenomenon directly linked to HbS polymerization. Patients with SCD face a range of complications. Acute complications include acute pain events, acute chest syndrome, meningitis, stroke, sickle hepatopathy, gallstones, papillary necrosis, splenic infarction, splenic sequestration, bone marrow infarction, osteomyelitis, post-hyphema glaucoma, septicemia, and priapism. Chronic complications involve multiple organ damage, such as functional asplenia retinopathy, pulmonary hypertension, cardiomegaly, diastolic Downloaded from jpet.aspetjournals.org at ASPET Journals on December 29, 2022

ABBREVIATIONS: cGMP, cyclic guanosine monophosphate; eNOS, endothelial nitric oxide synthase; FDA, US Food and Drug Administration; Hb, hemoglobin; HbS, hemoglobin S; nNOS, neuronal nitric oxide synthase; NO, nitric oxide; PDE5, phosphodiesterase type 5; ROS, reactive oxygen species; SCD, sickle cell disease; sGC, soluble guanylyl cyclase; TLR4, toll-like receptor 4.

This work was supported by the São Paulo Research Foundation [Grant 2017/08122-9].

No author has an actual or perceived conflict of interest with the contents of this article.

dx.doi.org/10.1124/jpet.123.001962.

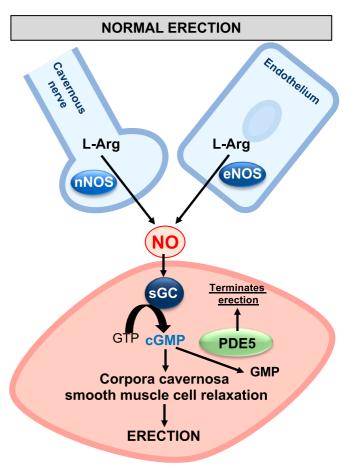
heart failure, and chronic kidney disease, contributing to increased morbidity and mortality. (Kavanagh et al., 2022). HbS polymerization also leads to hemolysis, severe hemolytic anemia, vaso-occlusive crises, and leg ulcers (Kato et al., 2018).

Priapism is a prolonged and persistent penile erection that occurs regardless of sexual stimulation or desire (Bivalacqua et al., 2022). There are two main types: ischemic priapism (low-flow) and non-ischemic priapism (high-flow) (Salonia et al., 2014). Ischemic priapism is characterized by reduced or absent intracorporeal blood flow, leading to painful erections (Salonia et al., 2014). This form of priapism is especially prevalent in men with SCD, with up to 48% of those affected experiencing episodes at some point in their lives (Arduini and Trovó de Marqui, 2018). The average age of onset is 15 years. Continuous and untreated episodes can cause irreversible damage to the erectile tissue, resulting in permanent erectile dysfunction (Arduini and Trovó de Margui, 2018). On the other hand, nonischemic priapism is typically the result of trauma affecting the arteries (Salonia et al., 2014). In individuals with SCD, ischemic priapism is the most common manifestation, accounting for 95% of cases (Hudnall et al., 2017). Of all priapism episodes, about 74% are of the "stuttering" (recurrent) type, lasting a few hours or less (Idris et al., 2020). These episodes can remit and recur with or without medical intervention (Idris et al., 2020). Notably, nearly half (40%) of patients with SCD who experience recurrent ischemic priapism also develop erectile dysfunction (Anele and Burnett, 2015).

For a long time, it was believed that priapism in SCD was solely due to the obstruction of venous outflow from the corpora cavernosa, a consequence of the vaso-occlusion caused by the interaction of sickled red blood cells with endothelial cells, leukocytes, and platelets (Burnett, 2003). However, experimental studies have revealed that priapism in SCD is also attributed to alterations in the mechanisms regulating erection, leading to excessive and persistent relaxation of the erectile tissue (Champion et al., 2005; Mi et al., 2008; Bivalacqua et al., 2009, 2013; Lagoda et al., 2013, 2014; Ning et al., 2014; Silva et al., 2016a,b; Musicki et al., 2018, 2021).

Nitric oxide (NO), recognized as a potent vasodilator, is crucial in regulating vascular tone and erectile function (MacDonald and Burnett, 2021). Over the past decade, numerous studies have suggested that the decrease in NO-cGMP bioavailability is closely linked to the onset of priapism associated with SCD (Champion et al., 2005; Bivalacqua et al., 2009, 2013; Lagoda et al., 2013, 2014; Silva et al., 2016a,b; Musicki et al., 2018, 2021). Moreover, restoring this bioavailability might be an effective therapeutic strategy. Notably, preclinical studies aimed at recovering NO-cGMP bioavailability have shown efficacy in treating priapism in SCD (Bivalacqua et al., 2013; Lagoda et al., 2014; Silva et al., 2016b; Musicki et al., 2018, 2021; Pereira et al., 2022). On the other hand, some preclinical studies have reported that excess plasma hemoglobin generated by intravascular hemolysis might limit the efficacy of specific NO donors, suggesting an NO resistance mechanism in the penis in SCD contexts (Pinheiro et al., 2022; Pereira et al., 2023).

This review focuses on the molecular mechanisms linked to the dysfunction of the NO-cyclic guanosine monophosphate (cGMP)-phosphodiesterase type 5 (PDE5) pathway implicated in SCD-associated priapism. We further discuss the impact of intravascular hemolysis on therapeutic approaches, present current



 $Fig. \ 1. \ Mechanism \ for \ NO-dependent \ penile \ erection.$

pharmacological strategies targeting the NO-cGMP-PDE5 pathway in the penis, and identify potential pharmacological targets for future priapism therapies.

Molecular Mechanisms of Penile Erection

A penile erection results from a complex interplay of vascular, neural, and hormonal factors (Andersson, 2011; MacDonald and Burnett, 2021). At the core of this process, NO acts as the primary mediator, with its release from the endothelium and penile nerve terminals essential for initiating an erection (Fig. 1). Two critical NO synthase (NOS) enzymes exist in the penis: endothelial (eNOS) and neuronal (nNOS) (MacDonald and Burnett, 2021). These enzymes are pivotal in starting and sustaining an erection, catalyzing the transformation of L-arginine into L-citrulline and NO. NO then diffuses into the smooth muscle cell and binds to the ferrous iron in the heme moiety of soluble guanylate cyclase (sGC-Fe²⁺), which in turn triggers the production of the secondary messenger, cGMP (Andersson, 2011). High concentrations of cGMP trigger the activation of cGMP-dependent protein kinase, which influences various proteins that mediate muscle relaxation, such as myosin light chain phosphatase and potassium channels (Andersson, 2011). This sequence of events reduces intracellular calcium levels, therefore causing smooth muscle relaxation and dilation of penile vessels, which facilitates an erection. cGMP production and its degradation by phosphodiesterase type 5 (PDE5) are essential to maintain balance in this process. PDE5 converts cGMP

Molecular Mechanisms of Priapism Pathophysiology

(MacDonald and Burnett, 2021).

Nitric Oxide Pathway Dysfunction in SCD-Associated Priapism Pathophysiology. Experimental research has highlighted that priapism related to SCD primarily arises due to decreased endothelial NO bioavailability and the subsequent downregulation of PDE5 function (Champion et al., 2005; Lagoda et al., 2013; Silva et al., 2016b) (Fig. 2). Studies involving eNOS knockout mice, combined eNOS and nNOS knockout mice, as well as transgenic SCD mice models, have demonstrated an amplified erectile response when subjected to electrical stimulation of the cavernous nerve, coupled with penile fibrotic alterations (Burnett et al., 2002; Champion et al., 2005; Bivalacqua et al., 2009; Silva et al., 2016b; Pinheiro et al., 2022). These studies deduced that the PDE5 downregulation originates from lowered basal cGMP levels. The reduced calcium-dependent constitutive activity of NOS is a pivotal factor in this cGMP-dependent PDE5 expression decline (Corbin et al., 2000). In transgenic SCD mice, this diminished eNOS activity is attributed to lessened phosphorylation

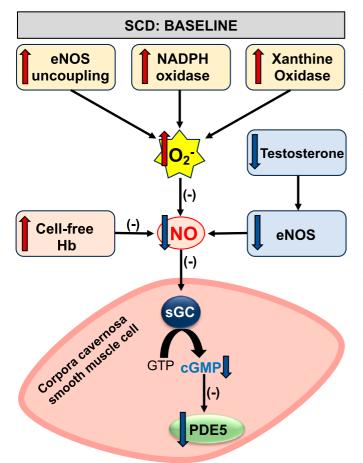


Fig. 2. Mechanisms of nitric oxide resistance in SCD. In SCD corpus cavernosum, baseline levels of endothelial NO and cGMP production are reduced due to decreased eNOS activation, increased superoxide anion production, testosterone deficiency, and the presence of cell-free Hb. Reduced cGMP levels subsequently lead to the downregulation of PDE5 expression and activity.

at Ser-1177, eNOS uncoupling, and a decrease in eNOS binding to the heat shock protein 90 (Musicki et al., 2011, 2012, 2014; Bivalacqua et al., 2013; Sopko et al., 2015; Silva et al., 2016a,b). A similar negative regulation of the eNOS protein was noted in the penile tissues of men with SCD (Lagoda et al., 2013).

The uncoupling of eNOS represents an important mechanism leading to endothelial dysfunction (Daiber et al., 2019). The concept of 'functional uncoupling of eNOS' refers to the phenomenon where, instead of oxidizing L-arginine, the enzyme redirects its electron transfer to the oxidation of molecular oxygen (Förstermann and Sessa, 2012). This results in the predominant production of superoxide anion (Daiber et al., 2019). Thus, the uncoupling of eNOS impairs its efficacy in generating NO and enhances its production of superoxide (Daiber et al., 2019). The mechanism of eNOS uncoupling is linked to the oxidation of the essential NOS cofactor, tetrahydrobiopterin (BH4), the depletion of its substrate L-arginine, the accumulation of endogenous methylarginines, and the S-glutathionylation of eNOS itself (Förstermann and Sessa, 2012).

The eNOS activity is modulated by several post-translational modifications, with phosphorylation being especially noteworthy (Fulton, 2016). This modification occurs at several sites, with Ser1177 being the most prominent. Scientific literature has widely agreed that the phosphorylation of eNOS at this site, mediated by the Akt, amplifies and activates its enzymatic function. The heat shock protein 90 protein, when associated with eNOS, plays a vital role in stabilizing the enzyme ensuring its active conformation (Fulton, 2016). However, in SCD mice, there is a significant reduction in the interaction between eNOS and heat shock protein 90 (Musicki et al., 2011). Concurrently, the phosphorylation of eNOS at the Ser1177 site by Akt is also diminished (Musicki et al., 2011, 2018).

In SCD, reduced endothelial NO bioavailability leads to decreased cGMP production in the penis (Champion et al., 2005; Bivalacqua et al., 2013; Pereira et al., 2022). cGMP is a positive potent regulator of PDE5 gene expression (Corbin et al., 2000; Lin et al., 2002). The reduction in baseline cGMP levels results in lower PDE5 protein expression in the smooth muscle of the corpora cavernosa (Champion et al., 2005; Bivalacqua et al., 2013; Sopko et al., 2015; Silva et al., 2016b). PDE5 protein expression has also decreased in patients with SCD who exhibit priapism (Lagoda et al., 2013). The reduction of baseline cGMP levels may also decrease PDE5 activity by reducing its phosphorylation at the Ser-92 site, a process mediated by cGMP-dependent protein kinase (Corbin et al., 2000; Musicki et al., 2018, 2021). With reduced PDE5 activity, cGMP excessively accumulates in the erectile tissue after sexual stimulation or during nocturnal erections. This accumulation causes an exaggerated relaxation of the smooth muscle in the corpora cavernosa, leading to priapism (Anele et al., 2015) (Fig. 3).

Mechanisms of Nitric Oxide Resistance in SCD

Intravascular Hemolysis. A hallmark of SCD is intravascular hemolysis (Reiter et al., 2002). This process leads to the rupture of red blood cells within blood vessels, releasing hemoglobin, arginase, and other cellular contents into the circulation (Reiter et al., 2002; Morris et al., 2005). Typically, patients with SCD exhibit a steady concentration of 4 μ M cell-free hemoglobin (Hb). However, this concentration can spike to 25 μ M

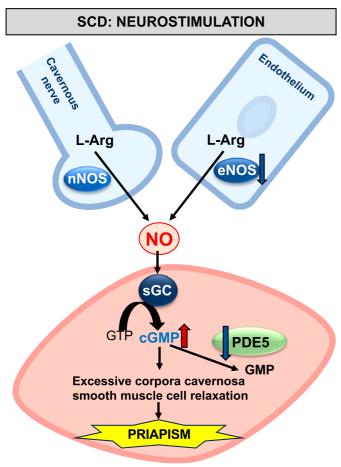


Fig. 3. NO signaling alterations in SCD-induced priapism. This illustration highlights the modifications in the NO signaling pathway in the corpus cavernosum during episodes of priapism associated with SCD.

during crisis episodes (Ballas and Marcolina, 2006). Notably, computational studies have suggested that cell-free Hb concentrations as low as 1 μ M can negatively impact NO bioavailability (Jeffers et al., 2006).

Under conditions where free hemoglobin is prevalent in the plasma or interstitial spaces, this rapidly reacts with NO (Reiter et al., 2002; Schaer et al., 2016). This reaction leads to the production of nitrate and the formation of methemoglobin, the oxidized form of hemoglobin, consequently reducing the bioavailability of NO (Reiter et al., 2002). An additional factor reducing NO bioavailability is the increased activity of plasma arginase, which metabolizes L-arginine, the primary substrate for NO synthesis (Morris et al., 2005). Clinical investigations have highlighted a significant positive correlation between priapism incidents and elevated plasma hemoglobin levels in men diagnosed with SCD (Nolan et al., 2005; Kato et al., 2006; Cita et al., 2016). Moreover, a recent study showcased that inducing intravascular hemolysis in healthy mice resulted in augmented cavernous smooth muscle relaxation governed by the NO-cGMP pathway, similar to that observed in SCD mice (Iacopucci et al., 2022).

Oxidative Stress. In SCD, the bioavailability of NO can also be compromised due to its interaction with reactive oxygen species (ROS) (Kim-Shapiro and Gladwin, 2018). The penis of SCD mice exhibits increased ROS production, which has been linked to various sources. High levels of xanthine oxidase activity have been identified in the plasma of SCD patients and the penises of SCD mice (Aslan et al., 2001; Bivalacqua et al., 2013). This enzyme catalyzes the conversion of hypoxanthine and xanthine into uric acid, generating superoxide (Schmidt et al., 2019). This superoxide rapidly combines with NO, leading to the formation of peroxynitrite and reducing the bioavailability of NO (Pacher et al., 2007).

In SCD, NADPH oxidase has also contributed to priapism through the production of superoxide (Musicki et al., 2012; Bivalacqua et al., 2013; Silva et al., 2016b; Pereira et al., 2022). In humans, seven known subtypes of NADPH oxidase are known, labeled NOX1-5 and DUOX1-2, each with distinct tissue distributions and activation mechanisms (Touyz et al., 2019). The NOX-2 protein comprises several subunits, including the membrane-bound gp91^{phox} (also referred to as Nox2) and p22phox, and cytosolic subunits like p47^{phox}, p67^{phox}, and p40^{phox}, and small GTPases (Rac1 and Rac2) (Touyz et al., 2019). The assembly of these subunits culminates in forming an active enzyme complex that catalyzes the transfer of electrons from cytosolic NADPH to molecular oxygen, generating superoxide as the primary reaction output (Zhang et al., 2020). An upregulation in the expression of various NADPH oxidase subunits, such as gp91^{phox}, p47^{phox}, and p67^{phox}, has been observed in the penises of men diagnosed with SCD and SCD mice (Musicki et al., 2012; Bivalacqua et al., 2013; Silva et al., 2016b; Pereira et al., 2022).

As previously highlighted, another factor contributing to the elevated production of superoxide in the SCD penis is uncoupled eNOS (Bivalacqua et al., 2013). Peroxynitrite, produced by its interaction with NO, is a potent oxidizing agent that damages proteins, lipids, and DNA and impairs cellular signaling pathways (Ayala et al., 2014; Pizzino et al., 2017). Peroxynitrite formation can also trigger lipid peroxidation, a chain reaction leading to the breakdown of membrane lipids, thus causing cell membrane damage and dysfunction (Ayala et al., 2014; Pizzino et al., 2017). Peroxynitrite reaction with tyrosine residues in proteins can result in the formation of 3-nitrotyrosine (Radi, 2013). This compound is recognized as a marker of protein damage from nitrosative stress, and its levels are elevated in penile tissue in SCD (Bivalacqua et al., 2022).

Therapeutic Interventions Based on Normalizing a Pathway NO-cGMP-PDE5

Pharmacological interventions should primarily target the pathophysiologic mechanisms of the disease rather than merely treating the symptoms. This approach enhances therapeutic effectiveness and contributes to minimizing undesired adverse effects. Considering the pivotal role of reduced endothelial NO bioavailability and the subsequent decrease in cGMP levels in the smooth muscle of the corpus cavernosum in the pathogenesis of priapism associated with sickle cell disease, therapeutic strategies that optimize eNOS function, increase the bioavailability of NO and cGMP, or modify subsequent NO targets emerge as promising and emerging treatment options.

PDE5 Inhibitors. Based on the NO-cGMP-PDE5 signaling pathway, PDE5 inhibitors reduce the intracellular degradation of cGMP in penile smooth muscle cells. Sildenafil citrate emerged as a revolution in sexual medicine, pioneering as a PDE5 inhibitor in treating male erectile dysfunction (Goldstein et al., 1998). Paradoxically, clinical and experimental studies have provided evidence for the pleiotropic use of sildenafil citrate in managing priapism associated with SCD (Burnett et al., 2006a,b, 2014).

Experimental studies have shown that treating with sildenafil for three weeks reversed the effects of oxidative/nitrosative stress in the penises of SCD mice (Bivalacqua et al., 2013; Musicki et al., 2014). This action is primarily due to the reversal of eNOS from its uncoupled state to the functionally coupled state (Bivalacqua et al., 2013; Musicki et al., 2014). This re-coupling of eNOS and the reduction in ROS production promotes the accumulation of cGMP. cGMP, in turn, provides positive feedback on the expression and activity of PDE5, restoring its normative function (Bivalacqua et al., 2013). PDE5 functions to degrade the surplus cGMP produced during neurostimulation efficiently (Bivalacqua et al., 2013). This process prevents excessive relaxation of the cavernous tissue, thereby controlling the occurrence of priapism (Bivalacqua et al., 2013). In clinical studies involving men with SCD, it was observed that prolonged treatment regimens with low doses of sildenafil, not associated with erection-stimulating conditions, reduced recurrent priapism episodes without compromising normal erectile ability (Burnett et al., 2006a,b, 2014).

NO-Donating Compounds. NO donors are compounds that spontaneously release NO or do so following metabolism. The enzymatic and/or non-enzymatic mechanisms that lead to NO release vary according to the specific class of the donor agent. Since the mid-1980s, the growing interest in NO physiology has driven the development of a wide range of new NO donors. Within this category, organic nitrate and nitrite esters are a significant class of NO donor drugs. They have been employed in cardiovascular therapeutics since the 19th century (Steinhorn et al., 2015).

As the reduction in NO bioavailability in the penis contributes to the development of priapism in SCD, pre-clinical studies have assessed the effect of NO donor treatment in SCD mice. Treatment of these animals with 3-(1,3-dioxoisoindolin-2-yl) benzyl nitrate (compound 4C) normalized the NO-cGMP-PDE5 pathway and reduced oxidative stress in the penis, reversing priapism (Silva et al., 2016b). The administration of the nitrosamine-based NO donor, 1,5-Bis-(dihexyl-N-nitrosoamino)-2, 4-dinitrobenzene (C6'), in the penile tissue of SCD mice for a week enhanced eNOS and PDE5 expression and decreased oxidative stress. However, its effect on priapic activity has yet to be determined (Lagoda et al., 2014).

The compound RVT-FxMe was synthesized by hybridizing the resveratrol molecule with the furoxan core, the latter being the structure that endows the compound with its ability to spontaneously donate NO or in the presence of thiol (Sorba et al., 1997; Dos Santos et al., 2012). A recent pre-clinical study assessed the effect of a two-week RVT-FxMe treatment on priapism in murine models exhibiting low NO-cGMP bioavailability, specifically in SCD mice and eNOS^{-/-} mice (Pinheiro et al., 2022). RVT-FxMe successfully normalized the NO-cGMP pathway in the corpora cavernosa and corrected the priapism phenotype in eNOS^{-/-} mice; however, it did not demonstrate the same effect in SCD mice (Pinheiro et al., 2022). A distinct characteristic of SCD mice is the excessive presence of free hemoglobin in the plasma, which inactivates the NO released by RVT-FxMe before it can activate the sGC in the penile tissue (Pinheiro et al., 2022).

Hydroxyurea, a potent ribonucleotide reductase inhibitor, marked a significant milestone in treating SCD. It was the pioneering drug therapy to receive approval from the US Food and Drug Administration (FDA) for managing and treating patients afflicted with this genetic condition (Steinberg et al., 2003). Its efficacy is attributed to its ability to elevate fetal hemoglobin levels, which acts as a deterrent against the polymerization of deoxygenated HbS (Charache et al., 1995). In addition, in patients with SCD, a single dose of hydroxyurea effectively elevated plasma nitrate and nitrite levels, as well as increased the total amount of nitrosylated hemoglobin (HbNO), suggesting enhanced intravascular and intraerythrocytic NO generation (Glover et al., 1999; Gladwin et al., 2002). However, the precise site and mechanism of hydroxyurea conversion into NO in vivo remains unclear. While hydroxyurea is commonly prescribed for male patients with SCD, only a few clinical studies have documented its beneficial impact on priapism in these individuals (Saad et al., 2004; Anele et al., 2014). A recent preclinical study found that hydroxyurea treatment did not modify the priapism phenotype in SCD mice (Pereira et al., 2023). This suggests that the NO generated by hydroxyurea could not effectively reach and activate the sGC in the smooth muscle cells of the corpus cavernosum, potentially due to excessive plasma hemoglobin or ROS in the penile tissue of these mice (Pereira et al., 2023).

NO donors with a nitrate-based molecular structure may hold an advantage over direct NO molecule donors under conditions of intravascular hemolysis (Lundberg et al., 2008; Silva et al., 2016b; Pinheiro et al., 2022). Previous research has indicated that Hb also serves as a nitrite reductase, facilitating the conversion of nitrite into NO during deoxygenation (Huang et al., 2005). Oral nitrate treatment has normalized voiding in sickle mice, with mechanisms likely tied to the upregulation of PDE5 activity (Musicki et al., 2019). The distinct effects of nitrate alone on priapism have not been investigated.

In summary, the challenges presented by the pathophysiology of SCD, such as elevated plasma hemoglobin that can interact with NO, indicate a state of resistance to NO. In general, these studies emphasize the need for therapeutic strategies that are not limited by plasma hemoglobin. The differential efficacy of various NO donors further accentuates the intricate complexity of this subject.

Haptoglobin. In patients with SCD, chronic intravascular hemolysis releases excess free hemoglobin into the plasma. This hemoglobin overwhelms and depletes haptoglobin, accumulating free hemoglobin in the circulation (Muller-Eberhard et al., 1968; Reiter et al., 2002; Santiago et al., 2018).

Under typical conditions, the plasma protein haptoglobin binds to free hemoglobin, preventing excessive accumulation in the bloodstream (Buehler et al., 2020). The reticuloendothelial system macrophages detect and absorb the hemoglobinhaptoglobin complex via the CD163 receptor, facilitating its breakdown and removal from circulation (Buehler et al., 2020). This process curtails the accumulation of free hemoglobin, thereby minimizing potential tissue damage and oxidative stress (Buehler et al., 2020).

Studies have shown that haptoglobin treatment inhibits vaso-occlusion in SCD mice and preserves vascular NO during hemolysis (Schaer et al., 2016; Shi et al., 2016; Belcher et al., 2018). Preclinical research indicates that haptoglobin reverses priapism in these mice by normalizing the eNOS-NO-PDE5cGMP pathway in the penis (Pereira et al., 2022). As such, haptoglobin therapy emerges as a promising avenue for addressing priapism linked to SCD. Clinical studies are essential to elucidate the extent of its therapeutic potential further and confirm its efficacy in treating priapism associated with SCD.

Testosterone. Testosterone, an androgenic steroid hormone, holds fundamental importance in erection physiology. It plays a pivotal role not only in the initial development of the penis but also in maintaining and ensuring the plasticity of the cavernous nerve that innervates it (Dandona and Rosenberg, 2010). Additionally, testosterone provides neuroprotection after potential injuries to the cavernous nerve (Podlasek et al., 2016). Testosterone and dihydrotestosterone play a significant role in relaxing penile arteries and cavernous tissue (Van den Broeck et al., 2020). In the presence of androgen deficiency, there is a noted decrease in the expressions and enzymatic activities of eNOS, nNOS, and PDE5, as well as an increased alpha-adrenergic response in the penis (Podlasek et al., 2016).

In men with SCD, hypogonadism is a common complication, affecting up to 25% of them (Taddesse et al., 2012). Clinical research has shown that primary hypogonadism, characterized by testicular insufficiency, is the main reason for this hormonal irregularity in these patients. Although less common, secondary hypogonadism has also been identified in individuals with SCD, particularly in men who exhibit more severe or advanced forms of the disease (Huang and Muneyyirci-Delale, 2017).

The Berkeley mouse model for SCD exhibits hypogonadism (Musicki et al., 2015). When subjected to testosterone replacement, there is a reduction in priapism, accompanied by the restoration of eNOS activity and the normalization of the expression and activity of PDE5 in the penile tissue (Musicki et al., 2018). The proper function of PDE5 in the corpora cavernosa prevents excessive accumulation of cGMP following neurostimulation, controlling priapic activity, as previously mentioned (Musicki et al., 2018). The enhanced eNOS activity in the penis due to testosterone supplementation is attributed to a non-genomic effect of testosterone on eNOS, involving the increased phosphorylation of eNOS at Ser-1177 (Musicki et al., 2018).

Hypogonadal adult men with SCD who were treated with long-acting testosterone undecanoate injections did not experience an increase in the occurrence of priapism but rather an improvement in sexual function (Morrison et al., 2013). Currently, it is believed that the administration of testosterone at physiologic levels does not induce priapism (Morrison et al., 2013). On the contrary, this treatment seems to reduce priapism due to normalizing molecular mechanisms that promote normal erectile responses (Morrison et al., 2015).

An experimental study in mice with SCD treated with FGIN-1-27, a ligand for the translocator protein (TSPO) that mobilizes cholesterol to the inner mitochondrial membrane, showed that it is possible to achieve eugonadal levels of serum testosterone without affecting intratesticular production (Musicki et al., 2021). As a result, a reduction in episodes of priapism was observed. On a molecular level, the activation of TSPO leads to the restoration of phosphodiesterase 5 activity and a reduction in oxidative stress-mediated by NADPH oxidase in the penis (Musicki et al., 2021). This study suggests that pharmacologic activation of TSPO is a promising therapeutic approach for treating hypogonadal men, especially SCD patients, without compromising fertility.

Promising Therapeutic Directions for the Future Based on Normalization of NO-cGMP Bioavailability

SecondGC Stimulators and Activators. Approximately two decades ago, the discovery and development of sGC stimulators and activators opened new horizons in the therapeutic approach to several conditions. These agents can directly act on sGC and promote cGMP synthesis (Sandner et al., 2021). These agents are categorized as stimulators (YC-1, BAY 41-2272, BAY 63-2521, BAY 41-8543, and BAY 60-4552) or activators (BAY 58-2667, BAY 60-2770, and HMR 1766) of sGC (Mónica and Antunes, 2018; Sandner et al., 2021). BAY 41-2272, for instance, is a sGC stimulator that acts independently of NO but is heme-dependent. It directly stimulates sGC and amplifies the sensitivity of enzymes to NO, subsequently raising cGMP levels through a NO-independent mechanism (Stasch et al., 2001; Priviero et al., 2005; Gur et al., 2010). BAY 41-2272 induces penile erection in rats (Bischoff et al., 2003) and promotes relaxation of cavernous tissues in vitro from rabbits, rats, and humans. Furthermore, chronic treatment with BAY 41-2272 improves erectile and ejaculatory dysfunctions in rats under conditions of chronic NO deficiency (Claudino et al., 2011; da Silva et al., 2012). Notably, in 2013, the FDA approved riociguat (BAY 63-2521) as a treatment modality for pulmonary hypertension.

Experimental evidence from in vitro studies has shown that sGC can exist in oxidized states and/or heme-free conditions (Stasch et al., 2006; Sandner et al., 2021). secondGC activators stand out as potential therapeutics due to their ability to activate sGC, mainly when the iron of the heme group is in an oxidized state (Fe^{3+} as opposed to Fe^{2+}) or with the complete absence of this heme group (Stasch et al., 2006). A previous study reported that BAY 60-2770 acts as a heme group mimetic. It is worth noting that the oxidation of the heme portion (Fe^{3+}) of sGC can render the enzyme insensitive to endogenous NO, thereby inhibiting the activation of the sGC-GMPccGMP-dependent protein kinase signaling pathway in various tissues (Stasch et al., 2006; Sandner et al., 2021). Experimental studies have shown that BAY 60-2770 displays potent erectile activity in rats and reverses erectile and micturition dysfunctions associated with obesity in mice (Lasker et al., 2013; Alexandre et al., 2014; Leiria et al., 2014; Silva et al., 2014).

In SCD mice, acute treatment with the sGC activator (BAY 54-6544) demonstrated superiority over the sGC stimulator (BAY 41-8543) in improving the endothelial function of pulmonary arteries (Potoka et al., 2018). Chronic treatment with the sGC activator (BAY 54-6544) improves endothelial function and reverses pulmonary hypertension without affecting systemic blood pressure in sickle cell mice (Potoka et al., 2018). Both the sGC activator (BAY 60-2770) and the sGC stimulator (BAY 41-2272) reduces vaso-occlusive events due to decreased leukocyte recruitment to the endothelium in sickle cell mice (Ferreira et al., 2020). BAY 41-2272, but not BAY 60-2770, increases the transcriptional expression of the γ -globin gene and the production of fetal hemoglobin in K562 erythroleukemic cell culture (Ferreira et al., 2020). Olinciguat, another sGC stimulator in focus, is currently under clinical study for use in patients with SCD (NCT03285178), reduces inflammation, vaso-occlusion, and nephropathy in sickle cell mice (Tchernychev et al., 2021).

Based on the presented studies, sGC activators and stimulators emerge as potential therapeutic alternatives for SCD. They can amplify cGMP-dependent signaling and induce the upregulation of PDE5, positioning them as a meaningful therapeutic strategy for preventing or treating priapism. Importantly, sGC activators and stimulators may have an advantage over NO donor compounds since they are not inactivated by plasma hemoglobin and excess ROS.

Hemopexin. Free hemoglobin in plasma or the interstitial space can rapidly oxidize, promptly releasing the heme group into the plasma (Schaer et al., 2013; Gladwin, 2016). Under physiologic conditions, hemopexin counteracts the accumulation of heme in plasma. Hemopexin binds to the heme group, forming a hemopexin-heme complex that is metabolized in hepatocytes (Smith and Morgan, 1979; Hvidberg et al., 2005). As a protective mechanism, the enzyme heme oxygenase-1 contributes to the enzymatic degradation of heme, thus restricting damage from hemolysis-related molecules. In patients and mice with SCD, elevated heme concentrations are released into the plasma, saturating hemopexin, whose levels decline rapidly and, thus, accumulate heme in plasma (Muller-Eberhard et al., 1968; Reiter et al., 2002; Gbotosho et al., 2021).

Plasma heme is a potent inducer of inflammation, known as a damage-associated molecular pattern (Figueiredo et al., 2007; Bozza and Jeney, 2020). In murine models of SCD, it was shown that heme can activate vascular endothelium through toll-like receptor 4 (TLR4). Activation of this receptor leads to the production of inflammatory mediators (interleukin-1, interleukin-6, interleukin-8), reactive oxygen species, and the release of von Willebrand factor and P-selectin, involved in the activation of blood coagulation and cell adhesion, favoring the occurrence of vascular stasis and vaso-occlusion (Belcher et al., 2014). Therefore, through TLR4 activation, free heme plays a fundamental role in the pathophysiology of SCD (Gbotosho et al., 2021). Experimental studies in corpus cavernosum of animal models of erectile dysfunction have shown that TLR4 activation contributes to alterations in erectile function (Nunes et al., 2017, 2018).

In mouse models with SCD, hemopexin has shown notable beneficial properties. Treatment with hemopexin decreased the release of p-selectin and von Willebrand factor and potentiated heme oxygenase-1 (Belcher et al., 2018). Moreover, studies have shown that hemopexin reduces these animals' vaso-occlusion and acute renal failure. In the aortas of these mice, hemopexin treatment promotes an increase in NOS activity and a reduction in both ROS and nitrosative stress. Hemopexin improves endothelial dysfunction, corrects cardiac alterations, and decreases mean arterial pressure (Vinchi et al., 2013). Due to its ability to enhance endothelial NO bioavailability and attenuate oxidative stress, hemopexin emerges as a promising therapeutic approach for treating priapism related to SCD.

In summary, hemopexin is vital in modulating the complications associated with SCD, particularly in mitigating the deleterious effects of free heme in plasma. Its beneficial properties, as evidenced in animal models, not only point to an ability to correct endothelial dysfunction but also suggest potential benefits in treating priapism related to SCD. The role of hemopexin in improving the bioavailability of endothelial NO and attenuating oxidative stress highlights it as a promising therapeutic approach. Future preclinical are needed to confirm vention in patients with SCD who have recurrent priapism. Antioxidants. Oxidative and nitrosative stress is pivotal in the pathogenesis of priapism associated with SCD (Musicki et al., 2012, 2014; Silva et al., 2016a; Pereira et al., 2022). Notably, L-glutamine, a precursor to the antioxidant glutathione, received FDA approval in 2017 for patients with SCD (Mullard, 2020). However, its efficacy in addressing priapism still needs to be determined. Other antioxidants, such as N-acetylcysteine and omega-3 fatty acids, studied in the context of SCD complications, have yet to establish their specific effects on priapism (Nur et al., 2012; Daak et al., 2013).

Apocynin, an inhibitor of NADPH oxidases, demonstrated promising effects when administered for four weeks in SCD mice by reducing ROS production and normalizing eNOS expression in the penis (Musicki et al., 2012). Similarly, the xanthine oxidase inhibitor attenuates ROS production in isolated penile tissues of these mice (Bivalacqua et al., 2013). However, the direct efficacy of these inhibitors on priapic activity requires further evaluation, underscoring the need for more comprehensive preclinical studies.

In summary, antioxidants are fascinating in treating SCD and its associated priapism. Given the intricate interplay between oxidative and nitrosative stress and the pathogenesis of priapism, antioxidants emerge as promising candidates for clinical exploration (Musicki et al., 2012, 2021; Bivalacqua et al., 2013; Lagoda et al., 2013, 2014; Silva et al., 2016b; Pereira et al., 2022). While findings from mouse studies are encouraging, it is paramount to conduct additional preclinical research to ensure the safety and effectiveness of these agents before their application in SCD patients. The FDA approval of L-glutamine and ongoing research on other antioxidants accentuate the importance of intensifying investigations into these compounds as potential therapeutic strategies for priapism in SCD.

Integrated Strategies to Elucidate the Pathobiology of Priapism in Sickle Cell Disease

Understanding the pathobiology of priapism in patients with SCD remains a significant challenge, limiting the development of effective treatments. An integrated approach, combining longitudinal clinical studies, translational research with animal models, and detailed molecular analyses, is essential to advance the understanding of this complex condition.

Longitudinal clinical studies following patients with SCD can provide valuable results on the progression of priapism, identifying risk factors and clinical correlations. Including a wide range of demographic and clinical data can help discern specific patterns and triggers of priapism. Concurrently, the use of transgenic mice for SCD is useful for investigating relevant molecular mechanisms, contributing to clinical applications. New molecular studies in human penile tissues may also contribute to a deeper understanding of changes in penile physiology, potentially revealing new therapeutic targets and intervention strategies.

Further expanding on this idea, it is becoming increasingly clear that there are likely multiple molecular derangements in the pathogenesis of priapism rather than a singular molecular mechanism. Our current understanding implicates irregularities in NO signaling, as well as various hematologic factors (Kato et al., 2018; Musicki and Burnett, 2020). Thus, recognizing a range of mechanisms in priapism is purposeful and exciting to research, and we may evolve toward personalized interventions based on biomarkers, but with selective cocktails of therapy in the future, not just a monotherapy.

Better understanding the pathobiology of priapism in SCD has direct implications for developing more effective therapies. Research in this area could lead to personalized interventions, potentially improving the quality of life of patients with SCD and reducing the risk of long-term complications. Thus, by expanding our understanding of the pathobiology of priapism in humans with SCD, we can expect significant advances in the management and treatment of priapism in patients with SCD, aligning scientific research with the urgent clinical needs of this population.

Conclusion

Priapism, associated with SCD, represents a significant clinical challenge as no treatment is available. Consistent experimental evidence indicates that the reduction in the bioavailability of NO-cGMP in the penis in SCD is the central change in the pathophysiology of priapism, with plasma hemoglobin, ROS, and endothelial dysfunction playing a fundamental role in contributing to the decrease in NO bioavailability. The emergence of new therapeutic agents focused on normalizing NO/cGMP signaling offers hope and direction for future interventions. Although PDE5 inhibitor therapy and testosterone replacement have shown promising results, recent preclinical research broadens the therapeutic horizon, suggesting opportunities for NO donors and haptoglobin treatment. However, it is essential to recognize potential limitations and the need for additional studies to validate these therapies.

Data Availability

This article contains no datasets generated or analyzed during the current study.

Authorship Contributions

Wrote or contributed to the writing of the manuscript: Pereira, Calmasini, Costa, Burnett, Silva.

References

- Alexandre EC, Leiria LO, Silva FH, Mendes-Silvério CB, Calmasini FB, Davel APC, Mónica FZ, De Nucci G, and Antunes E (2014) Soluble guanylyl cyclase (sGC) degradation and impairment of nitric oxide-mediated responses in urethra from obese mice: reversal by the sGC activator BAY 60-2770. J Pharmacol Exp Ther 349:2–9.
- Andersson K-E (2011) Mechanisms of penile erection and basis for pharmacological treatment of erectile dysfunction. *Pharmacol Rev* **63**:811–859.
- Anele UA and Burnett AL (2015) Erectile dysfunction after sickle cell disease-associated recurrent ischemic priapism: profile and risk factors. J Sex Med 12:713–719.
- Anele UA, Le BV, Resar LMS, and Burnett AL (2015) How I treat priapism. *Blood* **125**:3551–3558.
- Anele UA, Mack AK, Resar LMS, and Burnett AL (2014) Hydroxyurea therapy for priapism prevention and erectile function recovery in sickle cell disease: a case report and review of the literature. *Int Urol Nephrol* 46:1733–1736.
- Arduini GAO and Trovó de Marqui AB (2018) Prevalence and Characteristics of Priapism in Sickle Cell Disease. *Hemoglobin* 42:73-77.
- Aslan M, Ryan TM, Adler B, Townes TM, Parks DA, Thompson JA, Tousson A, Gladwin MT, Patel RP, Tarpey MM et al. (2001) Oxygen radical inhibition of nitric oxide-dependent vascular function in sickle cell disease. Proc Natl Acad Sci USA 98: 15215–15220.
- Ayala A, Muñoz MF, and Argüelles S (2014) Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. Oxid Med Cell Longev 2014:360438.
- Ballas SK and Marcolina MJ (2006) Hyperhemolysis during the evolution of uncomplicated acute painful episodes in patients with sickle cell anemia. *Transfusion* **46**:105–110.
- Belcher JD, Chen C, Nguyen J, Abdulla F, Zhang P, Nguyen H, Nguyen P, Killeen T, Miescher SM, Brinkman N et al. (2018) Haptoglobin and hemopexin inhibit vaso-

occlusion and inflammation in murine sickle cell disease: Role of heme oxygenase-1 induction. *PLoS One* **13**:e0196455.

- Belcher JD, Chen C, Nguyen J, Milbauer L, Abdulla F, Alayash AI, Smith A, Nath KA, Hebbel RP, and Vercellotti GM (2014) Heme triggers TLR4 signaling leading to endothelial cell activation and vaso-occlusion in murine sickle cell disease. *Blood* 123:377–390.
- Bischoff E, Schramm M, Straub A, Feurer A, and Stasch J-P (2003) BAY 41-2272: a stimulator of soluble guanylyl cyclase induces nitric oxide-dependent penile erection in vivo. Urology 61:464-467.
- Bivalacqua TJ, Allen BK, Brock GB, Broderick GA, Chou R, Kohler TS, Mulhall JP, Oristaglio J, Rahimi LL, Rogers ZR et al. (2022) The Diagnosis and Management of Recurrent Ischemic Priapism, Priapism in Sickle Cell Patients, and Non-Ischemic Priapism: An AUA/SMSNA Guideline. J Urol 208:43–52.
- Bivalacqua TJ, Musicki B, Hsu LL, Berkowitz DE, Champion HC, and Burnett AL (2013) Sildenafil citrate-restored eNOS and PDE5 regulation in sickle cell mouse penis prevents priapism via control of oxidative/nitrosative stress. *PLoS One* 8: e68028.
- Bivalacqua TJ, Musicki B, Hsu LL, Gladwin MT, Burnett AL, and Champion HC (2009) Establishment of a transgenic sickle-cell mouse model to study the pathophysiology of priapism. J Sex Med 6:2494-2504.
- Bozza MT and Jeney V (2020) Pro-inflammatory Actions of Heme and Other Hemoglobin-Derived DAMPs. Front Immunol 11:1323.
- Buehler PW, Humar R, and Schaer DJ (2020) Haptoglobin Therapeutics and Compartmentalization of Cell-Free Hemoglobin Toxicity. *Trends Mol Med* 26:683–697.
- Burnett AL (2003) Pathophysiology of Priapism: Dysregulatory Erection Physiology Thesis. J Urol **170**:26–34.
- Burnett AL, Anele UA, Trueheart IN, Strouse JJ, and Casella JF (2014) Randomized controlled trial of sildenafil for preventing recurrent ischemic priapism in sickle cell disease. *Am J Med* **127**:664–668.
- Burnett AL, Bivalacqua TJ, Champion HC, and Musicki B (2006a) Feasibility of the use of phosphodiesterase type 5 inhibitors in a pharmacologic prevention program for recurrent priapism. J Sex Med 3:1077–1084.
- Burnett AL, Bivalacqua TJ, Champion HC, and Musicki B (2006b) Long-term oral phosphodiesterase 5 inhibitor therapy alleviates recurrent priapism. Urology 67:1043-1048.
- Burnett AL, Chang AG, Crone JK, Huang PL, and Sezen SE (2002) Noncholinergic penile erection in mice lacking the gene for endothelial nitric oxide synthase. J Androl 23:92–97.
- Champion HC, Bivalacqua TJ, Takimoto E, Kass DA, and Burnett AL (2005) Phosphodiesterase-5A dysregulation in penile erectile tissue is a mechanism of priapism. *Proc Natl Acad Sci USA* **102**:1661–1666.
- Charache S, Terrin ML, Moore RD, Dover GJ, Barton FB, Eckert SV, McMahon RP, and Bonds DR; Investigators of the Multicenter Study of Hydroxyurea in Sickle Cell Anemia (1995) Effect of hydroxyurea on the frequency of painful crises in sickle cell anemia. N Engl J Med **332**:1317-1322.
- Cita K-C, Brureau L, Lemonne N, Billaud M, Connes P, Ferdinand S, Tressières B, Tarer V, Etienne-Julan M, Blanchet P et al. (2016) Men with Sickle Cell Anemia and Priapism Exhibit Increased Hemolytic Rate, Decreased Red Blood Cell Deformability and Increased Red Blood Cell Aggregate Strength. *PLoS One* **11**:e0154866.
- Claudino MA, da Silva FH, Mónica FZT, Rojas-Moscoso JA, De Nucci G, and Antunes E (2011) Long-term oral treatment with BAY 41-2272 ameliorates impaired corpus cavernosum relaxations in a nitric oxide-deficient rat model. *BJU Int* **108**:116–122.
- Corbin JD, Turko IV, Beasley A, and Francis SH (2000) Phosphorylation of phosphodiesterase-5 by cyclic nucleotide-dependent protein kinase alters its catalytic and allosteric cGMP-binding activities. *Eur J Biochem* **267**:2760–2767.
- da Silva FH, Claudino MA, Báu FR, Rojas-Moscoso JA, Mónica FZ, De Nucci G, and Antunes E (2012) Vas deferens smooth muscle responses to the nitric oxide-independent soluble guanylate cyclase stimulator BAY 41-2272. Eur J Pharmacol 688:49–55.
- Daak AA, Ghebremeskel K, Hassan Z, Attallah B, Azan HH, Elbashir MI, and Crawford M (2013) Effect of omega-3 (n-3) fatty acid supplementation in patients with sickle cell anemia: randomized, double-blind, placebo-controlled trial. Am J Clin Nutr 97:37–44.
- Daiber A, Xia N, Steven S, Oelze M, Hanf A, Kröller-Schön S, Münzel T, and Li H (2019) New Therapeutic Implications of Endothelial Nitric Oxide Synthase (eNOS) Function/Dysfunction in Cardiovascular Disease. *Int J Mol Sci* **20**:187.
- Dandona P and Rosenberg MT (2010) A practical guide to male hypogonadism in the primary care setting. Int J Clin Pract 64:682–696.
- Dos Santos JL, Lanaro C, Chelucci RC, Gambero S, Bosquesi PL, Reis JS, Lima LM, Cerecetto H, González M, Costa FF et al. (2012) Design, synthesis, and pharmacological evaluation of novel hybrid compounds to treat sickle cell disease symptoms. part II: furoxan derivatives. J Med Chem 55:7583–7592.
- Ferreira Jr WA, Chweih H, Lanaro C, Almeida CB, Brito PL, Gotardo EMF, Torres L, Miguel LI, Franco-Penteado CF, Leonardo FC et al. (2020) Beneficial Effects of Soluble Guanylyl Cyclase Stimulation and Activation in Sickle Cell Disease Are Amplified by Hydroxyurea: In Vitro and In Vivo Studies. J Pharmacol Exp Ther 374:469–478.
- Figueiredo RT, Fernandez PL, Mourao-Sa DS, Porto BN, Dutra FF, Alves LS, Oliveira MF, Oliveira PL, Graça-Souza AV, and Bozza MT (2007) Characterization of heme as activator of Toll-like receptor 4. J Biol Chem 282:20221–20229.
- Förstermann U and Sessa WC (2012) Nitric oxide synthases: regulation and function. Eur Heart J 33:829–837, 837a–837d.
- Fulton DJR (2016) Transcriptional and Posttranslational Regulation of eNOS in the Endothelium. Adv Pharmacol 77:29–64.
- Gbotosho OT, Kapetanaki MG, and Kato GJ (2021) The Worst Things in Life are Free: The Role of Free Heme in Sickle Cell Disease. *Front Immunol* **11**:561917.
- Gladwin MT (2016) Cardiovascular complications and risk of death in sickle-cell disease. Lancet **387**:2565–2574.
- Gladwin MT, Shelhamer JH, Ognibene FP, Pease-Fye ME, Nichols JS, Link B, Patel DB, Jankowski MA, Pannell LK, Schechter AN et al. (2002) Nitric oxide donor

- 116:436-444. Glover RE, Ivy ED, Orringer EP, Maeda H, and Mason RP (1999) Detection of nitrosyl hemoglobin in venous blood in the treatment of sickle cell anemia with hydroxyurea. *Mol Pharmacol* 55:1006-1010.
- Goldstein I, Lue TF, Padma-Nathan H, Rosen RC, Steers WD, and Wicker PA; Sildenafil Study Group (1998) Oral sildenafil in the treatment of erectile dysfunction. N Engl J Med 338:1397–1404.
- Gur S, Kadowitz PJ, and Hellstrom WJG (2010) Exploring the potential of NO-independent stimulators and activators of soluble guanylate cyclase for the medical treatment of erectile dysfunction. Curr Pharm Des 16:1619–1633.
- Huang AW and Muneyyirci-Delale O (2017) Reproductive endocrine issues in men with sickle cell anemia. Andrology 5:679–690.
- Huang Z, Shiva S, Kim-Shapiro DB, Patel RP, Ringwood LA, Irby CE, Huang KT, Ho C, Hogg N, Schechter AN et al. (2005) Enzymatic function of hemoglobin as a nitrite reductase that produces NO under allosteric control. J Clin Invest 115: 2099–2107.
- Hudnall M, Reed-Maldonado AB, and Lue TF (2017) Advances in the understanding of priapism. Transl Androl Urol 6:199–206.
- Hvidberg V, Maniecki MB, Jacobsen C, Højrup P, Møller HJ, and Moestrup SK (2005) Identification of the receptor scavenging hemopexin-heme complexes. *Blood* 106:2572–2579.
- Iacopucci APM, da Silva Pereira P, Pereira DA, Calmasini FB, Pittalà V, Reis LO, Burnett AL, Costa FF, and Silva FH (2022) Intravascular hemolysis leads to exaggerated corpus cavernosum relaxation: Implication for priapism in sickle cell disease. FASEB J 36:e22535.
- Idris IM, Abba A, Galadanci JA, Mashi SA, Hussaini N, Gumel SA, Burnett AL, and DeBaun MR (2020) Men with sickle cell disease experience greater sexual dysfunction when compared with men without sickle cell disease. *Blood Adv* 4:3277–3283.
- Jeffers A, Gladwin MT, and Kim-Shapiro DB (2006) Computation of plasma hemoglobin nitric oxide scavenging in hemolytic anemias. Free Radic Biol Med 41:1557–1565.
- Kato GJ, McGowan V, Machado RF, Little JA, Taylor 6th J, Morris CR, Nichols JS, Wang X, Poljakovic M, Morris Jr SM et al. (2006) Lactate dehydrogenase as a biomarker of hemolysis-associated nitric oxide resistance, priapism, leg ulceration, pulmonary hypertension, and death in patients with sickle cell disease. Blood 107:2279–2285.
- Kato GJ, Piel FB, Reid CD, Gaston MH, Ohene-Frempong K, Krishnamurti L, Smith WR, Panepinto JA, Weatherall DJ, Costa FF et al. (2018) Sickle cell disease. Nat Rev Dis Primers 4:18010.
- Kavanagh PL, Fasipe TA, and Wun T (2022) Sickle Cell Disease: A Review. JAMA ${\bf 328}{:}57{-}68.$
- Kim-Shapiro DB and Gladwin MT (2018) Nitric oxide pathology and therapeutics in sickle cell disease. *Clin Hemorheol Microcirc* 68:223–237.
- Lagoda G, Sezen SF, Cabrini MR, Musicki B, and Burnett AL (2013) Molecular analysis of erection regulatory factors in sickle cell disease associated priapism in the human penis. J Urol 189:762–768.
- Lagoda G, Sezen SF, Hurt KJ, Cabrini MR, Mohanty DK, and Burnett AL (2014) Sustained nitric oxide (NO)-releasing compound reverses dysregulated NO signal transduction in priapism. *FASEB J* 28:76–84.
- Lasker GF, Pankey EA, Frink TJ, Zeitzer JR, Walter KA, and Kadowitz PJ (2013) The sGC activator BAY 60-2770 has potent erectile activity in the rat. Am J Physiol Heart Circ Physiol 304:H1670–H1679.
- Leiria LO, Silva FH, Davel APC, Alexandre EC, Calixto MC, De Nucci G, Mónica FZ, and Antunes E (2014) The soluble guanylyl cyclase activator BAY 60-2770 ameliorates overactive bladder in obese mice. J Urol 191:539–547.
- Lin C-S, Chow S, Lau A, Tu R, and Lue TF (2002) Human PDE5A gene encodes three PDE5 isoforms from two alternate promoters. *Int J Impot Res* 14:15–24.
- Lundberg JO, Weitzberg E, and Gladwin MT (2008) The nitrate-nitrite-nitric oxide pathway in physiology and therapeutics. *Nat Rev Drug Discov* 7:156–167.
- MacDonald SM and Burnett AL (2021) Physiology of Erection and Pathophysiology of Erectile Dysfunction. Urol Clin North Am 48:513-525.
- Mi T, Abbasi S, Zhang H, Uray K, Chunn JL, Xia LW, Molina JG, Weisbrodt NW, Kellems RE, Blackburn MR et al. (2008) Excess adenosine in murine penile erectile tissues contributes to priapism via A2B adenosine receptor signaling. J Clin Invest 118:1491–1501.
- Mónica FZ and Antunes E (2018) Stimulators and activators of soluble guanylate cyclase for urogenital disorders. *Nat Rev Urol* **15**:42–54.
- Morris CR, Kato GJ, Poljakovic M, Wang X, Blackwelder WC, Sachdev V, Hazen SL, Vichinsky EP, Morris Jr SM, and Gladwin MT (2005) Dysregulated arginine metabolism, hemolysis-associated pulmonary hypertension, and mortality in sickle cell disease. JAMA 294:81–90.
- Morrison BF, Madden W, Clato-Day S, and Gabay L (2015) Testosterone Replacement Therapy in Adolescents With Sickle Cell Disease Reverses Hypogonadism Without Promoting Priapism: A Case Report. *Urol Case Rep* **3**:179–180.
- Morrison BF, Reid M, Madden W, and Burnett AL (2013) Testosterone replacement therapy does not promote priapism in hypogonadal men with sickle cell disease: 12month safety report. Andrology 1:576–582.
- Mullard A (2020) Sickle cell disease celebration. Nat Rev Drug Discov 19:10.
- Muller-Eberhard U, Javid J, Liem HH, Hanstein A, and Hanna M (1968) Plasma concentrations of hemopexin, haptoglobin and heme in patients with various hemolytic diseases. *Blood* 32:811–815.
- Musicki B, Anele UA, Campbell JD, Karakus S, Shiva S, Silva FH, and Burnett AL (2019) Dysregulated NO/PDE5 signaling in the sickle cell mouse lower urinary tract: Reversal by oral nitrate therapy. *Life Sci* 238:116922.
- Musicki B, Bivalacqua TJ, Champion HC, and Burnett AL (2014) Sildenafil promotes eNOS activation and inhibits NADPH oxidase in the transgenic sickle cell mouse penis. J Sex Med 11:424–430.
- Musicki B and Burnett AL (2020) Mechanisms underlying priapism in sickle cell disease: targeting and key innovations on the preclinical landscape. *Expert Opin Ther Targets* **24**:439–450.

- Musicki B, Champion HC, Hsu LL, Bivalacqua TJ, and Burnett AL (2011) Posttranslational inactivation of endothelial nitric oxide synthase in the transgenic sickle cell mouse penis. J Sex Med 8:419–426.
- Musicki B, Karakus S, Akakpo W, Silva FH, Liu J, Chen H, Zirkin BR, and Burnett AL (2018) Testosterone replacement in transgenic sickle cell mice controls priapic activity and upregulates PDE5 expression and eNOS activity in the penis. Andrology 6:184–191.
- Musicki B, Karakus S, La Favor JD, Chen H, Silva FH, Sturny M, Zirkin BR, and Burnett AL (2021) TSPO ligand FGIN-1-27 controls priapism in sickle cell mice via endogenous testosterone production. J Cell Physiol 236:3073–3082. 10.1002/jcp.30075.
- Musicki B, Liu T, Sezen SF, and Burnett AL (2012) Targeting NADPH oxidase decreases oxidative stress in the transgenic sickle cell mouse penis. J Sex Med 9: 1980–1987.
- Musicki B, Zhang Y, Chen H, Brown TR, Zirkin BR, and Burnett AL (2015) Mechanism of testosterone deficiency in the transgenic sickle cell mouse. *PLoS One* 10:e0128694.
- Ning C, Wen J, Zhang Y, Dai Y, Wang W, Zhang W, Qi L, Grenz A, Eltzschig HK, Blackburn MR et al. (2014) Excess adenosine A2B receptor signaling contributes to priapism through HIF-1 α mediated reduction of PDE5 gene expression. FASEB J 28:2725–2735.
- Nolan VG, Wyszynski DF, Farrer LA, and Steinberg MH (2005) Hemolysis-associated priapism in sickle cell disease. Blood 106:3264–3267.
- Nunes KP, Bomfim GF, Toque HA, Szasz T, and Clinton Webb R (2017) Toll-like receptor 4 (TLR4) impairs nitric oxide contributing to Angiotensin II-induced cavernosal dysfunction. *Life Sci* 191:219–226.
- Nunes KP, de Oliveira AA, Szasz T, Biancardi VC, and Webb RC (2018) Blockade of Toll-Like Receptor 4 Attenuates Erectile Dysfunction in Diabetic Rats. J Sex Med 15:1235–1245.
- Nur E, Brandjes DP, Teerlink T, Otten H-M, Oude Elferink RPJ, Muskiet F, Evers LM, ten Cate H, Biemond BJ, Duits AJ et al.; CURAMA study group (2012) N-acetylcysteine reduces oxidative stress in sickle cell patients. Ann Hematol 91:1097–1105.
- Pacher P, Beckman JS, and Liaudet L (2007) Nitric oxide and peroxynitrite in health and disease. *Physiol Rev* 87:315-424.
- Pereira DA, da Silva Pereira P, Silveira THR, Calmasini FB, Reis LO, Costa FF, and Silva FH (2023) Hydroxyurea does not reverse functional alterations of the nitric oxide-cGMP pathway associated with priapism phenotype in corpus cavernosum from sickle cell mouse. *PLOS ONE* 18:e0292706.
- Pereira PDS, Pereira DA, Calmasini FB, Reis LO, Brinkman N, Burnett AL, Costa FF, and Silva FH (2022) Haptoglobin treatment contributes to regulating nitric oxide signal and reduces oxidative stress in the penis: A preventive treatment for priapism in sickle cell disease. Front Physiol 13:961534.
- Pinheiro AK, Pereira DA, Dos Santos JL, Calmasini FB, Alexandre EC, Reis LO, Burnett AL, Costa FF, and Silva FH (2022) Resveratrol-nitric oxide donor hybrid effect on priapism in sickle cell and nitric oxide-deficient mouse. *PLoS One* 17:e0269310.
- Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, and Bitto A (2017) Oxidative Stress: Harms and Benefits for Human Health. Oxid Med Cell Longev 2017:8416763.
- Podlasek CA, Mulhall J, Davies K, Wingard CJ, Hannan JL, Bivalacqua TJ, Musicki B, Khera M, González-Cadavid NF, and Burnett 2nd AL (2016) Translational Perspective on the Role of Testosterone in Sexual Function and Dysfunction. J Sex Med 13:1183–1198.
- Potoka KP, Wood KC, Baust JJ, Bueno M, Hahn SA, Vanderpool RR, Bachman T, Mallampalli GM, Osei-Hwedieh DO, Schrott V et al. (2018) Nitric Oxide-Independent Soluble Guanylate Cyclase Activation Improves Vascular Function and Cardiac Remodeling in Sickle Cell Disease. Am J Respir Cell Mol Biol 58:636–647.
- Priviero FBM, Baracat JS, Teixeira CE, Claudino MA, De Nucci G, and Antunes E (2005) Mechanisms underlying relaxation of rabbit aorta by BAY 41-2272, a nitric oxide-independent soluble guanylate cyclase activator. *Clin Exp Pharmacol Physiol* 32:728–734.
- Radi R (2013) Protein tyrosine nitration: biochemical mechanisms and structural basis of functional effects. Acc Chem Res 46:550–559.
- Reiter CD, Wang X, Tanus-Santos JE, Hogg N, Cannon 3rd RO, Schechter AN, and Gladwin MT (2002) Cell-free hemoglobin limits nitric oxide bioavailability in sicklecell disease. *Nat Med* 8:1383–1389.
- Saad STO, Lajolo C, Gilli S, Marques Júnior JFC, Lima CS, Costa FF, and Arruda VR (2004) Follow-up of sickle cell disease patients with priapism treated by hydroxyurea. Am J Hematol 77:45–49.
- Salonia A, Eardley I, Giuliano F, Hatzichristou D, Moncada I, Vardi Y, Wespes E, and Hatzimouratidis K; European Association of Urology (2014) European Association of Urology guidelines on priapism. *Eur Urol* 65:480–489.
- Sandner P, Zimmer DP, Milne GT, Follmann M, Hobbs A, and Stasch J-P (2021) Soluble Guanylate Cyclase Stimulators and Activators. Handb Exp Pharmacol 264:355–394.
- Santiago RP, Guarda CC, Figueiredo CVB, Fiuza LM, Aleluia MM, Adanho CSA, Carvalho MOS, Pitanga TN, Zanette DL, Lyra IM et al. (2018) Serum haptoglobin and hemopexin levels are depleted in pediatric sickle cell disease patients. *Blood Cells Mol Dis* 72:34–36.
- Schaer CA, Deuel JW, Schildknecht D, Mahmoudi L, Garcia-Rubio I, Owczarek C, Schauer S, Kissner R, Banerjee U, Palmer AF et al. (2016) Haptoglobin Preserves Vascular Nitric Oxide Signaling during Hemolysis. Am J Respir Crit Care Med 198:1111-1122.
- Schaer DJ, Buehler PW, Alayash AI, Belcher JD, and Vercellotti GM (2013) Hemolysis and free hemoglobin revisited: exploring hemoglobin and hemin scavengers as a novel class of therapeutic proteins. *Blood* **121**:1276–1284.
 Schmidt HM, Kelley EE, and Straub AC (2019) The impact of xanthine oxidase (XO)
- Schmidt HM, Kelley EE, and Straub AC (2019) The impact of xanthine oxidase (XO) on hemolytic diseases. *Redox Biol* 21:101072.
- Shi PA, Choi E, Chintagari NR, Nguyen J, Guo X, Yazdanbakhsh K, Mohandas N, Alayash AI, Manci EA, Belcher JD et al. (2016) Sustained treatment of sickle cell mice with haptoglobin increases HO-1 and H-ferritin expression and decreases iron deposition in the kidney without improvement in kidney function. Br J Haematol 175:714–723.

212 Pereira et al.

- Silva FH, Claudino MA, Calmasini FB, Alexandre EC, Franco-Penteado C, Burnett AL, Antunes E, and Costa FF (2016a) Sympathetic Hyperactivity, Increased Tyrosine Hydroxylase and Exaggerated Corpus Cavernosum Relaxations Associated with Oxidative Stress Plays a Major Role in the Penis Dysfunction in Townes Sickle Cell Mouse. *PLoS One* 11:e0166291.
- Silva FH, Karakus S, Musicki B, Matsui H, Bivalacqua TJ, Dos Santos JL, Costa FF, and Burnett AL (2016b) Beneficial Effect of the Nitric Oxide Donor Compound 3-(1, 3-Dioxoisoindolin-2-yl)Benzyl Nitrate on Dysregulated Phosphodiesterase 5, NADPH Oxidase and Nitrosative Stress in the Sickle Cell Mouse Penis: Implication for Priapism Treatment. J Pharmacol Exp Ther 359:230–237 DOI: 10.1124/jpet.116.235473.
- Silva FH, Leiria LO, Alexandre EC, Davel APC, Mónica FZ, De Nucci G, and Antunes E (2014) Prolonged therapy with the soluble guanylyl cyclase activator BAY 60-2770 restores the erectile function in obese mice. J Sex Med 11:2661–2670.
- Smith A and Morgan WT (1979) Haem transport to the liver by haemopexin. Receptormediated uptake with recycling of the protein. *Biochem J* 182:47–54.
- Sopko NA, Matsui H, Hannan JL, Berkowitz D, Champion HC, Hsu LL, Musicki B, Burnett AL, and Bivalacqua TJ (2015) Subacute Hemolysis in Siddle Cell Mice Causes Priapism Secondary to NO Imbalance and PDE5 Dysregulation. J Sex Med 12:1878-1885. Sorba G, Medana C, Fruttero R, Cena C, Di Stilo A, Galli U, and Gasco A (1997) Wa-
- ter soluble furoxan derivatives as NO prodrugs. *J Med Chem* **40**:463–469. Stasch JP, Becker EM, Alonso-Alija C, Apeler H, Dembowsky K, Feurer A, Gerzer R,
- Stasch JP, Becker EM, Alonso-Anja C, Apeler H, Dembowsky K, Feurer A, Gerzer R, Minuth T, Perzborn E, Pleiss U et al. (2001) NO-independent regulatory site on soluble guanylate cyclase. *Nature* 410:212–215.
- Stasch J-P, Schmidt PM, Nedvetsky PI, Nedvetskaya TY, H S AK, Meurer S, Deile M, Taye A, Knorr A, Lapp H et al. (2006) Targeting the heme-oxidized nitric oxide receptor for selective vasodilatation of diseased blood vessels. J Clin Invest 116: 2552–2561.
- Steinberg MH, Barton F, Castro O, Pegelow CH, Ballas SK, Kutlar A, Orringer E, Bellevue R, Olivieri N, Eckman J et al. (2003) Effect of hydroxyurea on mortality and morbidity in adult sickle cell anemia: risks and benefits up to 9 years of treatment. JAMA 289:1645–1651.

- Steinhorn BS, Loscalzo J, and Michel T (2015) Nitroglycerin and Nitric Oxide-A Rondo of Themes in Cardiovascular Therapeutics. N Engl J Med **373**:277–280.
- Taddesse A, Woldie IL, Khana P, Swerdlow PS, Chu J-W, Abrams J, and Abou-Samra A-B (2012) Hypogonadism in patients with sickle cell disease: central or peripheral? *Acta Haematol* **128**:65–68 DOI: 10.1159/000337344.
- Tchernychev B, Li H, Lee SK, Gao X, Ramanarasimhaiah R, Liu G, Hall KC, Bernier SG, Jones JE, Feil S et al. (2021) Olinciguat, a stimulator of soluble guanylyl cyclase, attenuates inflammation, vaso-occlusion and nephropathy in mouse models of sickle cell disease. Br J Pharmacol 178:3463–3475.
- Telen MJ, Malik P, and Vercellotti GM (2019) Therapeutic strategies for sickle cell disease: towards a multi-agent approach. Nat Rev Drug Discov 18:139–158.
- Touyz RM, Anagnostopoulou A, Camargo LL, Rios FJ, and Montezano AC (2019) Vascular Biology of Superoxide-Generating NADPH Oxidase 5-Implications in Hypertension and Cardiovascular Disease. Antioxid Redox Signal 30:1027–1040.
- Van den Broeck T, Soebadi MA, Falter A, Raets L, Duponselle J, Lootsma J, Heintz A, Philtjens U, Hofkens L, Gonzalez-Viedma A et al. (2020) Testosterone Induces Relaxation of Human Corpus Cavernosum Tissue of Patients With Erectile Dysfunction. Sex Med 8:114–119.
- Vinchi F, De Franceschi L, Ghigo A, Townes T, Cimino J, Silengo L, Hirsch E, Altruda F, and Tolosano E (2013) Hemopexin therapy improves cardiovascular function by preventing heme-induced endothelial toxicity in mouse models of hemolytic diseases. *Circulation* 127:1317-1329.
- Zhang Y, Murugesan P, Huang K, and Cai H (2020) NADPH oxidases and oxidase crosstalk in cardiovascular diseases: novel therapeutic targets. Nat Rev Cardiol 17:170–194.

Address correspondence to: Dr. Fábio Henrique da Silva, Laboratory of Pharmacology, São Francisco University Medical School, Rua São Francisco, 218, Jardim São José, 2916-900, Bragança Paulista-SP, Brazil. E-mail: fabio.hsilva@usf.edu.br; fabiohsilva87@gmail.com